SUMMARY

(In accordance with 40 CFR Part 152, this summary is available
for public release after registration.)

STUDY TITLE

Magnitude of the Residue of 2,4-D and Quizalofop-P-ethyl in/on Herbicide Tolerant Field Corn Containing the Aryloxyalkanoate Dioxygenase-1 (ADD-1) Gene

DATA REQUIREMENTS

U.S. EPA Residue Chemistry Test Guidelines,
OPPTS 860.1500, Crop Field Trials
OPPTS 860.1520, Processed Food/Feed
PMRA Directive 98-02

STUDY DIRECTOR AND AUTHOR

John F. Culligan

STUDY COMPLETION DATE

08-Nov-2010

PERFORMING LABORATORIES

Field Phase Management and Test Facility
Ag Research Associates, LLC
1730 Denham Road
Sycamore, GA 31790

Analytical Phase
Dow AgroSciences LLC
Regulatory Sciences and Government Affairs-Indianapolis Lab
9330 Zionsville Road
Indianapolis, Indiana 46268

Processing Phase
GLP Technologies
22723 State Highway 6 South
Navasota, Texas 77868

STUDY NUMBER

ARA-09-15-10

Dow AgroSciences Study ID Number
090052

STUDY TITLE

Magnitude of the Residue of 2,4-D and Quizalofop-P-ethyl in/on Herbicide Tolerant Field Corn Containing the Aryloxyalkanoate Dioxygenase-1 (ADD-1) Gene

SUMMARY

The magnitude of residues of 2,4-D (2,4-dichlorophenoxyacetic acid) and quizalofop-P-ethyl (ethyl (R)-2-[4-(6-chloroquinoxalin-2-yloxy)phenoxy]propionate) in field corn forage, grain, stover, aspirated grain fractions and processed products was determined following application of 2,4-D and quizalofop-P-ethyl herbicides to hybrid field corn containing the inserted Aryloxyalkanoate Dioxygenase-1 (AAD-1) gene. Expression of the AAD-1 protein, which is encoded by the AAD-1 gene, results in a herbicide tolerance trait that increases the inherent tolerance of field corn to 2,4-D and provides tolerance to quizalofop-P-ethyl. Residue data in this study were produced with the use of field corn containing AAD-1 Event DAS-40278-9.

Field trials were established at 25 sites in the U.S. and Canada with two trials in each of NAFTA regions 1, 2, and 6, and nineteen trials in NAFTA region 5 (which includes 2 trials located in Canada). The corn was treated with three applications of 2,4-D with each application targeted at a rate of 1.0 pound acid equivalents per acre (lb ae/A), which is equivalent to 1,120 grams acid equivalents per hectare (g ae/ha) and one application of quizalofop-P-ethyl targeted at 0.082 pound active ingredient per acre (lb ai/A), which is equivalent to 92 grams active ingredient per hectare (g ai/ha). Additionally, at two trial sites, bulk grain samples were produced for use in generating aspirated grain fractions and processed products and these plots were treated using 2X rates (i.e. 2,4-D at a target rate of 2.0 lb ae/A and quizalofop-P-ethyl at a target rate of 0.164 lb ai/A). The three applications of 2,4-D were targeted as follows: (1) pre-emergence, (2) approximately 12 days before corn reached the V8 growth stage or 48 inch height, whichever came first, and (3) application to corn at approximately the V8 growth stage or 48 inch height. The single application of quizalofop-P-ethyl was targeted at approximately 5 days before corn reached the V8 growth stage or 48 inch height (i.e. approximately 5 days before the last

of three applications of 2,4-D). The corn was treated with 2,4-D using Weedar® 64 herbicide, a soluble concentrate (SL) formulation containing 2,4-D dimethylamine salt at 454 g ae/L. Quizalofop-P-ethyl was applied to corn using Assure® II herbicide, an emulsifiable concentrate (EC) formulation containing quizalofop-P-ethyl at 105 g ai/L. Non-ionic surfactant at approximately 0.25% (v/v) was included in the spray mixture with 2,4-D and crop oil concentrate at approximately 1.25% (v/v) was included in the spray mixture with quizalofop-P-ethyl. All postemergence treatments were applied broadcast with a boom sprayer over-the-top of the corn.

Forage RAC (raw agricultural commodity) samples were collected at each of two timings at all sites. The first sample collection timing for forage was set at approximately 40 days after the last application of 2,4-D, which at several locations involved sampling corn before the typical growth stage of late dough to early dent. Grain and stover raw agricultural commodity (RAC) samples were collected when plant maturity was consistent with that for normal commercial harvest. Decline samples of forage, grain and stover were collected at three trial sites. Bulk grain samples were collected from untreated control and treated plots at each of two trial sites. Aspirated grain fractions were separated from the bulk grain samples and the processed commodities of flour, grits, meal, refined oil (from both dry milling and wet milling), and starch were produced and samples were collected for residue analysis. Samples were analyzed for residues of 2,4-D and its metabolite 2,4-dichlorophenol (2,4-DCP) using one analytical method while a separate analytical method was used for determination of quizalofop-ethyl and its parent acid, quizalofop acid. Both analytical methods are liquid chromatography-mass spectroscopy/mass spectroscopy (LC/MS/MS) methods. The limit of detection (LOD) and limit of quantitation (LOQ) for each of the 4 analytes in all matrices from both methods was 0.003 ug/g and 0.01 ug/g, respectively.

Results of analysis of the samples from these trials for residues of 2,4-D, 2,4-DCP, quizalofop-ethyl and quizalofop acid are presented and discussed in the study report.

STUDY TITLE

Magnitude of the Residue of 2,4-D and Quizalofop-P-ethyl in/on Herbicide Tolerant Field Corn Containing the Aryloxyalkanoate Dioxygenase-1 (ADD-1) Gene

DATA REQUIREMENTS

U.S. EPA Residue Chemistry Test Guidelines,
OPPTS 860.1500, Crop Field Trials
OPPTS 860.1520, Processed Food/Feed
PMRA Directive 98-02

STUDY DIRECTOR AND AUTHOR

John F. Culligan

STUDY COMPLETION DATE

08-Nov-2010

PERFORMING LABORATORIES

Field Phase Management
Ag Research Associates, LLC
1730 Denham Road
Sycamore, GA 31790

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Processing Phase
GLP Technologies
22723 State Highway 6 South
Navasota, Texas 77868

STUDY NUMBER

ARA-09-15-10

Dow AgroSciences Study ID Number
090052

STATEMENT OF NO DATA CONFIDENTIALITY CLAIMS

Compounds: 2,4-dichlorophenoxyacetic acid (2,4-D) and quizalofop-P-ethyl

Study Title: Magnitude of the Residue of 2,4-D and Quizalofop-P-ethyl in/on Herbicide Tolerant Field Corn Containing the Aryloxyalkanoate Dioxygenase-1 (ADD-1) Gene

No claim of confidentiality, on any basis whatsoever, is made for any information contained in this document. I acknowledge that information not designated as within the scope of FIFRA sec. 10(d)(1)(A), (B), or (C) and which pertains to a registered or previously registered pesticide is not entitled to confidential treatment and may be released to the public, subject to the provisions regarding disclosure to multinational entities under FIFRA sec. 10(g).

Company: Dow AgroSciences LLC

Company Agent: Diego Fonseca

Title: Regulatory Manager

Signature: Fonseca G.

Date: 8 November 2010

THIS DATA MAY BE CONSIDERED CONFIDENTIAL IN COUNTRIES OUTSIDE THE UNITED STATES.

STATEMENT OF COMPLIANCE WITH GOOD LABORATORY PRACTICE STANDARDS

Title: Magnitude of the Residue of 2,4-D and Quizalofop-P-ethyl in/on Herbicide Tolerant Field Corn Containing the Aryloxyalkanoate Dioxygenase-1 (AAD-1) Gene

Study Initiation Date: 19-May-2009

I, the undersigned Study Director, hereby certify that this study was performed under my supervision according to the procedures described, and that this report represents a true and accurate record of the results and information obtained.

Excluding the exceptions listed below, the study described in this report was conducted to meet the requirements of Good Laboratory Practice Standards:

United States Environmental Protection Agency
Title 40 Code of Federal Regulations Part 160
FEDERAL REGISTER, August 17, 1989

1) Trial period weather data (all except Trials 19 and 20), irrigation records (for Trials 03, 09, 14, 15, 21 and 22), cultural practices (all except Trials 16, 22 and 23), maintenance pesticides (all Trials except 12, 13, 19, 20, 22 and 23), and fertilizer use (all except Trial 19) data were not collected and documented according to GLPS.
2) All Trials: historical weather data and pesticide history information were not collected and documented according to GLPS.
3) All Trials except 01 and 02: soil characterization data were not collected and documented according to GLPS.
4) All Trials except 03, 15, 16, 19, 24, and 25: sample weights were not measured and documented according to GLPS.
5) Trials 15 and 16: equipment maintenance logs, elevation, slope, distances, and GPS coordinates data were not collected and documented according to GLPS.
6) Trial 17: Maintenance equipment logs, GPS coordinate data, and max/min thermometer data were not collected and documented according to GLPS.
7) Trial 18: GPS coordinates data were not collected and documented according to GLPS.
8) Trial 19 and 20: Several data entries were not signed and dated at the time of entry. GPS coordinates data were not collected and documented according to GLPS and Google Earth (non-GLP compliant) was used for distances.

STATEMENT OF COMPLIANCE WITH GOOD LABORATORY PRACTICE STANDARDS
(continued)

9) Trial 25: On one occasion, data was not initialed and dated on the day it was recorded.

10) The test substance name was obscured on the label of the 2,4-D test substance bottle received at Trial 03 which made the test substance name unreadable. Other details on the label indicated that the correct test substance was being used.

Diego Fonseca
Sponsor
Dow AgroSciences LLC

Date: 8 November 2010

Diego Fonseca
Submitter
Dow AgroSciences LLC

Date: 8 November 2010

John F. Culligan
Study Director / Author
Ag Research Associates, LLC

Date: 08-Nov-2010

QUALITY ASSURANCE STATEMENT

Compounds: 2,4-dichlorophenoxyacetic acid (2,4-D) and quizalofop-P-ethyl

Study Title: Magnitude of the Residue of 2,4-D and Quizalofop-P-ethyl in/on Herbicide Tolerant Field Corn Containing the Aryloxyalkanoate Dioxygenase-1 (AAD-1) Gene

Study Number: ARA-09-15-10

Sponsor: Dow AgroSciences LLC **Sponsor Reference Number:** 090052

Study Initiation Date: 19-May-2009 **Study Completion Date:** 08-Nov-2010

This study was inspected/ audited and the findings reported to the Study Director and Management of Ag Research Associates, LLC in compliance with Good Laboratory Practice Standards, 40 CFR Part 160, on the following dates:

Date of Inspection	Subject of Audit/Inspection	Date Reported to Study Director and Management
10-Jun-2009	Trial 12 – Planting	14-Jun-2009
10-Jun-2009	Trial 25 - Application	02-Jul-2009
12-Jun-2009	Trial 03 – Application	16-Jun-2009
24-Jun-2009	Trial 17 - Application	03-Jul-2009
29-Jun-2009	Trial 16 – Planting/Application	24-Oct-2009
30-Jun-2009	Trial 23 - Planting	28-Jul-2009
07-Jul-2009	Trial 01 - Application	07-Jul-2009
08-Jul-2009	Trial 15 - Application	24-Oct-2009
08-Jul-2009	Trial 02 - Application	09-Jul-2009
13-Jul-2009	Trial 22 - Application	28-Jul-2009
16-Jul-2009	Trial 06- Application	20-Jul-2009
23-Jul-2009	Trial 19 - Application	10-Sep-2009
23-Jul-2009	Trial 20 - Application	10-Sep-2009
24-Aug-2009	Trial 24 - Sampling	08-Sep-2009
26-Aug-2009	Trial 07 - Sampling	11-Sep-2009
26-Aug-2009	Trial 25 - Sampling	08-Sep-2009
01-Sep-2009	Trial 14 - Sampling	31-Oct-2009

Date of Inspection	Subject of Audit/Inspection	Date Reported to Study Director and Management
14-Sep-2009	Trial 05 - Sampling	15-Sep-2009
30-Sep-2009	Trial 21 - Sampling	09-Oct-2009
20-Oct-2009	Trial 18 - Sampling	23-Oct-2009
02-Nov-2009	Trial 21 - Sampling	16-Nov-2009
17-Nov-2009	Trial 24 - Sampling	01-Dec-2009
24-Nov-2009	Trial 17 - Field Data/Summary	27-Nov-2009
30-Nov-2009	Trial 24 - Field Data/Summary	02-Dec-2009
05-Dec-2009	Trial 25 - Field Data/Summary	16-Dec-2009
17-Dec-2009	Trial 02 - Field Data	17-Dec-2009
21-Dec-2009	Trial 01 - Field Data	16-Jan-2010
04-Jan-2010	Trial 05 – Field Data/Summary	04-Jan-2010
12-Jan-2010	Trial 15 – Field Data/Summary	13-Jan-2010
13-Jan-2010	Trial 18 – Field Data/Summary	15-Jan-2010
16-Jan-2010	Trial 06 - Field Data	18-Jan-2010
17, 20-Jan-2010	Trial 03 - Field Data/Summary	21-Jan-2010
20, 21-Jan-2010	Trial 13 - Field Data/Summary	22-Jan-2010
22-Jan-2010	Trial 14 - Field Data/Summary	24-Jan-2010
22-Jan-2010	Trial 04 - Field Data/Summary	25-Jan-2010
22-Jan-2010	Trial 12 – Field Data/Summary	24-Jan-2010
31-Jan-2010	Trial 16 – Field Data/Summary	31-Jan-2010
10-Feb-2010	Trial 21 – Field Data	10-Feb-2010
16-Feb-2010	Trial 11 – Field Data/Summary	17-Feb-2010
18-Feb-2010	Trial 07 – Field Data	22-Feb-2010
22-Feb-2010	Trial 22 - Field Data	25-Feb-2010
02-Mar-2010	Trial 23 – Field Data	11-Mar-2010
17, 18-Mar-2010	Trial 19 - Field Data/Summary	18-Mar-2010
20-Mar-2010	Trial 20 – Field Data/Summary	20-Mar-2010
5 – 8-Apr-2010	Trial 10 – Field Data/Summary	12-Apr-2010
09-Apr-2010	Trial 08 – Field Data/Summary	12-Apr-2010
15-May-2010	Trial 09 – Field Data/Summary	17-May-2010
17, 18-May-2010	Trials 01and 02 – Field Summaries	19-May-2010
24-Jun-2010	Trial 21 – Field Summary	28-Jun-2010

Date of Inspection	Subject of Audit/Inspection	Date Reported to Study Director and Management
24-Jun-2010	Trial 22 – Field Summary	28-Jun-2010
24-Jun-2010	Trial 23 – Field Summary	28-Jun-2010
25-Jun-2010	Trial 06 – Field Summary	28-Jun-2010
02-Jul-2010	Trial 07 – Field Summary	02-Jul-2010
24-26-Oct-2010	Final Report	26-Oct-2010

Please see the Processing Summary (Appendix B) and the Analytical Summary (Appendix C) for the Quality Assurance Statements for those phases of the study.

The report and all records and raw data were audited and the report was found to be an accurate reflection of the study. All raw data will be maintained by Dow AgroSciences LLC, 9330 Zionsville Road, Indianapolis, IN 46268, in the Quality Assurance Unit archives.

Jane W. Brown
Quality Assurance
Ag Research Associates, LLC

08 Nov 2010
Date

CERTIFICATION OF AUTHENTICITY

We, the undersigned, hereby declare that this study was performed under our supervision according to the procedure described herein, and that this report provides a true and accurate record of the results obtained.

Study Director: ______________________ **Date:** 08-Nov-2010
John F. Culligan
Ag Research Associates, LLC

Approved By: ______________________ **Date:** 08-Nov-2010
Steve W. Rosser
Sponsor Study Monitor
Dow AgroSciences LLC

STUDY PERSONNEL

Study Title: Magnitude of the Residue of 2,4-D and Quizalofop-P-ethyl in/on Herbicide Tolerant Field Corn Containing the Aryloxyalkanoate Dioxygenase-1 (AAD-1) Gene

Study Director: John F. Culligan, Ag Research Associates, LLC
Principal Analyst: Brian M. Wendelburg, Dow AgroSciences LLC
Processing Principal Investigator: Dick L. Dusek, GLP Technologies
Other Study Personnel: S. E. Fisher, Dow AgroSciences LLC

Field Trial Sites

Trial #	Principal Field Investigator	Company	Facility Address
NJ-01 PA-02	Daniel Ramsdell	Crop Management Strategies	6273 Mountain Road Germansville, PA 18053
GA-03 GA-04	Chris Cromer	Ag Research Associates	1730 Denham Road Sycamore, GA 31790
IL-05	Sue Dorsey	SGS Alvey Ag Research	Route 2 Box 12 Wyoming, IL 61491
IL-06	Tim Boeker	SGS Alvey Ag Research	19300 Marydale RD Carlyle, IL 62231
IN-07	John Bailey	SGS Alvey Ag Research	1365 N Seip Rd. Rockville, IN 47872
MO-08 MO-09 IL-10	Matt Cordell	Ag Research Associates	1204 Fairview St. Lonoke, AR 72086
MI-11 OH-12 IN-13	Chad Harris	Ag Research Associates	2450 Hoagland Highway, Deerfield, MI 49238
NE-14	Matt Krause	Ag Research Associates	3605 N. Delaware Ave York, NE 68467
NE-15 KS-16	Denny Stamm	Ag Research Associates	3605 N. Delaware Ave York, NE 68467
IA-17	Jason Niekamp	Bennett Ag Research	1109 Ivy Avenue Richland, IA 52585
MO-18	David Bennett	Bennett Ag Research	1109 Ivy Avenue Richland, IA 52585
IA-19 MN-20	Kyle Johnson	Eurofins Agroscience Services	4614 Highway 63 Lime Springs, IA 52155
MO-21	Nathan Goldschmidt	Shoffner Farm Research, Inc. - North Station	4809 Highway FF Fisk, MO 63940
OK-22 TX-23	Tim Case	Ag Research Associates	89 Broadway Avenue Groom, TX 79039
ON-24 ON-25	Jamie Parnell	Vaughn Ag Research Services	1909 Maple Manor Rd. Branchton, ON N0B1L0

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GENERAL STUDY INFORMATION

Study Number
ARA-09-15-10

Study Title
Magnitude of the Residue of 2,4-D and Quizalofop-P-ethyl in/on Herbicide Tolerant Field Corn Containing the Aryloxyalkanoate Dioxygenase-1 (AAD-1) Gene

Study Director
John F. Culligan
Ag Research Associates, LLC
1730 Denham Road
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E-mail: jack.culligan@agresearchassociates.com

Test Substances
2,4-D and quizalofop-P-ethyl

Study Execution Dates
Study Initiation Date: 19-May-2009
Study Experimental Start Date: 04-Jun-2009
Study Experimental Termination Date: 30-Aug-2010
Study Completion Date: 08-Nov-2010

Performing Facilities

Test Facility: Ag Research Associates, LLC 1730 Denham Road Sycamore, GA 31790	Sponsor: Dow AgroSciences LLC 9330 Zionsville Road Indianapolis, Indiana 46268
Processing Phase: GLP Technologies 22723 State Hwy 6 South Navasota, Texas 77868	Analytical Phase: Dow AgroSciences LLC 9330 Zionsville Road Indianapolis, Indiana 46268

1.0 ABSTRACT

The magnitude of residues of 2,4-D (2,4-dichlorophenoxyacetic acid) and quizalofop-P-ethyl (ethyl (R)-2-[4-(6-chloroquinoxalin-2-yl oxy)phenoxy]propionate) in field corn forage, grain, stover, aspirated grain fractions and processed products was determined following application of 2,4-D and quizalofop-P-ethyl herbicides to hybrid field corn containing the inserted Aryloxyalkanoate Dioxygenase-1 (AAD-1) gene. Expression of the AAD-1 protein, which is encoded by the AAD-1 gene, results in a herbicide tolerance trait that increases the inherent tolerance of field corn to 2,4-D and provides tolerance to quizalofop-P-ethyl. Residue data in this study were produced with the use of field corn containing AAD-1 Event DAS-40278-9.

To determine maximum residue levels in the indicated corn matrices based on the proposed use patterns for 2,4-D and quizalofop-P-ethyl, field trials were established in which the corn was treated with three applications of 2,4-D with each application targeted at a rate of 1.0 pounds acid equivalents per acre (lb ae/A), which is equivalent to 1,120 grams acid equivalents per hectare (g ae/ha) and one application of quizalofop-P-ethyl targeted at 0.082 pounds active ingredient per acre (lb ai/A), which is equivalent to 92 grams active ingredient per hectare (g ai/ha). All applications of 2,4-D and quizalofop-P-ethyl were within 95% to 110% of the target application rates. The same plot of field corn was treated with both 2,4-D and quizalofop-P-ethyl. The three applications of 2,4-D were targeted as follows: (1) pre-emergence, (2) approximately 12 days before corn reached the V8 growth stage or 48 inch height, whichever came first, and (3) application to corn at approximately the V8 growth stage or 48 inch height. The single application of quizalofop-P-ethyl was targeted at approximately 5 days before corn reached the V8 growth stage or 48 inch height (i.e. approximately 5 days before the last of three applications of 2,4-D). The corn was treated with 2,4-D using Weedar® 64 herbicide, a soluble concentrate (SL) formulation containing 2,4-D dimethylamine salt at 454 g ae/L. Quizalofop-P-ethyl was applied to corn using Assure® II herbicide, an emulsifiable concentrate (EC)

formulation containing quizalofop-P-ethyl at 105 g ai./L. Non- ionic surfactant at approximately 0.25% (v/v) was included in the spray mixture with 2,4-D and crop oil concentrate at approximately 1.25% (v/v) was included in the spray mixture with quizalofop-P-ethyl. Other than the pre-emergence application of 2,4-D, the other applications of 2,4-D as well as the application of quizalofop-P-ethyl were made postemergence over-the-top of the corn. Treatments were applied broadcast with the use of a boom sprayer. Spray volumes ranged from 19.6 to 36.4 gallons/A.

Trials were conducted at 25 sites: two in NAFTA region 1, two in NAFTA region 2, nineteen in NAFTA region 5 (which includes 2 trials located in Canada), and two in NAFTA region 6. Each site had an untreated control plot that was planted with conventional field corn and a treated plot that was planted with field corn containing AAD-1 Event DAS-40278-9. (Each site also included a second treated plot that was planted with field corn containing AAD-1 Event DAS-40474-7, but samples from these plots were not analyzed due to a decision during the course of this study not to continue with commercial development of Event DAS-40474-7.) Two of the trial sites also included large plots that were treated with 2X rates of 2,4-D and quizalofop-P-ethyl (i.e. 2,4-D at approximately 2.0 lb ae/A (2240 g ae/ha) in each of 3 applications and quizalofop-P-ethyl at approximately 0.164 lb ai/A (184 g ai/ha)) to produce bulk grain samples for use in obtaining aspirated grain fractions and processed commodities. Rates higher than 2X were not used in plots for bulk grain samples due to concern over the potential for crop injury. Forage RAC (raw agricultural commodity) samples were collected at each of two timings at all sites. The first sample collection timing for forage was set at approximately 40 days after the last application of 2,4-D, which at several locations involved sampling corn before the typical growth stage of late dough to early dent. Samples were collected at this timing in order to generate residue data that could support a reduced PHI (pre-harvest interval) compared to sampling later when corn reached the more typical stage for forage harvest. The second sample collection timing for forage was targeted at the corn in the late dough to early dent stage. Grain and stover raw agricultural commodity (RAC) samples were collected when plant maturity was consistent with that for normal commercial harvest. Based on the last application of 2,4-D, the PHIs for the early forage harvest and "typical" forage harvest ranged from 27-42 days and 39-78 days, respectively. For grain and stover, the PHIs ranged from 73-139 days based on the last application of 2,4-D. Quizalofop-P-ethyl was applied slightly earlier than the final application of 2,4-D and the PHIs vary accordingly. Based on the application of quizalofop-P-ethyl, the PHIs for the early forage harvest and “typical” forage harvest ranged from 32-47 days and 44-83 days, respectively. For grain and stover, the PHIs ranged from 79-144 days based on the application of quizalofop-P-ethyl. In addition to RAC samples, decline samples of forage, grain and stover were collected at three trial sites. In addition to the RAC sample, which was targeted for collection at normal commercial harvest, decline samples of forage, grain and stover were each collected at approximately 7 and 14 days before the RAC sample collection and at approximately 7 and 14 days after the RAC sample collection. Bulk grain samples were collected from untreated control and treated plots at each of two trial sites. Aspirated grain fractions were separated from the bulk grain samples and the processed commodities of flour, grits, meal, refined oil (from both dry milling and wet milling) and starch were produced and samples were collected for residue analysis.

Samples were analyzed for residues of 2,4-D and its metabolite 2,4-dichlorophenol (2,4-DCP) using one analytical method while a separate analytical method was used for determination of quizalofop-ethyl and its parent acid, quizalofop acid. Both analytical methods are liquid chromatography-mass spectroscopy/mass spectroscopy (LC/MS/MS) methods. The limit of detection (LOD) and limit of quantitation (LOQ) for each of the 4 analytes in all matrices from both methods was 0.003 ug/g and 0.01 ug/g, respectively.

The current residue definition for 2,4-D does not include 2,4-DCP, but is based on only 2,4-D determined as the acid. However, 2,4-DCP was included as an analyte of interest because the herbicidal tolerance of AAD-1 corn to 2,4-D relies on increased plant metabolism through a pathway involving 2,4-DCP. Both quizalofop-ethyl and quizalofop acid were included as analytes because the current residue definition is for the combined residues of quizalofop-ethyl and quizalofop acid, all expressed as quizalofop-ethyl.

Residue data for 2,4-D and 2,4-DCP in RAC samples are summarized in the following table:

Analyte/ Commodity	Target Application Rate[a] lb ae/A (g ae/ha)	PHI[b] (days)	n	Residue Levels (ug/g)					
				Min.	Max.	Median[c]	Mean[c]	Std. Dev.[c]	HAFT[d]
2,4-D									
Early Forage	3 x 1.0 (~1120)	27-42	48	ND[e]	2.988	0.443	0.641	0.707	2.612
Forage	3 x 1.0 (~1120)	39-78	50	ND	4.373	0.252	0.697	1.008	3.124
Grain	3 x 1.0 (~1120)	73-139	48	ND	(0.004)[f/g]	ND	ND	NA[h]	(0.004)
Stover	3 x 1.0 (~1120)	73-139	50	ND	8.92	0.613	1.349	2.064	8.076
2,4-DCP									
Early Forage	3 x 1.0 (~1120)	27-42	48	ND	3.997	0.773	1.032	1.003	3.056
Forage	3 x 1.0 (~1120)	39-78	50	(0.004)	5.007	0.448	0.876	1.154	3.899
Grain	3 x 1.0 (~1120)	73-139	48	ND	(0.003)[f]	ND	ND	NA	ND
Stover	3 x 1.0 (~1120)	73-139	50	ND	5.842	0.453	0.911	1.224	5.377

[a] 2,4-D was applied as the first, second and fourth of four applications, quizalofop-P-ethyl was applied as the third application.

[b] PHI = Pre-Harvest Interval; days between last application of, 2,4-D and collection of field sample.

[c] For statistical purposes, ND has been given the value of zero.

[d] HAFT = Highest Average Field Trial

[e] ND = Not detected; less than the LOD (<0.003 µg/g)

[f] Values for 2,4-D in the replicate samples of grain from trial MO-08 were not included in this summary because one of two replicate samples is not believed to be representative of residues from the application pattern in this study and may have resulted from contamination. The MO-08 residue values of 2,4-D in replicate samples in grain from trial MO-08 are as follows: 0.067 ug/g, ND. The 2,4-DCP residues in the replicate samples from this site were both ND. For consistency, neither of the replicate values for 2,4-D or 2,4-DCP in grain from this trial site were included in the summary.

[g] Values in parentheses are greater than or equal to the limit of detection (0.003 ug/g), but less than the limit of quantitation (0.01 ug/g). Values are reported with a lower degree of confidence than values above the limit of quantitation

[h] NA = Not applicable

Residue data for quizalofop-ethyl and quizalofop acid in RAC samples are summarized in the following table.

Analyte/ Commodity	Target Application Rate[a] lb ai/A (g ai/ha)	PHI[b] (days)	n	Residue Levels (ug/g) Min.	Max.	Median[c]	Mean[c]	Std. Dev.[c]	HAFT[d]
Quizalofop-ethyl									
Early Forage	0.082 (92)	32-47	48	ND[e]	ND	ND	ND	NA[f]	ND
Forage	0.082 (92)	44-83	48	ND	ND[g]	ND	ND	NA	ND
Grain	0.082 (92)	79-144	50	ND	ND	ND	ND	NA	ND
Stover	0.082 (92)	79-144	50	ND	(0.004)[h]	ND	ND	NA	ND
Quizalofop acid									
Early Forage	0.082 (92)	32-47	48	ND	ND	ND	ND	NA	ND
Forage	0.082 (92)	44-83	48	ND	ND [h]	ND	ND	NA	ND
Grain	0.082 (92)	79-144	50	ND	(0.004)	ND	ND	NA	ND
Stover	0.082 (92)	79-144	50	ND	0.011	ND	ND	NA	0.011
Combined Residues of quizalofop-ethyl and quizalofop acid (expressed as quizalofop-ethyl)[i]									
Early Forage	0.082 (92)	32-47	48	ND	ND	ND	ND	NA	ND
Forage	0.082 (92)	44-83	48	ND	ND	ND	ND	NA	ND
Grain	0.082 (92)	79-144	50	ND	(0.004)	ND	ND	NA	ND
Stover	0.082 (92)	79-144	50	ND	0.016	ND	ND	NA	0.016

[a] Quizalofop-P-ethyl was applied as the third application only, and 2,4-D was applied as the first, second and fourth of four applications.

[b] PHI = Pre-Harvest Interval; days between the single application of quizalofop-P-ethyl and the collection of field samples.

[c] For statistical purposes, ND has been given the value of zero.

[d] HAFT = Highest Average Field Trial

[e] ND = Not detected; less than the LOD (<0.003 µg/g)

[f] NA = Not applicable

[g] Values for the replicate samples for forage / typical forage from trial TX-23 were not included in this summary because they are not believed to be representative of residues from the application pattern in this study and may have resulted from contamination. The TX-23 residue values for replicate samples in forage are as follows: quizalofop-ethyl: 0.197 ug/g and 0.099 ug/g; quizalofop acid: 0.167 ug/g and 0.113 ug/g

[h] Values in parentheses are greater than or equal to the limit of detection (0.003 ug/g), but less than the limit of quantitation (0.01 ug/g). Values are reported with a lower degree of confidence than values above the limit of quantitation

[i] Quizalofop acid residue values are converted to quizalofop-ethyl equivalents based on the molecular weight of the two compounds. Quizalofop-ethyl has a molecular weight of 372.8 and quizalofop acid has a molecular weight of 344.8. The factor to use to convert quizalofop acid to quizalofop-ethyl equivalents is calculated by dividing the molecular weight of quizalofop-ethyl by the molecular weight of quizalofop acid, which calculates to 1.08 (372.8/344.8 = 1.08). Therefore, the residue values for quizalofop acid are multiplied by 1.08 to convert them to quizalofop-ethyl equivalents. Once this is done then the values for quizalofop-ethyl and quizalofop acid expressed as quizalofop-ethyl equivalents are summed to provide the "combined" value.

The residue data from the decline trials showed a reasonably clear pattern of decline for 2,4-D and 2,4-DCP in corn forage over the 30 to 33-day collection intervals. A residue decline profile could not be established for 2,4-D or 2,4-DCP on corn grain as all residue values were below or just slightly above the limit of detection. The residue data from the decline trials showed a reasonably clear pattern of decline for 2,4-D and 2,4-DCP in stover over the 28 day collection interval in two of the three trials. The stover samples from the third decline trial did not exhibit decline over the five samplings as residue values remained relatively constant.

The residue decline profile could not be established for quizalofop-ethyl and quizalofop acid from the field corn forage or grain samples collected over the 30 to 33-day collection interval as nearly all of the forage samples yielded results below the limit of detection with just four data points slightly above the LOD. All grain samples were below the limit of detection. A clear pattern of decline for quizalofop-ethyl and quizalofop acid residues in field corn stover could not be established over the 28-day collection interval. All three trials yielded residue results for quizalofop-ethyl that were below the limit of detection. For quizalofop acid, one trial yielded non-detectable residue for all five sampling events, one trial had only 3 out of 10 samples from treated plots with detections slightly above the limit of detection, and one trial yielded residue that increased slightly (0.015 ug/g) over the five sampling intervals.

In the two processing trials, average residues in grain from the 2X rate treated plots when collected at the processor prior to separation of aspirated grain fractions (AGF) were (0.008) ug/g for 2,4-DCP in grain from the Nebraska (NE-14) trial site and were (0.003) ug/g for the combined residues of quizalofop-ethyl and quizalofop acid expressed as quizalofop-ethyl in grain also from the Nebraska trial site. However, 2,4-D was not detected in grain from either trial site and 2,4-DCP was not detected in grain from the Georgia trial site. Additionally, quizalofop-ethyl and quizalofop acid were not detected in grain from the Georgia trial site in samples collected at the processor prior to separation of AGF. After separating aspirated grain fractions and screening, grain was processed by dry milling to produce flour, grits, meal and oil, or by wet milling to produce oil and starch. There was no concentration of 2,4-D, 2,4-DCP, quizalofop-ethyl, or quizalofop acid residues from grain to flour, refined oil, starch, grits or meal. In the Nebraska trial (NE-14), there was no evidence of concentration of 2,4-D or 2,4-DCP into the AGF. However, in the Georgia trial, 2,4-D and 2,4-DCP were found in AGF at 0.021 ug/g and (0.006) ug/g, respectively, indicating a potential for concentration. A processing factor for concentration of 2,4-D or 2,4-DCP into AGF was not calculated since residues in grain from this site were below the limit of detection. There was no concentration of quizalofop-ethyl or quizalofop acid residues into the AGF from either trial site.

2.0 INTRODUCTION

2.1 Study Background Information

Dow AgroSciences has produced a novel herbicide tolerance trait in field corn that confers increased tolerance to certain phenoxy auxins, such as 2,4-D and aryloxyphenoxyproprionates, such as quizalofop-P-ethyl. To accomplish this, field corn has been modified by the insertion of the AAD-1 gene from *Sphingobium herbicidovorans*, a common soil bacterium, which encodes the aryloxyalkanoate dioxygenase (AAD-1) protein. The AAD-1 protein is an enzyme that metabolizes certain herbicides having an aryloxyalkanoate moiety, including 2,4-D and quizalofop-P. Expression of the AAD-1 protein results in a herbicide tolerance trait that provides access to significant new weed management technology for field corn by providing tolerance to quizalofop-P-ethyl and increasing the inherent level of tolerance to 2,4-D.

2,4-D is a commercially important herbicide that acts as a synthetic auxin. It is applied postemergence for control of a number of broadleaf weeds and is currently registered for use in a number of crops. Although currently registered for use in field corn, the usefulness of 2,4-D is limited by the potential for crop injury. In this study, 2,4-D was formulated as a dimethylamine (DMA) salt in Weedar 64 herbicide, a commercially available soluble concentrate (SL) formulation.

Quizalofop-P is a herbicide that acts as a acetyl coenzyme A carboxylase (ACCase) inhibitor. In this study, quizalofop-P acid was formulated as an ethyl ester (quizalofop-P-ethyl) in Assure II herbicide, a commercially available emulsifiable concentrate (EC) formulation. Once absorbed into plants, quizalofop-P-ethyl is rapidly deesterified to produce the herbicidally-active quizalofop-P acid. This herbicide controls many annual and perennial grass weeds and is currently registered for postemergence application in a number of broadleaf crops including canola, cotton, soybeans, sugar beets and sunflowers. Although conventional field corn is not tolerant to quizalofop-P, field corn that has been genetically modified to express the AAD-1 protein exhibits tolerance to this herbicide. Therefore, the herbicide tolerance trait that is conferred by expression of AAD-1 protein in field corn allows quizalofop-P-ethyl to be used as a selective postemergence herbicide in this crop.

2.2 Test Substance Background

The active ingredients in the test substances are 2,4-D and quizalofop-P-ethyl. 2,4-D [Weedar 64, a soluble liquid concentrate end-use product containing nominally 38.9% (w/w) acid equivalent (AE) of 2,4-dichlorophenoxyacetic acid, dimethylamine salt] is a broad-spectrum herbicide for the control of broadleaf weeds in a variety of crops. Quizalofop-P-ethyl [Assure II, an emulsifiable concentrate end-use product containing nominally 10.3% (w/w) of ethyl(R)-2-[4-(6-chloroquinoxalin-2-yl oxy) phenoxy] propionate] is a broad-spectrum herbicide for the control of grasses in a variety of crops.

2.3 Objective of the Study

The purpose of this study was to determine the magnitude of residues of 2,4-D and its metabolite 2,4-dichlorophenol (2,4-DCP), and quizalofop-ethyl and quizalofop acid in/on field corn raw agricultural commodities (forage, grain, and stover) and field corn processed fractions (aspirated grain fractions, starch, meal, grits, flour and refined oil) when herbicide tolerant transgenic hybrid lines of field corn containing the inserted AAD-1 gene in each of two Events, either AAD-1 Event DAS-40278-9 <u>or</u> AAD-1 Event DAS-40474-7, were treated with commercially available 2,4-D and quizalofop-P-ethyl herbicides. The data generated in this study are intended to satisfy data requirements for magnitude of the residues of 2,4-D and quizalofop-P-ethyl in field corn with the inserted AAD-1 gene. Although 2,4-DCP is not part of the residue definition for 2,4-D, it was included as an analyte of interest because the herbicidal tolerance of AAD-1 corn to 2,4-D relies on increased plant metabolism through a pathway involving DCP. See Appendix D for the study protocol and all its revisions (protocol amendments and deviations).

During the course of the study, the Sponsor discontinued development of the line containing AAD-1 Event DAS-40474-7. Samples were collected from both the twenty five RAC and two processing treated plots planted with this line but were not analyzed. Therefore, residue analysis of samples from treated plants and their processed fractions was carried out only for field corn with Event DAS-40278-9.

The study was conducted to meet data requirements for crop residue studies outlined in USEPA OPPTS Guidelines 860.1500, Crop Field Trials and OPPTS 860.1520 Processed Feed/Food, and Pesticide Management Regulatory Agency Directive 98-02 (References 1, 2, and 3, respectively), according to USEPA FIFRA GLPS (Reference 4).

2.4 Justification of Study Rate and Test System

Genetically modified hybrid field corn containing the AAD-1 gene was planted in treated plots in this study for use in producing residue data for 2,4-D and quizalofop-P-ethyl based on the proposed uses of these herbicides in field corn containing the inserted AAD-1 gene. Two transgenic hybrid lines of field corn each containing a different Event of the inserted AAD-1 gene were used in treated plots. One treated plot was planted with a hybrid containing AAD-1 Event DAS-40278-9 and a separate treated plot was planted with a hybrid containing AAD-1 Event DAS-40474-7. The source ID number for the AAD-1 Event DAS-40278-9 seed planted in trials in this study is ZQ08LQ656623 and a sample from these seeds was tested and confirmed positive for Event DAS-40278-9 (Reference 5). The source ID number for the AAD-1 Event DAS-40474-7 seed planted in trials in this study is ZQ08LQ656495 and a sample from these seeds was tested and confirmed positive for Event DAS-40474-7 (Reference 6). However, as documented in protocol amendment number 08, samples from plants with AAD-1 Event DAS-40474-7 were not analyzed for residues due to a business decision to discontinue the commercial development of this Event. Due to limited availability of seed with Event DAS-40278-9 or Event DAS-40474-7, the untreated control plots were planted with a hybrid field corn that had a genetic background similar to that used for the two transgenic hybrids, but lacking the inserted AAD-1 gene.

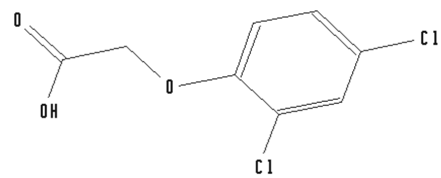
To determine the maximum residue levels in field corn raw agricultural commodities, 2,4-D and quizalofop-P-ethyl were applied at the intended maximum use rate, maximum number of applications and shortest re-treatment interval (considering the two postemergence applications of 2,4-D). Based on the proposed use of 2,4-D in field corn containing the AAD-1 gene, 2,4-D was applied three times to each treated plot with application timings targeted as follows: pre-emergence, 12 days (± 1 day) before the V-8 growth stage or 48 in. height and then a final application at the V-8 growth stage or 48 in. height (whichever occurred first). All applications of 2,4-D were targeted at a rate of 1120 g ae/ha (1.0 lb ae/acre) and were to include a non-ionic surfactant at approximately 0.25% (v/v). Based on the proposed use of quizalofop-P-ethyl in field corn containing the AAD-1 gene, it was applied only once at a target rate of 92 g ai./ha (0.082 lb ai./acre) with application timing targeted at 5 days (± 1 day) before the final application of 2,4-D at the V-8 growth stage or 48 in. height (whichever occurred first). Crop oil concentrate at approximately 1.25% (v/v) was to be included in the spray mixture with quizalofop-P-ethyl. In addition to the above, a 2X rate for each application of 2,4-D and for the application of quizalofop-P-ethyl was used at two trial locations where bulk grain samples were produced for use in generating processed fractions. A rate of 2,4-D or quizalofop-P-ethyl higher than 2X the proposed use rates was not applied to the plots used to generate grain for processing due to the potential for crop injury.

3.0 EXPERIMENTAL DESIGN

3.1 Test Substances

There were two test substances applied in this study. One test substance was Weedar 64 herbicide, a soluble concentrate (SL) formulation containing 2,4-D as the active ingredient formulated as 2,4-dichlorophenoxyacetic acid, dimethylamine salt (2,4-D, DMA salt). The formulation supplied for use in this study was identified as TSN026491-0010 (Reference 7) and contained 2,4-D at a concentration of 454 g ae/L (or 2,4-D, DMA salt at a concentration of 546 g ai./L). The Certificate of Analysis for this test substance is presented in Figure 1.

2,4-D acid nomenclature is summarized below:

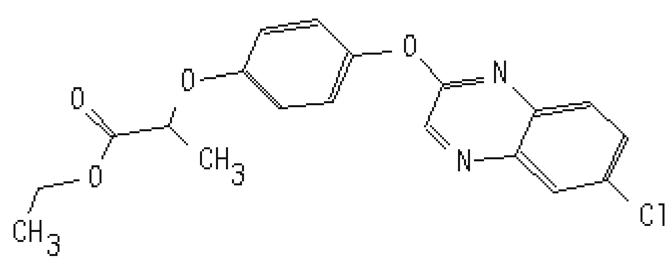
2,4-D chemical structure:	
Common Name:	2,4-D
IUPAC Name:	(2,4-dichlorophenoxy)acetic acid
CAS Name:	(2,4-dichlorophenoxy)acetic acid
CAS Number:	94-75-7

The physicochemical properties of 2,4-D acid are shown below.

Parameter	Value	Reference
Physical State at room temperature	Solid / crystalline	The Dow Chemical Company. "2,4-D Acid Physical and Chemical Characteristics", 1989.
Density (Relative / Specific Gravity)	1.601 g/mL at 20°C	Jones, J. "Determination of Relative Density for 2,4-D Acid Technical", Report No. NAFST-07-098, 2007, unpublished report of Dow AgroSciences LLC
Dissociation constant (pKa)	2.87	Martin, E. J. and T. J. Reading, "2,4-D; (2,4-Dichlorophenoxyacetic Acid) Dissociation Constant Determination", Report No. GHC-1974, DERBI No. 38798, 1987, unpublished report of Dow AgroSciences LLC
Water solubility (mean solubility at 25°C, mg/L)	20031 (buffered, pH 5) 23180 (buffered, pH 7) 34196 (buffered, pH 9)	Hopkins, D. L. "2,4-Dichlorophenoxyacetic Acid: Determination of the Water Solubility", Lab Project ID ES-994, DERBI No. 12662, 1987, unpublished report of Dow AgroSciences LLC
Vapor pressure	1.4×10^{-7} mm Hg at 25°C	Chakrabarti, A. "Vapor Pressure of 2,4-Dichlorophenoxyacetic Acid", Lab Project ID ML-AL 87-40047, DERBI No. 12628, 1987, unpublished report of Dow AgroSciences LLC
Octanol/water distribution coefficient - logD at 25°C	2.58 at pH 1 0.04 at pH 5 -0.91 at pH 7 -1.04 at pH 9	Bailey, R. E. "2,4-Dichlorophenoxyacetic Acid: Determination of Octanol/Water Partition Coefficient", Lab Project ID ES-DR-0002-2297-9, DERBI No. 12633, 1987, unpublished report of Dow AgroSciences LLC

A second test substance used in this study was Assure II herbicide, an emulsifiable concentrate (EC) formulation containing quizalofop-P acid formulated as quizalofop-P-ethyl. The formulation supplied for use in this study was identified as TSN020252-0004 (Reference 8) and contained quizalofop-P-ethyl at a concentration of 105 g ai./L. The Certificate of Analysis for this test substance is presented in Figure 2.

Quizalofop-P-ethyl nomenclature is summarized below:

Quizalofop-P-ethyl chemical structure:	
Common Name:	Quizalofop-P-ethyl
IUPAC Name:	ethyl (R)-2-[4-(6-chloroquinoxalin-2-yloxy)phenoxy]propionate
CAS Name:	propanoic acid, 2-[4-[(6-chloro-2-quinoxalinyl)oxy]phenoxy]-, ethyl ester, (2R)
CAS Number:	76578-14-8

The physicochemical properties of quizalofop-P-ethyl are shown below.

Parameter	Value	Reference
Molecular weight	372.8 g/mole (molecular wt. of the parent acid, quizalofop-P, is 344.8 g/mole)	The Pesticide Manual, A World Compendium, Fifteenth Edition. Ed. Tomlin, C.D.S., 2009. British Crop Production Council. Pages 1013-1015.
Physical State at room temperature	Solid / white crystalline	
Specific gravity / density	1.0223 g/mL at 20°C	
Water solubility (at 20°C)	0.61 mg/L	
Solvent solubility (at 20 - 23°C)	Acetone > 250 g/L Ethyl acetate > 250 g/L n-heptane = 7.2 1,2-Dichloroethane > 1000 g/L Methanol = 34.9 Xylene > 250 g/L	
Vapor pressure	1.1×10^{-4} mPa at 20°C	
Octanol/water partition coefficient - logP	4.61	

3.2 Study Site Information

The field portion of this study was conducted at twenty-five (25) sites in the United States and Canada from May 2009 (study initiation) to December 2009 (last field samples collected). The Principal Field Investigators and Field Testing Facilities responsible for conducting the field trials are listed in Appendix A, Table 3. The trials were located in areas where field corn is commonly grown commercially and were distributed among NAFTA regions in a manner that met US EPA and Canada PMRA requirements for the number and distribution of field corn residue trials. The field trials were distributed among NAFTA Regions as follows: two in Region 1 (New Jersey and Pennsylvania), two in Region 2 (Georgia), nineteen in Region 5, including Canada, (Missouri, Nebraska, Kansas, Iowa, Illinois, Indiana, Ohio, Michigan, Minnesota, and Ontario, Canada), and two in Region 6 (Oklahoma and Texas). Information concerning location of trial sites is given in Appendix A, Table 4.

Although there were instances at most sites in which temperatures and/or rainfall amounts varied appreciably from historical averages, the observations appear typical of year-to-year variation in weather conditions across the areas represented in this study. There was no indication that weather conditions negatively impacted the validity of the trials reported in this study. A summary of air temperatures during the test period and precipitation, as well as any irrigation, during the test period for each trial is presented in Appendix A, Table 36 and Appendix A, Table 37, respectively.

3.3 Test Site Plot Design

Among the total of 25 trials, there were 20 trials established to collect the RAC samples only. There were three decline trials established to collect RAC samples at one sampling interval and decline samples at four additional sampling intervals (five additional sampling intervals for forage if "early forage" is included) after the last application (RAC + Decline trials), and two trials were established to collect RAC samples as well as bulk grain for food/feed processing (RAC + Processing trials). The availability of field corn seed with the inserted AAD-1 Events was quite limited at the time this study was initiated. Due to the limited availability of corn seed with the specified AAD-1 Events, 2,4-D and quizalofop-P-ethyl were applied to the same treated plot rather than having a separate plot for application of each test substance. Additionally, control plots were planted with a conventional hybrid corn variety, Mycogen 2M746, having a genetic background similar to that used in the transgenic hybrid lines used in the treated plots, but lacking the inserted AAD-1 gene. The RAC and RAC + Decline trial sites had a control plot and two treated plots (one treated plot for each AAD-1 Event initially planned for the study). Decline samples were collected at the GA-04, MI-11, and OK-22 sites. The processing trials had five plots; one untreated plot to collect RAC and bulk samples, two 1X-rate treated plots to collect RAC samples, and two 2X-rate treated plots to collect the bulk grain sample for processing in order to assess residue levels in aspirated grain fractions and processed commodities. Bulk grain samples were collected from the control and exaggerated rate (2X) treated plots at the GA-03 and NE-14 sites.

The control plot was a minimum of 50 feet from the treated plots. The treated plot sizes at each trial location were large enough to collect all samples specified for that specific trial without sampling row ends or plot borders. Individual plot size as well as the planting date and soil types for each trial is presented in Appendix A, Table 5. Plant and row spacings used were representative of typical agronomic practices. Maintenance chemicals used on the trial sites during the study are shown in the Field Phase Summary in Appendix A, Table 6.

3.4 Application of Test Substances

There were four (4) broadcast herbicide applications with a boom sprayer to each treated plot, two applications of 2,4-D, followed by one application of quizalofop-P-ethyl, followed by a final application of 2,4-D to simulate a worst-case commercial herbicide treatment, based on the proposed use of 2,4-D and quizalofop-ethyl in corn containing the inserted AAD-1 gene. The two processing trials had an additional exaggerated rate (2X) treatment for each transgenic line from which bulk grain samples were collected for processing (5 total plots in RAC + processing trials).

The first application of 2,4-D was applied at pre-emergence. The fourth application (the last of the applications) was targeted to be an application of 2,4-D applied over-the-top to corn in approximately the V8 growth stage (or corn that was approximately 48 inches tall, whichever came first). The timings for Application No. 2 (the second application of 2,4-D) and Application No. 3 (the application of quizalofop-P-ethyl) were targeted to be scheduled relative to the date on which it was expected that the corn would be in approximately the V8 stage (or approximately 48 inches tall) and Application No. 4 was to be applied.

Application No. 2 (the second over-the-top application of 2,4-D) was targeted to be applied at 12 days (± 1 day) before Application No. 4. Application No. 3 (the over-the-top application of quizalofop-P-ethyl) was targeted to be applied at 5 days (±1 day) before Application No. 4. It was expected that the growth stage of the corn at the timing for Application No. 2 and Application No. 3 would vary somewhat among trial locations depending on how quickly the corn was expected to grow over the approximate 12 day interval between Application No. 2 and Application No. 4. Under average to good growing conditions, it was thought that corn would be at approximately the V4 growth stage at Application No. 2 and at approximately the V6 growth stage at Application No. 3. However, the timing for Application No. 2 and Application No. 3 was scheduled relative to the expected date for Application No. 4 rather than based on target growth stages. All applications that took place following crop emergence were foliar over-the-top applications. Application dates and crop growth stage at application are listed in Appendix A, Table 7.

A non-ionic adjuvant, such as X-77, at approximately 0.25% (v/v) was added in the spray mixture with each application of 2,4-D, except one application of 2,4-D at the IL-06 trial site where non-ionic adjuvant was added to the spray mixture at 0.18% (v/v). With the exception of Trial MO-08 (See Appendix D, Protocol Deviation 5, Item 4), a crop oil concentrate, such as Agri-Dex, at approximately 1.25% (v/v) was added in the spray mixture with quizalofop-P-ethyl.

The route of administration was a backpack, tractor-, or ATV-mounted boom sprayer with adequate agitation and output in order to simulate a typical commercial application. Application equipment used in these trials is described in Appendix A, Table 8. Spray pressure and nozzles were selected to provide a uniform broadcast spray avoiding drift and misting. Application equipment was calibrated no more than two days prior to the application event, but generally occurred on the day of application. A targeted spray volume equivalent to approximately 200 - 400 L/ha (~20 - 40 gallons/acre) for thorough coverage of the treated plot area was used for each application. Details concerning the spray volume used in each application in all trials are given in Appendix A, Table 7.

In this study the application rate for 2,4-D was expressed on an acid equivalent (ae) basis, so the target application rate was expressed in units of lb ae/A or g ae/ha. The application rate for quizalofop-P-ethyl was expressed on an active ingredient (ai.) basis, so the target application rate was in units of lb ai/A or g ai/ha.

In the RAC treated plots, there were two broadcast spray applications of 2,4-D, each at approximately 1.0 lb ae/A (~1120 g ae/ha), one application of quizalofop-P-ethyl at approximately 0.082 lb ai/acre (~92 g/ha), and one application of 2,4-D at approximately 1.0 lb ae/A (~1120 g ae/ha). All applications were within the range of 95 -110% of the target rates.

For the 2X-rate processing treated plots, there were an initial two broadcast spray applications of 2,4-D each at approximately 2.0 lb ae/A (~2240 g ae/ha), one application of quizalofop-P-ethyl at approximately 0.164 lb ai/A (~184 g/ha), and a final application of 2,4-D at approximately 2.0 lb ae/A (~2240 g acid equivalent/ha). Again, each application was within the range of 95 -110% of the target rates.

A summary of all application rates for each trial is presented in, Appendix A, Table 7.

3.5 Tank Mix Stability

A tank mix stability study was conducted with Weedar 64 herbicide, a commercially available product containing 2,4-D dimethylamine salt which was one of the test substances used in this study. Results of the tank mix study showed 2,4-D to be uniformly distributed and chemically stable in spray mixture for a period of 24 hours (Reference 9). Spray mixtures used in the trials in this study were applied within 3 hours of preparation and were typically applied within approximately 30-60 minutes of preparation. Therefore, the 2,4-D dimethylamine salt was expected to remain uniformly distributed and chemically stable in the spray mixtures used in this study from the time of preparation until applied.

A tank mix stability study was also conducted with Assure II herbicide, a commercially available product containing the active ingredient quizalofop-P-ethyl, which was the other test substance used in this study. Results of the study showed quizalofop-P-ethyl to be uniformly distributed and chemically stable in the spray mixture for a period of 24 hours (Reference 10). Spray mixtures with Assure II herbicide used in the trials in this study were applied within 3 hours of preparation and were typically applied within approximately 30-60 minutes of preparation. Therefore, the quizalofop-P-ethyl was expected to remain uniformly distributed and chemically stable in the spray mixtures used in this study from the time of preparation until applied.

3.6 Sample Handling and Preparation

For each raw agricultural commodity (RAC) at each site, a single, composite sample was taken from the control plot while duplicate, composite samples were taken independently from the treated plots. In all cases, sample collection was avoided near the ends of plot rows and in plot borders. Forage was collected at two growth stages, early and at the normal or “typical” timing. Grain and stover RAC samples were collected at normal harvest times. Additionally, at three trial sites samples were collected at five sampling events (six for forage) to evaluate residue decline. The developmental stage of the plants at RAC and decline sampling and the sampling dates are shown in the Field Phase Summary, Appendix A, (Tables 9-33). To distinguish among samples collected at various intervals for labeling purposes the samples of early forage were designated as “Forage 40 DAT” and typical forage was designated as ‘Forage 60 DAT” and these designations are included in Appendix A, Tables 9-33. When samples were collected at five intervals in the decline trials, the third sampling interval served as the normal commercial harvest timing so that there were two sampling points before normal commercial harvest and also two sampling points after normal commercial harvest. Sampling information for decline trials is displayed in Appendix A, Tables 12, 19 and 30. In the decline trials as in the RAC trials, the samples of early forage are designated as "Forage 40 DAT" and the samples of "typical" forage are listed as "Forage 60 DAT". With grain and stover samples, the samples collected in RAC and decline trials at normal commercial harvest timing are designated as "Grain 90 DAT" or "Stover 90 DAT". In all cases, the actual PHI for 2,4-D and for quizalofop-P-ethyl for each sample is listed in Table I.

Field corn forage RAC samples were collected at two timings. The first timing was targeted at 40 days (± 2 days) after the final application of 2,4-D at the V8 stage or 48 in. corn, whichever came first. As explained above, these samples were designated as "Forage 40 DAT". This sample timing was intended to be earlier than "typical" and the forage samples collected at this timing were expected to not yet have reached the late dough growth stage. "Early" forage samples were collected at 27-42 days after the last application of 2,4-D.

Field corn forage RAC samples were also targeted to be collected at a second timing which was a "typical" late-dough growth to early-dent stage. For this sample timing, the actual number of days after the final application of 2,4-D was dependent on the rate of corn maturation to reach the typical growth stage rather than a pre-assigned number of days after treatment. As explained above, these samples were designated as "Forage 60 DAT". These forage samples were collected at 39-78 days following the last application.

For each corn forage sample, 12 plants located randomly across the plot were cut approximately 1 to 2 inches above the ground. The 12 plants collected for each sample were then divided into 3 groups of 4 plants each. Each corn plant was cut into 3 approximately equal length sections with the leaves and ears attached (top, middle and bottom section). The top, middle, and bottom sections of the plants, respectively, from each of the three groups of four plants were then composited into a single corn forage sample. The composite samples contained a part of each of the 12 collected plants and include 4 bottom sections, 4 middle sections and 4 top sections with each section coming from a separate corn plant.

Field corn grain RAC samples consisted of the seed removed from the husked cob. Grain samples were collected once the plant and ears had senesced and dried and the corn first reached maturity suitable for commercial harvest. The grain samples were collected on the same day that the stover samples were collected. For the decline trials, grain was collected before, at and after typical maturity. Each RAC grain sample consisted of at least 1 kg from at least 12 plants from different areas across the plot. Each bulk grain sample was about 1000 pounds (~450 kg).

Field corn stover RAC consisted of the dried, mature plant stalk remaining after collection and removal of the grain or whole ear (cob + grain). Stover samples were collected on the same day as the grain. The stover and grain samples were collected when the plants reach maturity suitable for commercial harvest, which typically occurred when the plants had dried to approximately 80 to 85% dry matter. For the decline trials, stover was collected before, at and after typical maturity.

For each corn stover sample, 12 plants located randomly across the plot were cut approximately 1 to 2 inches above the ground and the ears were removed. The 12 plants collected for each sample were then divided into 3 groups of 4 plants each. Each corn plant was cut into 3 approximately equal lengths sections with the leaves attached (top, middle and bottom section). The top, middle, and bottom sections of the plants, respectively, from each of the three groups of four plants were composited into a single corn stover sample. The composite sample contained a part of each of the 12 collected plants and included 4 bottom sections, 4 middle sections and 4 top sections with each section coming from a separate corn plant.

Grain and stover RAC samples were collected at maturity which was 73-139 days after the last application of 2,4-D. Additionally, residue decline samples were collected at three trial locations at six sampling intervals for forage and five intervals for grain and stover which were representative of crop growth before, at, and after typical maturity. As explained above, the RAC samples at normal commercial maturity were designated as “Grain 90 DAT” or “Stover 90 DAT”.

All samples were assigned a unique Dow AgroSciences sample number that was used to track each sample throughout collection, receipt, preparation, storage, analysis, and reporting. RAC and decline samples were placed in sample bags with labeling and were either placed in frozen storage within four hours of sampling or stored in coolers with dry ice until placed in a freezer. The samples remained in frozen storage until shipped by freezer truck to Dow AgroSciences in Indianapolis for preparation and analysis. A summary of sample storage and shipping information from field facilities is shown in Appendix A, Table 34.

The samples of grain collected for processing were placed in labeled boxes, each containing approximately 17-35 lb of grain. The grain was stored frozen 8-21 days after collection until shipment by freezer truck to GLP Technologies in Navasota, TX where the grain was processed. The samples of grain were stored frozen at GLP Technologies until processing began. After collection, samples of grain, aspirated grain fractions and specified processed products were stored frozen at GLP Technologies and were later packed in dry ice and shipped frozen by overnight carrier to Dow AgroSciences in Indianapolis for preparation and analysis.

Samples were received frozen at Dow AgroSciences and logged into the DAS Regulatory Labs Information Management System (RLIMS). Each sample (with the exception of oil which needed no preparation) was blended with dry ice and ground using an RSI Robo Coupe bowl grinder with a 45-, 30-, or 3-L capacity bowl. After appropriate mixing, each sample was transferred to HDPE freezer containers and stored in temperature-monitored freezers at approximately -20 °C, except when removed to aliquot for analysis. All movements of samples at the Dow AgroSciences facility were documented within the RLIMS system.

Field sample analyses were conducted 18-May-2010 through 30-Aug-2010, as defined by the addition of extraction solution to samples. The dates of sampling, shipment from the field, receipt at Dow AgroSciences, and analysis for each sample group are summarized in Table 1 of the Analytical Phase Summary in Appendix C of this report.

3.7 Production of Aspirated Grain Fractions and Corn Processed Products

As discussed in Section 3.3, bulk grain samples for use in processing were produced at the GA-03 and NE-14 trial sites from untreated plots and from plots that were treated with 2,4-D and quizalofop-P-ethyl at 2X rates of 2,4-D and quizalofop-p-ethyl. Two grain samples were collected in the field prior to shipment to GLP Technologies in Navasota, TX for processing. Three sub-samples of unprocessed grain from the bulk samples (grain from the processor) were collected at the processing facility just prior to screening and separation of the aspirated grain fractions (AGF). Additionally, three sub-samples of unprocessed grain were also collected after the grain had been screened and AGF had been separated from the grain.

The grain samples were stored frozen at GLP Technologies until processing began. Samples of unprocessed grain were collected on the day processing began. Samples of aspirated grain fractions were collected from untreated and treated whole grain. After the grain was cleaned by aspiration and screening, samples of the cleaned grain were processed by either a dry milling process or a wet milling process. Samples of grits, flour, meal, and refined oil produced in the dry milling process were collected for residue analysis. Samples of starch and refined oil produced in the wet milling process were collected for residue analysis. Samples of aspirated grain fractions and processed fractions produced from grain collected from untreated and treated corn were stored frozen. As described in Section 3.6, these samples were shipped to Dow AgroSciences for preparation, if necessary, and residue analysis. Details of the corn processing at GLP Technologies are presented in the Processing Phase Summary in Appendix B.

3.8 Reference Materials/Analytical Standards

The following analytical standards were used as the reference material for calibration and fortification standards: 2,4-D, AGR275828, 99.5% pure; 2,4-DCP, AGR182992, 99% pure; quizalofop-ester, TSN106317, 99% pure; quizalofop acid, TSN106172, 96% pure. A 1000 µg/mL stock solution of each reference standard was prepared in 100 mL of methanol from a 0.1000-g weighed reference standard. A range of 2,4-D and 2,4-DCP mixed stock solutions was prepared in methanol by serial dilutions. A range of quizalofop-ethyl and quizalofop acid mixed stock solutions was prepared in methanol by serial dilutions The reagents 2,4-dichlorophenoxyacetic acid ($^{13}C_6$) , lot #SCHF-006, and 2,4-dichlorophenol ($^{13}C_6$) , lot #SCCJ-003, were used as stable isotope internal standards in the study. The stable isotopes used in this study required evaporative removal of the solvent they were delivered in to allow stock preparation. A stock solution of each stable isotope was prepared in a methanol/water (50:50) solution, and a further dilution containing both internal standards was prepared in a methanol/2N HCl (50:50) solution. A range of mixed calibration standards of 2,4-D and 2,4-DCP, containing 10 ng/mL of the 2,4-dichlorophenoxyacetic acid ($^{13}C_6$) and 2,4-dichlorophenol ($^{13}C_6$) internal standards, was prepared in a methanol/2N HCl (50:50) solution. A range of mixed calibration standards containing quizalofop-ethyl and quizalofop acid was prepared in a methanol/2N HCl (50:50) solution. Common and chemical names, molecular formulae, and the nominal masses for the analytes and related compounds are shown in Appendix C, Table 2.

3.9 Analytical Methodology

One analytical method was used for the determination of 2,4-dichlorophenoxyacetic acid (2,4-D) and 2,4-dichlorophenol (2,4-DCP), while a separate analytical method was used for the determination of quizalofop-ethyl and quizalofop acid in corn commodities. The two methods are outlined in detail in the Analytical Phase Summary in Appendix C.

The limit of detection (LOD) and limit of quantitation (LOQ) for all analytes on all matrices were established at the initiation of the analytical phase of the study at 0.003 µg/g and 0.01 µg/g, respectively, for both methods.

3.10 Calculations

Calculations for the determination of all analytes were performed using linear regression with 1/x weighting. A series of calibration standards were included with the samples in each chromatographic run. The detector response was determined by measuring peak areas for the analyte and internal standard in the standard and sample solution chromatograms. Quantitation ratios were then calculated using the analyte and internal standard peak areas. The equation for the calibration curve was calculated using the analyte concentration of the standards and the corresponding quantitation ratios.

The concentrations in the field and recovery samples were then calculated along with the average percent recovery for the fortified samples in the same analytical set. The equations used for the calculations along with example calculations are presented in Appendix C.

3.11 Statistical Analysis

The statistics used for this study were linear with 1/x weighting regression analysis, means, medians, standard deviations, relative standard deviations, and coefficients of determination.

4.0 RESULTS AND DISCUSSION

4.1 Storage Stability

The samples from this study were stored frozen for up to 376 days from the date of sampling to analysis. The maximum storage interval from collection until analysis was 376 days for forage, 335 days for stover, and 320 days for grain. A summary of the storage intervals is in Appendix C, Table 1. Frozen storage stability of 2,4-D in corn forage, fodder (stover) and grain has previously been evaluated and no significant degradation was found for a period of 385 days, which was the maximum period evaluated (Reference 11). A frozen storage stability study with 2,4-DCP, quizalofop-ethyl and quizalofop acid in corn commodities was initiated in 2010 and is currently in progress (Dow AgroSciences study ID 101560). Results from this frozen storage stability study were not available at the time this study report was finalized and will be reported separately. Samples were stored in temperature-monitored freezers at Dow AgroSciences at approximately -20 ºC and a summary of field storage conditions for samples is given in Appendix A, Table 34.

4.2 Analytical Method Performance

The efficiency of the analytical method was determined at the time of analysis of each set of samples by fortifying the appropriate control matrix with the analyte(s) of concern and analyzing according to the analyte-specific method. An unfortified control matrix was also included in each set as well. Fortified recoveries were analyzed over a range of 0.01 to 20 µg/g for 2,4-D and 2,4-DCP and over a range of 0.01 to 1 µg/g for quizalofop-ethyl and quizalofop acid. The detailed concurrent method recovery data is presented in Appendix C, Table 3 and the summarized concurrent method recovery data is presented in Appendix C, Table 4. Typical chromatograms from the analyses can also be found in Appendix C. Method performance was demonstrated, with concurrent recoveries across fortification levels and matrices ranging from 77-109% for 2,4-D, 61-112% for 2,4-DCP, 72-116% for quizalofop-ethyl, and 75-143% for quizalofop acid. Several individual and averaged concurrent recovery values, while outside a commonly accepted range of 70-120%, were deemed acceptable due to consistency across all recovery samples of a given matrix.

4.3 Residue Results

The analytical data to support these residue data are presented in the Analytical Phase Summary report (Appendix C). Residue results are expressed as uncorrected (i.e. not corrected for concurrent recoveries) and reported concentrations. The reported concentration is the uncorrected value with the LOD and LOQ applied. The residue concentration was reported as ND (Not Detected) if the uncorrected concentration was below the LOD (0.003 ug/g). The residue concentration was reported as the uncorrected concentration in parenthesis, ( ), or brackets, [ ], if the concentration was less than the LOQ (0.01 ug/g) but greater than or equal to the LOD (0.003 ug/g). The residue concentration was reported as the uncorrected concentration if the uncorrected concentration was equal to or greater than the LOQ. Residue concentrations reported between the LOD and LOQ have a higher degree of uncertainty than values above the LOQ.

4.3.1 2,4-D and 2,4-DCP Residues in RAC and Decline Samples

Residues of 2,4-D and 2,4-DCP in field corn forage, grain and stover after three applications of 2,4-D (preemergence, and two postemergence over-the-top applications at approximately the V4-V5 growth stage and 12 days later at approximately the V8 stage) with each application at a target rate of 1.0 lb ae/A (1120 g ae/ha) applied with non-ionic surfactant at approximately 0.25% (v/v) are presented in Tables II, IV and VI, respectively. Results for a single grain sample from the MO-08 trial site are highly divergent from the rest of the data set. With corn grain at the MO-08 trial site, one of the two replicate samples from the treated plot had non-detectable residues (ND) of 2,4-D, whereas the other replicate sample had a 2,4-D residue of 0.067 ug/g. Out of the 25 trial sites / 50 grain samples from treated plots, this grain sample is the only one with residue above the LOQ and there were only 3 other grain samples from treated plots that had detectable residues (i.e. near the LOD of 0.003 ug/g) (see Figure 3 for a graphic distribution of residue results in corn grain). No specific explanation was found for the anomalous residue in this sample. However, given that 46 out of 50 other grain samples had 2,4-D residues that were ND and that three of the remaining four samples had residues that were near the LOD of 0.003 ug/g, the single value of 0.067 ug/g in one replicate sample when the other replicate sample has ND residue strongly suggests that the 2,4-D residue reported in this sample is the result of contamination. It is not believed to be a true representation of a 2,4-D residue level in grain that resulted from the application pattern for 2,4-D used in this study. Residues of 2,4-DCP were not detected in either of the two samples of grain from the treated plot at the MO-08 trial site. Therefore, the results for 2,4-D analysis in grain samples from the treated plot at the MO-08 trial site are not considered reliable and neither of the replicate values for 2,4-D in grain have been included in the table that provides a summary of residues for 2,4-D (Table XVIII).

Untreated samples of forage and grain were found to have no detectable residues of 2,4-D, but some of the untreated samples had detectable levels of 2,4-DCP, although all were at or below the LOQ of 0.01 ug/g. A few untreated samples of stover had detectable residues of 2,4-D, but none were above the LOQ of 0.01 ug/g. Many of the untreated samples of stover had detectable residues of 2,4-DCP and three of the samples had 2,4-DCP residues that were above the LOQ of 0.01 ug/g. No specific explanation could be determined for residues of 2,4-D found in certain untreated samples of stover or 2,4-DCP found in some untreated samples of forage, grain, and stover. It is assumed that residues in these samples resulted from contamination.

A summary of all residue data for 2,4-D and 2,4-DCP in corn forage, grain and stover RAC samples is presented in Table XVIII, which includes minimum, maximum, mean, median, standard deviation, and Highest Average Field Trial (HAFT) residue values for RAC samples in the 25 trials. As discussed above, 2,4-D residues in grain were not included in this summary for grain from the MO-08 trial location since those residues are thought to be unreliable. As shown in Table XVIII, the HAFT residues in early field corn forage at 27-42 days after the last application were 2.612 µg/g and 3.056 µg/g for 2,4-D and 2,4-DCP, respectively. The HAFT residues in typical field corn forage at 39-78 days after the last application were 3.124 µg/g and 3.899 µg/g for 2,4-D and 2,4-DCP, respectively. The HAFT residues in field corn grain at 73-139 days after the last application were (0.004) µg/g (based on excluding the 2,4-D values in grain from trial site MO-08 or the HAFT would be 0.034 µg/g if trial site MO-08 grain is included) and ND for 2,4-D and 2,4-DCP, respectively. The HAFT residues in field corn stover at 73-139 days after the last application were 8.076 µg/g and 5.377 µg/g for 2,4-D and 2,4-DCP, respectively. A summary of the PHIs for 2,4-D and 2,4-DCP in corn forage, grain and stover by trial site is presented in Table I. Residue data for 2,4-D and 2,4-DCP from this study are evaluated with regard to potential human dietary exposure and existing U.S. tolerances for 2,4-D in a separate report by Stagg, et.al. (Reference 12).

A summary of residue data for 2,4-D and 2,4-DCP from the three decline trials (GA-04, MI-11 and OK-22) is presented in Tables III (forage), V (grain), and VII (stover). The residue data from the decline trials field corn forage samples showed a reasonably clear pattern of decline for 2,4-D and 2,4-DCP over the six sampling events during the 30- to 33-day collection intervals. A residue decline profile could not be established for 2,4-D or 2,4-DCP on corn grain as all residue values determined were below or just slightly above the limit of detection. The residue data from the decline trials field corn stover samples showed a reasonably clear pattern of decline for 2,4-D and 2,4-DCP over the five sampling events during the 28-day collection interval in two of the three trials (Trials GA-04 and OK-22). The stover samples from the third decline trial (Trial MI-11) did not exhibit decline over the five samplings as residue values remained relatively constant. See Figures 4, 5, and 6 for a graphic representation of the decline of 2,4-D and 2,4-DCP residues of forage and stover in RAC samples collected from the three decline trials, GA-04, MI-11, and OK-22, respectively.

4.3.2 Quizalofop-ethyl and Quizalofop Acid Residues in RAC and Decline Samples

The residues of quizalofop-ethyl and quizalofop acid in field corn forage, grain and stover after a single postemergence over-the-top application of quizalofop-P-ethyl at a target rate of 0.082 lb ai./A (92 g ai./ha) with crop oil concentrate in the spray mixture at approximately 1.25% (v/v) applied at approximately the V6-V7 growth stage are presented in Tables X, XII, XIV, respectively. Since the date of application of quizalofop-P-ethyl is a few days earlier than the final application of 2,4-D, the PHIs for quizalofop-ethyl and quizalofop acid in samples in this study are slightly different than those presented for 2,4-D. A summary of the PHIs for quizalofop-ethyl and quizalofop acid in corn forage, grain and stover by trial site is presented in Table I.

Forage results from the "typical" sample timing at the TX-23 trial site are questionable. The forage samples collected at the "typical" sample timing at the TX-23 trial site had quizalofop-ethyl and quizalofop acid residues at approximately 10x to 20x the LOQ. The single postemergence over-the-top application of quizalofop-P-ethyl took place in the TX-23 trial on 19-Jul-2009 and the "early" forage sample was collected 32 days later on 20-Aug-2009. Residues of quizalofop-ethyl and quizalofop acid were not detected (ND) in either of the replicate samples of early forage from the treated plot. However, 12 days later on 01-Sep-2009, the "typical" growth stage / timing forage samples were collected and at this time both replicates of forage samples from the treated plot had residues of quizalofop-ethyl and quizalofop acid that were approximately 10X – 20X the LOQ of 0.01 ug/g. All other "early" and "typical" forage samples from the 24 other trial sites / 96 samples had non-detectable residues of quizalofop-ethyl and quizalofop acid.

Although a thorough review of the field data from this trial did not reveal any possible causes for the aberrant residue results, these data points are highly questionable and are not considered representative of residues expected in forage following the application pattern in this study.

Residues of both "early" and "typical" forage from treated plots at all other trials sites (24 trials / 96 samples) had no detectable residues of quizalofop-ethyl or quizalofop acid and forage samples collected from the TX-23 trial site 12 days earlier had no detectable (ND) residues of quizalofop-ethyl or quizalofop acid, which strongly suggests that these residues did not result from the application pattern for quizalofop-P-ethyl used in this study. It is thought that the residues in these samples are likely a result of contamination or other occurrence that did not result from the proposed use pattern for quizalofop-P-ethyl in field corn containing the AAD-1 gene. Given the previous discussion, the results from quizalofop-ethyl and quizalofop acid analysis in the "typical" forage samples from the treated plot at the TX-23 trial site are not considered reliable and neither of the replicate sample values for quizalofop-ethyl or quizalofop-acid have been included in the table that provides a summary of residues for quizalofop-ethyl and quizalofop acid (Table XIX).

One sample of forage from an untreated plot had a detectable level of quizalofop-ethyl, but it was less than the LOQ of 0.01 ug/g. One sample of grain had a detectable level of quizalofop-ethyl and quizalofop acid, but the levels were less than the LOQ of 0.01 ug/g. All other untreated samples of forage and grain, and all untreated samples of stover had no detected (ND) residues of quizalofop-ethyl or quizalofop acid. Although no explanation could be determined, it is assumed that residues in the one untreated forage sample and the one untreated grain sample resulted from contamination.

A summary of all residue data for quizalofop-ethyl and quizalofop acid in corn forage, grain and stover RAC samples is presented in Table XIX, which includes minimum, maximum, mean, median, standard deviation, and HAFT residue values for RAC samples in the 25 trials. As discussed above, quizalofop-ethyl and quizalofop acid residues in "typical" forage were not included in this summary for forage from the TX-23 trial location since those residues are thought to be unreliable. As shown in Table XIX, in both "early" forage at 32-47 days after application and "typical" forage at 44-83 days after application (if the TX-23 "typical" forage data points that are suspected to be due to contamination are excluded) residues of quizalofop-ethyl and quizalofop acid were not detected (ND,<0.003 ug/g). Residues in field corn grain at 79-144 days after the last application were less than the limit of quantitation (<0.01 μg/g) for quizalofop acid and were below the limit of detection (<0.003 μg/g) for quizalofop-ethyl. However, the HAFT in grain for the combined residues of quizalofop-ethyl and quizalofop expressed as quizalofop-ethyl is ND (< 0.003 μg/g). The HAFT residues in field corn stover at 79-144 days after the last application for the combined residues of quizalofop-ethyl and quizalofop expressed as quizalofop-ethyl is 0.016 μg/g .

A summary of the residue data for quizalofop-ethyl and quizalofop acid from the three decline trials is presented in Tables XI (forage), XIII (grain), and XV (stover). The residue decline profile could not be established for quizalofop-ethyl and quizalofop acid from the field corn forage or grain samples collected over the 30- to 33-day collection interval as nearly all of the forage samples yielded results below the limit of detection with just four data points slightly above the LOD. All grain samples were below the limit of detection. A clear pattern of decline for quizalofop-ethyl and quizalofop acid residues in field corn stover could not be established over the 28-day collection interval. All three trials yielded residue results for quizalofop-ethyl that were below the limit of detection. For quizalofop acid, one trial yielded non-detectable residue for all five sampling events, one trial had only 3 out of 10 samples from treated plots with detections slightly above the limit of detection, and one trial yielded residue that increased slightly (0.015 ug/g) over the five sampling intervals.

4.3.3 Residue Results from Processing Trials

For both the GA-03 and NE-14 trial sites, residues of 2,4-D in unprocessed grain (from the processor prior to separation of AGF) were below the limit of detection. For the GA-03 trial site, one of the three subsamples of grain collected before separation of AGF had 2,4-DCP residues at the limit of detection, but 2,4-DCP residues in the two other subsamples were below the limit of detection. Residues of 2,4-DCP in all three subsamples of grain from trial GA-03 collected after separation of AGF were below the limit of detection.

For the NE-14 trial site, all three subsamples of grain collected at the processor before screening and separation of AGF had residues of 2,4-DCP with the an average residue value for 2,4-DCP in the three subsamples of (0.008) ug/g. However, after screening and separation of AGF, 2,4-DCP was below the limit of detection in all three sub-samples of grain. In the NE-14 trial, there was no evidence of concentration of 2,4-D or 2,4-DCP into the aspirated grain fractions. Since 2,4-DCP was found in grain from the NE-14 trial at an average concentration of 0.008 ug/g before screening and separation of AGF, but residues of 2,4-DCP were below the limit of detection in the AGF, it is possible that the 2,4-DCP residues were associated with larger fragments / plant debris that were separated by screening and were not included as part of the AGF.

In the GA-03 trial, although residues of 2,4-D and 2,4-DCP were below the limit of detection in grain, the AGF separated from this grain had residues of 2,4-D and 2,4-DCP of 0.021 ug/g and (0.006) ug/g, respectively, indicating some concentration of residues of both compounds into the AGF. However, a processing factor was not calculated since no residues were detected in the grain prior to separation of the AGF. Residues of 2,4-D and 2,4-DCP were below the limit of detection in all samples of flour, grits, meal, starch and oil (produced by wet-milling and dry-milling) produced from grain in exaggerated rate (2X) treated plots from the GA-03 and NE-14 trial sites, indicating no concentration of 2,4-D or 2,4-DCP residues into these processed fractions. (See Tables VIII and IX and a summary in Table XX.)

No residues of quizalofop-ethyl or quizalofop acid above the limit of detection were found in unprocessed grain from the bulk grain samples (grain from the processor) collected at GA-03 trial site. Although residues of quizalofop-ethyl and quizalofop acid were below the limit of detection in two of the three subsamples collected from the bulk treated grain sample from the NE-14 trial site, residues of quizalofop-ethyl and quizalofop acid were detected at levels below the LOQ, (0.004) ug/g and (0.005) ug/g, respectively, in one subsample of grain prior to screening and separation of AGF, which resulted in an average value for the combined residues of quizalofop-ethyl and quizalofop acid of (0.003) ug/g. However, after screening and removal of AGF from the grain, residues of quizalofop-ethyl and quizalofop acid were below the limit of detection in all three subsamples.

Residues of quizalofop-ethyl and quizalofop acid were below the limit of detection in all samples of flour, grits, meal, starch and oil (produced by wet-milling and dry-milling) produced from grain in exaggerated rate (2X) treated plots from the GA-03 and NE-14 trial sites, except for one of two replicate samples of starch from the NE-14 trial site in which quizalofop-ethyl was found at (0.005) ug/g, which is a concentration below the LOQ (see Tables XVI and XVII). Given the low level of residue in the one of two starch samples from the NE-14 trial site and that low-level detection of quizalofop-ethyl were found in untreated control samples of meal and refined oil, it is possible that the detection of quizalofop-ethyl in starch in one sample was due to contamination. In starch, the average of the combined residues of quizalofop-ethyl and quizalofop acid expressed as quizalofop-ethyl is ND (<0.003 ug/g).

In consideration of all results for quizalofop-ethyl and quizalofop acid in grain samples collected from the exaggerated rate (2X) treated plots and the subsequent processed fractions, it does not appear that there is an appreciable potential for concentration of these compounds into AGF or processed fractions of field corn. A summary of processing residue data for the combined residues of quizalofop-ethly and quizalofop acid expressed as quizalofop-ethyl is presented in Table XX.

5.0 CONCLUSIONS

The agronomic and environmental conditions experienced during the course of this study are representative of typical farming practices; therefore, the residue values obtained would likely not be exceeded for the use patterns evaluated in this study.

6.0 RETENTION OF RECORDS

The final report, protocol and any amendments and deviations and all original field raw data and records generated except for facility raw data, such as calibration logs and chemical and freezer temperature logs, will be archived temporarily at Ag Research Associates, LLC, 1730 Denham Road, Sycamore, GA 31790 until study completion. Shortly following study completion all original study records, with the exception of facility raw data, will be transferred to the Sponsor's archives at Dow AgroSciences LLC, 9330 Zionsville Road, Indianapolis, Indiana 46268 for final archive. Facility records are being maintained by the corresponding facilities. Verified copies of facility records may be requested by the Sponsor.

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Tables

TABLE I - Summary of Pre-Harvest Intervals (PHI) Following the Last Application of Each Test Substance

	2,4-D			**Quizalofop-P-ethyl**		
Trial ID	**Early Forage PHI (days)**	**Typical Forage PHI (days)**	**Grain and Stover PHI (days)**	**Early Forage PHI (days)**	**Typical Forage PHI (days)**	**Grain and Stover PHI (days)**
NJ-01	38	66	98	44	71	111
PA-02	42	56	107	47	61	112
GA-03	38	43	73	43	48	79
GA-04*	30	46	76	34	50	80
IL-05	40	69	135	46	73	139
IL-06	38	63	118	44	69	124
IN-07	42	61	139	47	66	144
MO-08	40	47	104	45	52	109
MO-09	41	48	105	46	53	110
IL-10	40	43	100	45	48	105
MI-11*	40	60	90	45	64	95
OH-12	40	55	90	45	60	95
IN-13	40	60	90	45	65	95
NE-14	No sample	57	134	No sample	63	140
NE-15	42	66	83	47	71	94
KS-16	38	64	94	43	69	98
IA-17	38	60	103	43	65	108
MO-18	40	53	100	46	59	106
IA-19	41	68	121	46	73	126
MN-20	40	68	121	45	73	126
MO-21	42	57	90	47	62	95
OK-22*	30	49	79	35	54	84
TX-23	27	39	107	32	44	112
ON-24	41	78	126	46	83	131
ON-25	39	68	123	44	73	128
PHI Range:	27-42	39-78	73-139	32-47	44-83	79-144
DECLINE*						
GA-04	30, 32, 39, 46, 53, 60		62, 69, 76, 83, 90	34, 36, 43, 50, 57, 64		66, 73, 80, 87, 94
MI-11	40, 46, 53, 59, 66, 73		76, 83, 90, 97, 104	45, 51, 58, 64, 71, 78		81, 88, 95, 102,109
OK-22	30, 37, 43, 49, 55, 61		65, 71, 79, 86, 93	35, 42, 48, 54, 60, 66		70, 76, 84, 91,98

* Decline trial

TABLE II - Residues of 2,4-D and 2,4-DCP in Corn Forage

State/ Trial No.	NAFTA Region	AAD-1 Event[a] DAS-	RAC	PHI[b] (days)	Plot ID	Sample ID ARA-09-15-10-	2,4-D (µg/g)	2,4-D Avg[c] (µg/g)	2,4-DCP (µg/g)	2,4-DCP Avg[c] (µg/g)
NJ-01	1	------	Forage	NA	T1	001-0001A2	ND[d]	------	ND	------
NJ-01	1	40278-9	Forage	38	T2	001-0002A2	2.236	------	1.778	------
NJ-01	1	40278-9	Forage	38	T2	001-0003A2	2.988	2.612	2.190	1.984
NJ-01	1	------	Forage	NA	T1	002-0001A2	ND	------	ND	------
NJ-01	1	40278-9	Forage	66	T2	002-0002A2	3.353	------	1.982	------
NJ-01	1	40278-9	Forage	66	T2	002-0003A2	2.069	2.711	1.366	1.674
PA-02	1	------	Forage	NA	T1	005-0001A2	ND	------	ND	------
PA-02	1	40278-9	Forage	42	T2	005-0002A2	2.227	------	3.997	------
PA-02	1	40278-9	Forage	42	T2	005-0003A2	1.566	1.896	2.115	3.056
PA-02	1	------	Forage	NA	T1	006-0001A2	ND	------	[0.007][e]	------
PA-02	1	40278-9	Forage	56	T2	006-0002A2	1.775	------	3.102	------
PA-02	1	40278-9	Forage	56	T2	006-0003A2	1.805	1.790	2.689	2.895
GA-03	2	------	Forage	N/A	T1	009-0001A2	ND	------	ND	------
GA-03	2	40278-9	Forage	38	T2	009-0002A2	0.616	------	0.395	------
GA-03	2	40278-9	Forage	38	T2	009-0003A2	0.688	0.652	0.551	0.473
GA-03	2	------	Forage	N/A	T1	010-0001A2	ND	------	[0.004]	------
GA-03	2	40278-9	Forage	43	T2	010-0002A2	0.336	------	0.169	------
GA-03	2	40278-9	Forage	43	T2	010-0003A2	0.481	0.408	0.174	0.171
GA-04	2	------	Forage	N/A	T1	014-0001A2	ND	------	ND	------
GA-04	2	40278-9	Forage	30	T2	014-0002A2	0.888	------	0.692	------
GA-04	2	40278-9	Forage	30	T2	014-0003A2	1.168	1.028	0.927	0.809
GA-04	2	------	Forage	N/A	T1	017-0001A2	ND	------	ND	------
GA-04	2	40278-9	Forage	46	T2	017-0002A2	[0.006][e]	------	[0.005]	------
GA-04	2	40278-9	Forage	46	T2	017-0003A2	ND	[0.003][f]	[0.004]	[0.005]
IL-05	5	------	Forage	N/A	T1	030-0001A3	ND	------	ND	------

State/ Trial No.	NAFTA Region	AAD-1 Event[a] DAS-	RAC	PHI[b] (days)	Plot ID	Sample ID ARA-09-15-10-	2,4-D (µg/g)	2,4-D Avg[c] (µg/g)	2,4-DCP (µg/g)	2,4-DCP Avg[c] (µg/g)
IL-05	5	40278-9	Forage	40	T2	030-0002A3	0.449	------	0.423	------
IL-05	5	40278-9	Forage	40	T2	030-0003A3	0.303	0.376	0.478	0.450
IL-05	5	------	Forage	N/A	T1	031-0001A2	ND	------	ND	------
IL-05	5	40278-9	Forage	69	T2	031-0002A2	0.571	------	0.614	------
IL-05	5	40278-9	Forage	69	T2	031-0003A2	0.903	0.737	1.199	0.906
IL-06	5	------	Forage	N/A	T1	034-0001A2	ND	------	ND	------
IL-06	5	40278-9	Forage	38	T2	034-0002A2	0.436	------	0.889	------
IL-06	5	40278-9	Forage	38	T2	034-0003A2	0.469	0.453	0.848	0.868
IL-06	5	------	Forage	N/A	T1	035-0001A2	ND	------	ND	------
IL-06	5	40278-9	Forage	63	T2	035-0002A2	0.223	------	0.437	------
IL-06	5	40278-9	Forage	63	T2	035-0003A2	0.219	0.221	0.528	0.482
IN-07	5	------	Forage	N/A	T1	038-0001A2	ND	------	ND	------
IN-07	5	40278-9	Forage	42	T2	038-0002A2	1.054	------	1.914	------
IN-07	5	40278-9	Forage	42	T2	038-0003A2	1.297	1.176	2.115	2.014
IN-07	5	------	Forage	N/A	T1	039-0001A2	ND	------	ND	------
IN-07	5	40278-9	Forage	61	T2	039-0002A2	1.577	------	2.431	------
IN-07	5	40278-9	Forage	61	T2	039-0003A2	0.595	1.086	0.987	1.709
M0-08	5	------	Forage	N/A	T1	042-0001A2	ND	------	[0.005]	------
M0-08	5	40278-9	Forage	40	T2	042-0002A2	0.010	------	0.0115	------
M0-08	5	40278-9	Forage	40	T2	042-0003A2	ND	[0.005]	[0.003]	[0.007]
M0-08	5	------	Forage	N/A	T1	043-0001A2	ND	------	[0.006]	------
M0-08	5	40278-9	Forage	47	T2	043-0002A2	0.064	------	0.084	------
M0-08	5	40278-9	Forage	47	T2	043-0003A2	0.030	0.047	0.052	0.068
MO-09	5	------	Forage	N/A	T1	046-0001A2	ND	------	ND	------
MO-09	5	40278-9	Forage	41	T2	046-0002A2	1.597	------	1.685	------

State/ Trial No.	NAFTA Region	AAD-1 Event[a] DAS-	RAC	PHI[b] (days)	Plot ID	Sample ID ARA-09-15-10-	2,4-D (µg/g)	2,4-D Avg[c] (µg/g)	2,4-DCP (µg/g)	2,4-DCP Avg[c] (µg/g)
MO-09	5	40278-9	Forage	41	T2	046-0003A2	0.968	1.282	1.472	1.579
MO-09	5	------	Forage	N/A	T1	047-0001A2	ND	------	[0.004]	------
MO-09	5	40278-9	Forage	48	T2	047-0002A2	1.285	------	1.559	------
MO-09	5	40278-9	Forage	48	T2	047-0003A2	0.628	0.957	1.005	1.282
IL-10	5	------	Forage	N/A	T1	050-0001A2	ND	------	[0.009]	------
IL-10	5	40278-9	Forage	40	T2	050-0002A2	ND	------	[0.005]	------
IL-10	5	40278-9	Forage	40	T2	050-0003A2	0.084	0.042	0.113	0.059
IL-10	5	------	Forage	N/A	T1	051-0001A2	ND	------	[0.005]	------
IL-10	5	40278-9	Forage	43	T2	051-0002A2	0.018	------	0.029	------
IL-10	5	40278-9	Forage	43	T2	051-0003A2	0.024	0.021	0.025	0.027
MI-11	5	------	Forage	N/A	T1	054-0001A2	ND	------	[0.008]	------
MI-11	5	40278-9	Forage	40	T2	054-0002A2	1.475	------	3.005	------
MI-11	5	40278-9	Forage	40	T2	054-0003A2	1.599	1.537	2.673	2.839
MI-11	5	------	Forage	N/A	T1	057-0001A2	ND	------	[0.003]	------
MI-11	5	40278-9	Forage	60	T2	057-0002A2	3.167	------	4.147	------
MI-11	5	40278-9	Forage	60	T2	057-0003A2	2.026	2.596	3.204	3.676
OH-12	5	------	Forage	N/A	T1	070-0001A2	ND	------	ND	------
OH-12	5	40278-9	Forage	40	T2	070-0002A2	0.814	------	1.179	------
OH-12	5	40278-9	Forage	40	T2	070-0003A2	0.894	0.854	1.566	1.372
OH-12	5	------	Forage	N/A	T1	071-0001A2	ND	------	ND	------
OH-12	5	40278-9	Forage	55	T2	071-0002A2	0.346	------	0.564	------
OH-12	5	40278-9	Forage	55	T2	071-0003A2	0.222	0.284	0.751	0.658
IN-13	5	------	Forage	N/A	T1	074-0001A2	ND	------	[0.003]	------
IN-13	5	40278-9	Forage	40	T2	074-0002A2	1.831	------	3.233	------
IN-13	5	40278-9	Forage	40	T2	074-0003A2	0.609	1.220	2.008	2.620

State/ Trial No.	NAFTA Region	AAD-1 Event[a] DAS-	RAC	PHI[b] (days)	Plot ID	Sample ID ARA-09-15-10-	2,4-D (µg/g)	2,4-D Avg[c] (µg/g)	2,4-DCP (µg/g)	2,4-DCP Avg[c] (µg/g)
IN-13	5	------	Forage	N/A	T1	075-0001A2	ND	------	ND	------
IN-13	5	40278-9	Forage	60	T2	075-0002A2	1.339	------	2.792	------
IN-13	5	40278-9	Forage	60	T2	075-0003A2	2.445	1.892	5.007	3.899
NE-14	5	------	Forage	N/A	T1	079-0001A2	ND	------	ND	------
NE-14	5	40278-9	Forage	57	T2	079-0002A2	0.041	------	0.164	------
NE-14	5	40278-9	Forage	57	T2	079-0003A2	0.082	0.061	0.287	0.225
NE-15	5	------	Forage	N/A	T1	083-0001A2	ND	------	ND	------
NE-15	5	40278-9	Forage	42	T2	083-0002A2	0.022	------	0.012	------
NE-15	5	40278-9	Forage	42	T2	083-0003A2	[0.007]	0.015	0.047	0.030
NE-15	5	------	Forage	N/A	T1	084-0001A2	ND	------	ND	------
NE-15	5	40278-9	Forage	66	T2	084-0002A2	[0.005]	------	0.038	------
NE-15	5	40278-9	Forage	66	T2	084-0003A2	ND	ND	0.016	0.027
KS-16	5	------	Forage	N/A	T1	087-0001A2	ND	------	[0.007]	------
KS-16	5	40278-9	Forage	38	T2	087-0002A2	0.035	------	0.078	------
KS-16	5	40278-9	Forage	38	T2	087-0003A2	[0.008]	0.022	0.027	0.052
KS-16	5	------	Forage	N/A	T1	088-0001A2	ND	------	[0.004]	------
KS-16	5	40278-9	Forage	64	T2	088-0002A2	ND	------	0.016	------
KS-16	5	40278-9	Forage	64	T2	088-0003A2	0.013	[0.006]	0.023	0.020
IA-17	5	------	Forage	N/A	T1	091-0001A3	ND	------	ND	------
IA-17	5	40278-9	Forage	38	T2	091-0002A3	0.352	------	0.897	------
IA-17	5	40278-9	Forage	38	T2	091-0003A3	0.493	0.422	1.157	1.027
IA-17	5	------	Forage	N/A	T1	092-0001A2	ND	------	ND	------
IA-17	5	40278-9	Forage	60	T2	092-0002A2	0.160	------	0.459	------
IA-17	5	40278-9	Forage	60	T2	092-0003A2	0.211	0.186	0.665	0.562
MO-18	5	------	Forage	N/A	T1	095-0001A3	ND	------	0.010	------

State/ Trial No.	NAFTA Region	AAD-1 Event[a] DAS-	RAC	PHI[b] (days)	Plot ID	Sample ID ARA-09-15-10-	2,4-D (µg/g)	2,4-D Avg[c] (µg/g)	2,4-DCP (µg/g)	2,4-DCP Avg[c] (µg/g)
MO-18	5	40278-9	Forage	40	T2	095-0002A3	[0.005]	------	0.014	------
MO-18	5	40278-9	Forage	40	T2	095-0003A3	ND	ND	[0.003]	[0.008]
MO-18	5	------	Forage	N/A	T1	096-0001A2	ND	------	ND	------
MO-18	5	40278-9	Forage	53	T2	096-0002A2	0.012	------	0.040	------
MO-18	5	40278-9	Forage	53	T2	096-0003A2	[0.006]	[0.009]	0.016	0.028
IA-19	5	------	Forage	N/A	T1	099-0001A2	ND	------	[0.007]	------
IA-19	5	40278-9	Forage	41	T2	099-0002A2	0.273	------	1.547	------
IA-19	5	40278-9	Forage	41	T2	099-0003A2	0.523	0.398	2.152	1.850
IA-19	5	------	Forage	N/A	T1	100-0001A2	ND	------	ND	------
IA-19	5	40278-9	Forage	68	T2	100-0002A2	0.308	------	1.301	------
IA-19	5	40278-9	Forage	68	T2	100-0003A2	0.185	0.247	0.846	1.074
MN-20	5	------	Forage	N/A	T1	103-0001A2	ND	------	ND	------
MN-20	5	40278-9	Forage	40	T2	103-0002A2	0.480	------	1.496	------
MN-20	5	40278-9	Forage	40	T2	103-0003A2	0.732	0.606	2.211	1.854
MN-20	5	------	Forage	N/A	T1	104-0001A3	ND	------	ND	------
MN-20	5	40278-9	Forage	68	T2	104-0002A2	0.293	------	1.019	------
MN-20	5	40278-9	Forage	68	T2	104-0003A2	0.293	0.293	0.812	0.916
MO-21	5	------	Forage	N/A	T1	107-0001A3	ND	------	ND	------
MO-21	5	40278-9	Forage	42	T2	107-0002A2	0.331	------	0.323	------
MO-21	5	40278-9	Forage	42	T2	107-0003A2	0.377	0.354	0.345	0.334
MO-21	5	------	Forage	N/A	T1	108-0001A2	ND	------	ND	------
MO-21	5	40278-9	Forage	57	T2	108-0002A2	0.616	------	0.369	------
MO-21	5	40278-9	Forage	57	T2	108-0003A2	0.281	0.448	0.185	0.277
OK-22	6	------	Forage	N/A	T1	111-0001A2	ND	------	[0.003]	------
OK-22	6	40278-9	Forage	30	T2	111-0002A2	0.021	------	0.032	------

State/ Trial No.	NAFTA Region	AAD-1 Event[a] DAS-	RAC	PHI[b] (days)	Plot ID	Sample ID ARA-09-15-10-	2,4-D (µg/g)	2,4-D Avg[c] (µg/g)	2,4-DCP (µg/g)	2,4-DCP Avg[c] (µg/g)
OK-22	6	40278-9	Forage	30	T2	111-0003A2	0.253	0.137	0.130	0.081
OK-22	6	------	Forage	N/A	T1	114-0001A2	ND	------	ND	------
OK-22	6	40278-9	Forage	49	T2	114-0002A2	0.295	------	0.058	------
OK-22	6	40278-9	Forage	49	T2	114-0003A2	0.213	0.254	0.056	0.057
TX-23	6	------	Forage	N/A	T1	127-0001A2	ND	------	ND	------
TX-23	6	40278-9	Forage	27	T2	127-0002A2	ND	------	ND	------
TX-23	6	40278-9	Forage	27	T2	127-0003A2	ND	ND	ND	ND
TX-23	6	------	Forage	N/A	T1	128-0001A2	ND	------	ND	------
TX-23	6	40278-9	Forage	39	T2	128-0002A2	4.373	------	1.163	------
TX-23	6	40278-9	Forage	39	T2	128-0003A2	1.876	3.124	0.777	0.970
ON-24	5	------	Forage	N/A	T1	131-0001A2	ND	------	[0.003]	------
ON-24	5	40278-9	Forage	41	T2	131-0002A2	0.202	------	0.697	------
ON-24	5	40278-9	Forage	41	T2	131-0003A2	0.230	0.216	0.921	0.809
ON-24	5	------	Forage	N/A	T1	132-0001A3	ND	------	ND	------
ON-24	5	40278-9	Forage	78	T2	132-0002A2	0.013	------	0.0404	------
ON-24	5	40278-9	Forage	78	T2	132-0003A2	0.027	0.020	0.0886	0.065
ON-25	5	------	Forage	N/A	T1	135-0001A3	ND	------	ND	------
ON-25	5	40278-9	Forage	39	T2	135-0002A2	0.073	------	0.604	------
ON-25	5	40278-9	Forage	39	T2	135-0003A2	0.066	0.070	0.568	0.586
ON-25	5	------	Forage	N/A	T1	136-0001A3	ND	------	ND	------
ON-25	5	40278-9	Forage	68	T2	136-0002A2	0.030	------	0.200	------
ON-25	5	40278-9	Forage	68	T2	136-0003A2	0.030	0.030	0.275	0.237

[a] Untreated control plots (T1) were planted with the commercial variety Mycogen 2M746, which lacked the inserted AAD-1 gene, but had a genetic background similar to that used for the two non-commercial transgenic hybrids in treated plots (T2).
[b] PHI = Pre Harvest Interval (actual days after the last application of 2,4-D)
[c] Averages presented in this column are of treated samplings (T2 samples) only at each trial's sampling event
[d] ND = Not Detected (below the limit of detection of 0.003 µg/g)
[e] [ ] = Residue values greater than or equal to the limit of detection of 0.003 µg/g but less than the limit of quantitation of 0.01 µg/g. Values are reported with a lower degree of confidence than values above the limit of quantitation.
[f] For the purposes of calculating an average, "ND" is given the value of zero.

TABLE III – Decline Sampling Residues of 2,4-D and 2,4-DCP in Corn Forage

State/ Trial No.	NAFTA Region	AAD-1 Event[a] DAS-	RAC	PHI[b] (days)	Plot ID	Sample ID ARA-09-15-10-	2,4-D (µg/g)	2,4-D Avg[c] (µg/g)	2,4-DCP (µg/g)	2,4-DCP Avg[c] (µg/g)
GA-04	2	------	Forage	N/A	T1	014-0001A2	ND[d]	------	ND	------
GA-04	2	40278-9	Forage	30	T2	014-0002A2	0.888	------	0.692	------
GA-04	2	40278-9	Forage	30	T2	014-0003A2	1.168	1.028	0.927	0.809
GA-04	2	------	Forage	N/A	T1	015-0001A2	ND	------	ND	------
GA-04	2	40278-9	Forage	32	T2	015-0002A2	0.521	------	0.471	------
GA-04	2	40278-9	Forage	32	T2	015-0003A2	0.655	0.588	0.523	0.497
GA-04	2	------	Forage	N/A	T1	016-0001A2	ND	------	ND	------
GA-04	2	40278-9	Forage	39	T2	016-0002A2	0.447	------	0.394	------
GA-04	2	40278-9	Forage	39	T2	016-0003A2	0.124	0.286	0.010	0.247
GA-04	2	------	Forage	N/A	T1	017-0001A2	ND	------	ND	------
GA-04	2	40278-9	Forage	46	T2	017-0002A2	[0.006][e]	------	[0.005]	------
GA-04	2	40278-9	Forage	46	T2	017-0003A2	ND	[0.003][f]	[0.004]	[0.005]
GA-04	2	------	Forage	N/A	T1	018-0001A2	ND	------	ND	------
GA-04	2	40278-9	Forage	53	T2	018-0002A2	ND	------	ND	------
GA-04	2	40278-9	Forage	53	T2	018-0003A2	ND	ND	[0.004]	ND
GA-04	2	------	Forage	N/A	T1	019-0001A2	ND	------	ND	------
GA-04	2	40278-9	Forage	60	T2	019-0002A2	ND	------	[0.005]	------
GA-04	2	40278-9	Forage	60	T2	019-0003A2	ND	ND	[0.006]	[0.005]
MI-11	5	------	Forage	N/A	T1	054-0001A2	ND	------	[0.008]	------
MI-11	5	40278-9	Forage	40	T2	054-0002A2	1.475	------	3.005	------
MI-11	5	40278-9	Forage	40	T2	054-0003A2	1.599	1.537	2.673	2.839
MI-11	5	------	Forage	N/A	T1	055-0001A2	ND	------	[0.004]	------
MI-11	5	40278-9	Forage	46	T2	055-0002A2	0.712	------	1.471	------
MI-11	5	40278-9	Forage	46	T2	055-0003A2	0.913	0.812	2.144	1.807

State/ Trial No.	NAFTA Region	AAD-1 Event[a] DAS-	RAC	PHI[b] (days)	Plot ID	Sample ID ARA-09-15-10-	2,4-D (µg/g)	2,4-D Avg[c] (µg/g)	2,4-DCP (µg/g)	2,4-DCP Avg[c] (µg/g)
MI-11	5	------	Forage	N/A	T1	056-0001A3	ND	------	ND	------
MI-11	5	40278-9	Forage	53	T2	056-0002A3	1.419	------	2.137	------
MI-11	5	40278-9	Forage	53	T2	056-0003A3	2.510	1.965	4.084	3.111
MI-11	5	------	Forage	N/A	T1	057-0001A2	ND	------	[0.003]	------
MI-11	5	40278-9	Forage	60	T2	057-0002A2	3.167	------	4.147	------
MI-11	5	40278-9	Forage	60	T2	057-0003A2	2.026	2.596	3.204	3.676
MI-11	5	------	Forage	N/A	T1	058-0001A2	ND	------	[0.003]	------
MI-11	5	40278-9	Forage	67	T2	058-0002A2	1.937	------	3.273	------
MI-11	5	40278-9	Forage	67	T2	058-0003A2	2.115	2.026	4.291	3.782
MI-11	5	------	Forage	N/A	T1	059-0001A2	ND	------	ND	------
MI-11	5	40278-9	Forage	74	T2	059-0002A2	0.891	------	1.587	------
MI-11	5	40278-9	Forage	74	T2	059-0003A2	0.827	0.859	1.742	1.665
OK-22	6	------	Forage	N/A	T1	111-0001A2	ND	------	[0.003]	------
OK-22	6	40278-9	Forage	30	T2	111-0002A2	0.021	------	0.032	------
OK-22	6	40278-9	Forage	30	T2	111-0003A2	0.253	0.137	0.130	0.081
OK-22	6	------	Forage	N/A	T1	112-0001A2	ND	------	ND	------
OK-22	6	40278-9	Forage	37	T2	112-0002A2	0.252	------	0.094	------
OK-22	6	40278-9	Forage	37	T2	112-0003A2	0.273	0.262	0.073	0.083
OK-22	6	------	Forage	N/A	T1	113-0001A2	ND	------	[0.005]	------
OK-22	6	40278-9	Forage	43	T2	113-0002A2	0.187	------	0.079	------
OK-22	6	40278-9	Forage	43	T2	113-0003A2	1.095	0.641	0.207	0.143
OK-22	6	------	Forage	N/A	T1	114-0001A2	ND	------	ND	------
OK-22	6	40278-9	Forage	49	T2	114-0002A2	0.295	------	0.058	------
OK-22	6	40278-9	Forage	49	T2	114-0003A2	0.213	0.254	0.056	0.057

State/ Trial No.	NAFTA Region	AAD-1 Event[a] DAS-	RAC	PHI[b] (days)	Plot ID	Sample ID ARA-09-15-10-	2,4-D (µg/g)	2,4-D Avg[c] (µg/g)	2,4-DCP (µg/g)	2,4-DCP Avg[c] (µg/g)
OK-22	6	------	Forage	N/A	T1	115-0001A2	ND	------	[0.004]	------
OK-22	6	40278-9	Forage	55	T2	115-0002A2	0.192	------	0.032	------
OK-22	6	40278-9	Forage	55	T2	115-0003A2	0.107[g]	0.149	0.037[g]	0.035
OK-22	6	------	Forage	N/A	T1	116-0001A2	ND	------	[0.005]	------
OK-22	6	40278-9	Forage	61	T2	116-0002A2	0.059	------	0.036	------
OK-22	6	40278-9	Forage	61	T2	116-0003A2	0.084	0.071	0.030	0.033

[a] Untreated control plots (T1) were planted with the commercial variety Mycogen 2M746, which lacked the inserted AAD-1 gene, but had a genetic background similar to that used for the two non-commercial transgenic hybrids in treated plots (T2).

[b] PHI = Pre Harvest Interval (actual days after the last application of 2,4-D)

[c] Averages presented in this column are of treated samplings (T2 samples) only at each trial's sampling event

[d] ND = Not Detected (below the limit of detection of 0.003 µg/g)

[e] [ ] = Residue values greater than or equal to the limit of detection of 0.003 µg/g but less than the limit of quantitation of 0.01 µg/g. Values are reported with a lower degree of confidence than values above the limit of quantitation.

[f] For the purposes of calculating an average, "ND" is given the value of zero.

[g] Value is an average of six analyses which were conducted to ensure consistent performance of the method. The individual analytical values are given in detail in the analytical summary.

TABLE IV - Residues of 2,4-D and 2,4-DCP in Corn Grain

State/ Trial No.	NAFTA Region	AAD-1 Event[a] DAS-	RAC	PHI[b] (days)	Plot ID	Sample ID ARA-09-15-10-	2,4-D (µg/g)	2,4-D Avg[c] (µg/g)	2,4-DCP (µg/g)	2,4-DCP Avg[c] (µg/g)
NJ-01	1	------	Grain	NA	T1	003-0001A2	ND[d]	------	ND	------
NJ-01	1	40278-9	Grain	98	T2	003-0002A2	ND	------	ND	------
NJ-01	1	40278-9	Grain	98	T2	003-0003A2	ND	ND	ND	ND
PA-02	1	------	Grain	NA	T1	007-0001A2	ND	------	ND	------
PA-02	1	40278-9	Grain	107	T2	007-0002A2	ND	------	ND	------
PA-02	1	40278-9	Grain	107	T2	007-0003A2	ND	ND	ND	ND
GA-03	2	------	Grain	N/A	T1	011-0001A2	ND	------	ND	------
GA-03	2	40278-9	Grain	73	T2	011-0002A2	ND	------	ND	------
GA-03	2	40278-9	Grain	73	T2	011-0003A2	ND	ND	ND	ND
GA-04	2	------	Grain	N/A	T1	024-0001A2	ND	------	ND	------
GA-04	2	40278-9	Grain	76	T2	024-0002A2	ND	------	ND	------
GA-04	2	40278-9	Grain	76	T2	024-0003A2	ND	ND	ND	ND
IL-05	5	------	Grain	N/A	T1	032-0001A2	ND	------	ND	------
IL-05	5	40278-9	Grain	135	T2	032-0002A2	ND	------	ND	------
IL-05	5	40278-9	Grain	135	T2	032-0003A2	ND	ND	ND	ND
IL-06	5	------	Grain	N/A	T1	036-0001A2	ND	------	ND	------
IL-06	5	40278-9	Grain	118	T2	036-0002A2	ND	------	ND	------
IL-06	5	40278-9	Grain	118	T2	036-0003A2	ND	ND	ND	ND
IN-07	5	------	Grain	N/A	T1	040-0001A2	ND	------	ND	------
IN-07	5	40278-9	Grain	139	T2	040-0002A2	ND	------	ND	------
IN-07	5	40278-9	Grain	139	T2	040-0003A2	ND	ND	ND	ND
MO-08	5	------	Grain	N/A	T1	044-0001A2	ND	------	ND	------
MO-08	5	40278-9	Grain	104	T2	044-0002A4	0.067	------	ND	------
MO-08	5	40278-9	Grain	104	T2	044-0003A4	ND	0.034[e]	ND	ND
MO-09	5	------	Grain	N/A	T1	048-0001A2	ND	------	ND	------

State/ Trial No.	NAFTA Region	AAD-1 Event[a] DAS-	RAC	PHI[b] (days)	Plot ID	Sample ID ARA-09-15-10-	2,4-D (µg/g)	2,4-D Avg[c] (µg/g)	2,4-DCP (µg/g)	2,4-DCP Avg[c] (µg/g)
MO-09	5	40278-9	Grain	105	T2	048-0002A2	ND	------	ND	------
MO-09	5	40278-9	Grain	105	T2	048-0003A2	ND	ND	ND	ND
IL-10	5	------	Grain	N/A	T1	052-0001A2	ND	------	[0.007][f]	------
IL-10	5	40278-9	Grain	100	T2	052-0002A2	ND	------	ND	------
IL-10	5	40278-9	Grain	100	T2	052-0003A2	ND	ND	ND	ND
MI-11	5	------	Grain	N/A	T1	064-0001A2	ND	------	ND	------
MI-11	5	40278-9	Grain	90	T2	064-0002A2	ND	------	ND	------
MI-11	5	40278-9	Grain	90	T2	064-0003A2	ND	ND	[0.003]	ND
OH-12	5	------	Grain	N/A	T1	072-0001A2	ND	------	ND	------
OH-12	5	40278-9	Grain	90	T2	072-0002A2	[0.004]	------	ND	------
OH-12	5	40278-9	Grain	90	T2	072-0003A2	ND	ND	ND	ND
IN-13	5	------	Grain	N/A	T1	076-0001A2	ND	------	ND	------
IN-13	5	40278-9	Grain	90	T2	076-0002A2	ND	------	ND	------
IN-13	5	40278-9	Grain	90	T2	076-0003A2	ND	ND	ND	ND
NE-14	5	------	Grain	N/A	T1	080-0001A2	ND	------	ND	------
NE-14	5	40278-9	Grain	134	T2	080-0002A2	ND	------	ND	------
NE-14	5	40278-9	Grain	134	T2	080-0003A2	ND	ND	ND	ND
NE-15	5	------	Grain	N/A	T1	085-0001A2	ND	------	ND	------
NE-15	5	40278-9	Grain	83	T2	085-0002A2	ND	------	ND	------
NE-15	5	40278-9	Grain	83	T2	085-0003A2	ND	ND	ND	ND
KS-16	5	------	Grain	N/A	T1	089-0001A2	ND	------	[0.003]	------
KS-16	5	40278-9	Grain	94	T2	089-0002A2	ND	------	ND	------
KS-16	5	40278-9	Grain	94	T2	089-0003A2	ND	ND	ND	ND
IA-17	5	------	Grain	N/A	T1	093-0001A2	ND	------	ND	------
IA-17	5	40278-9	Grain	103	T2	093-0002A2	ND	------	ND	------

State/ Trial No.	NAFTA Region	AAD-1 Event[a] DAS-	RAC	PHI[b] (days)	Plot ID	Sample ID ARA-09-15-10-	2,4-D (µg/g)	2,4-D Avg[c] (µg/g)	2,4-DCP (µg/g)	2,4-DCP Avg[c] (µg/g)
IA-17	5	40278-9	Grain	103	T2	093-0003A2	ND	ND	ND	ND
MO-18	5	------	Grain	N/A	T1	097-0001A2	ND	------	ND	------
MO-18	5	40278-9	Grain	100	T2	097-0002A2	ND	------	ND	------
MO-18	5	40278-9	Grain	100	T2	097-0003A2	ND	ND	ND	ND
IA-19	5	------	Grain	N/A	T1	101-0001A2	ND	------	ND	------
IA-19	5	40278-9	Grain	121	T2	101-0002A2	ND	------	ND	------
IA-19	5	40278-9	Grain	121	T2	101-0003A2	ND	ND	ND	ND
MN-20	5	------	Grain	N/A	T1	105-0001A3	ND	------	ND	------
MN-20	5	40278-9	Grain	121	T2	105-0002A3	ND	------	ND	------
MN-20	5	40278-9	Grain	121	T2	105-0003A3	ND	ND	ND	ND
MO-21	5	------	Grain	N/A	T1	109-0001A2	ND	------	ND	------
MO-21	5	40278-9	Grain	90	T2	109-0002A2	ND	------	ND	------
MO-21	5	40278-9	Grain	90	T2	109-0003A2	ND	ND	ND	ND
OK-22	6	------	Grain	N/A	T1	121-0001A2	ND	------	ND	------
OK-22	6	40278-9	Grain	79	T2	121-0002A2	[0.004]	------	ND	------
OK-22	6	40278-9	Grain	79	T2	121-0003A2	[0.004]	[0.004]	ND	ND
TX-23	6	------	Grain	N/A	T1	129-0001A2	ND	------	ND	------
TX-23	6	40278-9	Grain	107	T2	129-0002A2	ND	------	ND	------
TX-23	6	40278-9	Grain	107	T2	129-0003A2	ND	ND	ND	ND
ON-24	5	------	Grain	N/A	T1	133-0001A2	ND	------	ND	------
ON-24	5	40278-9	Grain	126	T2	133-0002A2	ND	------	ND	------
ON-24	5	40278-9	Grain	126	T2	133-0003A2	ND	ND	ND	ND

State/ Trial No.	NAFTA Region	AAD-1 Event[a] DAS-	RAC	PHI[b] (days)	Plot ID	Sample ID ARA-09-15-10-	2,4-D (µg/g)	2,4-D Avg[c] (µg/g)	2,4-DCP (µg/g)	2,4-DCP Avg[c] (µg/g)
ON-25	5	------	Grain	N/A	T1	137-0001A2	ND	------	ND	------
ON-25	5	40278-9	Grain	123	T2	137-0002A2	ND	------	ND	------
ON-25	5	40278-9	Grain	123	T2	137-0003A2	ND	ND	ND	ND

[a] Untreated control plots (T1) were planted with the commercial variety Mycogen 2M746, which lacked the inserted AAD-1 gene, but had a genetic background similar to that used for the two non-commercial transgenic hybrids in treated plots (T2).

[b] PHI = Pre Harvest Interval (actual days after the last application of 2,4-D)

[c] Averages presented in this column are of treated samplings (T2 samples) only at each trial's sampling event

[d] ND = Not Detected (below the limit of detection of 0.003 µg/g)

[e] For the purposes of calculating an average, "ND" is given the value of zero.

[f] [ ] = Residue values greater than or equal to the limit of detection of 0.003 µg/g but less than the limit of quantitation of 0.01 µg/g. Values are reported with a lower degree of confidence than values above the limit of quantitation.

TABLE V – Decline Sampling Residues of 2,4-D and 2,4-DCP in Corn Grain

State/ Trial No.	NAFTA Region	AAD-1 Event[a] DAS-	RAC	PHI[b] (days)	Plot ID	Sample ID ARA-09-15-10-	2,4-D (µg/g)	2,4-D Avg[c] (µg/g)	2,4-DCP (µg/g)	2,4-DCP Avg[c] (µg/g)
GA-04	2	------	Grain	N/A	T1	020-0001A44	ND[d]	------	ND	------
GA-04	2	40278-9	Grain	62	T2	020-0002A2	ND	------	ND	------
GA-04	2	40278-9	Grain	62	T2	020-0003A2	ND	ND	ND	ND
GA-04	2	------	Grain	N/A	T1	022-0001A2	ND	------	ND	------
GA-04	2	40278-9	Grain	69	T2	022-0002A2	ND	------	ND	------
GA-04	2	40278-9	Grain	69	T2	022-0003A2	ND	ND	ND	ND
GA-04	2	------	Grain	N/A	T1	024-0001A2	ND	------	ND	------
GA-04	2	40278-9	Grain	76	T2	024-0002A2	ND	------	ND	------
GA-04	2	40278-9	Grain	76	T2	024-0003A2	ND	ND	ND	ND
GA-04	2	------	Grain	N/A	T1	026-0001A2	ND	------	ND	------
GA-04	2	40278-9	Grain	83	T2	026-0002A2	ND	------	ND	------
GA-04	2	40278-9	Grain	83	T2	026-0003A2	ND	ND	ND	ND
GA-04	2	------	Grain	N/A	T1	028-0001A2	ND	------	ND	------
GA-04	2	40278-9	Grain	90	T2	028-0002A2	ND	------	ND	------
GA-04	2	40278-9	Grain	90	T2	028-0003A2	ND	ND	ND	ND
MI-11	5	------	Grain	N/A	T1	060-0001A2	ND	------	ND	------
MI-11	5	40278-9	Grain	76	T2	060-0002A2	ND	------	[0.004][e]	------
MI-11	5	40278-9	Grain	76	T2	060-0003A2	ND	ND	[0.004]	[0.004]
MI-11	5	------	Grain	N/A	T1	062-0001A2	ND	------	ND	------
MI-11	5	40278-9	Grain	83	T2	062-0002A2	ND	------	[0.003]	------
MI-11	5	40278-9	Grain	83	T2	062-0003A2	ND	ND	[0.003]	[0.003]
MI-11	5	------	Grain	N/A	T1	064-0001A2	ND	------	ND	------
MI-11	5	40278-9	Grain	90	T2	064-0002A2	ND	------	ND	------
MI-11	5	40278-9	Grain	90	T2	064-0003A2	ND	ND	[0.003]	ND

State/ Trial No.	NAFTA Region	AAD-1 Event[a] DAS-	RAC	PHI[b] (days)	Plot ID	Sample ID ARA-09-15-10-	2,4-D (µg/g)	2,4-D Avg[c] (µg/g)	2,4-DCP (µg/g)	2,4-DCP Avg[c] (µg/g)
MI-11	5	------	Grain	N/A	T1	066-0001A2	ND	------	ND	------
MI-11	5	40278-9	Grain	97	T2	066-0002A2	ND	------	[0.004]	------
MI-11	5	40278-9	Grain	97	T2	066-0003A2	ND	ND	[0.004]	[0.004]
MI-11	5	------	Grain	N/A	T1	068-0001A2	ND	------	ND	------
MI-11	5	40278-9	Grain	104	T2	068-0002A2	ND	------	ND	------
MI-11	5	40278-9	Grain	104	T2	068-0003A2	ND	ND	[0.004]	ND[f]
OK-22	6	------	Grain	N/A	T1	117-0001A2	ND	------	ND	------
OK-22	6	40278-9	Grain	65	T2	117-0002A2	ND	------	ND	------
OK-22	6	40278-9	Grain	65	T2	117-0003A2	ND	ND	ND	ND
OK-22	6	------	Grain	N/A	T1	119-0001A2	ND	------	ND	------
OK-22	6	40278-9	Grain	71	T2	119-0002A2	ND	------	ND	------
OK-22	6	40278-9	Grain	71	T2	119-0003A2	ND	ND	ND	ND
OK-22	6	------	Grain	N/A	T1	121-0001A2	ND	------	ND	------
OK-22	6	40278-9	Grain	79	T2	121-0002A2	[0.004]	------	ND	------
OK-22	6	40278-9	Grain	79	T2	121-0003A2	[0.004]	[0.004]	ND	ND
OK-22	6	------	Grain	N/A	T1	123-0001A2	ND	------	ND	------
OK-22	6	40278-9	Grain	86	T2	123-0002A2	ND	------	ND	------
OK-22	6	40278-9	Grain	86	T2	123-0003A2	ND	ND	ND	ND
OK-22	6	------	Grain	N/A	T1	125-0001A2	ND	------	ND	------
OK-22	6	40278-9	Grain	93	T2	125-0002A2	ND	------	ND	------
OK-22	6	40278-9	Grain	93	T2	125-0003A2	ND	ND	ND	ND

[a] Untreated control plots (T1) were planted with the commercial variety Mycogen 2M746, which lacked the inserted AAD-1 gene, but had a genetic background similar to that used for the two non-commercial transgenic hybrids in treated plots (T2).
[b] PHI = Pre Harvest Interval (actual days after the last application of 2,4-D)
[c] Averages presented in this column are of treated samplings (T2 samples) only at each trial's sampling event
[d] ND = Not Detected (below the limit of detection of 0.003 µg/g)
[e] [ ] = Residue values greater than or equal to the limit of detection of 0.003 µg/g but less than the limit of quantitation of 0.01 µg/g. Values are reported with a lower degree of confidence than values above the limit of quantitation.
[f] For the purposes of calculating an average, "ND" is given the value of zero.

TABLE VI - Residues of 2,4-D and 2,4-DCP in Corn Stover

State/ Trial No.	NAFTA Region	AAD-1 Event[a] DAS-	RAC	PHI[b] (days)	Plot ID	Sample ID ARA-09-15-10-	2,4-D (µg/g)	2,4-D Avg[c] (µg/g)	2,4-DCP (µg/g)	2,4-DCP Avg[c] (µg/g)
NJ-01	1	------	Stover	NA	T1	004-0001A2	[0.004][d]	------	ND[e]	------
NJ-01	1	40278-9	Stover	98	T2	004-0002A2	8.191	------	2.241	------
NJ-01	1	40278-9	Stover	98	T2	004-0003A2	5.877	7.034	1.557	1.899
PA-02	1	------	Stover	NA	T1	008-0001A2	ND	------	ND	------
PA-02	1	40278-9	Stover	107	T2	008-0002A2	2.695	------	1.692	------
PA-02	1	40278-9	Stover	107	T2	008-0003A2	1.627	2.161	1.208	1.450
GA-03	2	------	Stover	N/A	T1	013-0001A2	ND	------	ND	------
GA-03	2	40278-9	Stover	73	T2	013-0002A2	0.324	------	0.207	------
GA-03	2	40278-9	Stover	73	T2	013-0003A2	0.162	0.243	0.072	0.139
GA-04	2	------	Stover	N/A	T1	025-0001A2	ND	------	ND	------
GA-04	2	40278-9	Stover	76	T2	025-0002A2	1.372	------	0.522	------
GA-04	2	40278-9	Stover	76	T2	025-0003A2	1.082	1.227	0.476	0.499
IL-05	5	------	Stover	N/A	T1	033-0001A3	ND	------	[0.003]	------
IL-05	5	40278-9	Stover	135	T2	033-0002A3	0.154	------	0.148	------
IL-05	5	40278-9	Stover	135	T2	033-0003A3	0.105	0.130	0.094	0.121
IL-06	5	------	Stover	N/A	T1	037-0001A2	ND	------	ND	------
IL-06	5	40278-9	Stover	118	T2	037-0002A2	0.277	------	0.168	------
IL-06	5	40278-9	Stover	118	T2	037-0003A2	0.486	0.382	0.310	0.239
IN-07	5	------	Stover	N/A	T1	041-0001A2	[0.004]	------	0.012	------
IN-07	5	40278-9	Stover	139	T2	041-0002A2	1.443	------	1.278	------
IN-07	5	40278-9	Stover	139	T2	041-0003A2	0.623	1.033	0.554	0.916
MO-08	5	------	Stover	N/A	T1	045-0001A2	ND	------	ND	------
MO-08	5	40278-9	Stover	104	T2	045-0002A2,3,4	0.018[f]	------	[0.007][f]	------
MO-08	5	40278-9	Stover	104	T2	045-0003A2	0.026	0.022	0.019	0.013

State/ Trial No.	NAFTA Region	AAD-1 Event[a] DAS-	RAC	PHI[b] (days)	Plot ID	Sample ID ARA-09-15-10-	2,4-D (µg/g)	2,4-D Avg[c] (µg/g)	2,4-DCP (µg/g)	2,4-DCP Avg[c] (µg/g)
MO-09	5	------	Stover	N/A	T1	049-0001A2	[0.008]	------	ND	------
MO-09	5	40278-9	Stover	105	T2	049-0002A2	1.185	------	0.741	------
MO-09	5	40278-9	Stover	105	T2	049-0003A2	2.464	1.825	1.136	0.938
IL-10	5	------	Stover	N/A	T1	053-0001A2	[0.006]	------	0.011	------
IL-10	5	40278-9	Stover	100	T2	053-0002A2	2.540	------	1.306	------
IL-10	5	40278-9	Stover	100	T2	053-0003A2	0.914	1.727	0.491	0.899
MI-11	5	------	Stover	N/A	T1	065-0001A2	ND	------	[0.005]	------
MI-11	5	40278-9	Stover	90	T2	065-0002A2	7.233	------	4.912	------
MI-11	5	40278-9	Stover	90	T2	065-0003A2	8.920	8.076	5.842	5.377
OH-12	5	------	Stover	N/A	T1	073-0001A2	ND	------	[0.006]	------
OH-12	5	40278-9	Stover	90	T2	073-0002A2	1.676	------	1.187	------
OH-12	5	40278-9	Stover	90	T2	073-0003A2	2.422	2.049	1.768	1.478
IN-13	5	------	Stover	N/A	T1	077-0001A2	ND	------	[0.007]	------
IN-13	5	40278-9	Stover	90	T2	077-0002A2	3.101	------	3.175	------
IN-13	5	40278-9	Stover	90	T2	077-0003A2	3.134	3.118	3.032	3.103
NE-14	5	------	Stover	N/A	T1	082-0001A2	ND	------	[0.005]	------
NE-14	5	40278-9	Stover	134	T2	082-0002A2	0.405	------	0.398	------
NE-14	5	40278-9	Stover	134	T2	082-0003A2	0.250	0.328	0.247	0.323
NE-15	5	------	Stover	N/A	T1	086-0001A2	ND	------	[0.005]	------
NE-15	5	40278-9	Stover	83	T2	086-0002A2	[0.004]	------	[0.007]	------
NE-15	5	40278-9	Stover	83	T2	086-0003A2	0.011	0.007	0.032	0.019
KS-16	5	------	Stover	N/A	T1	090-0001A2	ND	------	[0.006]	------
KS-16	5	40278-9	Stover	94	T2	090-0002A2	0.027	------	0.024	------
KS-16	5	40278-9	Stover	94	T2	090-0003A2	0.030	0.028	0.035	0.029
IA-17	5	------	Stover	N/A	T1	094-0001A2	ND	------	[0.003]	------

State/ Trial No.	NAFTA Region	AAD-1 Event[a] DAS-	RAC	PHI[b] (days)	Plot ID	Sample ID ARA-09-15-10-	2,4-D (µg/g)	2,4-D Avg[c] (µg/g)	2,4-DCP (µg/g)	2,4-DCP Avg[c] (µg/g)
IA-17	5	40278-9	Stover	103	T2	094-0002A2	0.159	------	0.191	------
IA-17	5	40278-9	Stover	103	T2	094-0003A2	0.092	0.125	0.127	0.159
MO-18	5	------	Stover	N/A	T1	098-0001A2	ND	------	[0.005]	------
MO-18	5	40278-9	Stover	100	T2	098-0002A2	ND	------	ND	------
MO-18	5	40278-9	Stover	100	T2	098-0003A2	ND	ND	[0.004]	ND[g]
IA-19	5	------	Stover	N/A	T1	102-0001A2	ND	------	ND	------
IA-19	5	40278-9	Stover	121	T2	102-0002A2	1.311	------	2.447	------
IA-19	5	40278-9	Stover	121	T2	102-0003A2	0.755	1.033	1.576	2.011
MN-20	5	------	Stover	N/A	T1	106-0001A2	ND	------	[0.003]	------
MN-20	5	40278-9	Stover	121	T2	106-0002A2	1.196	------	1.716	------
MN-20	5	40278-9	Stover	121	T2	106-0003A2	0.875	1.035	1.159	1.438
MO-21	5	------	Stover	N/A	T1	110-0001A2	ND	------	[0.007]	------
MO-21	5	40278-9	Stover	90	T2	110-0002A2	0.794	------	0.271	------
MO-21	5	40278-9	Stover	90	T2	110-0003A2	0.769	0.781	0.305	0.288
OK-22	6	------	Stover	N/A	T1	122-0001A2	ND	------	ND	------
OK-22	6	40278-9	Stover	79	T2	122-0002A2	0.177	------	0.035	------
OK-22	6	40278-9	Stover	79	T2	122-0003A2	0.176	0.177	0.033	0.034
TX-23	6	------	Stover	N/A	T1	130-0001A3	ND	------	[0.006]	------
TX-23	6	40278-9	Stover	107	T2	130-0002A2	0.602	------	0.229	------
TX-23	6	40278-9	Stover	107	T2	130-0003A2	0.874	0.738	0.517	0.373
ON-24	5	------	Stover	N/A	T1	134-0001A3	ND	------	[0.006]	------
ON-24	5	40278-9	Stover	126	T2	134-0002A2	0.231	------	0.314	------
ON-24	5	40278-9	Stover	126	T2	134-0003A2	0.356	0.293	0.430	0.372
ON-25	5	------	Stover	N/A	T1	138-0001A3	ND	------	ND	------
ON-25	5	40278-9	Stover	123	T2	138-0002A2	0.127	------	0.618	------

State/ Trial No.	NAFTA Region	AAD-1 Event[a] DAS-	RAC	PHI[b] (days)	Plot ID	Sample ID ARA-09-15-10-	2,4-D (µg/g)	2,4-D Avg[c] (µg/g)	2,4-DCP (µg/g)	2,4-DCP Avg[c] (µg/g)
ON-25	5	40278-9	Stover	123	T2	138-0003A2	0.170	0.149	0.674	0.646

[a] Untreated control plots (T1) were planted with the commercial variety Mycogen 2M746, which lacked the inserted AAD-1 gene, but had a genetic background similar to that used for the two non-commercial transgenic hybrids in treated plots (T2).

[b] PHI = Pre Harvest Interval (actual days after the last application of 2,4-D)

[c] Averages presented in this column are of treated samplings (T2 samples) only at each trial's sampling event

[d] [ ] = Residue values greater than or equal to the limit of detection of 0.003 µg/g but less than the limit of quantitation of 0.01 µg/g. Values are reported with a lower degree of confidence than values above the limit of quantitation.

[e] ND = Not Detected (below the limit of detection of 0.003 µg/g)

[f] Average of three analyses.

[g] For the purposes of calculating an average, "ND" is given the value of zero.

TABLE VII – Decline Sampling Residues of 2,4-D and 2,4-DCP in Corn Stover

State/ Trial No.	NAFTA Region	AAD-1 Event[a] DAS-	RAC	PHI[b] (days)	Plot ID	Sample ID ARA-09-15-10-	2,4-D (µg/g)	2,4-D Avg[c] (µg/g)	2,4-DCP (µg/g)	2,4-DCP Avg[c] (µg/g)
GA-04	2	------	Stover	N/A	T1	021-0001A9	ND[d]	------	ND	------
GA-04	2	40278-9	Stover	62	T2	021-0002A2	2.862	------	1.139	------
GA-04	2	40278-9	Stover	62	T2	021-0003A2	2.639	2.750	0.799	0.969
GA-04	2	------	Stover	N/A	T1	023-0001A2	ND	------	[0.004][e]	------
GA-04	2	40278-9	Stover	69	T2	023-0002A2	0.785	------	0.365	------
GA-04	2	40278-9	Stover	69	T2	023-0003A2	0.862	0.824	0.368	0.367
GA-04	2	------	Stover	N/A	T1	025-0001A2	ND	------	ND	------
GA-04	2	40278-9	Stover	76	T2	025-0002A2	1.372	------	0.522	------
GA-04	2	40278-9	Stover	76	T2	025-0003A2	1.082	1.227	0.476	0.499
GA-04	2	------	Stover	N/A	T1	027-0001A2	[0.003]	------	[0.003]	------
GA-04	2	40278-9	Stover	83	T2	027-0002A2	1.327	------	0.555	------
GA-04	2	40278-9	Stover	83	T2	027-0003A2	1.293	1.310	0.537	0.546
GA-04	2	------	Stover	N/A	T1	029-0001A2	ND	------	ND	------
GA-04	2	40278-9	Stover	90	T2	029-0002A2	0.878	------	0.432	------
GA-04	2	40278-9	Stover	90	T2	029-0003A2	1.095	0.986	0.599	0.515
MI-11	5	------	Stover	N/A	T1	061-0001A2	ND	------	[0.005]	------
MI-11	5	40278-9	Stover	76	T2	061-0002A2	3.546	------	3.180	------
MI-11	5	40278-9	Stover	76	T2	061-0003A2	5.979	4.762	5.476	4.328
MI-11	5	------	Stover	N/A	T1	063-0001A2	ND	------	[0.008]	------
MI-11	5	40278-9	Stover	83	T2	063-0002A2	4.080	------	3.231	------
MI-11	5	40278-9	Stover	83	T2	063-0003A2	6.616	5.348	6.121	4.676
MI-11	5	------	Stover	N/A	T1	065-0001A2	ND	------	[0.005]	------
MI-11	5	40278-9	Stover	90	T2	065-0002A2	7.233	------	4.912	------
MI-11	5	40278-9	Stover	90	T2	065-0003A2	8.920	8.076	5.842	5.377

State/ Trial No.	NAFTA Region	AAD-1 Event[a] DAS-	RAC	PHI[b] (days)	Plot ID	Sample ID ARA-09-15-10-	2,4-D (µg/g)	2,4-D Avg[c] (µg/g)	2,4-DCP (µg/g)	2,4-DCP Avg[c] (µg/g)
MI-11	5	------	Stover	N/A	T1	067-0001A3	ND	------	[0.006]	------
MI-11	5	40278-9	Stover	97	T2	067-0002A2	4.735	------	3.871	------
MI-11	5	40278-9	Stover	97	T2	067-0003A2	7.051	5.893	4.437	4.154
MI-11	5	------	Stover	N/A	T1	069-0001A2	ND	------	0.015	------
MI-11	5	40278-9	Stover	104	T2	069-0002A2	5.798	------	3.769	------
MI-11	5	40278-9	Stover	104	T2	069-0003A2	3.228	4.513	2.608	3.188
OK-22	6	------	Stover	N/A	T1	118-0001A2	ND	------	[0.003]	------
OK-22	6	40278-9	Stover	65	T2	118-0002A2	0.287	------	0.058	------
OK-22	6	40278-9	Stover	65	T2	118-0003A2	0.277	0.282	0.055	0.057
OK-22	6	------	Stover	N/A	T1	120-0001A2	ND	------	[0.009]	------
OK-22	6	40278-9	Stover	71	T2	120-0002A2	0.258	------	0.043	------
OK-22	6	40278-9	Stover	71	T2	120-0003A2	0.164	0.211	0.033	0.038
OK-22	6	------	Stover	N/A	T1	122-0001A2	ND	------	ND	------
OK-22	6	40278-9	Stover	79	T2	122-0002A2	0.177	------	0.035	------
OK-22	6	40278-9	Stover	79	T2	122-0003A2	0.176	0.177	0.033	0.034
OK-22	6	------	Stover	N/A	T1	124-0001A2	ND	------	ND	------
OK-22	6	40278-9	Stover	86	T2	124-0002A2	0.209	------	0.040	------
OK-22	6	40278-9	Stover	86	T2	124-0003A2	0.188	0.199	0.031	0.036
OK-22	6	------	Stover	N/A	T1	126-0001A3	ND	------	[0.003]	------
OK-22	6	40278-9	Stover	93	T2	126-0002A2	0.096	------	0.022	------
OK-22	6	40278-9	Stover	93	T2	126-0003A2	0.099	0.097	0.018	0.020

[a] Untreated control plots (T1) were planted with the commercial variety Mycogen 2M746, which lacked the inserted AAD-1 gene, but had a genetic background similar to that used for the two non-commercial transgenic hybrids in treated plots (T2).

[b] PHI = Pre Harvest Interval (actual days after the last application of 2,4-D)

[c] Averages presented in this column are of treated samplings (T2 samples) only at each trial's sampling event

[d] ND = Not Detected (below the limit of detection of 0.003 µg/g)

[e] [ ] = Residue values greater than or equal to the limit of detection of 0.003 µg/g but less than the limit of quantitation of 0.01 µg/g. Values are reported with a lower degree of confidence than values above the limit of quantitation.

TABLE VIII -

Residues of 2,4-D and 2,4-DCP in Field Corn Processed Commodities (Trial GA-03, NAFTA Region 2)

AAD-1 Event[a] DAS-	Commodity	PHI[b] (days)	Plot ID	Sample ID ARA-09-15-10-	2,4-D (µg/g)	2,4-D Avg[c] (µg/g)	2,4-DCP (µg/g)	2,4-DCP Avg[c] (µg/g)
------	Grain, from the field	N/A	T1	011-0001A2	ND[d]	------	ND	------
40278-9	Grain, from the field	73	T4	011-0006A2	ND	------	ND	------
40278-9	Grain, from the field	73	T4	011-0007A2	ND	ND	ND	ND
------	Grain, from the processor prior to AGF generation	N/A	T1	139-0001A2	ND	------	ND	------
------	Grain, from the processor prior to AGF generation	N/A	T1	139-0002A2	ND	------	ND	------
------	Grain, from the processor prior to AGF generation	N/A	T1	139-0003A2	ND	ND	ND	ND
40278-9	Grain, from the processor prior to AGF generation	73	T4	139-0004A3	ND	------	[0.003][e]	------
40278-9	Grain, from the processor prior to AGF generation	73	T4	139-0005A2	ND	------	ND	------
40278-9	Grain, from the processor prior to AGF generation	73	T4	139-0006A2	ND	ND	ND	ND
------	Grain, from the processor after AGF generation	N/A	T1	140-0001A2	ND	------	ND	------
------	Grain, from the processor after AGF generation	N/A	T1	140-0002A2	ND	------	ND	------
------	Grain, from the processor after AGF generation	N/A	T1	140-0003A2	ND	ND	ND	ND
40278-9	Grain, from the processor after AGF generation	73	T4	140-0004A3	ND	------	ND	------
40278-9	Grain, from the processor after AGF generation	73	T4	140-0005A2	ND	------	ND	------
40278-9	Grain, from the processor after AGF generation	73	T4	140-0006A2	ND	ND	ND	ND

AAD-1 Event[a] DAS-	Commodity	PHI[b] (days)	Plot ID	Sample ID ARA-09-15-10-	2,4-D (μg/g)	2,4-D Avg[c] (μg/g)	2,4-DCP (μg/g)	2,4-DCP Avg[c] (μg/g)
------	Starch	N/A	T1	142-0001A30	ND	------	ND	------
------	Starch	N/A	T1	142-0002A2	ND	ND	ND	ND
40278-9	Starch	N/A	T4	142-0003A3	ND	------	ND	------
40278-9	Starch	N/A	T4	142-0004A4	ND	ND	ND	ND
------	Oil, refined (wet milling)	N/A	T1	143-0001A30	ND	------	ND	------
------	Oil, refined (wet milling)	N/A	T1	143-0002A2	ND	ND	ND	ND
40278-9	Oil, refined (wet milling)	N/A	T4	143-0003A3	ND	------	ND	------
40278-9	Oil, refined (wet milling)	N/A	T4	143-0004A3	ND	ND	ND	ND
------	Oil, refined (dry milling)	N/A	T1	147-0001A30	ND	------	ND	------
------	Oil, refined (dry milling)	N/A	T1	147-0002A2	ND	ND	ND	ND
40278-9	Oil, refined (dry milling)	N/A	T4	147-0003A3	ND	------	ND	------
40278-9	Oil, refined (dry milling)	N/A	T4	147-0004A3	ND	ND	ND	ND
------	Meal	N/A	T1	144-0001A22	ND	------	ND	------
------	Meal	N/A	T1	144-0002A2	ND	ND	ND	ND
40278-9	Meal	N/A	T4	144-0003A2	ND	------	ND	------
40278-9	Meal	N/A	T4	144-0004A2	ND	ND	ND	ND
------	Grits	N/A	T1	145-0001A22	ND	------	ND	------
------	Grits	N/A	T1	145-0002A2	ND	ND	ND	ND
40278-9	Grits	N/A	T4	145-0003A2	ND	------	ND	------
40278-9	Grits	N/A	T4	145-0004A2	ND	ND	ND	ND
------	Flour	N/A	T1	146-0001A3	ND	------	ND	------
------	Flour	N/A	T1	146-0002A3	ND	ND	ND	ND
40278-9	Flour	N/A	T4	146-0003A3	ND	------	ND	------
40278-9	Flour	N/A	T4	146-0004A3	ND	ND	ND	ND

AAD-1 Event[a] DAS-	Commodity	PHI[b] (days)	Plot ID	Sample ID ARA-09-15-10-	2,4-D (µg/g)	2,4-D Avg[c] (µg/g)	2,4-DCP (µg/g)	2,4-DCP Avg[c] (µg/g)
------	Aspirated grain fractions	N/A	T1	141-0001A2	ND	------	ND	------
40278-9	Aspirated grain fractions	N/A	T4	141-0002A2	0.021	------	[0.006]	------

[a] Untreated control plots (T1) were planted with the commercial variety Mycogen 2M746, which lacked the inserted AAD-1 gene, but had a genetic background similar to that used for the two non-commercial transgenic hybrids in treated plots (T2).

[b] PHI = Pre Harvest Interval (actual days after the last application of 2,4-D)

[c] Averages presented in this column are for each untreated (T1) and treated (T4)

[d] ND = Not Detected (below the limit of detection of 0.003 µg/g)

[e] [ ] = Residue values greater than or equal to the limit of detection of 0.003 µg/g but less than the limit of quantitation of 0.01 µg/g. Values are reported with a lower degree of confidence than values above the limit of quantitation.

TABLE IX –

Residues of 2,4-D and 2,4-DCP in Field Corn Processed Commodities (Trial NE-14, NAFTA Region 5)

AAD-1 Event[a] DAS-	Commodity	PHI[b] (days)	Plot ID	Sample ID ARA-09-15-10-	2,4-D (µg/g)	2,4-D Avg[c] (µg/g)	2,4-DCP (µg/g)	2,4-DCP Avg[c] (µg/g)
------	Grain, from the field	N/A	T1	080-0001A2	ND[d]	------	ND	------
40278-9	Grain, from the field	134	T4	080-0006A2	ND	------	ND	------
40278-9	Grain, from the field	134	T4	080-0007A2	ND	ND	ND	ND
------	Grain, from the processor prior to AGF generation	N/A	T1	148-0001A2	ND	------	ND	------
------	Grain, from the processor prior to AGF generation	N/A	T1	148-0002A2	ND	------	ND	------
------	Grain, from the processor prior to AGF generation	N/A	T1	148-0003A2	ND	ND	ND	ND
40278-9	Grain, from the processor prior to AGF generation	134	T4	148-0004A3	ND	------	0.014	------
40278-9	Grain, from the processor prior to AGF generation	134	T4	148-0005A2	ND	------	[0.007][e]	------
40278-9	Grain, from the processor prior to AGF generation	134	T4	148-0006A2	ND	ND	[0.004]	[0.008]
------	Grain, from the processor after AGF generation	N/A	T1	149-0001A2	ND	------	ND	------
------	Grain, from the processor after AGF generation	N/A	T1	149-0002A2	ND	------	ND	------
------	Grain, from the processor after AGF generation	N/A	T1	149-0003A2	ND	ND	ND	ND
40278-9	Grain, from the processor after AGF generation	134	T4	149-0004A3	ND	------	ND	------
40278-9	Grain, from the processor after AGF generation	134	T4	149-0005A2	ND	------	ND	------

AAD-1 Event[a] DAS-	Commodity	PHI[b] (days)	Plot ID	Sample ID ARA-09-15-10-	2,4-D (µg/g)	2,4-D Avg[c] (µg/g)	2,4-DCP (µg/g)	2,4-DCP Avg[c] (µg/g)
40278-9	Grain, from the processor after AGF generation	134	T4	149-0006A2	ND	ND	ND	ND
------	Starch	N/A	T1	151-0001A2	ND	------	ND	------
------	Starch	N/A	T1	151-0002A2	ND	ND	ND	ND
40278-9	Starch	N/A	T4	151-0003A4	ND	------	ND	------
40278-9	Starch	N/A	T4	151-0004A4	ND	ND	ND	ND
------	Oil, refined (wet milling)	N/A	T1	152-0001A2	ND	------	ND	------
------	Oil, refined (wet milling)	N/A	T1	152-0002A2	ND	ND	ND	ND
40278-9	Oil, refined (wet milling)	N/A	T4	152-0003A3	ND	------	ND	------
40278-9	Oil, refined (wet milling)	N/A	T4	152-0004A3	ND	ND	ND	ND
------	Oil, refined (dry milling)	N/A	T1	156-0001A2	ND	------	ND	------
------	Oil, refined (dry milling)	N/A	T1	156-0002A2	ND	ND	ND	ND
40278-9	Oil, refined (dry milling)	N/A	T4	156-0003A3	ND	------	ND	------
40278-9	Oil, refined (dry milling)	N/A	T4	156-0004A3	ND	ND	ND	ND
------	Meal	N/A	T1	153-0001A2	ND	------	ND	------
------	Meal	N/A	T1	153-0002A2	ND	ND	ND	ND
40278-9	Meal	N/A	T4	153-0003A2	ND	------	ND	------
40278-9	Meal	N/A	T4	153-0004A2	ND	ND	ND	ND
------	Grits	N/A	T1	154-0001A32	ND	------	ND	------
------	Grits	N/A	T1	154-0002A2	ND	ND	ND	ND
40278-9	Grits	N/A	T4	154-0003A2	ND	------	ND	------
40278-9	Grits	N/A	T4	154-0004A2	ND	ND	ND	ND
------	Flour	N/A	T1	155-0001A22	ND	------	ND	------
------	Flour	N/A	T1	155-0002A2	ND	ND	ND	ND
40278-9	Flour	N/A	T4	155-0003A2	ND	------	ND	------

AAD-1 Event[a] DAS-	Commodity	PHI[b] (days)	Plot ID	Sample ID ARA-09-15-10-	2,4-D (µg/g)	2,4-D Avg[c] (µg/g)	2,4-DCP (µg/g)	2,4-DCP Avg[c] (µg/g)
40278-9	Flour	N/A	T4	155-0004A2	ND	ND	ND	ND
------	Aspirated grain fractions	N/A	T1	150-0001A22	ND	------	ND	------
40278-9	Aspirated grain fractions	N/A	T4	150-0002A2	ND	------	ND	------

[a] Untreated control plots (T1) were planted with the commercial variety Mycogen 2M746, which lacked the inserted AAD-1 gene, but had a genetic background similar to that used for the two non-commercial transgenic hybrids in treated plots (T2).

[b] PHI = Pre Harvest Interval (actual days after the last application of 2,4-D)

[c] Averages presented in this column are for each untreated (T1) and treated (T4)

[d] ND = Not Detected (below the limit of detection of 0.003 µg/g)

[e] [ ] = Residue values greater than or equal to the limit of detection of 0.003 µg/g but less than the limit of quantitation of 0.01 µg/g. Values are reported with a lower degree of confidence than values above the limit of quantitation.

TABLE X - Residues of Quizalofop Acid and Quizalofop-ethyl in Corn Forage

State/ Trial No.	NAFTA Region	AAD-1 Event[a] DAS-	RAC	PHI[b] (days)	Plot ID	Sample ID ARA-09-15-10-	Quizalofop acid (µg/g)		Quizalofop acid (as quizalofop-ethyl equivalents)[c]		Quizalofop-ethyl (µg/g)		Total Residue[d] (µg/g)	
								Avg[e]		Avg[e]		Avg[e]		Avg[e]
NJ-01	1	------	Forage	NA	T1	001-0001A1	ND[f]	------	ND	------	ND	------	ND	------
NJ-01	1	40278-9	Forage	44	T2	001-0002A1	ND	------	ND	------	ND	------	ND	------
NJ-01	1	40278-9	Forage	44	T2	001-0003A1	ND	ND	ND	ND	ND	ND	ND	ND
NJ-01	1	------	Forage	NA	T1	002-0001A1	ND	------	ND	------	ND	------	ND	------
NJ-01	1	40278-9	Forage	71	T2	002-0002A1	ND	------	ND	------	ND	------	ND	------
NJ-01	1	40278-9	Forage	71	T2	002-0003A1	ND	ND	ND	ND	ND	ND	ND	ND
PA-02	1	------	Forage	NA	T1	005-0001A1	ND	------	ND	------	ND	------	ND	------
PA-02	1	40278-9	Forage	47	T2	005-0002A1	ND	------	ND	------	ND	------	ND	------
PA-02	1	40278-9	Forage	47	T2	005-0003A1	ND	ND	ND	ND	ND	ND	ND	ND
PA-02	1	------	Forage	NA	T1	006-0001A1	ND	------	ND	------	ND	------	ND	------
PA-02	1	40278-9	Forage	61	T2	006-0002A1	ND	------	ND	------	ND	------	ND	------
PA-02	1	40278-9	Forage	61	T2	006-0003A1	ND	ND	ND	ND	ND	ND	ND	ND
GA-03	2	------	Forage	N/A	T1	009-0001A1	ND	------	ND	------	ND	------	ND	------
GA-03	2	40278-9	Forage	43	T2	009-0002A1	ND	------	ND	------	ND	------	ND	------
GA-03	2	40278-9	Forage	43	T2	009-0003A1	ND	ND	ND	ND	ND	ND	ND	ND
GA-03	2	------	Forage	N/A	T1	010-0001A1	ND	------	ND	------	ND	------	ND	------

State/ Trial No.	NAFTA Region	AAD-1 Event[a] DAS-	RAC	PHI[b] (days)	Plot ID	Sample ID ARA-09-15-10-	Quizalofop acid (µg/g)		Quizalofop acid (as quizalofop-ethyl equivalents)[c]		Quizalofop-ethyl (µg/g)		Total Residue[d] (µg/g)	
								Avg[e]		Avg[e]		Avg[e]		Avg[e]
GA-03	2	40278-9	Forage	48	T2	010-0002A1	ND	------	ND	------	ND	------	ND	------
GA-03	2	40278-9	Forage	48	T2	010-0003A1	ND	ND	ND	ND	ND	ND	ND	ND
GA-04	2	------	Forage	N/A	T1	014-0001A1	ND	------	ND	------	ND	------	ND	------
GA-04	2	40278-9	Forage	34	T2	014-0002A1	ND	------	ND	------	ND	------	ND	------
GA-04	2	40278-9	Forage	34	T2	014-0003A1	ND	ND	ND	ND	ND	ND	ND	ND
GA-04	2	------	Forage	N/A	T1	017-0001A1	ND	------	ND	------	ND	------	ND	------
GA-04	2	40278-9	Forage	50	T2	017-0002A1	ND	------	ND	------	ND	------	ND	------
GA-04	2	40278-9	Forage	50	T2	017-0003A1	ND	ND	ND	ND	ND	ND	ND	ND
IL-05	5	------	Forage	N/A	T1	030-0001A1	ND	------	ND	------	ND	------	ND	------
IL-05	5	40278-9	Forage	46	T2	030-0002A1	ND	------	ND	------	ND	------	ND	------
IL-05	5	40278-9	Forage	46	T2	030-0003A1	ND	ND	ND	ND	ND	ND	ND	ND
IL-05	5	------	Forage	N/A	T1	031-0001A1	ND	------	ND	------	ND	------	ND	------
IL-05	5	40278-9	Forage	73	T2	031-0002A1	ND	------	ND	------	ND	------	ND	------
IL-05	5	40278-9	Forage	73	T2	031-0003A1	ND	ND	ND	ND	ND	ND	ND	ND
IL-06	5	------	Forage	N/A	T1	034-0001A1	ND	------	ND	------	ND	------	ND	------
IL-06	5	40278-9	Forage	44	T2	034-0002A1	ND	------	ND	------	ND	------	ND	------
IL-06	5	40278-9	Forage	44	T2	034-0003A1	ND	ND	ND	ND	ND	ND	ND	ND

State/ Trial No.	NAFTA Region	AAD-1 Event[a] DAS-	RAC	PHI[b] (days)	Plot ID	Sample ID ARA-09-15-10-	Quizalofop acid (µg/g)		Quizalofop acid (as quizalofop-ethyl equivalents)[c]		Quizalofop-ethyl (µg/g)		Total Residue[d] (µg/g)	
								Avg[e]		Avg[e]		Avg[e]		Avg[e]
IL-06	5	------	Forage	N/A	T1	035-0001A1	ND	------	ND	------	ND	------	ND	------
IL-06	5	40278-9	Forage	69	T2	035-0002A1	ND	------	ND	------	ND	------	ND	------
IL-06	5	40278-9	Forage	69	T2	035-0003A1	ND	ND	ND	ND	ND	ND	ND	ND
IN-07	5	------	Forage	N/A	T1	038-0001A1	ND	------	ND	------	ND	------	ND	------
IN-07	5	40278-9	Forage	47	T2	038-0002A1	ND	------	ND	------	ND	------	ND	------
IN-07	5	40278-9	Forage	47	T2	038-0003A1	ND	ND	ND	ND	ND	ND	ND	ND
IN-07	5	------	Forage	N/A	T1	039-0001A1	ND	------	ND	------	ND	------	ND	------
IN-07	5	40278-9	Forage	66	T2	039-0002A1	ND	------	ND	------	ND	------	ND	------
IN-07	5	40278-9	Forage	66	T2	039-0003A1	ND	ND	ND	ND	ND	ND	ND	ND
MO-08	5	------	Forage	N/A	T1	042-0001A1	ND	------	ND	------	ND	------	ND	------
MO-08	5	40278-9	Forage	45	T2	042-0002A1	ND	------	ND	------	ND	------	ND	------
MO-08	5	40278-9	Forage	45	T2	042-0003A1	ND	ND	ND	ND	ND	ND	ND	ND
MO-08	5	------	Forage	N/A	T1	043-0001A1	ND	------	ND	------	ND	------	ND	------
MO-08	5	40278-9	Forage	52	T2	043-0002A1	ND	------	ND	------	ND	------	ND	------
MO-08	5	40278-9	Forage	52	T2	043-0003A1	ND	ND	ND	ND	ND	ND	ND	ND
MO-09	5	------	Forage	N/A	T1	046-0001A1	ND	------	ND	------	ND	------	ND	------
MO-09	5	40278-9	Forage	46	T2	046-0002A1	ND	------	ND	------	ND	------	ND	------

State/ Trial No.	NAFTA Region	AAD-1 Event[a] DAS-	RAC	PHI[b] (days)	Plot ID	Sample ID ARA-09-15-10-	Quizalofop acid (µg/g)		Quizalofop acid (as quizalofop-ethyl equivalents)[c]		Quizalofop-ethyl (µg/g)		Total Residue[d] (µg/g)	
								Avg[e]		Avg[e]		Avg[e]		Avg[e]
MO-09	5	40278-9	Forage	46	T2	046-0003A1	ND	ND	ND	ND	ND	ND	ND	ND
MO-09	5	------	Forage	N/A	T1	047-0001A1	ND	------	ND	------	ND	------	ND	------
MO-09	5	40278-9	Forage	53	T2	047-0002A1	ND	------	ND	------	ND	------	ND	------
MO-09	5	40278-9	Forage	53	T2	047-0003A1	ND	ND	ND	ND	ND	ND	ND	ND
IL-10	5	------	Forage	N/A	T1	050-0001A1	ND	------	ND	------	ND	------	ND	------
IL-10	5	40278-9	Forage	45	T2	050-0002A1	ND	------	ND	------	ND	------	ND	------
IL-10	5	40278-9	Forage	45	T2	050-0003A1	ND	ND	ND	ND	ND	ND	ND	ND
IL-10	5	------	Forage	N/A	T1	051-0001A1	ND	------	ND	------	ND	------	ND	------
IL-10	5	40278-9	Forage	48	T2	051-0002A1	ND	------	ND	------	ND	------	ND	------
IL-10	5	40278-9	Forage	48	T2	051-0003A1	ND	ND	ND	ND	ND	ND	ND	ND
MI-11	5	------	Forage	N/A	T1	054-0001A1	ND	------	ND	------	ND	------	ND	------
MI-11	5	40278-9	Forage	45	T2	054-0002A1	ND	------	ND	------	ND	------	ND	------
MI-11	5	40278-9	Forage	45	T2	054-0003A1	ND	ND	ND	ND	ND	ND	ND	ND
MI-11	5	------	Forage	N/A	T1	057-0001A1	ND	------	ND	------	ND	------	ND	------
MI-11	5	40278-9	Forage	64	T2	057-0002A1	ND	------	ND	------	ND	------	ND	------
MI-11	5	40278-9	Forage	64	T2	057-0003A1	ND	ND	ND	ND	ND	ND	ND	ND
OH-12	5	------	Forage	N/A	T1	070-0001A1	ND	------	ND	------	ND	------	ND	------

State/ Trial No.	NAFTA Region	AAD-1 Event[a] DAS-	RAC	PHI[b] (days)	Plot ID	Sample ID ARA-09-15-10-	Quizalofop acid (µg/g)		Quizalofop acid (as quizalofop-ethyl equivalents)[c]		Quizalofop-ethyl (µg/g)		Total Residue[d] (µg/g)	
								Avg[e]		Avg[e]		Avg[e]		Avg[e]
OH-12	5	40278-9	Forage	45	T2	070-0002A1	ND	------	ND	------	ND	------	ND	------
OH-12	5	40278-9	Forage	45	T2	070-0003A1	ND	ND	ND	ND	ND	ND	ND	ND
OH-12	5	------	Forage	N/A	T1	071-0001A1	ND	------	ND	------	ND	------	ND	------
OH-12	5	40278-9	Forage	60	T2	071-0002A1	ND	------	ND	------	ND	------	ND	------
OH-12	5	40278-9	Forage	60	T2	071-0003A1	ND	ND	ND	ND	ND	ND	ND	ND
IN-13	5	------	Forage	N/A	T1	074-0001A1	ND	------	ND	------	ND	------	ND	------
IN-13	5	40278-9	Forage	45	T2	074-0002A1	ND	------	ND	------	ND	------	ND	------
IN-13	5	40278-9	Forage	45	T2	074-0003A1	ND	ND	ND	ND	ND	ND	ND	ND
IN-13	5	------	Forage	N/A	T1	075-0001A1	ND	------	ND	------	ND	------	ND	------
IN-13	5	40278-9	Forage	65	T2	075-0002A1	ND	------	ND	------	ND	------	ND	------
IN-13	5	40278-9	Forage	65	T2	075-0003A1	ND	ND	ND	ND	ND	ND	ND	ND
NE-14	5	------	Forage	N/A	T1	079-0001A1	ND	------	ND	------	ND	------	ND	------
NE-14	5	40278-9	Forage	63	T2	079-0002A1	ND	------	ND	------	ND	------	ND	------
NE-14	5	40278-9	Forage	63	T2	079-0003A1	ND	ND	ND	ND	ND	ND	ND	ND
NE-15	5	------	Forage	N/A	T1	083-0001A1	ND	------	ND	------	ND	------	ND	------
NE-15	5	40278-9	Forage	47	T2	083-0002A1	ND	------	ND	------	ND	------	ND	------
NE-15	5	40278-9	Forage	47	T2	083-0003A1	ND	ND	ND	ND	ND	ND	ND	ND

State/ Trial No.	NAFTA Region	AAD-1 Event[a] DAS-	RAC	PHI[b] (days)	Plot ID	Sample ID ARA-09-15-10-	Quizalofop acid (µg/g)		Quizalofop acid (as quizalofop-ethyl equivalents)[c]		Quizalofop-ethyl (µg/g)		Total Residue[d] (µg/g)	
								Avg[e]		Avg[e]		Avg[e]		Avg[e]
NE-15	5	------	Forage	N/A	T1	084-0001A1	ND	------	ND	------	ND	------	ND	------
NE-15	5	40278-9	Forage	71	T2	084-0002A1	ND	------	ND	------	ND	------	ND	------
NE-15	5	40278-9	Forage	71	T2	084-0003A1	ND	ND	ND	ND	ND	ND	ND	ND
KS-16	5	------	Forage	N/A	T1	087-0001A1	ND	------	ND	------	ND	------	ND	------
KS-16	5	40278-9	Forage	43	T2	087-0002A1	ND	------	ND	------	ND	------	ND	------
KS-16	5	40278-9	Forage	43	T2	087-0003A1	ND	ND	ND	ND	ND	ND	ND	ND
KS-16	5	------	Forage	N/A	T1	088-0001A1	ND	------	ND	------	ND	------	ND	------
KS-16	5	40278-9	Forage	69	T2	088-0002A1	ND	------	ND	------	ND	------	ND	------
KS-16	5	40278-9	Forage	69	T2	088-0003A1	ND	ND	ND	ND	ND	ND	ND	ND
IA-17	5	------	Forage	N/A	T1	091-0001A1	ND	------	ND	------	ND	------	ND	------
IA-17	5	40278-9	Forage	43	T2	091-0002A1	ND	------	ND	------	ND	------	ND	------
IA-17	5	40278-9	Forage	43	T2	091-0003A1	ND	ND	ND	ND	ND	ND	ND	ND
IA-17	5	------	Forage	N/A	T1	092-0001A1	ND	------	ND	------	ND	------	ND	------
IA-17	5	40278-9	Forage	65	T2	092-0002A1	ND	------	ND	------	ND	------	ND	------
IA-17	5	40278-9	Forage	65	T2	092-0003A1	ND	ND	ND	ND	ND	ND	ND	ND
MO-18	5	------	Forage	N/A	T1	095-0001A1	ND	------	ND	------	ND	------	ND	------
MO-18	5	40278-9	Forage	46	T2	095-0002A1	ND	------	ND	------	ND	------	ND	------

State/ Trial No.	NAFTA Region	AAD-1 Event[a] DAS-	RAC	PHI[b] (days)	Plot ID	Sample ID ARA-09-15-10-	Quizalofop acid (µg/g)		Quizalofop acid (as quizalofop-ethyl equivalents)[c]		Quizalofop-ethyl (µg/g)		Total Residue[d] (µg/g)	
								Avg[e]		Avg[e]		Avg[e]		Avg[e]
MO-18	5	40278-9	Forage	46	T2	095-0003A1	ND	ND	ND	ND	ND	ND	ND	ND
MO-18	5	------	Forage	N/A	T1	096-0001A1	ND	------	ND	------	ND	------	ND	------
MO-18	5	40278-9	Forage	59	T2	096-0002A1	ND	------	ND	------	ND	------	ND	------
MO-18	5	40278-9	Forage	59	T2	096-0003A1	ND	ND	ND	ND	ND	ND	ND	ND
IA-19	5	------	Forage	N/A	T1	099-0001A1	ND	------	ND	------	ND	------	ND	------
IA-19	5	40278-9	Forage	46	T2	099-0002A1	ND	------	ND	------	ND	------	ND	------
IA-19	5	40278-9	Forage	4	T2	099-0003A1	ND	ND	ND	ND	ND	ND	ND	ND
IA-19	5	------	Forage	N/A	T1	100-0001A1	ND	------	ND	------	ND	------	ND	------
IA-19	5	40278-9	Forage	73	T2	100-0002A1	ND	------	ND	------	ND	------	ND	------
IA-19	5	40278-9	Forage	73	T2	100-0003A1	ND	ND	ND	ND	ND	ND	ND	ND
MN-20	5	------	Forage	N/A	T1	103-0001A1	ND	------	ND	------	ND	------	ND	------
MN-20	5	40278-9	Forage	45	T2	103-0002A1	ND	------	ND	------	ND	------	ND	------
MN-20	5	40278-9	Forage	45	T2	103-0003A1	ND	ND	ND	ND	ND	ND	ND	ND
MN-20	5	------	Forage	N/A	T1	104-0001A1	ND	------	ND	------	ND	------	ND	------
MN-20	5	40278-9	Forage	73	T2	104-0002A1	ND	------	ND	------	ND	------	ND	------
MN-20	5	40278-9	Forage	73	T2	104-0003A1	ND	ND	ND	ND	ND	ND	ND	ND
MO-21	5	------	Forage	N/A	T1	107-0001A1	ND	------	ND	------	ND	------	ND	------

State/ Trial No.	NAFTA Region	AAD-1 Event[a] DAS-	RAC	PHI[b] (days)	Plot ID	Sample ID ARA-09-15-10-	Quizalofop acid (µg/g)		Quizalofop acid (as quizalofop-ethyl equivalents)[c]		Quizalofop-ethyl (µg/g)		Total Residue[d] (µg/g)	
								Avg[e]		Avg[e]		Avg[e]		Avg[e]
MO-21	5	40278-9	Forage	47	T2	107-0002A1	ND	------	ND	------	ND	------	ND	------
MO-21	5	40278-9	Forage	47	T2	107-0003A1	ND	ND	ND	ND	ND	ND	ND	ND
MO-21	5	------	Forage	N/A	T1	108-0001A1	ND	------	ND	------	ND	------	ND	------
MO-21	5	40278-9	Forage	62	T2	108-0002A1	ND	------	ND	------	ND	------	ND	------
MO-21	5	40278-9	Forage	62	T2	108-0003A1	ND	ND	ND	ND	ND	ND	ND	ND
OK-22	6	------	Forage	N/A	T1	111-0001A1	ND	------	ND	------	ND	------	ND	------
OK-22	6	40278-9	Forage	35	T2	111-0002A1	ND	------	ND	------	ND	------	ND	------
OK-22	6	40278-9	Forage	35	T2	111-0003A1	ND	ND	ND	ND	ND	ND	ND	ND
OK-22	6	------	Forage	N/A	T1	114-0001A1	ND	------	ND	------	ND	------	ND	------
OK-22	6	40278-9	Forage	54	T2	114-0002A1	ND	------	ND	------	ND	------	ND	------
OK-22	6	40278-9	Forage	54	T2	114-0003A1	ND	ND	ND	ND	ND	ND	ND	ND
TX-23	6	------	Forage	N/A	T1	127-0001A1	ND	------	ND	------	ND	------	ND	------
TX-23	6	40278-9	Forage	32	T2	127-0002A1	ND	------	ND	------	ND	------	ND	------
TX-23	6	40278-9	Forage	32	T2	127-0003A1	ND	ND	ND	ND	ND	ND	ND	ND
TX-23	6	------	Forage	N/A	T1	128-0001A1	ND	------	ND	------	ND	------	ND	------
TX-23	6	40278-9	Forage	44	T2	128-0002A1	0.167	------	0.180	------	0.197	------	0.377	------
TX-23	6	40278-9	Forage	44	T2	128-0003A1	0.113	0.140	0.122	0.151	0.099	0.148	0.221	0.299

State/ Trial No.	NAFTA Region	AAD-1 Event[a] DAS-	RAC	PHI[b] (days)	Plot ID	Sample ID ARA-09-15-10-	Quizalofop acid (µg/g)		Quizalofop acid (as quizalofop-ethyl equivalents)[c]		Quizalofop-ethyl (µg/g)		Total Residue[d] (µg/g)	
								Avg[e]		Avg[e]		Avg[e]		Avg[e]
ON-24	5	------	Forage	N/A	T1	131-0001A1	ND	------	ND	------	ND	------	ND	------
ON-24	5	40278-9	Forage	46	T2	131-0002A1	ND	------	ND	------	ND	------	ND	------
ON-24	5	40278-9	Forage	46	T2	131-0003A1	ND	ND	ND	ND	ND	ND	ND	ND
ON-24	5	------	Forage	N/A	T1	132-0001A1	ND	------	ND	------	ND	------	ND	------
ON-24	5	40278-9	Forage	83	T2	132-0002A1	ND	------	ND	------	ND	------	ND	------
ON-24	5	40278-9	Forage	83	T2	132-0003A1	ND	ND	ND	ND	ND	ND	ND	ND
ON-25	5	------	Forage	N/A	T1	135-0001A1	ND	------	ND	------	ND	------	ND	------
ON-25	5	40278-9	Forage	44	T2	135-0002A1	ND	------	ND	------	ND	------	ND	------
ON-25	5	40278-9	Forage	44	T2	135-0003A1	ND	ND	ND	ND	ND	ND	ND	ND
ON-25	5	------	Forage	N/A	T1	136-0001A1	ND	------	ND	------	ND	------	ND	------
ON-25	5	40278-9	Forage	73	T2	136-0002A1	ND	------	ND	------	ND	------	ND	------
ON-25	5	40278-9	Forage	73	T2	136-0003A1	ND	ND	ND	ND	ND	ND	ND	ND

[a] Untreated control plots (T1) were planted with the commercial variety Mycogen 2M746, which lacked the inserted AAD-1 gene, but had a genetic background similar to that used for the two non-commercial transgenic hybrids in treated plots (T2).
[b] PHI = Pre Harvest Interval (actual days after the single application of quizalofop-P-ethyl)
[c] The quizalofop acid residue values are converted to quizalofop-ethyl equivalents based on the molecular weight of the two compounds. Quizalofop-ethyl has a molecular weight of 372.8 and quizalofop acid has a molecular weight of 344.8. The factor to use to convert quizalofop acid to quizalofop-ethyl equivalents is calculated by dividing the molecular weight of quizalofop-ethyl by the molecular weight of quizalofop acid, which calculates to 1.08 (372.8/344.8 = 1.08). Therefore, the residue values for quizalofop acid are multiplied by 1.08 to convert them to quizalofop-ethyl equivalents.
[d] Total residue = sum of quizalofop acid (expressed as quizalofop-ethyl equivalents) and quizalofop-ethyl.
[e] Averages presented in this column are of treated samplings (T2 samples) only at each trial's sampling event.
[f] ND = Not Detected (below the limit of detection of 0.003 µg/g)

TABLE XI – Decline Sampling Residues of Quizalofop Acid and Quizalofop-ethyl in Corn Forage

State/ Trial No.	NAFTA Region	AAD-1 Event[a] DAS-	RAC	PHI[b] (days)	Plot ID	Sample ID ARA-09-15-10-	Quizalofop acid (µg/g)		Quizalofop acid (as quizalofop-ethyl equivalents)[c]		Quizalofop-ethyl (µg/g)		Total Residue[d] (µg/g)	
								Avg[e]		Avg[e]		Avg[e]		Avg[e]
GA-04	2	------	Forage	N/A	T1	014-0001A1	ND[f]	------	ND	------	ND	------	ND	------
GA-04	2	40278-9	Forage	34	T2	014-0002A1	ND	------	ND	------	ND	------	ND	------
GA-04	2	40278-9	Forage	34	T2	014-0003A1	ND	ND	ND	ND	ND	ND	ND	ND
GA-04	2	------	Forage	N/A	T1	015-0001A1	ND	------	ND	------	ND	------	ND	------
GA-04	2	40278-9	Forage	36	T2	015-0002A1	ND	------	ND	------	ND	------	ND	------
GA-04	2	40278-9	Forage	36	T2	015-0003A1	ND	ND	ND	ND	[0.004][g]	ND[h]	[0.004]	ND
GA-04	2	------	Forage	N/A	T1	016-0001A1	ND	------	ND	------	ND	------	ND	------
GA-04	2	40278-9	Forage	43	T2	016-0002A1	ND	------	ND	------	ND	------	ND	------
GA-04	2	40278-9	Forage	43	T2	016-0003A1	ND	ND	ND	ND	ND	ND	ND	ND
GA-04	2	------	Forage	N/A	T1	017-0001A1	ND	------	ND	------	ND	------	ND	------
GA-04	2	40278-9	Forage	50	T2	017-0002A1	ND	------	ND	------	ND	------	ND	------
GA-04	2	40278-9	Forage	50	T2	017-0003A1	ND	ND	ND	ND	ND	ND	ND	ND
GA-04	2	------	Forage	N/A	T1	018-0001A1	ND	------	ND	------	ND	------	ND	------
GA-04	2	40278-9	Forage	57	T2	018-0002A1	ND	------	ND	------	ND	------	ND	------
GA-04	2	40278-9	Forage	57	T2	018-0003A1	ND	ND	ND	ND	ND	ND	ND	ND
GA-04	2	------	Forage	N/A	T1	019-0001A1	ND	------	ND	------	ND	------	ND	------

State/ Trial No.	NAFTA Region	AAD-1 Event[a] DAS-	RAC	PHI[b] (days)	Plot ID	Sample ID ARA-09-15-10-	Quizalofop acid (µg/g)		Quizalofop acid (as quizalofop-ethyl equivalents)[c]		Quizalofop-ethyl (µg/g)		Total Residue[d] (µg/g)	
								Avg[e]		Avg[e]		Avg[e]		Avg[e]
GA-04	2	40278-9	Forage	64	T2	019-0002A1	ND	------	ND	------	ND	------	ND	------
GA-04	2	40278-9	Forage	64	T2	019-0003A1	ND	ND	ND	ND	ND	ND	ND	ND
MI-11	5	------	Forage	N/A	T1	054-0001A1	ND	------	ND	------	ND	------	ND	------
MI-11	5	40278-9	Forage	45	T2	054-0002A1	ND	------	ND	------	ND	------	ND	------
MI-11	5	40278-9	Forage	45	T2	054-0003A1	ND	ND	ND	ND	ND	ND	ND	ND
MI-11	5	------	Forage	N/A	T1	055-0001A1	ND	------	ND	------	ND	------	ND	------
MI-11	5	40278-9	Forage	51	T2	055-0002A1	ND	------	ND	------	ND	------	ND	------
MI-11	5	40278-9	Forage	51	T2	055-0003A1	ND	ND	ND	ND	ND	ND	ND	ND
MI-11	5	------	Forage	N/A	T1	056-0001A1	ND	------	ND	------	ND	------	ND	------
MI-11	5	40278-9	Forage	58	T2	056-0002A1	ND	------	ND	------	ND	------	ND	------
MI-11	5	40278-9	Forage	58	T2	056-0003A1	ND	ND	ND	ND	ND	ND	ND	ND
MI-11	5	------	Forage	N/A	T1	057-0001A1	ND	------	ND	------	ND	------	ND	------
MI-11	5	40278-9	Forage	64	T2	057-0002A1	ND	------	ND	------	ND	------	ND	------
MI-11	5	40278-9	Forage	64	T2	057-0003A1	ND	ND	ND	ND	ND	ND	ND	ND
MI-11	5	------	Forage	N/A	T1	058-0001A1	ND	------	ND	------	ND	------	ND	------
MI-11	5	40278-9	Forage	71	T2	058-0002A1	ND	------	ND	------	ND	------	ND	------

State/ Trial No.	NAFTA Region	AAD-1 Event[a] DAS-	RAC	PHI[b] (days)	Plot ID	Sample ID ARA-09-15-10-	Quizalofop acid (µg/g)		Quizalofop acid (as quizalofop-ethyl equivalents)[c]		Quizalofop-ethyl (µg/g)		Total Residue[d] (µg/g)	
								Avg[e]		Avg[e]		Avg[e]		Avg[e]
MI-11	5	40278-9	Forage	71	T2	058-0003A1	ND	ND	ND	ND	ND	ND	ND	ND
MI-11	5	------	Forage	N/A	T1	059-0001A1	ND	------	ND	------	ND	------	ND	------
MI-11	5	40278-9	Forage	78	T2	059-0002A1	ND	------	ND	------	ND	------	ND	------
MI-11	5	40278-9	Forage	78	T2	059-0003A1	ND	ND	ND	ND	ND	ND	ND	ND
OK-22	6	------	Forage	N/A	T1	111-0001A1	ND	------	ND	------	ND	------	ND	------
OK-22	6	40278-9	Forage	35	T2	111-0002A1	ND	------	ND	------	ND	------	ND	------
OK-22	6	40278-9	Forage	35	T2	111-0003A1	ND	ND	ND	ND	ND	ND	ND	ND
OK-22	6	------	Forage	N/A	T1	112-0001A1	ND	------	ND	------	[0.003]	------	[0.003]	------
OK-22	6	40278-9	Forage	42	T2	112-0002A1	[0.003]	------	[0.003]	------	ND	------	[0.003]	------
OK-22	6	40278-9	Forage	42	T2	112-0003A1	ND	ND	ND	ND	ND	ND	ND	ND
OK-22	6	------	Forage	N/A	T1	113-0001A1	ND	------	ND	------	ND	------	ND	------
OK-22	6	40278-9	Forage	48	T2	113-0002A1	ND	------	ND	------	ND	------	ND	------
OK-22	6	40278-9	Forage	48	T2	113-0003A1	[0.004]	ND	[0.004]	ND	ND	ND	[0.004]	ND
OK-22	6	------	Forage	N/A	T1	114-0001A1	ND	------	ND	------	ND	------	ND	------
OK-22	6	40278-9	Forage	54	T2	114-0002A1	ND	------	ND	------	ND	------	ND	------
OK-22	6	40278-9	Forage	54	T2	114-0003A1	ND	ND	ND	ND	ND	ND	ND	ND

State/ Trial No.	NAFTA Region	AAD-1 Event[a] DAS-	RAC	PHI[b] (days)	Plot ID	Sample ID ARA-09-15-10-	Quizalofop acid (µg/g)		Quizalofop acid (as quizalofop-ethyl equivalents)[c]		Quizalofop-ethyl (µg/g)		Total Residue[d] (µg/g)	
								Avg[e]		Avg[e]		Avg[e]		Avg[e]
OK-22	6	------	Forage	N/A	T1	115-0001A1	ND	------	ND	------	ND	------	ND	------
OK-22	6	40278-9	Forage	60	T2	115-0002A1	ND	------	ND	------	ND	------	ND	------
OK-22	6	40278-9	Forage	60	T2	115-0003A1	ND	ND	ND	ND	ND	ND	ND	ND
OK-22	6	------	Forage	N/A	T1	116-0001A1	ND	------	ND	------	ND	------	ND	------
OK-22	6	40278-9	Forage	66	T2	116-0002A1	ND	------	ND	------	ND	------	ND	------
OK-22	6	40278-9	Forage	66	T2	116-0003A1	ND	ND	ND	ND	ND	ND	ND	ND

[a] Untreated control plots (T1) were planted with the commercial variety Mycogen 2M746, which lacked the inserted AAD-1 gene, but had a genetic background similar to that used for the two non-commercial transgenic hybrids in treated plots (T2).

[b] PHI = Pre Harvest Interval (actual days after the single application of quizalofop-P-ethyl)

[c] The quizalofop acid residue values are converted to quizalofop-ethyl equivalents based on the molecular weight of the two compounds. Quizalofop-ethyl has a molecular weight of 372.8 and quizalofop acid has a molecular weight of 344.8. The factor to use to convert quizalofop acid to quizalofop-ethyl equivalents is calculated by dividing the molecular weight of quizalofop-ethyl by the molecular weight of quizalofop acid, which calculates to 1.08 (372.8/344.8 = 1.08). Therefore, the residue values for quizalofop acid are multiplied by 1.08 to convert them to quizalofop-ethyl equivalents.

[d] Total residue = sum of quizalofop acid (expressed as quizalofop-ethyl equivalents) and quizalofop-ethyl.

[e] Averages presented in this column are of treated samplings (T2 samples) only at each trial's sampling event.

[f] ND = Not Detected (below the limit of detection of 0.003 µg/g)

[g] [ ] = Residue values greater than or equal to the limit of detection of 0.003 µg/g but less than the limit of quantitation of 0.01 µg/g. Values are reported with a lower degree of confidence than values above the limit of quantitation.

[h] For the purposes of calculating an average, "ND" is given the value of zero.

TABLE XII - Residues of Quizalofop Acid and Quizalofop-ethyl in Corn Grain

State/ Trial No.	NAFTA Region	AAD-1 Event[a] DAS-	RAC	PHI[b] (days)	Plot ID	Sample ID ARA-09-15-10-	Quizalofop acid (µg/g)		Quizalofop acid (as quizalofop-ethyl equivalents)[c]		Quizalofop-ethyl (µg/g)		Total Residue[d] (µg/g)	
								Avg[e]		Avg[e]		Avg[e]		Avg[e]
NJ-01	1	------	Grain	NA	T1	003-0001A1	ND[f]	------	ND	------	ND	------	ND[d]	------
NJ-01	1	40278-9	Grain	111	T2	003-0002A1	ND	------	ND	------	ND	------	ND	------
NJ-01	1	40278-9	Grain	111	T2	003-0003A1	ND	ND	ND	ND	ND	ND	ND	ND
PA-02	1	------	Grain	NA	T1	007-0001A1	ND	------	ND	------	ND	------	ND	------
PA-02	1	40278-9	Grain	112	T2	007-0002A1	ND	------	ND	------	ND	------	ND	------
PA-02	1	40278-9	Grain	112	T2	007-0003A1	ND	ND	ND	ND	ND	ND	ND	ND
GA-03	2	------	Grain	N/A	T1	011-0001A1	ND	------	ND	------	ND	------	ND	------
GA-03	2	40278-9	Grain	79	T2	011-0002A1	ND	------	ND	------	ND	------	ND	------
GA-03	2	40278-9	Grain	79	T2	011-0003A1	ND	ND	ND	ND	ND	ND	ND	ND
GA-04	2	------	Grain	N/A	T1	024-0001A1	ND	------	ND	------	ND	------	ND	------
GA-04	2	40278-9	Grain	80	T2	024-0002A1	ND	------	ND	------	ND	------	ND	------
GA-04	2	40278-9	Grain	80	T2	024-0003A1	ND	ND	ND	ND	ND	ND	ND	ND
IL-05	5	------	Grain	N/A	T1	032-0001A1	ND	------	ND	------	ND	------	ND	------
IL-05	5	40278-9	Grain	139	T2	032-0002A1	ND	------	ND	------	ND	------	ND	------
IL-05	5	40278-9	Grain	139	T2	032-0003A1	ND	ND	ND	ND	ND	ND	ND	ND
IL-06	5	------	Grain	N/A	T1	036-0001A1	ND	------	ND	------	ND	------	ND	------

State/ Trial No.	NAFTA Region	AAD-1 Event[a] DAS-	RAC	PHI[b] (days)	Plot ID	Sample ID ARA-09-15-10-	Quizalofop acid (µg/g)		Quizalofop acid (as quizalofop-ethyl equivalents)[c]		Quizalofop-ethyl (µg/g)		Total Residue[d] (µg/g)	
								Avg[e]		Avg[e]		Avg[e]		Avg[e]
IL-06	5	40278-9	Grain	124	T2	036-0002A1	ND	------	ND	------	ND	------	ND	------
IL-06	5	40278-9	Grain	124	T2	036-0003A1	[0.004][g]	ND[h]	[0.004]	ND	ND	ND	[0.004]	ND
IN-07	5	------	Grain	N/A	T1	040-0001A1	ND	------	ND	------	ND	------	ND	------
IN-07	5	40278-9	Grain	144	T2	040-0002A1	ND	------	ND	------	ND	------	ND	------
IN-07	5	40278-9	Grain	144	T2	040-0003A1	ND	ND	ND	ND	ND	ND	ND	ND
MO-08	5	------	Grain	N/A	T1	044-0001A1	ND	------	ND	------	ND	------	ND	------
MO-08	5	40278-9	Grain	109	T2	044-0002A1	ND	------	ND	------	ND	------	ND	------
MO-08	5	40278-9	Grain	109	T2	044-0003A1	ND	ND	ND	ND	ND	ND	ND	ND
MO-09	5	------	Grain	N/A	T1	048-0001A1	ND	------	ND	------	ND	------	ND	------
MO-09	5	40278-9	Grain	110	T2	048-0002A1	ND	------	ND	------	ND	------	ND	------
MO-09	5	40278-9	Grain	110	T2	048-0003A1	ND	ND	ND	ND	ND	ND	ND	ND
IL-10	5	------	Grain	N/A	T1	052-0001A1	ND	------	ND	------	ND	------	ND	------
IL-10	5	40278-9	Grain	105	T2	052-0002A1	ND	------	ND	------	ND	------	ND	------
IL-10	5	40278-9	Grain	105	T2	052-0003A1	ND	ND	ND	ND	ND	ND	ND	ND
MI-11	5	------	Grain	N/A	T1	064-0001A1	ND	------	ND	------	ND	------	ND	------
MI-11	5	40278-9	Grain	95	T2	064-0002A1	ND	------	ND	------	ND	------	ND	------
MI-11	5	40278-9	Grain	95	T2	064-0003A1	ND	ND	ND	ND	ND	ND	ND	ND

State/ Trial No.	NAFTA Region	AAD-1 Event[a] DAS-	RAC	PHI[b] (days)	Plot ID	Sample ID ARA-09-15-10-	Quizalofop acid (µg/g)		Quizalofop acid (as quizalofop-ethyl equivalents)[c]		Quizalofop-ethyl (µg/g)		Total Residue[d] (µg/g)	
								Avg[e]		Avg[e]		Avg[e]		Avg[e]
OH-12	5	------	Grain	N/A	T1	072-0001A1	ND	------	ND	------	ND	------	ND	------
OH-12	5	40278-9	Grain	95	T2	072-0002A1	ND	------	ND	------	ND	------	ND	------
OH-12	5	40278-9	Grain	95	T2	072-0003A1	ND	ND	ND	ND	ND	ND	ND	ND
IN-13	5	------	Grain	N/A	T1	076-0001A1	ND	------	ND	------	ND	------	ND	------
IN-13	5	40278-9	Grain	95	T2	076-0002A1	ND	------	ND	------	ND	------	ND	------
IN-13	5	40278-9	Grain	95	T2	076-0003A1	ND	ND	ND	ND	ND	ND	ND	ND
NE-14	5	------	Grain	N/A	T1	080-0001A1	ND	------	ND	------	ND	------	ND	------
NE-14	5	40278-9	Grain	140	T2	080-0002A1	ND	------	ND	------	ND	------	ND	------
NE-14	5	40278-9	Grain	140	T2	080-0003A1	ND	ND	ND	ND	ND	ND	ND	ND
NE-15	5	------	Grain	N/A	T1	085-0001A1	ND	------	ND	------	ND	------	ND	------
NE-15	5	40278-9	Grain	94	T2	085-0002A1	ND	------	ND	------	ND	------	ND	------
NE-15	5	40278-9	Grain	94	T2	085-0003A1	ND	ND	ND	ND	ND	ND	ND	ND
KS-16	5	------	Grain	N/A	T1	089-0001A1	ND	------	ND	------	ND	------	ND	------
KS-16	5	40278-9	Grain	98	T2	089-0002A1	ND	------	ND	------	ND	------	ND	------
KS-16	5	40278-9	Grain	98	T2	089-0003A1	ND	ND	ND	ND	ND	ND	ND	ND
IA-17	5	------	Grain	N/A	T1	093-0001A1	ND	------	ND	------	ND	------	ND	------
IA-17	5	40278-9	Grain	108	T2	093-0002A1	ND	------	ND	------	ND	------	ND	------

State/ Trial No.	NAFTA Region	AAD-1 Event[a] DAS-	RAC	PHI[b] (days)	Plot ID	Sample ID ARA-09-15-10-	Quizalofop acid (µg/g)		Quizalofop acid (as quizalofop-ethyl equivalents)[c]		Quizalofop-ethyl (µg/g)		Total Residue[d] (µg/g)	
								Avg[e]		Avg[e]		Avg[e]		Avg[e]
IA-17	5	40278-9	Grain	108	T2	093-0003A1	ND	ND	ND	ND	ND	ND	ND	ND
MO-18	5	------	Grain	N/A	T1	097-0001A1	ND	------	ND	------	ND	------	ND	------
MO-18	5	40278-9	Grain	106	T2	097-0002A1	ND	------	ND	------	ND	------	ND	------
MO-18	5	40278-9	Grain	106	T2	097-0003A1	ND	ND	ND	ND	ND	ND	ND	ND
IA-19	5	------	Grain	N/A	T1	101-0001A1	ND	------	ND	------	ND	------	ND	------
IA-19	5	40278-9	Grain	126	T2	101-0002A1	ND	------	ND	------	ND	------	ND	------
IA-19	5	40278-9	Grain	126	T2	101-0003A1	ND	ND	ND	ND	ND	ND	ND	ND
MN-20	5	------	Grain	N/A	T1	105-0001A1	ND	------	ND	------	ND	------	ND	------
MN-20	5	40278-9	Grain	126	T2	105-0002A1	ND	------	ND	------	ND	------	ND	------
MN-20	5	40278-9	Grain	126	T2	105-0003A1	ND	ND	ND	ND	ND	ND	ND	ND
MO-21	5	------	Grain	N/A	T1	109-0001A1	ND	------	ND	------	ND	------	ND	------
MO-21	5	40278-9	Grain	95	T2	109-0002A1	ND	------	ND	------	ND	------	ND	------
MO-21	5	40278-9	Grain	95	T2	109-0003A1	ND	ND	ND	ND	ND	ND	ND	ND
OK-22	6	------	Grain	N/A	T1	121-0001A1	ND	------	ND	------	ND	------	ND	------
OK-22	6	40278-9	Grain	84	T2	121-0002A1	ND	------	ND	------	ND	------	ND	------
OK-22	6	40278-9	Grain	84	T2	121-0003A1	ND	ND	ND	ND	ND	ND	ND	ND
TX-23	6	------	Grain	N/A	T1	129-0001A1	ND	------	ND	------	ND	------	ND	------

State/ Trial No.	NAFTA Region	AAD-1 Event[a] DAS-	RAC	PHI[b] (days)	Plot ID	Sample ID ARA-09-15-10-	Quizalofop acid (µg/g)		Quizalofop acid (as quizalofop-ethyl equivalents)[c]		Quizalofop-ethyl (µg/g)		Total Residue[d] (µg/g)	
								Avg[e]		Avg[e]		Avg[e]		Avg[e]
TX-23	6	40278-9	Grain	112	T2	129-0002A1	ND	------	ND	------	ND	------	ND	------
TX-23	6	40278-9	Grain	112	T2	129-0003A1	ND	ND	ND	ND	ND	ND	ND	ND
ON-24	5	------	Grain	N/A	T1	133-0001A1	ND	------	ND	------	ND	------	ND	------
ON-24	5	40278-9	Grain	131	T2	133-0002A1	ND	------	ND	------	ND	------	ND	------
ON-24	5	40278-9	Grain	131	T2	133-0003A1	ND	ND	ND	ND	ND	ND	ND	ND
ON-25	5	------	Grain	N/A	T1	137-0001A1	[0.006]	------	[0.006]	------	[0.003]	------	[0.009]	-----
ON-25	5	40278-9	Grain	128	T2	137-0002A1	ND	------	ND	------	ND	------	ND	------
ON-25	5	40278-9	Grain	128	T2	137-0003A1	ND	ND	ND	ND	ND	ND	ND	ND

[a] Untreated control plots (T1) were planted with the commercial variety Mycogen 2M746, which lacked the inserted AAD-1 gene, but had a genetic background similar to that used for the two non-commercial transgenic hybrids in treated plots (T2).

[b] PHI = Pre Harvest Interval (actual days after the single application of quizalofop-P-ethyl)

[c] The quizalofop acid residue values are converted to quizalofop-ethyl equivalents based on the molecular weight of the two compounds. Quizalofop-ethyl has a molecular weight of 372.8 and quizalofop acid has a molecular weight of 344.8. The factor to use to convert quizalofop acid to quizalofop-ethyl equivalents is calculated by dividing the molecular weight of quizalofop-ethyl by the molecular weight of quizalofop acid, which calculates to 1.08 (372.8/344.8 = 1.08). Therefore, the residue values for quizalofop acid are multiplied by 1.08 to convert them to quizalofop-ethyl equivalents.

[d] Total residue = sum of quizalofop acid (expressed as quizalofop-ethyl equivalents) and quizalofop-ethyl.

[e] Averages presented in this column are of treated samplings (T2 samples) only at each trial's sampling event.

[f] ND = Not Detected (below the limit of detection of 0.003 µg/g)

[g] [ ] = Residue values greater than or equal to the limit of detection of 0.003 µg/g but less than the limit of quantitation of 0.01 µg/g. Values are reported with a lower degree of confidence than values above the limit of quantitation.

[h] For the purposes of calculating an average, "ND" is given the value of zero.

TABLE XIII – Decline Sampling Residues of Quizalofop Acid and Quizalofop-ethyl in Corn Grain

State/ Trial No.	NAFTA Region	AAD-1 Event[a] DAS-	RAC	PHI[b] (days)	Plot ID	Sample ID ARA-09-15-10-	Quizalofop acid (µg/g)		Quizalofop acid (as quizalofop-ethyl equivalents)[c]		Quizalofop-ethyl (µg/g)		Total Residue[d] (µg/g)	
								Avg[e]		Avg[e]		Avg[e]		Avg[e]
GA-04	2	------	Grain	N/A	T1	020-0001A2	ND[f]	------	ND	------	ND	------	ND	------
GA-04	2	40278-9	Grain	66	T2	020-0002A1	ND	------	ND	------	ND	------	ND	------
GA-04	2	40278-9	Grain	66	T2	020-0003A1	ND	ND	ND	ND	ND	ND	ND	ND
GA-04	2	------	Grain	N/A	T1	022-0001A1	ND	------	ND	------	ND	------	ND	------
GA-04	2	40278-9	Grain	73	T2	022-0002A1	ND	------	ND	------	ND	------	ND	------
GA-04	2	40278-9	Grain	73	T2	022-0003A1	ND	ND	ND	ND	ND	ND	ND	ND
GA-04	2	------	Grain	N/A	T1	024-0001A1	ND	------	ND	------	ND	------	ND	------
GA-04	2	40278-9	Grain	80	T2	024-0002A1	ND	------	ND	------	ND	------	ND	------
GA-04	2	40278-9	Grain	80	T2	024-0003A1	ND	ND	ND	ND	ND	ND	ND	ND
GA-04	2	------	Grain	N/A	T1	026-0001A1	ND	------	ND	------	ND	------	ND	------
GA-04	2	40278-9	Grain	87	T2	026-0002A1	ND	------	ND	------	ND	------	ND	------
GA-04	2	40278-9	Grain	87	T2	026-0003A1	ND	ND	ND	ND	ND	ND	ND	ND
GA-04	2	------	Grain	N/A	T1	028-0001A1	ND	------	ND	------	ND	------	ND	------
GA-04	2	40278-9	Grain	94	T2	028-0002A1	ND	------	ND	------	ND	------	ND	------
GA-04	2	40278-9	Grain	94	T2	028-0003A1	ND	ND	ND	ND	ND	ND	ND	ND
MI-11	5	------	Grain	N/A	T1	060-0001A1	ND	------	ND	------	ND	------	ND	------

State/ Trial No.	NAFTA Region	AAD-1 Event[a] DAS-	RAC	PHI[b] (days)	Plot ID	Sample ID ARA-09-15-10-	Quizalofop acid (µg/g)		Quizalofop acid (as quizalofop-ethyl equivalents)[c]		Quizalofop-ethyl (µg/g)		Total Residue[d] (µg/g)	
								Avg[e]		Avg[e]		Avg[e]		Avg[e]
MI-11	5	40278-9	Grain	81	T2	060-0002A1	ND	------	ND	------	ND	------	ND	------
MI-11	5	40278-9	Grain	81	T2	060-0003A1	ND	ND	ND	ND	ND	ND	ND	ND
MI-11	5	------	Grain	N/A	T1	062-0001A1	ND	------	ND	------	ND	------	ND	------
MI-11	5	40278-9	Grain	88	T2	062-0002A1	ND	------	ND	------	ND	------	ND	------
MI-11	5	40278-9	Grain	88	T2	062-0003A1	ND	ND	ND	ND	ND	ND	ND	ND
MI-11	5	------	Grain	N/A	T1	064-0001A1	ND	------	ND	------	ND	------	ND	------
MI-11	5	40278-9	Grain	95	T2	064-0002A1	ND	------	ND	------	ND	------	ND	------
MI-11	5	40278-9	Grain	95	T2	064-0003A1	ND	ND	ND	ND	ND	ND	ND	ND
MI-11	5	------	Grain	N/A	T1	066-0001A1	ND	------	ND	------	ND	------	ND	------
MI-11	5	40278-9	Grain	102	T2	066-0002A1	ND	------	ND	------	ND	------	ND	------
MI-11	5	40278-9	Grain	102	T2	066-0003A1	ND	ND	ND	ND	ND	ND	ND	ND
MI-11	5	------	Grain	N/A	T1	068-0001A1	ND	------	ND	------	ND	------	ND	------
MI-11	5	40278-9	Grain	109	T2	068-0002A1	ND	------	ND	------	ND	------	ND	------
MI-11	5	40278-9	Grain	109	T2	068-0003A1	ND	ND	ND	ND	ND	ND	ND	ND
OK-22	6	------	Grain	N/A	T1	117-0001A1	ND	------	ND	------	ND	------	ND	------
OK-22	6	40278-9	Grain	70	T2	117-0002A1	ND	------	ND	------	ND	------	ND	------
OK-22	6	40278-9	Grain	70	T2	117-0003A1	ND	ND	ND	ND	ND	ND	ND	ND

State/ Trial No.	NAFTA Region	AAD-1 Event[a] DAS-	RAC	PHI[b] (days)	Plot ID	Sample ID ARA-09-15-10-	Quizalofop acid (µg/g)		Quizalofop acid (as quizalofop-ethyl equivalents)[c]		Quizalofop-ethyl (µg/g)		Total Residue[d] (µg/g)	
								Avg[e]		Avg[e]		Avg[e]		Avg[e]
OK-22	6	------	Grain	N/A	T1	119-0001A1	ND	------	ND	------	ND	------	ND	------
OK-22	6	40278-9	Grain	76	T2	119-0002A1	ND	------	ND	------	ND	------	ND	------
OK-22	6	40278-9	Grain	76	T2	119-0003A1	ND	ND	ND	ND	ND	ND	ND	ND
OK-22	6	------	Grain	N/A	T1	121-0001A1	ND	------	ND	------	ND	------	ND	------
OK-22	6	40278-9	Grain	84	T2	121-0002A1	ND	------	ND	------	ND	------	ND	------
OK-22	6	40278-9	Grain	84	T2	121-0003A1	ND	ND	ND	ND	ND	ND	ND	ND
OK-22	6	------	Grain	N/A	T1	123-0001A1	ND	------	ND	------	ND	------	ND	------
OK-22	6	40278-9	Grain	91	T2	123-0002A1	ND	------	ND	------	ND	------	ND	------
OK-22	6	40278-9	Grain	91	T2	123-0003A1	ND	ND	ND	ND	ND	ND	ND	ND
OK-22	6	------	Grain	N/A	T1	125-0001A1	ND	------	ND	------	ND	------	ND	------
OK-22	6	40278-9	Grain	98	T2	125-0002A1	ND	------	ND	------	ND	------	ND	------
OK-22	6	40278-9	Grain	98	T2	125-0003A1	ND	ND	ND	ND	ND	ND	ND	ND

[a] Untreated control plots (T1) were planted with the commercial variety Mycogen 2M746, which lacked the inserted AAD-1 gene, but had a genetic background similar to that used for the two non-commercial transgenic hybrids in treated plots (T2).

[b] PHI = Pre Harvest Interval (actual days after the single application of quizalofop-P-ethyl)

[c] The quizalofop acid residue values are converted to quizalofop-ethyl equivalents based on the molecular weight of the two compounds. Quizalofop-ethyl has a molecular weight of 372.8 and quizalofop acid has a molecular weight of 344.8. The factor to use to convert quizalofop acid to quizalofop-ethyl equivalents is calculated by dividing the molecular weight of quizalofop-ethyl by the molecular weight of quizalofop acid, which calculates to 1.08 (372.8/344.8 = 1.08). Therefore, the residue values for quizalofop acid are multiplied by 1.08 to convert them to quizalofop-ethyl equivalents.

[d] Total residue = sum of quizalofop acid (expressed as quizalofop-ethyl equivalents) and quizalofop-ethyl.

[e] Averages presented in this column are of treated samplings (T2 samples) only at each trial's sampling event.

[f] ND = Not Detected (below the limit of detection of 0.003 µg/g)

TABLE XIV - Residues of Quizalofop Acid and Quizalofop-ethyl in Corn Stover

State/ Trial No.	NAFTA Region	AAD-1 Event[a] DAS-	RAC	PHI[b] (days)	Plot ID	Sample ID ARA-09-15-10-	Quizalofop acid (µg/g)		Quizalofop acid (as quizalofop-ethyl equivalents)[c]		Quizalofop-ethyl (µg/g)		Total Residue[d] (µg/g)	
								Avg[e]		Avg[e]		Avg[e]		Avg[e]
NJ-01	1	------	Stover	NA	T1	004-0001A1	ND[f]	------	ND	------	ND	------	ND[d]	------
NJ-01	1	40278-9	Stover	111	T2	004-0002A1	ND	------	ND	------	ND	------	ND	------
NJ-01	1	40278-9	Stover	111	T2	004-0003A1	ND	ND	ND	ND	ND	ND	ND	ND
PA-02	1	------	Stover	NA	T1	008-0001A1	ND	------	ND	------	ND	------	ND	------
PA-02	1	40278-9	Stover	112	T2	008-0002A1	ND	------	ND	------	ND	------	ND	------
PA-02	1	40278-9	Stover	112	T2	008-0003A1	ND	ND	ND	ND	ND	ND	ND	ND
GA-03	2	------	Stover	N/A	T1	013-0001A1	ND	------	ND	------	ND	------	ND	------
GA-03	2	40278-9	Stover	79	T2	013-0002A1	ND	------	ND	------	ND	------	ND	------
GA-03	2	40278-9	Stover	79	T2	013-0003A1	ND	ND	ND	ND	ND	ND	ND	ND
GA-04	2	------	Stover	N/A	T1	025-0001A1	ND	------	ND	------	ND	------	ND	------
GA-04	2	40278-9	Stover	80	T2	025-0002A1	[0.003][g]	------	[0.003]	------	ND	------	[0.003]	------
GA-04	2	40278-9	Stover	80	T2	025-0003A1	ND	ND[h]	ND	ND	ND	ND	ND	ND
IL-05	5	------	Stover	N/A	T1	033-0001A1	ND	------	ND	------	ND	------	ND	------
IL-05	5	40278-9	Stover	139	T2	033-0002A1	ND	------	ND	------	ND	------	ND	------
IL-05	5	40278-9	Stover	139	T2	033-0003A1	ND	ND	ND	ND	ND	ND	ND	ND
IL-06	5	------	Stover	N/A	T1	037-0001A1	ND	------	ND	------	ND	------	ND	------

State/ Trial No.	NAFTA Region	AAD-1 Event[a] DAS-	RAC	PHI[b] (days)	Plot ID	Sample ID ARA-09-15-10-	Quizalofop acid (µg/g)		Quizalofop acid (as quizalofop-ethyl equivalents)[c]		Quizalofop-ethyl (µg/g)		Total Residue[d] (µg/g)	
								Avg[e]		Avg[e]		Avg[e]		Avg[e]
IL-06	5	40278-9	Stover	124	T2	037-0002A1	ND	------	ND	------	ND	------	ND	------
IL-06	5	40278-9	Stover	124	T2	037-0003A1	ND	ND	ND	ND	ND	ND	ND	ND
IN-07	5	------	Stover	N/A	T1	041-0001A1	ND	------	ND	------	ND	------	ND	------
IN-07	5	40278-9	Stover	144	T2	041-0002A1	ND	------	ND	------	ND	------	ND	------
IN-07	5	40278-9	Stover	144	T2	041-0003A1	ND	ND	ND	ND	ND	ND	ND	ND
MO-08	5	------	Stover	N/A	T1	045-0001A1	ND	------	ND	------	ND	------	ND	------
MO-08	5	40278-9	Stover	109	T2	045-0002A1	ND	------	ND	------	ND	------	ND	------
MO-08	5	40278-9	Stover	109	T2	045-0003A1	ND	ND	ND	ND	ND	ND	ND	ND
MO-09	5	------	Stover	N/A	T1	049-0001A1	ND	------	ND	------	ND	------	ND	------
MO-09	5	40278-9	Stover	110	T2	049-0002A1	ND	------	ND	------	ND	------	ND	------
MO-09	5	40278-9	Stover	110	T2	049-0003A1	ND	ND	ND	ND	ND	ND	ND	ND
IL-10	5	------	Stover	N/A	T1	053-0001A1	ND	------	ND	------	ND	------	ND	------
IL-10	5	40278-9	Stover	105	T2	053-0002A1	ND	------	ND	------	ND	------	ND	------
IL-10	5	40278-9	Stover	105	T2	053-0003A1	ND	ND	ND	ND	ND	ND	ND	ND
MI-11	5	------	Stover	N/A	T1	065-0001A1	ND	------	ND	------	ND	------	ND	------
MI-11	5	40278-9	Stover	95	T2	065-0002A1	0.01	------	0.011	------	ND	------	0.011	------
MI-11	5	40278-9	Stover	95	T2	065-0003A1	[0.006]	[0.008]	[0.006]	[0.009]	ND	ND	[0.006]	[0.009]

State/ Trial No.	NAFTA Region	AAD-1 Event[a] DAS-	RAC	PHI[b] (days)	Plot ID	Sample ID ARA-09-15-10-	Quizalofop acid (µg/g)		Quizalofop acid (as quizalofop-ethyl equivalents)[c]		Quizalofop-ethyl (µg/g)		Total Residue[d] (µg/g)	
								Avg[e]		Avg[e]		Avg[e]		Avg[e]
OH-12	5	------	Stover	N/A	T1	073-0001A1	ND	------	ND	------	ND	------	ND	------
OH-12	5	40278-9	Stover	95	T2	073-0002A1	ND	------	ND	------	ND	------	ND	------
OH-12	5	40278-9	Stover	95	T2	073-0003A1	[0.004]	ND	[0.004]	ND	ND	ND	[0.004]	ND
IN-13	5	------	Stover	N/A	T1	077-0001A1	ND	------	ND	------	ND	------	ND	------
IN-13	5	40278-9	Stover	95	T2	077-0002A1	[0.003]	------	[0.003]	------	ND	------	[0.003]	------
IN-13	5	40278-9	Stover	95	T2	077-0003A1	ND	ND	ND	ND	ND	ND	ND	ND
NE-14	5	------	Stover	N/A	T1	082-0001A1	ND	------	ND	------	ND	------	ND	------
NE-14	5	40278-9	Stover	140	T2	082-0002A1	ND	------	ND	------	ND	------	ND	------
NE-14	5	40278-9	Stover	140	T2	082-0003A1	ND	ND	ND	ND	ND	ND	ND	ND
NE-15	5	------	Stover	N/A	T1	086-0001A1	ND	------	ND	------	ND	------	ND	------
NE-15	5	40278-9	Stover	94	T2	086-0002A1	ND	------	ND	------	ND	------	ND	------
NE-15	5	40278-9	Stover	94	T2	086-0003A1	ND	ND	ND	ND	ND	ND	ND	ND
KS-16	5	------	Stover	N/A	T1	090-0001A1	ND	------	ND	------	ND	------	ND	------
KS-16	5	40278-9	Stover	98	T2	090-0002A1	ND	------	ND	------	ND	------	ND	------
KS-16	5	40278-9	Stover	98	T2	090-0003A1	ND	ND	ND	ND	ND	ND	ND	ND
IA-17	5	------	Stover	N/A	T1	094-0001A1	ND	------	ND	------	ND	------	ND	------
IA-17	5	40278-9	Stover	108	T2	094-0002A1	ND	------	ND	------	ND	------	ND	------

State/ Trial No.	NAFTA Region	AAD-1 Event[a] DAS-	RAC	PHI[b] (days)	Plot ID	Sample ID ARA-09-15-10-	Quizalofop acid (µg/g)		Quizalofop acid (as quizalofop-ethyl equivalents)[c]		Quizalofop-ethyl (µg/g)		Total Residue[d] (µg/g)	
								Avg[e]		Avg[e]		Avg[e]		Avg[e]
IA-17	5	40278-9	Stover	108	T2	094-0003A1	ND	ND	ND	ND	ND	ND	ND	ND
MO-18	5	------	Stover	N/A	T1	098-0001A1	ND	------	ND	------	ND	------	ND	------
MO-18	5	40278-9	Stover	106	T2	098-0002A1	ND	------	ND	------	ND	------	ND	------
MO-18	5	40278-9	Stover	106	T2	098-0003A1	ND	ND	ND	ND	ND	ND	ND	ND
IA-19	5	------	Stover	N/A	T1	102-0001A1	ND	------	ND	------	ND	------	ND	------
IA-19	5	40278-9	Stover	126	T2	102-0002A1	ND	------	ND	------	ND	------	ND	------
IA-19	5	40278-9	Stover	126	T2	102-0003A1	ND	ND	ND	ND	ND	ND	ND	ND
MN-20	5	------	Stover	N/A	T1	106-0001A1	ND	------	ND	------	ND	------	ND	------
MN-20	5	40278-9	Stover	126	T2	106-0002A1	ND	------	ND	------	ND	------	ND	------
MN-20	5	40278-9	Stover	126	T2	106-0003A1	ND	ND	ND	ND	ND	ND	ND	ND
MO-21	5	------	Stover	N/A	T1	110-0001A1	ND	------	ND	------	ND	------	ND	------
MO-21	5	40278-9	Stover	95	T2	110-0002A1	ND	------	ND	------	ND	------	ND	------
MO-21	5	40278-9	Stover	95	T2	110-0003A1	ND	ND	ND	ND	ND	ND	ND	ND
OK-22	6	------	Stover	N/A	T1	122-0001A1	ND	------	ND	------	ND	------	ND	------
OK-22	6	40278-9	Stover	84	T2	122-0002A1	ND	------	ND	------	ND	------	ND	------
OK-22	6	40278-9	Stover	84	T2	122-0003A1	ND	ND	ND	ND	ND	ND	ND	ND
TX-23	6	------	Stover	N/A	T1	130-0001A1	ND	------	ND	------	ND	------	ND	------

State/ Trial No.	NAFTA Region	AAD-1 Event[a] DAS-	RAC	PHI[b] (days)	Plot ID	Sample ID ARA-09-15-10-	Quizalofop acid (µg/g)		Quizalofop acid (as quizalofop-ethyl equivalents)[c]		Quizalofop-ethyl (µg/g)		Total Residue[d] (µg/g)	
								Avg[e]		Avg[e]		Avg[e]		Avg[e]
TX-23	6	40278-9	Stover	112	T2	130-0002A1	0.010	------	0.011	------	[0.004]	------	0.015	------
TX-23	6	40278-9	Stover	112	T2	130-0003A1	0.011	0.011	0.012	0.012	[0.004]	[0.004]	0.016	0.016
ON-24	5	------	Stover	N/A	T1	134-0001A1	ND	------	ND	------	ND	------	ND	------
ON-24	5	40278-9	Stover	131	T2	134-0002A1	ND	------	ND	------	ND	------	ND	------
ON-24	5	40278-9	Stover	131	T2	134-0003A1	ND	ND	ND	ND	ND	ND	ND	ND
ON-25	5	------	Stover	N/A	T1	138-0001A1	ND	------	ND	------	ND	------	ND	------
ON-25	5	40278-9	Stover	128	T2	138-0002A1	ND	------	ND	------	ND	------	ND	------
ON-25	5	40278-9	Stover	128	T2	138-0003A1	ND	ND	ND	ND	[0.004]	ND	[0.004]	ND

[a] Untreated control plots (T1) were planted with the commercial variety Mycogen 2M746, which lacked the inserted AAD-1 gene, but had a genetic background similar to that used for the two non-commercial transgenic hybrids in treated plots (T2).

[b] PHI = Pre Harvest Interval (actual days after the single application of quizalofop-P-ethyl)

[c] The quizalofop acid residue values are converted to quizalofop-ethyl equivalents based on the molecular weight of the two compounds. Quizalofop-ethyl has a molecular weight of 372.8 and quizalofop acid has a molecular weight of 344.8. The factor to use to convert quizalofop acid to quizalofop-ethyl equivalents is calculated by dividing the molecular weight of quizalofop-ethyl by the molecular weight of quizalofop acid, which calculates to 1.08 (372.8/344.8 = 1.08). Therefore, the residue values for quizalofop acid are multiplied by 1.08 to convert them to quizalofop-ethyl equivalents.

[d] Total residue = sum of quizalofop acid (expressed as quizalofop-ethyl equivalents) and quizalofop-ethyl.

[e] Averages presented in this column are of treated samplings (T2 samples) only at each trial's sampling event.

[f] ND = Not Detected (below the limit of detection of 0.003 µg/g)

[g] [ ] = Residue values greater than or equal to the limit of detection of 0.003 µg/g but less than the limit of quantitation of 0.01 µg/g. Values are reported with a lower degree of confidence than values above the limit of quantitation.

[h] For the purposes of calculating an average, "ND" is given the value of zero.

TABLE XV – Decline Sampling Residues of Quizalofop Acid and Quizalofop-ethyl in Corn Stover

State/ Trial No.	NAFTA Region	AAD-1 Event[a] DAS-	RAC	PHI[b] (days)	Plot ID	Sample ID ARA-09-15-10-	Quizalofop acid (µg/g)		Quizalofop acid (as quizalofop-ethyl equivalents)[c]		Quizalofop-ethyl (µg/g)		Total Residue[d] (µg/g)	
								Avg[e]		Avg[e]		Avg[e]		Avg[e]
GA-04	2	------	Stover	N/A	T1	021-0001A1	ND[f]	------	ND	------	ND	------	ND	------
GA-04	2	40278-9	Stover	66	T2	021-0002A1	[0.004][g]	------	[0.004]	------	ND	------	[0.004]	------
GA-04	2	40278-9	Stover	66	T2	021-0003A1	[0.004]	[0.004]	[0.004]	[0.004]	ND	ND	[0.004]	[0.004]
GA-04	2	------	Stover	N/A	T1	023-0001A1	ND	------	ND	------	ND	------	ND	------
GA-04	2	40278-9	Stover	73	T2	023-0002A1	ND	------	ND	------	ND	------	ND	------
GA-04	2	40278-9	Stover	73	T2	023-0003A1	ND	ND	ND	ND	ND	ND	ND	ND
GA-04	2	------	Stover	N/A	T1	025-0001A1	ND	------	ND	------	ND	------	ND	------
GA-04	2	40278-9	Stover	80	T2	025-0002A1	[0.003]	------	[0.003]	------	ND	------	[0.003]	------
GA-04	2	40278-9	Stover	80	T2	025-0003A1	ND	ND[h]	ND	ND	ND	ND	ND	ND
GA-04	2	------	Stover	N/A	T1	027-0001A1	ND	------	ND	------	ND	------	ND	------
GA-04	2	40278-9	Stover	87	T2	027-0002A1	ND	------	ND	------	ND	------	ND	------
GA-04	2	40278-9	Stover	87	T2	027-0003A1	ND	ND	ND	ND	ND	ND	ND	ND
GA-04	2	------	Stover	N/A	T1	029-0001A1	ND	------	ND	------	ND	------	ND	------
GA-04	2	40278-9	Stover	94	T2	029-0002A1	ND	------	ND	------	ND	------	ND	------
GA-04	2	40278-9	Stover	94	T2	029-0003A1	ND	ND	ND	ND	ND	ND	ND	ND
MI-11	5	------	Stover	N/A	T1	061-0001A1	ND	------	ND	------	ND	------	ND	------

State/ Trial No.	NAFTA Region	AAD-1 Event[a] DAS-	RAC	PHI[b] (days)	Plot ID	Sample ID ARA-09-15-10-	Quizalofop acid (µg/g)		Quizalofop acid (as quizalofop-ethyl equivalents)[c]		Quizalofop-ethyl (µg/g)		Total Residue[d] (µg/g)	
								Avg[e]		Avg[e]		Avg[e]		Avg[e]
MI-11	5	40278-9	Stover	81	T2	061-0002A1	[0.004]	------	[0.004]	------	ND	------	[0.004]	------
MI-11	5	40278-9	Stover	81	T2	061-0003A1	[0.004]	[0.004]	[0.004]	[0.004]	ND	ND	[0.004]	[0.004]
MI-11	5	------	Stover	N/A	T1	063-0001A1	ND	------	ND	------	ND	------	ND	------
MI-11	5	40278-9	Stover	88	T2	063-0002A1	[0.004]	------	[0.004]	------	ND	------	[0.004]	------
MI-11	5	40278-9	Stover	88	T2	063-0003A1	[0.004]	[0.004]	[0.004]	[0.004]	ND	ND	[0.004]	[0.004]
MI-11	5	------	Stover	N/A	T1	065-0001A1	ND	------	ND	------	ND	------	ND	------
MI-11	5	40278-9	Stover	95	T2	065-0002A1	0.010	------	0.011	------	ND	------	0.011	------
MI-11	5	40278-9	Stover	95	T2	065-0003A1	[0.006]	[0.008]	[0.006]	[0.009]	ND	ND	[0.006]	[0.009]
MI-11	5	------	Stover	N/A	T1	067-0001A1	ND	------	ND	------	ND	------	ND	------
MI-11	5	40278-9	Stover	102	T2	067-0002A1	[0.006]	------	[0.006]	------	ND	------	[0.006]	------
MI-11	5	40278-9	Stover	102	T2	067-0003A1	[0.009]	[0.008]	0.010	[0.008]	ND	ND	0.010	[0.008]
MI-11	5	------	Stover	N/A	T1	069-0001A1	ND	------	ND	------	ND	------	ND	------
MI-11	5	40278-9	Stover	109	T2	069-0002A1	[0.005]	------	[0.005]	------	ND	------	[0.005]	------
MI-11	5	40278-9	Stover	109	T2	069-0003A1	0.015	0.010	0.016	0.011	ND	ND	0.016	0.011
OK-22	6	------	Stover	N/A	T1	118-0001A1	ND	------	ND	------	ND	------	ND	------
OK-22	6	40278-9	Stover	70	T2	118-0002A1	ND	------	ND	------	ND	------	ND	------
OK-22	6	40278-9	Stover	70	T2	118-0003A1	ND	ND	ND	ND	ND	ND	ND	ND

State/ Trial No.	NAFTA Region	AAD-1 Event[a] DAS-	RAC	PHI[b] (days)	Plot ID	Sample ID ARA-09-15-10-	Quizalofop acid (µg/g)		Quizalofop acid (as quizalofop-ethyl equivalents)[c]		Quizalofop-ethyl (µg/g)		Total Residue[d] (µg/g)	
								Avg[e]		Avg[e]		Avg[e]		Avg[e]
OK-22	6	------	Stover	N/A	T1	120-0001A1	ND	------	ND	------	ND	------	ND	------
OK-22	6	40278-9	Stover	76	T2	120-0002A1	ND	------	ND	------	ND	------	ND	------
OK-22	6	40278-9	Stover	76	T2	120-0003A1	ND	ND	ND	ND	ND	ND	ND	ND
OK-22	6	------	Stover	N/A	T1	122-0001A1	ND	------	ND	------	ND	------	ND	------
OK-22	6	40278-9	Stover	84	T2	122-0002A1	ND	------	ND	------	ND	------	ND	------
OK-22	6	40278-9	Stover	84	T2	122-0003A1	ND	ND	ND	ND	ND	ND	ND	ND
OK-22	6	------	Stover	N/A	T1	124-0001A1	ND	------	ND	------	ND	------	ND	------
OK-22	6	40278-9	Stover	91	T2	124-0002A1	ND	------	ND	------	ND	------	ND	------
OK-22	6	40278-9	Stover	91	T2	124-0003A1	ND	ND	ND	ND	ND	ND	ND	ND
OK-22	6	------	Stover	N/A	T1	126-0001A1	ND	------	ND	------	ND	------	ND	------
OK-22	6	40278-9	Stover	98	T2	126-0002A1	ND	------	ND	------	ND	------	ND	------
OK-22	6	40278-9	Stover	98	T2	126-0003A1	ND	ND	ND	ND	ND	ND	ND	ND

[a] Untreated control plots (T1) were planted with the commercial variety Mycogen 2M746, which lacked the inserted AAD-1 gene, but had a genetic background similar to that used for the two non-commercial transgenic hybrids in treated plots (T2).

[b] PHI = Pre Harvest Interval (actual days after the single application of quizalofop-P-ethyl)

[c] The quizalofop acid residue values are converted to quizalofop-ethyl equivalents based on the molecular weight of the two compounds. Quizalofop-ethyl has a molecular weight of 372.8 and quizalofop acid has a molecular weight of 344.8. The factor to use to convert quizalofop acid to quizalofop-ethyl equivalents is calculated by dividing the molecular weight of quizalofop-ethyl by the molecular weight of quizalofop acid, which calculates to 1.08 (372.8/344.8 = 1.08). Therefore, the residue values for quizalofop acid are multiplied by 1.08 to convert them to quizalofop-ethyl equivalents.

[d] Total residue = sum of quizalofop acid (expressed as quizalofop-ethyl equivalents) and quizalofop-ethyl.

[e] Averages presented in this column are of treated samplings (T2 samples) only at each trial's sampling event.

[f] ND = Not Detected (below the limit of detection of 0.003 µg/g)

[g] [ ] = Residue values greater than or equal to the limit of detection of 0.003 µg/g but less than the limit of quantitation of 0.01 µg/g. Values are reported with a lower degree of confidence than values above the limit of quantitation.

[h] For the purposes of calculating an average, "ND" is given the value of zero.

TABLE XVI -

Residues of Quizalofop Acid and Quizalofop-ethyl in Field Corn Processed Commodities (Trial GA-03, NAFTA Region 2)

AAD-1 Event[a] DAS-	Commodity	PHI[b] (days)	Plot ID	Sample ID ARA-09-15-10-	Quizalofop acid (µg/g)		Quizalofop acid (as quizalofop-ethyl equivalents)[c]		Quizalofop-ethyl (µg/g)		Total Residue[d] (µg/g)	
						Avg[e]		Avg[e]		Avg[e]		Avg[e]
------	Grain, from the field	N/A	T1	011-0001A1	ND[f]	------	ND[e]	------	ND	------	ND	------
40278-9	Grain, from the field	79	T4	011-0006A1	ND	------	ND	------	ND	------	ND	------
40278-9	Grain, from the field	79	T4	011-0007A1	[0.003][g]	ND[h]	[0.003]	ND	ND	ND	[0.003]	ND
------	Grain, from the processor prior to AGF generation	N/A	T1	139-0001A1	ND	------	ND	------	ND	------	ND	------
------	Grain, from the processor prior to AGF generation	N/A	T1	139-0002A1	ND	------	ND	------	ND	------	ND	------
------	Grain, from the processor prior to AGF generation	N/A	T1	139-0003A1	ND	ND	ND	ND	ND	ND	ND	ND
40278-9	Grain, from the processor prior to AGF generation	79	T4	139-0004A1	ND	------	ND	------	ND	------	ND	------
40278-9	Grain, from the processor prior to AGF generation	79	T4	139-0005A1	ND	------	ND	------	ND	------	ND	------
40278-9	Grain, from the processor prior to AGF generation	79	T4	139-0006A1	ND	ND	ND	ND	ND	ND	ND	ND
------	Grain, from the processor after AGF generation	N/A	T1	140-0001A1	ND	------	ND	------	ND	------	ND	------

AAD-1 Event[a] DAS-	Commodity	PHI[b] (days)	Plot ID	Sample ID ARA-09-15-10-	Quizalofop acid (µg/g)		Quizalofop acid (as quizalofop-ethyl equivalents)[c]		Quizalofop-ethyl (µg/g)		Total Residue[d] (µg/g)	
						Avg[e]		Avg[e]		Avg[e]		Avg[e]
------	Grain, from the processor after AGF generation	N/A	T1	140-0002A1	ND	------	ND	------	ND	------	ND	------
------	Grain, from the processor after AGF generation	N/A	T1	140-0003A1	ND	ND	ND	ND	ND	ND	ND	ND
40278-9	Grain, from the processor after AGF generation	79	T4	140-0004A1	ND	------	ND	------	ND	------	ND	------
40278-9	Grain, from the processor after AGF generation	79	T4	140-0005A1	ND	------	ND	------	ND	------	ND	------
40278-9	Grain, from the processor after AGF generation	79	T4	140-0006A1	ND	ND	ND	ND	ND	ND	ND	ND
------	Starch	N/A	T1	142-0001A9	ND	------	ND	------	ND	------	ND	------
------	Starch	N/A	T1	142-0002A1	ND	ND	ND	ND	ND	ND	ND	ND
40278-9	Starch	N/A	T4	142-0003A1	ND	------	ND	------	ND	------	ND	------
40278-9	Starch	N/A	T4	142-0004A1	ND	ND	ND	ND	ND	ND	ND	ND
------	Oil, refined (wet milling)	N/A	T1	143-0001A9	ND	------	ND	------	ND	------	ND	------
------	Oil, refined (wet milling)	N/A	T1	143-0002A1	ND	ND	ND	ND	[0.003]	ND	ND	ND
40278-9	Oil, refined (wet milling)	N/A	T4	143-0003A1	ND	------	ND	------	ND	------	ND	------

AAD-1 Event[a] DAS-	Commodity	PHI[b] (days)	Plot ID	Sample ID ARA-09-15-10-	Quizalofop acid (µg/g)		Quizalofop acid (as quizalofop-ethyl equivalents)[c]		Quizalofop-ethyl (µg/g)		Total Residue[d] (µg/g)	
						Avg[e]		Avg[e]		Avg[e]		Avg[e]
40278-9	Oil, refined (wet milling)	N/A	T4	143-0004A1	ND	ND	ND	ND	ND	ND	ND	ND
------	Oil, refined (dry milling)	N/A	T1	147-0001A1	ND	------	ND	------	ND	------	ND	------
------	Oil, refined (dry milling)	N/A	T1	147-0001A9	ND	ND	ND	ND	ND	ND	ND	ND
40278-9	Oil, refined (dry milling)	N/A	T4	147-0003A1	ND	------	ND	------	ND	------	ND	------
40278-9	Oil, refined (dry milling)	N/A	T4	147-0004A1	ND	ND	ND	ND	ND	ND	ND	ND
------	Meal	N/A	T1	144-0001A1	ND	------	ND	------	ND	------	ND	------
------	Meal	N/A	T1	144-0001A9	ND	ND	ND	ND	ND	ND	ND	ND
40278-9	Meal	N/A	T4	144-0003A1	ND	------	ND	------	ND	------	ND	------
40278-9	Meal	N/A	T4	144-0004A1	ND	ND	ND	ND	ND	ND	ND	ND
------	Grits	N/A	T1	145-0001A9	ND	------	ND	------	ND	------	ND	------
------	Grits	N/A	T1	145-0002A1	ND	ND	ND	ND	ND	ND	ND	ND
40278-9	Grits	N/A	T4	145-0003A1	ND	------	ND	------	ND	------	ND	------
40278-9	Grits	N/A	T4	145-0004A1	ND	ND	ND	ND	ND	ND	ND	ND
------	Flour	N/A	T1	146-0001A1	ND	------	ND	------	ND	------	ND	------
------	Flour	N/A	T1	146-0002A1	ND	ND	ND	ND	ND	ND	ND	ND
40278-9	Flour	N/A	T4	146-0003A1	ND	------	ND	------	ND	------	ND	------

AAD-1 Event[a] DAS-	Commodity	PHI[b] (days)	Plot ID	Sample ID ARA-09-15-10-	Quizalofop acid (µg/g)		Quizalofop acid (as quizalofop-ethyl equivalents)[c]		Quizalofop-ethyl (µg/g)		Total Residue[d] (µg/g)	
						Avg[e]		Avg[e]		Avg[e]		Avg[e]
40278-9	Flour	N/A	T4	146-0004A1	ND	ND	ND	ND	ND	ND	ND	ND
------	Aspirated grain fractions	N/A	T1	141-0001A1	ND	------	ND	------	ND	------	ND	------
40278-9	Aspirated grain fractions	N/A	T4	141-0002A1	ND	------	ND	------	ND	------	ND	------

[a] Untreated control plots (T1) were planted with the commercial variety Mycogen 2M746, which lacked the inserted AAD-1 gene, but had a genetic background similar to that used for the two non-commercial transgenic hybrids in treated plots (T2).

[b] PHI = Pre Harvest Interval (actual days after the last application of quizalofop-P-ethyl)

[c] The quizalofop acid residue values are converted to quizalofop-ethyl equivalents based on the molecular weight of the two compounds. Quizalofop-ethyl has a molecular weight of 372.8 and quizalofop acid has a molecular weight of 344.8. The factor to use to convert quizalofop acid to quizalofop-ethyl equivalents is calculated by dividing the molecular weight of quizalofop-ethyl by the molecular weight of quizalofop acid, which calculates to 1.08 (372.8/344.8 = 1.08). Therefore, the residue values for quizalofop acid are multiplied by 1.08 to convert them to quizalofop-ethyl equivalents.

[d] Total residue = sum of quizalofop acid (expressed as quizalofop-ethyl equivalents) and quizalofop-ethyl.

[e] Averages presented in this column are of treated samplings (T2 samples) only at each trial's sampling event.

[f] ND = Not Detected (below the limit of detection of 0.003 µg/g)

[g] [ ] = Residue values greater than or equal to the limit of detection of 0.003 µg/g but less than the limit of quantitation of 0.01 µg/g. Values are reported with a lower degree of confidence than values above the limit of quantitation.

[h] For the purposes of calculating an average, "ND" is given the value of zero.

TABLE XVII –

Residues of Quizalofop Acid and Quizalofop-ethyl in Field Corn Processed Commodities (Trial NE-14, NAFTA Region 5)

AAD-1 Event[a] DAS-	Commodity	PHI[b] (days)	Plot ID	Sample ID ARA-09-15-10-	Quizalofop acid (µg/g)		Quizalofop acid (as quizalofop-ethyl equivalents)[c]		Quizalofop-ethyl (µg/g)		Total Residue[d] (µg/g)	
						Avg[e]		Avg[e]		Avg[e]		Avg[e]
------	Grain, from the field	N/A	T1	080-0001A1	ND[f]	------	ND	------	ND	------	ND	------
40278-9	Grain, from the field	140	T4	080-0006A1	ND	------	ND	------	ND	------	ND	------
40278-9	Grain, from the field	140	T4	080-0007A1	ND	ND	ND	ND	ND	ND	ND	ND
------	Grain, from the processor prior to AGF generation	N/A	T1	148-0001A1	ND	------	ND	------	ND	------	ND	------
------	Grain, from the processor prior to AGF generation	N/A	T1	148-0002A1	ND	------	ND	------	ND	------	ND	------
------	Grain, from the processor prior to AGF generation	N/A	T1	148-0003A1	ND	ND	ND	ND	ND	ND	ND	ND
40278-9	Grain, from the processor prior to AGF generation	140	T4	148-0004A1	ND	------	ND	------	ND	------	ND	------
40278-9	Grain, from the processor prior to AGF generation	140	T4	148-0005A1	[0.005][g]	------	[0.005]	------	[0.004]	------	[0.009]	------
40278-9	Grain, from the processor prior to AGF generation	140	T4	148-0006A1	ND	ND[h]	ND	ND	ND	ND	ND	[0.003]
------	Grain, from the processor after AGF generation	N/A	T1	149-0001A1	ND	------	ND	------	ND	------	ND	------

AAD-1 Event[a] DAS-	Commodity	PHI[b] (days)	Plot ID	Sample ID ARA-09-15-10-	Quizalofop acid (µg/g)		Quizalofop acid (as quizalofop-ethyl equivalents)[c]		Quizalofop-ethyl (µg/g)		Total Residue[d] (µg/g)	
						Avg[e]		Avg[e]		Avg[e]		Avg[e]
------	Grain, from the processor after AGF generation	N/A	T1	149-0002A1	ND	------	ND	------	ND	------	ND	------
------	Grain, from the processor after AGF generation	N/A	T1	149-0003A1	ND	ND	ND	ND	ND	ND	ND	ND
40278-9	Grain, from the processor after AGF generation	140	T4	149-0004A1	ND	------	ND	------	ND	------	ND	------
40278-9	Grain, from the processor after AGF generation	140	T4	149-0005A1	ND	------	ND	------	ND	------	ND	------
40278-9	Grain, from the processor after AGF generation	140	T4	149-0006A1	ND	ND	ND	ND	ND	ND	ND	ND
------	Starch	N/A	T1	151-0001A1	ND	------	ND	------	ND	------	ND	------
------	Starch	N/A	T1	151-0002A1	ND	ND	ND	ND	ND	ND	ND	ND
40278-9	Starch	N/A	T4	151-0003A1	ND	------	ND	------	ND	------	ND	------
40278-9	Starch	N/A	T4	151-0004A1	ND	ND	ND	ND	[0.005][e]	ND	[0.005][e]	ND
------	Oil, refined (wet milling)	N/A	T1	152-0001A1	ND	------	ND	------	ND	------	ND	------
------	Oil, refined (wet milling)	N/A	T1	152-0002A1	ND	ND	ND	ND	ND	ND	ND	ND
40278-9	Oil, refined (wet milling)	N/A	T4	152-0003A1	ND	------	ND	------	ND	------	ND	------

AAD-1 Event[a] DAS-	Commodity	PHI[b] (days)	Plot ID	Sample ID ARA-09-15-10-	Quizalofop acid (µg/g)		Quizalofop acid (as quizalofop-ethyl equivalents)[c]		Quizalofop-ethyl (µg/g)		Total Residue[d] (µg/g)	
						Avg[e]		Avg[e]		Avg[e]		Avg[e]
40278-9	Oil, refined (wet milling)	N/A	T4	152-0004A1	ND	ND	ND	ND	ND	ND	ND	ND
------	Oil, refined (dry milling)	N/A	T1	156-0001A1	ND	------	ND	------	ND	------	ND	------
------	Oil, refined (dry milling)	N/A	T1	156-0002A1	ND	ND	ND	ND	[0.006]	[0.003]	[0.006]	[0.003]
40278-9	Oil, refined (dry milling)	N/A	T4	156-0003A1	ND	------	ND	------	ND	------	ND	------
40278-9	Oil, refined (dry milling)	N/A	T4	156-0004A1	ND	ND	ND	ND	ND	ND	ND	ND
------	Meal	N/A	T1	153-0001A1	ND	------	ND	------	ND	------	ND	------
------	Meal	N/A	T1	153-0002A1	ND	ND	ND	ND	[0.003]	ND	[0.003]	ND
40278-9	Meal	N/A	T4	153-0003A1	ND	------	ND	------	ND	------	ND	------
40278-9	Meal	N/A	T4	153-0004A1	ND	ND	ND	ND	ND	ND	ND	ND
------	Grits	N/A	T1	154-0001A1	ND	------	ND	------	ND	------	ND	------
------	Grits	N/A	T1	154-0002A1	ND	ND	ND	ND	ND	ND	ND	ND
40278-9	Grits	N/A	T4	154-0003A1	ND	------	ND	------	ND	------	ND	------
40278-9	Grits	N/A	T4	154-0004A1	ND	ND	ND	ND	ND	ND	ND	ND
------	Flour	N/A	T1	155-0001A9	ND	------	ND	------	ND	------	ND	------
------	Flour	N/A	T1	155-0002A1	ND	ND	ND	ND	ND	ND	ND	ND

AAD-1 Event[a] DAS-	Commodity	PHI[b] (days)	Plot ID	Sample ID ARA-09-15-10-	Quizalofop acid (µg/g)		Quizalofop acid (as quizalofop-ethyl equivalents)[c]		Quizalofop-ethyl (µg/g)		Total Residue[d] (µg/g)	
						Avg[e]		Avg[e]		Avg[e]		Avg[e]
40278-9	Flour	N/A	T4	155-0003A1	ND	------	ND	------	ND	------	ND	------
40278-9	Flour	N/A	T4	155-0004A1	ND	ND	ND	ND	ND	ND	ND	ND
------	Aspirated grain fractions	N/A	T1	150-0001A1	ND	------	ND	------	ND	------	ND	------
40278-9	Aspirated grain fractions	N/A	T4	150-0002A1	ND	------	ND	------	ND	------	ND	------

[a] Untreated control plots (T1) were planted with the commercial variety Mycogen 2M746, which lacked the inserted AAD-1 gene, but had a genetic background similar to that used for the two non-commercial transgenic hybrids in treated plots (T2).

[b] PHI = Pre Harvest Interval (actual days after the last application of quizalofop-P-ethyl)

[c] The quizalofop acid residue values are converted to quizalofop-ethyl equivalents based on the molecular weight of the two compounds. Quizalofop-ethyl has a molecular weight of 372.8 and quizalofop acid has a molecular weight of 344.8. The factor to use to convert quizalofop acid to quizalofop-ethyl equivalents is calculated by dividing the molecular weight of quizalofop-ethyl by the molecular weight of quizalofop acid, which calculates to 1.08 (372.8/344.8 = 1.08). Therefore, the residue values for quizalofop acid are multiplied by 1.08 to convert them to quizalofop-ethyl equivalents.

[d] Total residue = sum of quizalofop acid (expressed as quizalofop-ethyl equivalents) and quizalofop-ethyl.

[e] Averages presented in this column are of treated samplings (T2 samples) only at each trial's sampling event.

[f] ND = Not Detected (below the limit of detection of 0.003 µg/g)

[g] [ ] = Residue values greater than or equal to the limit of detection of 0.003 µg/g but less than the limit of quantitation of 0.01 µg/g. Values are reported with a lower degree of confidence than values above the limit of quantitation.

[h] For the purposes of calculating an average, "ND" is given the value of zero.

TABLE XVIII – Summary of Residues of 2,4-D and 2,4-DCP in Field Corn Raw Agricultural Commodities

Analyte/ Commodity	Target Application Rate[a] lb ae/A (g ae/ha)	PHI[b] (days)	n	Residue Levels (ug/g)					
				Min.	Max.	Median[c]	Mean[c]	Std. Dev.[c]	HAFT[d]
2,4-D									
Early Forage	3 x 1.0 (~1120)	27-42	48	ND[e]	2.988	0.443	0.641	0.707	2.612
Forage	3 x 1.0 (~1120)	39-78	50	ND	4.373	0.252	0.697	1.008	3.124
Grain	3 x 1.0 (~1120)	73-139	48	ND	(0.004)[f/g]	ND	ND	NA[h]	(0.004)
Stover	3 x 1.0 (~1120)	73-139	50	ND	8.92	0.613	1.349	2.064	8.076
2,4-DCP									
Early Forage	3 x 1.0 (~1120)	27-42	48	ND	3.997	0.773	1.032	1.003	3.056
Forage	3 x 1.0 (~1120)	39-78	50	(0.004)	5.007	0.448	0.876	1.154	3.899
Grain	3 x 1.0 (~1120)	73-139	48	ND	(0.003)[f]	ND	ND	NA	ND
Stover	3 x 1.0 (~1120)	73-139	50	ND	5.842	0.453	0.911	1.224	5.377

[a] 2,4-D was applied as the first, second and fourth of four applications, quizalofop-P-ethyl was applied as the third application.

[b] PHI = Pre-Harvest Interval; days between last application of, 2,4-D and collection of field sample.

[c] For statistical purposes, ND has been given the value of zero.

[d] HAFT = Highest Average Field Trial

[e] ND = Not detected; less than the LOD (<0.003 µg/g)

[f] Values for 2,4-D in the replicate samples of grain from trial MO-08 were not included in this summary because one of two replicate samples is not believed to be representative of residues from the application pattern in this study and may have resulted from contamination. The MO-08 residue values of 2,4-D in replicate samples in grain from trial MO-08 are as follows: 0.067 ug/g, ND. The 2,4-DCP residues in the replicate samples from this site were both ND. For consistency, neither of the replicate values for 2,4-D or 2,4-DCP in grain from this trial site were included in the summary.

[g] Values in parentheses are greater than or equal to the limit of detection (0.003 ug/g) by less than the limit of quantitation (0.01 ug/g). These values are reported with a lower degree of confidence than values above the limit of quantitation.

[h] NA = Not applicable

TABLE XIX - Summary of Residues of Quizalofop Acid and Quizalofop-ethyl in Field Corn Raw Agricultural Commodities

Analyte/ Commodity	Target Application Rate[a] lb ai/A (g ai/ha)	PHI[b] (days)	n	Residue Levels (ug/g) Min.	Max.	Median[c]	Mean[c]	Std. Dev.[c]	HAFT[d]
Quizalofop-ethyl									
Early Forage	0.082 (92)	32-47	48	ND[e]	ND	ND	ND	NA[f]	ND
Forage	0.082 (92)	44-83	48	ND	ND[g]	ND	ND	NA	ND
Grain	0.082 (92)	79-144	50	ND	ND	ND	ND	NA	ND
Stover	0.082 (92)	79-144	50	ND	(0.004)[h]	ND	ND	NA	ND
Quizalofop acid									
Early Forage	0.082 (92)	32-47	48	ND	ND	ND	ND	NA	ND
Forage	0.082 (92)	44-83	48	ND	ND [h]	ND	ND	NA	ND
Grain	0.082 (92)	79-144	50	ND	(0.004)	ND	ND	NA	ND
Stover	0.082 (92)	79-144	50	ND	0.011	ND	ND	NA	0.011
Combined Residues of quizalofop-ethyl and quizalofop acid (expressed as quizalofop-ethyl)[i]									
Early Forage	0.082 (92)	32-47	48	ND	ND	ND	ND	NA	ND
Forage	0.082 (92)	44-83	48	ND	ND	ND	ND	NA	ND
Grain	0.082 (92)	79-144	50	ND	(0.004)	ND	ND	NA	ND
Stover	0.082 (92)	79-144	50	ND	0.016	ND	ND	NA	0.016

[a] Quizalofop-P-ethyl was applied as the third application only, and 2,4-D was applied as the first, second and fourth of four applications.

[b] PHI = Pre-Harvest Interval; days between the single application of quizalofop-P-ethyl and the collection of field samples.

[c] For statistical purposes, ND has been given the value of zero.

[d] HAFT = Highest Average Field Trial

[e] ND = Not detected; less than the LOD (<0.003 µg/g)

[f] NA = Not applicable

[g] Values for the replicate samples for forage / typical forage from trial TX-23 were not included in this summary because they are not believed to be representative of residues from the application pattern in this study and may have resulted from contamination. The TX-23 residue values for replicate samples in forage are as follows: quizalofop-ethyl: 0.197 ug/g and 0.099 ug/g; quizalofop acid: 0.167 ug/g and 0.113 ug/g

[h] Values displayed in brackets or parenthesis indicate a concentration < LOQ (0.010 µg/g) and≥ LOD (0.003 µg/g) . These values are reported with a lower degree of confidence than values above the limit of quantitation.

[i] Quizalofop acid residue values are converted to quizalofop-ethyl equivalents based on the molecular weight of the two compounds. Quizalofop-ethyl has a molecular weight of 372.8 and quizalofop acid has a molecular weight of 344.8. The factor to use to convert quizalofop acid to quizalofop-ethyl equivalents is calculated by dividing the molecular weight of quizalofop-ethyl by the molecular weight of quizalofop acid, which calculates to 1.08 (372.8/344.8 = 1.08). Therefore, the residue values for quizalofop acid are multiplied by 1.08 to convert them to quizalofop-ethyl equivalents. Once this is done then the values for quizalofop-ethyl and quizalofop acid expressed as quizalofop-ethyl equivalents are summed to provide the "combined" value.

TABLE XX - Summary of Residues in Field Corn Processed Commodities

Field Corn Processed Fraction	**Average Residues (µg/g)**					
	2,4-D	2-4-DCP	Combined Quizalofop Residues[a]	2,4-D Processing factor	2,4-DCP Processing factor	Quizalofop Processing factor
Trial GA-03						
Grain[b]	ND[c]	ND	ND	NA[d]	NA	NA
Starch	ND	ND	ND	NCF[e]	NCF	NCF
Oil, refined (wet milling)	ND	ND	ND	NCF	NCF	NCF
Oil, refined (dry milling)	ND	ND	ND	NCF	NCF	NCF
Meal	ND	ND	ND	NCF	NCF	NCF
Grits	ND	ND	ND	NCF	NCF	NCF
Flour	ND	ND	ND	NCF	NCF	NCF
AGF	0.021	[0.006][f]	ND	NCF[g]	NCF	NCF
Trial NE-14						
Grain[a]	ND	[0.008]	[0.003]	NA	NA	NA
Starch	ND	ND	ND	NCF	NCF	NCF
Oil, refined (wet milling)	ND	ND	ND	NCF	NCF	NCF
Oil, refined (dry milling)	ND	ND	ND	NCF	NCF	NCF
Meal	ND	ND	ND	NCF	NCF	NCF
Grits	ND	ND	ND	NCF	NCF	NCF
Flour	ND	ND	ND	NCF	NCF	NCF
AGF	ND	ND	ND	NCF	NCF	NCF

[1] Quizalofop acid residue values are converted to quizalofop-ethyl equivalents based on the molecular weight of the two compounds. The factor to use to convert quizalofop acid to quizalofop-ethyl equivalents is calculated by dividing the molecular weight of quizalofop-ethyl by the molecular weight of quizalofop acid, which calculates to 1.08 (372.8/344.8 = 1.08). Therefore, the residue values for quizalofop acid are multiplied by 1.08 to convert them to quizalofop-ethyl equivalents. Once this is done then the values for quizalofop-ethyl and quizalofop acid expressed as quizalofop-ethyl equivalents are summed to provide the "combined" value.

[b] Grain residue values used for calculating processing factors are the average values of the grain samples collected at the processor prior to separation of Aspirated Grain Fractions.

[c] ND = Not detected; less than the LOD (<0.003 µg/g)

[d] NA = Not applicable

[e] NCF = No Concentration Factor

[f] Values displayed in brackets or parenthesis indicate a concentration < LOQ (0.010 µg/g) and ≥ LOD (0.003 µg/g). These values are reported with a lower degree of confidence than values above the limit of quantitation.

[g] Since no residue was detected in the grain, no processing factor is calculated even though residues were found in the aspirated grain fractions.

Figures

Figure 1: Weedar 64 - Certificate of Analysis

REPORT **FA&PC NUMBER** 09-205129

CERTIFICATE OF ANALYSIS FOR TEST/REFERENCE/CONTROL SUBSTANCES

TITLE OBJECTIVE: Determination of purity and/or identity of the following test/reference/control substance for use in a study.

	TEST/REFERENCE/CONTROL SUBSTANCE:		
TEST SUBSTANCE NO:	TSN026491-0010		
LOT NO:	ILG-C-004		
DESCRIPTION:	Weedar 64 Broadleaf Herbicide		
	End Use Product		
REFERENCE SUBSTANCE USED:	2,4-D, AGR275828	PURITY:	99.5%

INITIATION DATE: 6 February 2009

METHODS USED:

PURITY:	IDENTIFICATION:
HPLC, density	Retention time

RESULTS and CONCLUSIONS:

X **INITIAL DETERMINATION:**
47.1%w/w (546g/L) 2,4-Dichlorophenoxyacetic acid, dimethylamine salt (a.i.)
39.1%w/w (454g/L) 2,4-Dichlorophenoxyacetic acid (a.e.)

X **IDENTITY:**
The retention time of the sample component was consistent with the retention time of the analytical standard.

X **OTHER:**
density = 1.1601 g/mL at 20°C

RE-CERTIFICATION DATE: 12 February, 2011

CALCULATIONS:

Area Normalized: N.A. Internal Standard: N.A. External Standard: X
Other (explain): g/L = Wt% x density x 10
AI = AE /AE factor (AE factor = 0.830)

STUDY DIRECTOR SIGNATURE: Rose M. Nelson
Rose M. Nelson

STUDY COMPLETION DATE: February 19, 2009

PEER REVIEWER SIGNATURE: Cynthia Crouse

DATE: 19-February-2009

TESTING FACILITY ADDRESS:
Dow AgroSciences LLC
Crop Protection R&D Analytical/Product Chemistry Center of Expertise
9330 Zionsville Road
Indianapolis, Indiana 46268

All raw data and retainer samples associated with this study will be archived in the testing facility archive. Only descriptive statistics were used unless otherwise noted in the results. This study was conducted in accordance with the Good Laboratory Practice Standard, 40 CFR Part 160.135 (b).

Figure 2: Assure II - Certificate of Analysis

REPORT **FA&PC NUMBER** 09-205013

CERTIFICATE OF ANALYSIS FOR TEST/REFERENCE/CONTROL SUBSTANCES

TITLE OBJECTIVE: Determination of purity and/or identity of the following test/reference/control substance for use in a study.

	TEST/REFERENCE/CONTROL SUBSTANCE:		
TEST SUBSTANCE NO:	TSN020252-0004		
LOT NO:	APR06YZ059		
DESCRIPTION:	ASSURE II		
	End Use Product		
REFERENCE SUBSTANCE USED:	Quizalofop P-ethyl, TSN106169	PURITY:	98%

INITIATION DATE: 6 February 2009

METHODS USED:

PURITY: GC, density

IDENTIFICATION: Retention time

RESULTS and CONCLUSIONS:

X **INITIAL DETERMINATION:**
10.3%w/w (105g/L) quizalofop P-ethyl

X **IDENTITY:**
The retention time of the sample component was consistent with the retention time of the analytical standard.

X **OTHER:**
density = 1.0223 g/mL at 20°C

RE-CERTIFICATION DATE: 05 March, 2011

CALCULATIONS:

Area Normalized: N.A. Internal Standard: N.A. External Standard: X
Other (explain): g/L = Wt% x density x 10

STUDY DIRECTOR SIGNATURE: Rose M. Nelson
Rose M. Nelson

STUDY COMPLETION DATE: March 16, 2009

PEER REVIEWER SIGNATURE: Beth Koors

DATE: March 16, 2009

TESTING FACILITY ADDRESS:
Dow AgroSciences LLC
Crop Protection R&D Analytical/Product Chemistry Center of Expertise
9330 Zionsville Road
Indianapolis, Indiana 46268

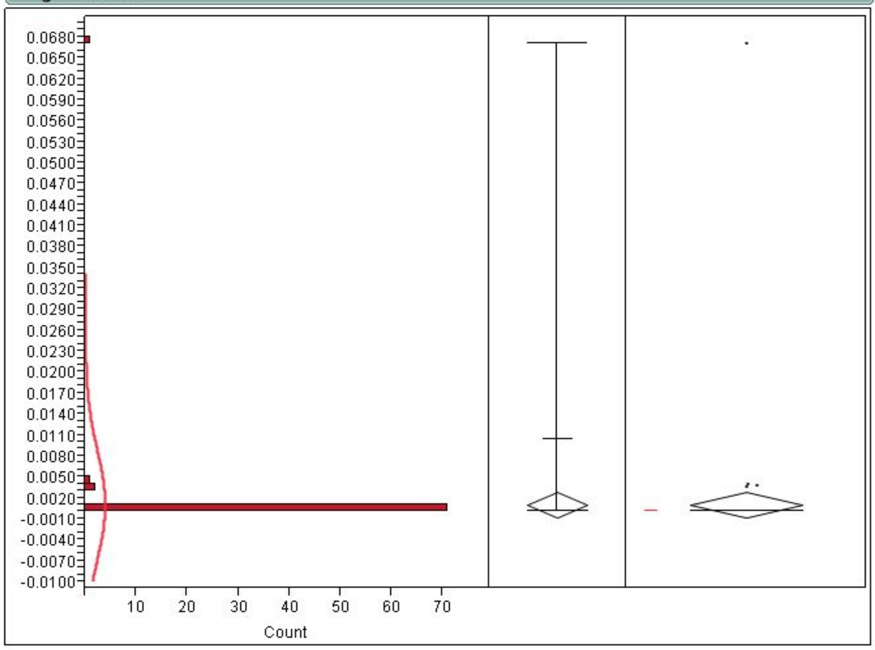
All raw data and retainer samples associated with this study will be archived in the testing facility archive. Only descriptive statistics were used unless otherwise noted in the results. This study was conducted in accordance with the Good Laboratory Practice Standard, 40 CFR Part 160.135 (b), except that the quizalofop standard was obtained from a commercial supplier and the GLP status is unknown.

ARA-09-15-10
MO-18 DB 1/12/10

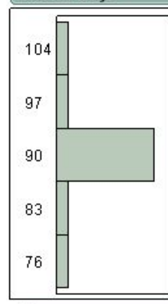
Figure 3: Distribution of 2,4-D Residue Results in Field Corn Grain

Distributions

single residue



event days



Normal(0.00105,0.00777)

Frequencies

Level	Count	Prob
76	6	0.08000
83	6	0.08000
90	51	0.68000
97	6	0.08000
104	6	0.08000
Total	75	1.00000

N Missing 0
5 Levels

Quantiles

100.0%	maximum	0.0671
99.5%		0.0671
97.5%		0.01031
90.0%		0
75.0%	quartile	0
50.0%	median	0
25.0%	quartile	0
10.0%		0
2.5%		0
0.5%		0
0.0%	minimum	0

Moments

Mean	0.0010453
Std Dev	0.0077661
Std Err Mean	0.0008968
Upper 95% Mean	0.0028321
Lower 95% Mean	-0.000741
N	75

Fitted Normal

Parameter Estimates

Type	Parameter	Estimate	Lower 95%	Upper 95%
Location	μ	0.0010453	-0.000741	0.0028321
Dispersion	σ	0.0077661	0.0066914	0.0092552

-2log(Likelihood) = -516.857379958465

Figure 4 - Decline of Forage and Stover Residues of 2,4-D and 2,4-DCP from Trial GA-04

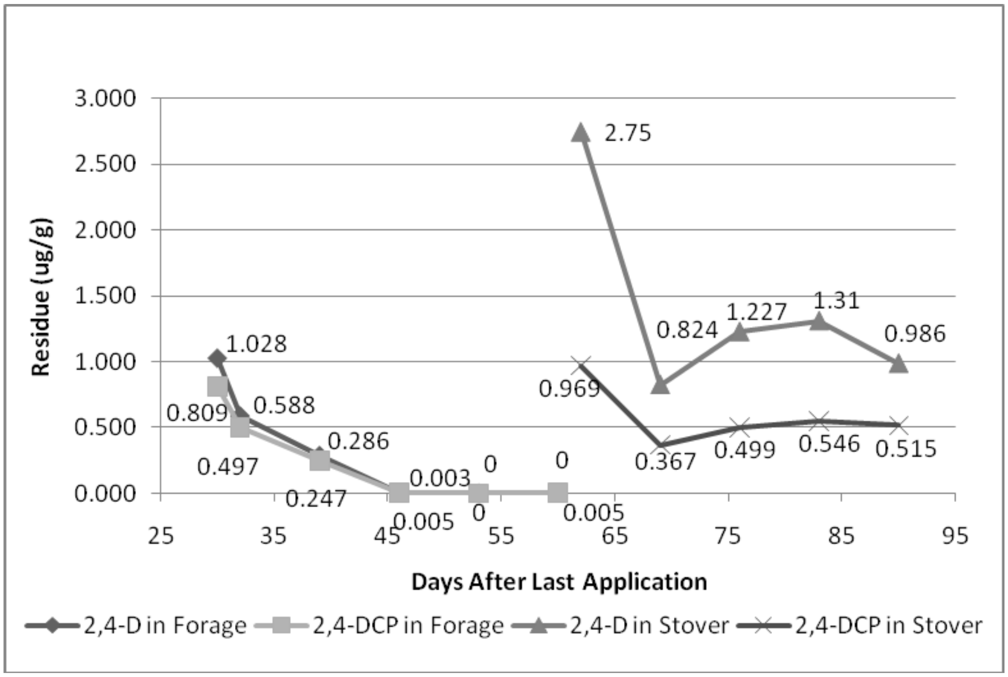


Figure 5 - Decline of Forage and Stover Residues of 2,4-D and 2,4-DCP from Trial MI-11

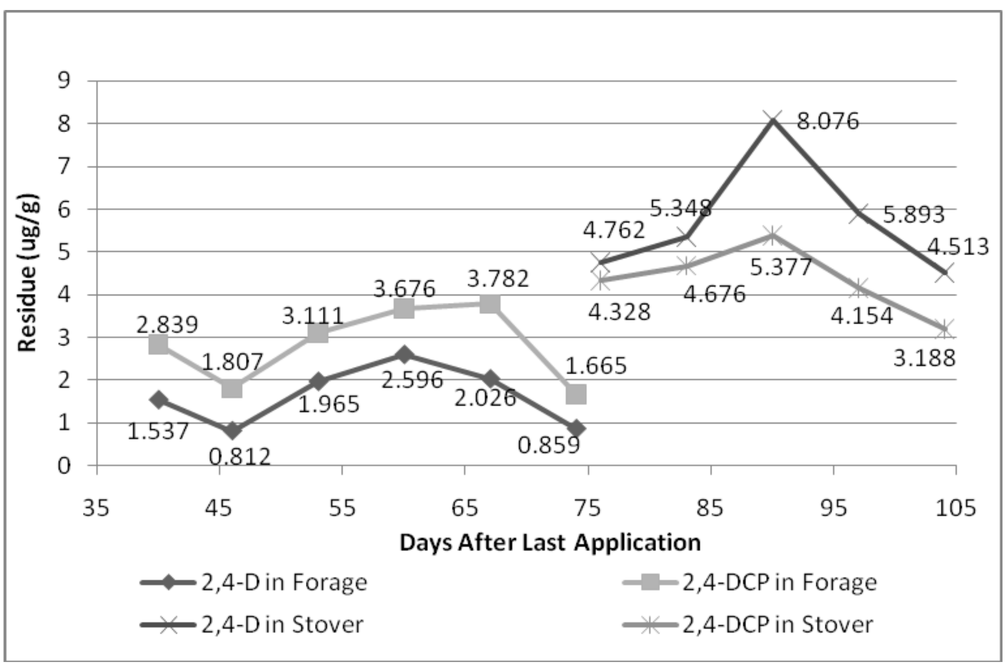
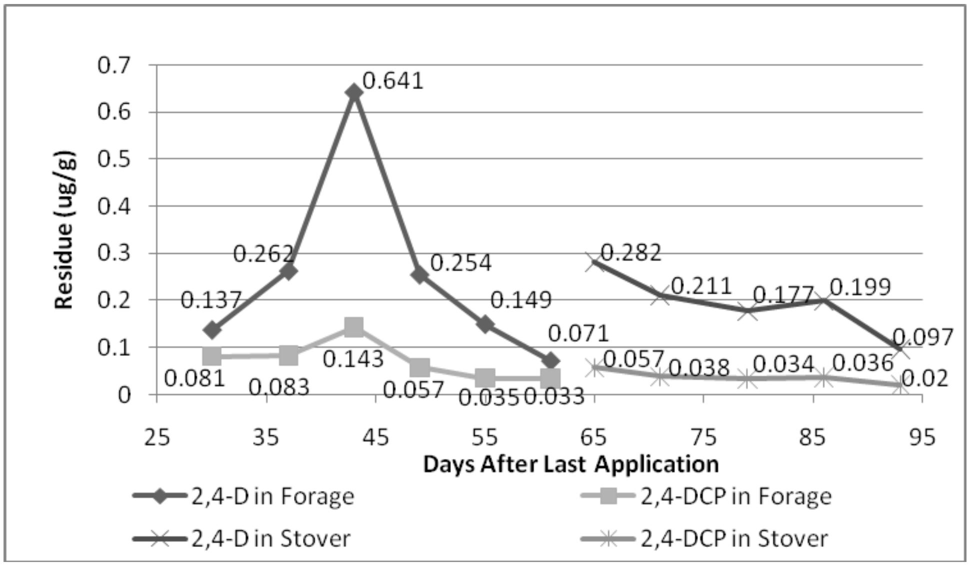


Figure 6 - Decline of Forage and Stover Residues of 2,4-D and 2,4-DCP from Trial OK-22



APPENDICES

Appendix A – Field Phase Summary

FIELD PHASE SUMMARY

Study Title:
Magnitude of the Residue of 2,4-D and Quizalofop-P-ethyl in/on Herbicide Tolerant Field Corn Containing the Aryloxyalkanoate Dioxygenase-1 (ADD-1) Gene

Study Number
ARA-09-15-10

Authors
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Sponsor
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Indianapolis, Indiana 46268

DAS Project Number
090052

1. TEST SUBSTANCE

The test substances for this study were Weedar 64 Broadleaf Herbicide and Assure II. Weedar 64 is a soluble concentrate (SL) formulation containing active ingredient (2,4-dichlorophenoxyacetic acid dimethylamine salt or 2,4-D dimethylamine (DMA) salt) at 454 g ae (acid equivalent)/L. Assure II is an emulsifiable concentrate (EC) formulation containing active ingredient (ethyl(R)-2-[4-(6-chloroquinoxalin-2-yl-oxy)-phenoxy] propionate or Quizalofop-P-ethyl) at 105 g ai/L. Both test substances are commercially available products. The test substances were applied in this study as a broadcast application four times during the season to corn plants. The first two and fourth applications were 2,4-D and the third application was quizalofop-P-ethyl (See Tables 1 and 2).

Table 1: Test Substance Identification Summary (2, 4-D)

Test Substance Common Name	2,4-D
Trade Name:	Weedar 64 Broadleaf Herbicide
Formulation Batch Number:	TSN026491-0010
Lot Number:	ILG-C-004
Tested Formulation and Type:	SL (Soluble Liquid Concentrate)
Nominal Concentration	454 g AE/L (acid equivalent)
GLP Certified Active Ingredient:[1]	2,4-Dichlorophenoxyacetic acid, dimethylamine salt
Density	1.1601 g/ml at 20°C
Date of Initial Analysis:	06 February 2009
Date of Re-analysis:	12 February 2011
Expiration Date:	12 February 2011

1. Actual assay results by Certificate of Analysis performed by the Sponsor

Table 2: Test Substance Identification Summary (Quizalofop-P-ethyl)

Test Substance Common Name	Quizalofop-P-ethyl
Trade Name:	Assure II Herbicide
Formulation Batch Number:	TSN020252-0004
Lot Number:	APR06YZ059
Tested Formulation and Type:	EC (Emulsifiable Concentrate)
Nominal Concentration	105 g/L (a.i., active ingredient)
GLP Certified Active Ingredient:[1]	Ethyl(R)-2-[4-(6-chloroquinoxalin-2-yl oxy)-phenoxyl propionate
Density	1.0223 g.ml at 20°C
Date of Initial Analysis:	06 February 2009
Date of Re-analysis:	05 March 2011
Expiration Date:	05 March 2011

1. Actual assay results by Certificate of Analysis performed by the Sponsor.

2. FIELD FACILITIES AND PERSONNEL

2.1. Field Trial Conduct and Management

The purpose of this study was to determine the magnitude of residues of 2,4-D and quizalofop-P-ethyl herbicides in/on field corn raw agricultural commodities (forage, grain, and stover) and processed fractions (aspirated grain fractions, starch, meal, grits, flour, refined oil from wet and dry process) when a transgenic hybrid corn line containing the inserted AAD-1 gene, a genetic modification added to increase tolerance to certain herbicides is treated with commercially available 2,4-D and quizalofop-P-ethyl herbicides.

Field corn was selected as the test system to establish the level of residues in a commercially viable transgenic herbicide tolerant line of field corn in order to meet requirements in OPPTS 860.1500. Trial locations were in NAFTA regions 1, 2, 5 (United States and Canada), and 6 which are major regions where field corn is commonly grown. The field corn in these trials was grown using cultural practices typical of those used in the region to produce a commercial crop.

The field data and crop samples generated were based on the requirements of the USEPA Pesticide Assessment Guideline OPPTS 860.1500 (Reference 1), OPPTS 860.1520 Processed Feed/Food (Reference 2), and Canadian Pesticide Management Regulatory Agency Directive 98-02, Section 9 (Reference 3). This study was conducted according to USEPA FIFRA Good Laboratory Practice Standards, Title 40 Code of Federal Regulations Part 160 (Reference 4) with exceptions noted on the GLP Compliance page of this report.

The overall field phase was managed by Ag Research Associates, LLC (ARA), including Study Director, Study Director Management, and Lead Quality Assurance. Twenty-five field trials were conducted in/on field corn. All field trials were conducted under the direction of the ARA Study Director. See Table 3 for PFIs responsible for the individual field trials.

2.2. Field Site Locations

The Sponsor and Study Director identified appropriate test sites in the Regions and States where field corn is commonly grown. Principal Field Investigators (PFI) were identified for each trial location (Table 3) and the PFI identified a suitable trial site (Table 4). The test sites were chosen to be relatively uniform in soil characteristics, were well-drained, and had proper levels of nutrients. The trials received agronomic management typical for the location to produce a commercial crop (such as fertilizer, pesticides, and irrigation) as needed.

The treated and control plots at each trial location were clearly identified with a unique trial number and treatment designation. Test sites were not located where 2,4-D or quizalofop-P-ethyl had been applied within the previous 18 months from the planting date. Test sites were located to avoid any inadvertent contamination to/from an adjacent agricultural field, residential, or commercial area. Buffer zones were required to avoid cross contamination due to spray drift or run-off. A 3-year pesticide use history for each trial site was established to avoid and identify possible contamination during residue analyses.

Table 3: Principal Field Investigators (PFIs) and Field Testing Facilities

Trial #	PFI	Company	Facility Address
NJ-01	Daniel Ramsdell	Crop Management Strategies	6273 Mountain Road Germansville, PA 18053
PA-02	Daniel Ramsdell	Crop Management Strategies	6273 Mountain Road Germansville, PA 18053
GA-03	Chris Cromer	Ag Research Associates	1730 Denham Road, Sycamore, GA 31790
GA-04	Chris Cromer	Ag Research Associates	1730 Denham Road, Sycamore, GA 31790
IL-05	Sue Dorsey	SGS Alvey Ag Research	Rt 2 Box 12 Wyoming, IL 61491
IL-06	Tim Boeker	SGS Alvey Ag Research	19300 Marydale RD Carlyle, IL 62231
IN-07	John Bailey	SGS Alvey Ag Research	1365 N Seip Rd. Rockville, IN 47872
MO-08	Matt Cordell	Ag Research Associates	1204 Fairview St. Lonoke, AR 72086
MO-09	Matt Cordell	Ag Research Associates	1204 Fairview St. Lonoke, AR 72086
IL-10	Matt Cordell	Ag Research Associates	1204 Fairview St. Lonoke, AR 72086
MI-11	Chad Harris	Ag Research Associates	2450 Hoagland Highway, Deerfield, MI 49238
OH-12	Chad Harris	Ag Research Associates	2450 Hoagland Highway, Deerfield, MI 49238
IN-13	Chad Harris	Ag Research Associates	2450 Hoagland Highway, Deerfield, MI 49238
NE-14	Matt Krause	Ag Research Associates	3605 N. Delaware Ave York, NE 68467
NE-15	Denny Stamm	Ag Research Associates	3605 N. Delaware Ave, York, NE 68467
KS-16	Denny Stamm	Ag Research Associates	3605 N. Delaware Ave, York, NE 68467
IA-17	Jason Niekamp	Bennett Ag Research	1109 Ivy Avenue, Richland, IA 52585
MO-18	David Bennett	Bennett Ag Research	1109 Ivy Avenue, Richland, IA 52585
IA-19	Kyle Johnson	Eurofins Agroscience Services	4614 HWY 63 Lime Springs, IA 52155
MN-20	Kyle Johnson	Eurofins Agroscience Services	4614 HWY 63 Lime Springs, IA 52155
MO-21	Nathan Goldschmidt	Shoffner Farm Research, Inc. - North Station	4809 Hwy FF Fisk, MO 63940
OK-22	Tim Case	Ag Research Associates	89 Broadway Avenue Groom, TX 79039
TX-23	Tim Case	Ag Research Associates	89 Broadway Avenue Groom, TX 79039
ON-24	Jamie Parnell	Vaughn Ag Research Services	1909 Maple Manor Rd. Branchton, ON N0B1L0
ON-25	Jamie Parnell	Vaughn Ag Research Services	1909 Maple Manor Rd. Branchton, ON N0B1L0

Table 4: Field Trial Site Locations

Trial #	Nearest Town	County	State	NAFTA Region
NJ-01	Baptistown	Hunterdon	NJ	1
PA-02	Germansville	Lehigh	PA	1
GA-03	Sycamore	Turner	GA	2
GA-04	Chula	Tift	GA	2
IL-05	Bradford	Bureau	IL	5
IL-06	Carlyle	Clinton	IL	5
IN-07	Rockville	Parke	IN	5
MO-08	Jasper	Jasper	MO	5
MO-09	Montrose	Henry	MO	5
IL-10	Rushville	Schuyler	IL	5
MI-11	Deerfield	Lenawee	MI	5
OH-12	Metamora	Fulton	OH	5
IN-13	Shipshewana	Lagrange	IN	5
NE-14	York	York	NE	5
NE-15	Stromsburg	Polk	NE	5
KS-16	Munden	Republic	KS	5
IA-17	Richland	Jefferson	IA	5
MO-18	La Plata	Adair	MO	5
IA-19	Spillville	Winneshiek	IA	5
MN-20	Cherry Grove	Filmore	MN	5
MO-21	Fisk	Stoddard	MO	5
OK-22	Madill	Marshall	OK	6
TX-23	Pilot Point	Denton	TX	6
ON-24	Branchton	Waterloo	Ontario	5
ON-25	Dutton	Elgin	Ontario	5

3. TEST SYSTEM, TRIAL LAYOUT, TRIAL MAINTENANCE

3.1 Test System

Field corn was selected as the test system to establish the level of residues in a commercially viable transgenic herbicide tolerant line of corn in order to meet the requirements in OPPTS 860.1500. Two transgenic hybrid lines of field corn each containing a different Event of the inserted AAD-1 gene were used in treated plots. One treated plot was planted with a hybrid containing AAD-1 Event DAS-40278-9 and a separate treated plot was planted with a hybrid containing AAD-1 Event DAS-40474-7. However, although the field trials were completed and samples were collected from plants with Event DAS-40278-9 and Event DAS-40474-7, the samples from plants containing Event DAS-40474-7 were not analyzed for residues due to a business decision by the Sponsor to discontinue the commercial development of this Event. Due to limited availability of seed with Event DAS-40278-9 or Event DAS-40474-7, the untreated plots were planted with the field corn variety Mycogen 2M746, which is a hybrid field corn having a genetic background similar to that used for the two transgenic hybrids, but lacking the inserted AAD-1 gene.

Trials were conducted in 25 locations: two in Region 1, two in Region 2, seventeen in Region 5 (United States), two in Canada Region 5, and two in Region 6 (United States). These regions are where field corn is commonly grown. The field corn grown in these regions represents typical cultural practices, climate, and environment where 2,4-D and quizalofop-P-ethyl are expected to be used.

Samples collected in this study involved the collection of raw agricultural commodities of field corn forage, field corn grain, and field corn stover.

At each of four sampling intervals for each RAC trial, one composite sample was collected from the untreated plot and two independently collected composite RAC samples were collected from each treated plot. At each of four sampling intervals for each RAC + processing trial, one composite sample was collected from the untreated plot and two independently collected composite RAC samples were collected from both treated plots 2 and 3. In addition, one bulk grain sample was collected from each of the control plot, treatment 4 plot and treatment 5 plot. At each of five sampling intervals for each decline + RAC trial, one composite sample was collected from the untreated plot and two independently collected composite RAC samples were collected from each treated plot. "Pre-forage" whole plants, (sample timing was intended to be earlier than "typical" with sample not expected to have yet reached the late dough growth stage) field corn forage, field corn grain, and field corn stover were collected. Typically, each sample weighed at least 1 kilogram and represented at least 12 plants from different areas within each plot.

Plants selected for sampling were selected randomly from within the plot. For the forage and stover samples, the composite sample contained a part (~1/3 of a stalk) of each of 12 collected plants and included 4 bottom segments, 4 middle segments, and 4 top segments with each segment coming from a different corn plant.

3.2. Field Trial Layout

Each RAC and decline + RAC trial consisted of one control plot and two treated plots. Each RAC + processing trial consisted of one control plot and four treated plots. The plot area was large enough to yield the required number of samples specified in the protocol and to allow the test substances to be applied accurately and in a manner that represents a commercial application technique. Plot size was determined by the PFI following guidelines in the protocol.

An accurate plot diagram(s) of the plot layout with dimensions was recorded in the field trial notebook. The treated and control plots were clearly identified with a unique number. Labeled stakes or other means were used to allow ease of location and prevent workers or equipment uses that were not under the direction of the PFI. The control plot was located to minimize the potential for contamination from the treated plot. There was an untreated buffer zone of at least 50 feet between the untreated and treated plots at each trial location and an untreated buffer zone of at least 25 feet between treated plots at each trial location. Test sites were located to avoid any inadvertent contamination from an adjacent agricultural field, residential, or commercial area. See Table 5 below for detailed plot information.

All test plots were maintained in accordance with prevailing commercial agronomic practices for field corn. All operations were performed to the control plot first to avoid potential contamination.

The plots were maintained relatively free of weeds, insects, and pathogens. Any applications of maintenance chemicals were made to all plots (control and treated). Fertilization and irrigation were used as needed to produce a crop typical of a commercial crop. All cultivation operations, irrigation, maintenance pesticide applications, and fertilization were documented in the field trial notebook.

Table 5: Trial Layout Summary

Trial #	Trt #	Plot Size (ft)	AAD-1 Event [a]	Planting Date	Soil Type
NJ-01	1	25 X 25	---	05 Jul 2009	Loam
	2	25 X 30	DAS-40278-9	05 Jul 2009	Loam
	3	25 X 30	DAS-40474-7	05 Jul 2009	Loam
PA-02	1	15 X 40	---	08 Jun 2009	Clay loam
	2	20 X 40	DAS-40278-9	08 Jun 2009	Clay loam
	3	20 X 40	DAS-40474-7	08 Jun 2009	Clay loam
GA-03	1 (UTC)	150 X 96	---	09 Jun 2009	Tifton Loamy Sand
	2 (TRT 1)	50 X 24	DAS-40278-9	09 Jun 2009	Tifton Loamy Sand
	3 (TRT 2)	50 X 24	DAS-40474-7	09 Jun 2009	Tifton Loamy Sand
	4 (TRT 1)	150 X 96	DAS-40278-9	09 Jun 2009	Tifton Loamy Sand
	5 (TRT 2)	150 X 96	DAS-40474-7	09 Jun 2009	Tifton Loamy Sand
GA-04	1 (UTC)	100 X 48	---	10 Jun 2009	Tifton Loamy Sand
	2 (TRT)	100 X 48	DAS-40278-9	10 Jun 2009	Tifton Loamy Sand
	3 (TRT)	100 X 48	DAS-40474-7	10 Jun 2009	Tifton Loamy Sand
IL-05	1	20 X 50	---	04 Jun 2009	Sable Silt Loam
	2	20 X 50	DAS-40278-9	04 Jun 2009	Sable Silt Loam
	3	20 X 50	DAS-40474-7	04 Jun 2009	Sable Silt Loam
IL-06	1	10 X 100	---	26 Jun 2009	Hoyleton Darmstadt silt loam
	2	10 X 100	DAS-40278-9	26 Jun 2009	Hoyleton Darmstadt silt loam
	3	10 X 100	DAS-40474-7	26 Jun 2009	Hoyleton Darmstadt silt loam
IN-07	1	20 X 50	---	08 Jun 2009	Reesville Silt Loam
	2	20 X 50	DAS-40278-9	08 Jun 2009	Reesville Silt Loam
	3	20 X 50	DAS-40474-7	08 Jun 2009	Reesville Silt Loam
MO-08	1	20 X 75	---	03 Jul 2009	Newtonia-Eldorodo silt loam
	2	20 X 75	DAS-40278-9	03 Jul 2009	Newtonia-Eldorodo silt loam
	3	20 X 75	DAS-40474-7	03 Jul 2009	Newtonia-Eldorodo silt loam

Table 5: Trial Layout Summary

Trial #	Trt #	Plot Size (ft)	AAD-1 Event [a]	Planting Date	Soil Type
MO-09	1	50 X 20	---	28 Jun 2009	Hartwell Silt Loam
	2	50 X 20	DAS-40278-9	28 Jun 2009	Hartwell Silt Loam
	3	50 X 20	DAS-40474-7	28 Jun 2009	Hartwell Silt Loam
IL-10	1	75 X 20	---	06 Jul 2009	Rozetta Silt Loam
	2	75 X 20	DAS-40278-9	06 Jul 2009	Rozetta Silt Loam
	3	75 X 20	DAS-40474-7	06 Jul 2009	Rozetta Silt Loam
MI-11	1	60 X 40	---	13 Jun 2009	Lenawee silty clay loam
	2	60 X 40	DAS-40278-9	13 Jun 2009	Lenawee silty clay loam
	3	60 X 40	DAS-40474-7	13 Jun 2009	Lenawee silty clay loam
OH-12	1	50 X 20	---	10 Jun 2009	Hoytville clay loam
	2	50 X 20	DAS-40278-9	10 Jun 2009	Hoytville clay loam
	3	50 X 20	DAS-40474-7	10 Jun 2009	Hoytville clay loam
IN-13	1	30 X 25	---	15 Jun 2009	Homer, Sebewa sandy loam
	2	30 X 25	DAS-40278-9	15 Jun 2009	Homer, Sebewa sandy loam
	3	30 X 25	DAS-40474-7	15 Jun 2009	Homer, Sebewa sandy loam
NE-14	1	120 X 125	---	01 Jun 2009	Hastings Silt Loam
	2	20 X 50	DAS-40278-9	01 Jun 2009	Hastings Silt Loam
	3	20 X 50	DAS-40474-7	01 Jun 2009	Hastings Silt Loam
	4	120 X 125	DAS-40278-9	01 Jun 2009	Hastings Silt Loam
	5	120 X 125	DAS-40474-7	01 Jun 2009	Hastings Silt Loam
NE-15	1	15 X 50	---	22 Jun 2009	Hastings Silt Loam
	2	20 X 66	DAS-40278-9	22 Jun 2009	Hastings Silt Loam
	3	20 X 66	DAS-40474-7	22 Jun 2009	Hastings Silt Loam
KS-16	1	20 X 50	---	29 Jun 2009	Crete Silt Loam
	2	20 X 66	DAS-40278-9	29 Jun 2009	Crete Silt Loam
	3	20 X 66	DAS-40474-7	29 Jun 2009	Crete Silt Loam
IA-17	1	20 X 50	---	05 Jun 2009	Taintor silt loam
	2	20 X 50	DAS-40278-9	05 Jun 2009	Taintor silt loam
	3	20 X 50	DAS-40474-7	05 Jun 2009	Taintor silt loam
MO-18	1	20 X 50	---	05 Jun 2009	Abco Silt Loam
	2	20 X 50	DAS-40278-9	05 Jun 2009	Abco Silt Loam
	3	20 X 50	DAS-40474-7	05 Jun 2009	Abco Silt Loam
IA-19	1	50 X 20	---	02 Jun 2009	Sandy Loam
	2	50 X 20	DAS-40278-9	02 Jun 2009	Sandy Loam
	3	50 X 20	DAS-40474-7	02 Jun 2009	Sandy Loam
MN-20	1	50 X 20	---	01 Jun 2009	Loam
	2	50 X 20	DAS-40278-9	01 Jun 2009	Loam
	3	50 X 20	DAS-40474-7	01 Jun 2009	Loam
MO-21	1	20 X 50	---	30 Jun 2009	Calhoun Silt loam
	2	20 X 50	DAS-40278-9	30 Jun 2009	Calhoun Silt loam
	3	20 X 50	DAS-40474-7	30 Jun 2009	Calhoun Silt loam
OK-22	1	50 X 40	---	27 Jun 2009	Konisl fine sandy loam
	2	50 X 40	DAS-40278-9	27 Jun 2009	Konisl fine sandy loam
	3	50 X 40	DAS-40474-7	27 Jun 2009	Konisl fine sandy loam
TX-23	1	50 X 20	---	28 Jun 2009	Wilson clay loam
	2	50 X 20	DAS-40278-9	28 Jun 2009	Wilson clay loam
	3	50 X 20	DAS-40474-7	28 Jun 2009	Wilson clay loam

Table 5: Trial Layout Summary

Trial #	Trt #	Plot Size (ft)	AAD-1 Event [a]	Planting Date	Soil Type
ON-24	1	24.6 X 9.8	---	03 Jun 2009	Brant silt loam
	2	49.2 X 9.8	DAS-40278-9	03 Jun 2009	Brant silt loam
	3	49.2 X 9.8	DAS-40474-7	03 Jun 2009	Brant silt loam
ON-25	1	24.6 X 9.8	---	05 Jun 2009	silt loam
	2	49.2 X 9.8	DAS-40278-9	05 Jun 2009	silt loam
	3	49.2 X 9.8	DAS-40474-7	05 Jun 2009	silt loam

[a] Untreated control plots (Trt. 1) were planted to the commercial variety Mycogen 2M746, which lacked the inserted AAD-1 gene, but had a genetic background similar to that used for the two non-commercial transgenic hybrids in treated plots (Treatment nos. 2-5). Treatments 2 and 4 were planted to corn containing AAD-1 Event DAS-40278-9, while treatments 3 and 5 were planted to corn containing AAD-1 Event DAS-40474-7.

3.3. Trial Site and Crop Maintenance

For all trials, the field corn plants were grown and maintained in a healthy, commercially representative state using standard agricultural practices to yield commercially representative field corn. As needed, the plots were maintained relatively free of weeds using a registered herbicides at label rates that did not contain 2,4-D or other phenoxy-type herbicides. Disease and insects were controlled with registered fungicides and insecticides at label rates as needed. Typical maintenance chemicals were applied during the trial period as noted in Table 6. Irrigation and fertilizer was applied as needed during the growing season to maintain a viable crop. Tilling of rows and work areas, and hand weeding if done, was conducted in the control plot first to avoid potential contamination.

Table 6: Plot Maintenance Chemicals Summary

Trial #	Application Date	Maintenance Product Name	Application Rate	Purpose
NJ-01	07 Jul 2009	Dual	1.5 pt/A	Herbicide
	07 Jul 2009	Atrazine	1.25 qt/A	Herbicide
	07 Jul 2009	Roundup	2.0 pt/A	Herbicide
PA-02	08 Jun 2009	Dual	1.5 pt/A	Herbicide
	08 Jun 2009	Atrazine	1.25 qt/A	Herbicide
GA-03	01 Jul 2009	Karate	2 oz/A	Insecticide
	10 Jul 2009	Brigade	8 oz/A	Insecticide
	20 Jul 2009	Karate	4 oz/A	Insecticide
	27 Jul 2009	Lannate	1.5 pts/A	Insecticide
	08 Aug 2009	Brigade	6 oz/A	Insecticide
GA-04	01 Jul 2009	Karate	2 oz/A	Insecticide
	10 Jul 2009	Brigade	8 oz/A	Insecticide
	20 Jul 2009	Karate	4 oz/A	Insecticide
	27 Jul 2009	Lannate	1.5 pts/A	Insecticide
	08 Aug 2009	Brigade	6 oz/A	Insecticide
IL-05	04 Jun 2009	Lexar 3.7EC	3.5 qt/A	Herbicide

Table 6: Plot Maintenance Chemicals Summary

Trial #	Application Date	Maintenance Product Name	Application Rate	Purpose
IL-06	26 June 2009	Bicep II Mag.	3 pt/A	Herbicide
	29 Jul 2009	Warrior	3 oz/A	Insecticide
	13 Aug 2009	Warrior	3 oz/A	Insecticide
IN-07	09 Jun 2009	Bicep II Magnum	1.5 qt./A	Herbicide
	09 Jun 2009	Princep	1.0 lb/A	Herbicide
	09 Jun 2009	Callisto	6.0 fl. oz/A	Herbicide
MO-08	10 Jul 2009	Atrazine	1.0 lb ai/A	Herbicide
	10 Jul 2009	Steadfast	0.75 oz/A	Herbicide
	10 Jul 2009	Tracer	3.0 oz/A	Insecticide
MO-09	28 Jun 2009	Glyphosphate	1.0 lb ai/A	Herbicide
	28 Jun 2009	Metolachlor	1.0 pt prod/A	Herbicide
	10 Jul 2009	Atrazine	2.0 pt prod/A	Herbicide
	10 Jul 2009	Tracer	4.0 oz prod/A	Insecticide
IL-10	06 Jul 2009	Glyphosphate	1.0 lb ai/A	Herbicide
	06 Jul 2009	Metolachlor	1.3 pt prod/A	Herbicide
	25 Jul 2009	Atrazine	1.0 lb ai/A	Herbicide
	25 Jul 2009	Steadfast	0.75 oz prod/A	Herbicide
MI-11	No Maintenance Chemicals Applied			
OH-12	No Maintenance Chemicals Applied			
IN-13	No Maintenance Chemicals Applied			
NE-14	04 Jun 2009	Lexar	2.78 lb ai/A	Herbicide
	24 Jul 2009	Headline	0.10 lb ai/A	Fungicide
	01 Aug 2009	Mustang Max	0.10 lb ai/A	Insecticide
NE-15	22 Jun 2009	Dual II Magnum	1.3 pt prdt/A	Herbicide

Table 6: Plot Maintenance Chemicals Summary

Trial #	Application Date	Maintenance Product Name	Application Rate	Purpose
KS-16	29 Jun 2009	Dual II Magnum	1.3 pt prdt/A	Herbicide
	29 Jun 2009	Touchdown Total	24 oz Prdt/A	Herbicide
	29 Jun 2009	AMS	17 lbs Prdt/100 gal	Herbicide
IA-17	20 May 2009	Charger Max	1.3 lb ai/A	Herbicide
	20 May 2009	Atrazine	1.6 lb ai/A	Herbicide
	05 Jun 2009	Force 3G	0.12 lb ai/A	Insecticide
MO-18	09 Jun 2009	Degree Xtra	3.5 qt/A	Herbicide
	09 Jun 2009	Atrazine 90DF	1.0 lb/A	Herbicide
IA-19	No Maintenance Chemicals Applied			
MN-20	No Maintenance Chemicals Applied			
MO-21	14 Jul 2009	Aatrex	1.5 qt/A	Herbicide
	14 Jul 2009	Dual Magnum	1 pt/A	Herbicide
	29 Jul 2009	Steadfast	0.75 oz/A	Herbicide
	29 Jul 2009	Karate Z	1.5 oz/A	Insecticide
	08 Aug 2009	Karate Z	1.5 oz/A	Insecticide
	19 Aug 2009	Karate Z	1.5 oz/A	Insecticide
OK-22	No Maintenance Chemicals Applied			
TX-23	No Maintenance Chemicals Applied			
ON-24	27 May 2009	Dual II Magnum	1.75 L/Ha	Herbicide
	27 May 2009	Callisto	0.3 L/Ha	Herbicide
	27 May 2009	Aatrex 480	2.5 L/Ha	Herbicide
ON-25	11 Jun 2009	Dual II Magnum	1.75 L/Ha	Herbicide
	11 Jun 2009	Callisto	0.3 L/Ha	Herbicide
	11 Jun 2009	Aatrex 480	3.1 L/Ha	Herbicide

4. TEST SUBSTANCE APPLICATION

For the RAC and decline + RAC trials, treated plots received two broadcast spray applications of 2,4-D at approximately 1.0 pound acid equivalent per acre (lb ae/A) one at preemergence and one foliar. A third broadcast application was made of quizalofop-P-ethyl and was a foliar application at an approximate rate of 0.082 pounds active ingredient per acre (lb ai/A). A fourth broadcast application was made of 2,4-D and was a foliar application at an approximate rate of 1.0 lb ae/A. For the RAC + processing trials, treatments 2 and 3 were the same as for the RAC and decline + RAC trials, but two additional treatments (Treatments 4 and 5) were included and here the 2,4-D and quizalofop-P-ethyl were applied at exaggerated 2x rates. Treatments 4 and 5 received two broadcast spray applications at approximately 2.0 pound acid equivalent per acre (lb ae/A) one at preemergence and one foliar. A third broadcast application was made of quizalofop-P-ethyl and was a foliar application at an approximate rate of 0.164 pounds active ingredient per acre (lb ai/A). A fourth broadcast application was made of 2,4-D and was a foliar application at an approximate rate of 2.0 lb ae/A. Applications were made to the treated plot using calibrated ground sprayer equipment and typical agricultural practices and, according to established Standard Operating Procedures (SOPs). All applications were made using a target spray volume of 20-40 gallons per acre (GPA) with 2,4-D applications including a non-ionic surfactant (NIS), such as X-77, in the spray mixture at approximately 0.25% (v/v) and quizalofop-P-ethyl applications including a crop oil concentrate (COC), such as Agri-Dex, in the spray mixture at approximately 1.25% (v/v).

As indicated above, in the first application 2,4-D was applied preemergence. The second and third applications were timed to be 12 days (+/-1 day) and 5 days (+/-1 day) before the fourth application, with the fourth application targeted at the V8 growth stage. Application of the test substance was as uniform as possible to the entire treated plot area and spray passes did not overlap. The untreated plot received applications of the maintenance chemicals used at each location, but did not receive any applications of the test substance. The spray equipment was calibrated prior to each application. Refer to Table 7 below for application summary information and Table 8 for details of the application equipment and pattern.

Table 7: Application Summary

Trial No.	Plot ID	No. of Apps	Application Dates	Spray interval (days)	Test Substance	Rate (a) (lb ae/A or lb ai/A)	Additive & Rate (v/v) (b)	Spray Volume (GPA)	Crop Height (inches)	Crop Growth Stage (c)
NJ-01	T2	4	07 Jul 2009	N/A	2,4-D	1.033	NIS @ 0.25%	33.6	0	Pre Emerge
			30 Jul 2009	23	2,4-D	1.043	NIS @ 0.25%	31.26	22	V5
			04 Aug 2009	5	Quizalofop-P-ethyl	0.0837	COC @ 1.25%	33.09	48 [b]	V8
			10 Aug 2009	6	2,4-D	1.02	NIS @ 0.25%	35.64	48 [b]	V8
	T3	4	07 Jul 2009	N/A	2,4-D	1.0206	NIS @ 0.25%	33.2	0	Pre Emerge
			30 Jul 2009	23	2,4-D	1.02	NIS @ 0.25%	30.58	22	V5
			04 Aug 2009	5	Quizalofop-P-ethyl	0.0838	COC @ 1.25%	33.13	48 [b]	V8
			10 Aug 2009	6	2,4-D	1.04	NIS @ 0.25%	36.37	48 [b]	V8
PA-02	T2	4	08 Jun 2009	N/A	2,4-D	1.03	NIS @ 0.25%	23.15	0	Pre Emerge
			08 Jul 2009	30	2,4-D	1.026	NIS @ 0.25%	23.18	18 - 22	5-6 leaf stage
			15 Jul 2009	7	Quizalofop-P-ethyl	0.0844	COC @ 1.25%	23.1	30	V6
			20 Jul 2009	5	2,4-D	1.05	NIS @ 0.25%	23.69	36 - 42	V8
	T3	4	08 Jun 2009	N/A	2,4-D	1.02	NIS @ 0.25%	22.8	0	Pre Emerge
			08 Jul 2009	30	2,4-D	1.01	NIS @ 0.25%	22.82	18 - 22	5-6 leaf stage
			15 Jul 2009	7	Quizalofop-P-ethyl	0.0848	COC @ 1.25%	23.2	30	V6
			20 Jul 2009	5	2,4-D	1.03	NIS @ 0.25%	23.38	36 - 42	V8

Table 7: Application Summary (continued)

Trial No.	Plot ID	No. of Apps	Application Dates	Spray interval (days)	Test Substance	Rate (a) (lb ae/A or lb ai/A)	Additive & Rate (v/v) (b)	Spray Volume (GPA)	Crop Height (inches)	Crop Growth Stage
GA03	T2	4	12 Jun 2009	N/A	2,4-D	1.0	NIS @ 0.25%	22.4	0	Pre Emerge
			09 Jul 2009	27	2,4-D	1.0	NIS @ 0.25%	22.6	38 - 40	BBCH 18
			15 Jul 2009	6	Quizalofop-P-ethyl	0.082	COC @ 1.25%	22.1	42 - 44	V8
			20 Jul 2009	5	2,4-D	1.0	NIS @ 0.25%	21.8	44 - 47	V8
	T3	4	12 Jun 2009	N/A	2,4-D	1.0	NIS @ 0.25%	22.8	0	Pre Emerge
			09 Jul 2009	27	2,4-D	1.0	NIS @ 0.25%	22.6	38 - 40	BBCH 18
			15 Jul 2009	6	Quizalofop-P-ethyl	0.082	COC @ 1.25%	22.2	42 - 44	V8
			20 Jul 2009	5	2,4-D	1.0	NIS @ 0.25%	21.7	44 - 47	V8
	T4	4	12 Jun 2009	N/A	2,4-D	2.0	NIS @ 0.25%	22.6	0	Pre Emerge
			09 Jul 2009	27	2,4-D	2.0	NIS @ 0.25%	22.6	38 - 40	BBCH 18
			15 Jul 2009	6	Quizalofop-P-ethyl	0.164	COC @ 1.25%	22.0	42 - 44	V8
			20 Jul 2009	5	2,4-D	2.0	NIS @ 0.25%	21.8	42 - 47	V8
	T5	4	12 Jun 2009	N/A	2,4-D	2.0	NIS @ 0.25%	22.6	0	Pre Emerge
			09 Jul 2009	27	2,4-D	2.0	NIS @ 0.25%	22.6	38 - 40	BBCH 18
			15 Jul 2009	6	Quizalofop-P-ethyl	0.164	COC @ 1.25%	22.0	42 - 44	V8
			20 Jul 2009	5	2,4-D	2.0	NIS @ 0.25%	21.8	42 - 47	V8

Table 7: Application Summary (continued)

Trial No.	Plot ID	No. of Apps	Application Dates	Spray interval (days)	Test Substance	Rate (a) (lb ae/A or lb ai/A)	Additive & Rate (v/v) (b)	Spray Volume (GPA)	Crop Height (inches)	Crop Growth Stage
GA04	T2	4	12 Jun 2009	N/A	2,4-D	1.0	NIS @ 0.25%	22.7	0	Pre Emerge
			20 Jul 2009	38	2,4-D	1.0	NIS @ 0.25%	21.7	36 - 40	BBCH 18
			27 Jul 2009	7	Quizalofop-P-ethyl	0.082	COC @ 1.25%	21.7	40 - 44	V8
			31 Jul 2009	4	2,4-D	1.0	NIS @ 0.25%	21.9	46 - 50	V8
	T3	4	12 Jun 2009	N/A	2,4-D	1.0	NIS @ 0.25%	22.7	0	Pre Emerge
			20Jul 2009	38	2,4-D	1.0	NIS @ 0.25%	21.7	36 - 40	BBCH 18
			27 Jul 2009	7	Quizalofop-P-ethyl	0.082	COC @ 1.25%	21.7	40 - 44	V8
			31 Jul 2009	4	2,4-D	1.0	NIS @ 0.25%	22.0	46 - 50	V8
IL-05	T2	4	05 Jun 2009	N/A	2,4-D	1.0	NIS @ 0.25%	22.9	0	Pre Emerge
			26 Jun 2009	21	2,4-D	1.0	NIS @ 0.25%	20.8	14	BBCH 14
			03 Jul 2009	7	Quizalofop-P-ethyl	0.080	COC @ 1.25%	20.3	17	BBCH 16
			09 Jul 2009	4	2,4-D	1.0	NIS @ 0.25%	20.7	30 - 32	V8
	T3	4	05 Jun 2009	N/A	2,4-D	1.0	NIS @ 0.25%	22.9	0	Pre Emerge
			26 Jun 2009	21	2,4-D	1.0	NIS @ 0.25%	20.8	14	BBCH 14
			03 Jul 2009	7	Quizalofop-P-ethyl	0.080	COC @ 1.25%	20.3	17	BBCH 16
			09 Jul 2009	4	2,4-D	1.0	NIS @ 0.25%	20.7	30 - 32	V8

Table 7: Application Summary (continued)

Trial No.	Plot ID	No. of Apps	Application Dates	Spray interval (days)	Test Substance	Rate (a) (lb ae/A or lb ai/A)	Additive & Rate (v/v) (b)	Spray Volume (GPA)	Crop Height (inches)	Crop Growth Stage
IL-06	T2	4	26 Jun 2009	N/A	2,4-D	1.0	NIS @ 0.18%	35.3	0	Pre Emerge
			16 Jul 2009	20	2,4-D	0.99	NIS @ 0.25%	25.4	20	V4
			22 Jul 2009	6	Quizalofop-P-ethyl	0.083	COC @ 1.25%	24.6	28 - 30	V6
			28 Jul 209	6	2,4-D	1.01	NIS @ 0.25%	26.3	36 - 38	V8
	T3	4	26 Jun 2009	N/A	2,4-D	0.998	NIS @ 0.18%	35.2	0	Pre Emerge
			16 Jul 2009	20	2,4-D	0.99	NIS @ 0.25%	25.5	20	V4
			22 Jul 2009	6	Quizalofop-P-ethyl	0.083	COC @ 1.25%	24.6	28 - 30	V6
			28 Jul 209	6	2,4-D	1.01	NIS @ 0.25%	26.3	36 - 38	V8
IN-07	T2	4	09 Jun 2009	N/A	2,4-D	0.99	NIS @ 0.25%	22.54	0	Pre Emerge
			02 Jul 2009	23	2,4-D	1.03	NIS @ 0.25%	23.38	18 - 22	V5
			10 Jul 2009	8	Quizalofop-P-ethyl	0.083	COC @ 1.25%	22.38	28 - 32	V6-V7
			15 Jul 2009	5	2,4-D	1.01	NIS @ 0.25%	23.46	36 - 40	V8
	T3	4	09 Jun 2009	N/A	2,4-D	0.99	NIS @ 0.25%	22.54	0	Pre Emerge
			02 Jul 2009	23	2,4-D	1.03	NIS @ 0.25%	23.38	18 - 22	V5
			10 Jul 2009	8	Quizalofop-P-ethyl	0.083	COC @ 1.25%	22.38	28 - 32	V6-V7
			15 Jul 2009	5	2,4-D	1.01	NIS @ 0.25%	23.46	36 - 40	V8

Table 7: Application Summary (continued)

Trial No.	Plot ID	No. of Apps	Application Dates	Spray interval (days)	Test Substance	Rate (a) (lb ae/A or lb ai/A)	Additive & Rate (v/v) (b)	Spray Volume (GPA)	Crop Height (inches)	Crop Growth Stage
MO-08	T2	4	03 Jul 2009	N/A	2,4-D	1.03	NIS @ 0.25%	20.6	0	BBCH 00
			28 Jul 2009	25	2,4-D	0.99	NIS @ 0.25%	19.9	36	BBCH 13
			04 Aug 2009	7	Quizalofop-P-ethyl	0.083	NIS @ 1.25%	19.7	38	BBCH 16
			09 Aug 2009	5	2,4-D	1.01	NIS @ 0.25%	20	40	BBCH 18
	T3	4	03 Jul 2009	N/A	2,4-D	1.00	NIS @ 0.25%	20.3	0	BBCH 00
			28 Jul 2009	25	2,4-D	0.99	NIS @ 0.25%	19.8	36	BBCH 13
			04 Aug 2009	7	Quizalofop-P-ethyl	0.082	NIS @ 1.25%	20.1	38	BBCH 16
			09 Aug 2009	5	2,4-D	1.02	NIS @ 0.25%	20.2	40	BBCH 18
MO-09	T2	4	28 Jun 2009	N/A	2,4-D	0.987	NIS @ 0.25%	20.1	37	BBCH 13
			27 Jul 2009	29	2,4-D	0.99	NIS @ 0.25%	20.1	39	BBCH 16
			03 Aug 2009	7	Quizalofop-P-ethyl	0.081	COC @ 1.25%	19.9	40	BBCH 18
			08 Aug 2009	5	2,4-D	0.971	NIS @ 0.25%	19.9	46	BBCH 18
	T3	4	28 Jun 2009	N/A	2,4-D	0.987	NIS @ 0.25%	20.1	37	BBCH 13
			27 Jul 2009	29	2,4-D	1.0	NIS @ 0.25%	20.2	39	BBCH 16
			03 Aug 2009	7	Quizalofop-P-ethyl	0.082	COC @ 1.25%	20.3	40	BBCH 18
			08 Aug 2009	5	2,4-D	0.976	NIS @ 0.25%	20.0	46	BBCH 18

Table 7: Application Summary (continued)

Trial No.	Plot ID	No. of Apps	Application Dates	Spray interval (days)	Test Substance	Rate (a) (lb ae/A or lb ai/A)	Additive & Rate (v/v) (b)	Spray Volume (GPA)	Crop Height (inches)	Crop Growth Stage
IL-10	T2	4	06 Jul 2009	N/A	2,4-D	1.015	NIS @ 0.25%	20.09	0	BBCH 00
			01 Aug 2009	26	2,4-D	1.00	NIS @ 0.25%	20.56	36	BBCH 14
			08 Aug 2009	7	Quizalofop-P-ethyl	0.078	COC @ 1.25%	19.8	35	BBCH 16
			13 Aug 2009	5	2,4-D	0.96	NIS @ 0.25%	20.2	41	BBCH 18
	T3	4	06 Jul 2009	N/A	2,4-D	1.04	NIS @ 0.25%	20.61	0	BBCH 00
			01 Aug 2009	26	2,4-D	1.01	NIS @ 0.25%	20.79	36	BBCH 14
			08 Aug 2009	7	Quizalofop-P-ethyl	0.077	COC @ 1.25%	19.6	35	BBCH 16
			13 Aug 2009	5	2,4-D	0.95	NIS @ 0.25%	19.9	41	BBCH 18
MI-11	T2	4	17 Jun 2009	N/A	2,4-D	1.0	NIS @ 0.25%	23.0	0	BBCH 00
			25 Jul 2009	38	2,4-D	1.0	NIS @ 0.25%	23.9	35 - 40	BBCH-30
			31 Jul 2009	6	Quizalofop-P-ethyl	0.082	COC @ 1.25%	22.6	45 - 48	BBCH 39
			05 Aug 2009	5	2,4-D	1.0	NIS @ 0.25%	22.8	48 - 50	BBCH 45
	T3	4	17 Jun 2009	N/A	2,4-D	1.0	NIS @ 0.25%	23	0	BBCH 00
			25 Jul 2009	38	2,4-D	1.00	NIS @ 0.25%	23.8	35 - 40	BBCH 30
			31 Jul 2009	6	Quizalofop-P-ethyl	0.082	COC @ 1.25%	22.8	45 -48	BBCH 39
			05 Aug 2009	5	2,4-D	1.0	NIS @ 0.25%	22.7	48 - 50	BBCH 45

Table 7: Application Summary (continued)

Trial No.	Plot ID	No. of Apps	Application Dates	Spray interval (days)	Test Substance	Rate (a) (lb ae/A or lb ai/A)	Additive & Rate (v/v) (b)	Spray Volume (GPA)	Crop Height (inches)	Crop Growth Stage
OH-12	T2	4	14 Jun 2009	N/A	2,4-D	1.0	NIS @ 0.25%	23.1	0	BBCH 00
			27 Jul 2009	43	2,4-D	1.0	NIS @ 0.25%	25.2	30 – 36	BBCH 18
			03 Aug 2009	7	Quizalofop-P-ethyl	0.081	COC @ 1.25%	23.1	38 - 46	BBCH 30
			08 Aug 2009	5	2,4-D	1.0	NIS @ 0.25%	22.5	46 - 48	BBCH 35
	T3	4	14 Jun 2009	N/A	2,4-D	1.0	NIS @ 0.25%	23.1	0	BBCH 00
			27 Jul 2009	43	2,4-D	1.0	NIS @ 0.25%	25.2	30 - 36	BBCH 18
			03 Aug 2009	7	Quizalofop-P-ethyl	0.082	COC @ 1.25%	23.2	38 - 46	BBCH 30
			08 Aug 2009	5	2,4-D	1.0	NIS @ 0.25%	22.4	46 - 48	BBCH 35
IN-13	T2	4	15 Jun 2009	N/A	2,4-D	1.0	NIS @ 0.25%	22.7	0	BBCH 00
			25 Jul 2009	40	2,4-D	1.0	NIS @ 0.25%	25.2	36 - 40	BBCH 39-40
			31 Jul 2009	6	Quizalofop-P-ethyl	0.081	COC @ 1.25%	23.1	44 - 48	BBCH 41-42
			05 Aug 2009	5	2,4-D	1.0	NIS @ 0.25%	22.9	49 - 52	BBCH 43-44
	T3	4	15 Jun 2009	N/A	2,4-D	1.0	NIS @ 0.25%	22.6	0	BBCH 00
			25 Jul 2009	40	2,4-D	1.0	NIS @ 0.25%	25.2	36 - 40	BBCH 39-40
			31 Jul 2009	6	Quizalofop-P-ethyl	0.082	COC @ 1.25%	23.2	44 - 48	BBCH 41-42
			05 Aug 2009	5	2,4-D	1.0	NIS @ 0.25%	22.9	49 - 52	BBCH 43-44

Table 7: Application Summary (continued)

Trial No.	Plot ID	No. of Apps	Application Dates	Spray interval (days)	Test Substance	Rate (a) (lb ae/A or lb ai/A)	Additive & Rate (v/v) (b)	Spray Volume (GPA)	Crop Height (inches)	Crop Growth Stage
NE-14	T2	4	04 Jun 2009	N/A	2,4-D	0.99	NIS @ 0.25%	25.93	0	BBCH 00
			24 Jun 2009	20	2,4-D	1.01	NIS @ 0.25%	31.78	8 - 12	BBCH 14
			30 Jun 2009	6	Quizalofop-P-ethyl	0.082	COC @ 1.25%	31.78	18 - 20	BBCH 16
			06 Jul 2009	6	2,4-D	1.01	NIS @ 0.25%	31.65	28 - 33	BBCH 18
	T3	4	04 Jun 2009	N/A	2,4-D	1.01	NIS @ 0.25%	26.35	0	BBCH 00
			24 Jun 2009	20	2,4-D	1.01	NIS @ 0.25%	31.68	8 - 12	BBCH 14
			30 Jun 2009	6	Quizalofop-P-ethyl	0.081	COC @ 1.25%	31.45	18 - 20	BBCH 16
			06 Jul 2009	6	2,4-D	1.01	NIS @ 0.25%	31.72	28 - 33	BBCH 18
	T4	4	04 Jun 2009	N/A	2,4-D	2.02	NIS @ 0.25%	26.34	0	BBCH 00
			24 Jun 2009	20	2,4-D	1.98	NIS @ 0.25%	31.38	8 - 12	BBCH 14
			30 Jun 2009	6	Quizalofop-P-ethyl	0.163	COC @ 1.25%	31.33	18 - 20	BBCH 16
			06 Jul 2009	6	2,4-D	2.00	NIS @ 0.25%	31.58	28 - 33	BBCH 18
	T5	4	04 Jun 2009	N/A	2,4-D	2.01	NIS @ 0.25%	26.09	0	BBCH 00
			24 Jun 2009	20	2,4-D	1.99	NIS @ 0.25%	31.40	8 - 12	BBCH 14
			30 Jun 2009	6	Quizalofop-P-ethyl	0.163	COC @ 1.25%	31.32	18 - 20	BBCH 16
			06 Jul 2009	6	2,4-D	2.01	NIS @ 0.25%	31.60	28 - 33	BBCH 18

Table 7: Application Summary (continued)

Trial No.	Plot ID	No. of Apps	Application Dates	Spray interval (days)	Test Substance	Rate (a) (lb ae/A or lb ai/A)	Additive & Rate (v/v) (b)	Spray Volume (GPA)	Crop Height (inches)	Crop Growth Stage
NE-15	T2	4	23 Jun 2009	N/A	2,4-D	1.01	NIS @ 0.25%	20.16	0	BBCH 00
			08 Jul 2009	15	2,4-D	1.01	NIS @ 0.25%	20.26	7 - 10	BBCH 13-14
			15 Jul 2009	7	Quizalofop-P-ethyl	0.08	COC @ 1.25%	20.32	15 - 19	BBCH 15-16
			20 Jul 2009	5	2,4-D	1.00	NIS @ 0.25%	20.17	18 - 25	BBCH 17-18
	T3	4	23 Jun 2009	N/A	2,4-D	1.00	NIS @ 0.25%	20.01	0	BBCH 00
			08 Jul 2009	15	2,4-D	1.00	NIS @ 0.25%	20.16	7 - 10	BBCH 13-14
			15 Jul 2009	7	Quizalofop-P-ethyl	0.08	COC @ 1.25%	20.38	15 - 19	BBCH 15-16
			20 Jul 2009	5	2,4-D	1.00	NIS @ 0.25%	20.15	18 - 25	BBCH 17-18
KS-16	T2	4	29 Jun 2009	N/A	2,4-D	1.02	NIS @ 0.25%	23.1	0	BBCH 00
			15 Jul 2009	16	2,4-D	0.99	NIS @ 0.25%	19.7	10 - 13	BBCH 14-15
			22 Jul 2009	7	Quizalofop-P-ethyl	0.082	COC @ 1.25%	20.1	15 - 22	BBCH 17-18
			27 Jul 2009	5	2,4-D	1.004	NIS @ 0.25%	19.6	23 - 35	BBCH 18-19
	T3	4	29 Jun 2009	N/A	2,4-D	1.02	NIS @ 0.25%	23.2	0	BBCH 00
			15 Jul 2009	16	2,4-D	1.00	NIS @ 0.25%	19.8	10 - 13	BBCH 14-15
			22 Jul 2009	7	Quizalofop-P-ethyl	0.083	COC @ 1.25%	20.4	15 - 22	BBCH 17-18
			27 Jul 2009	5	2,4-D	1.004	NIS @ 0.25%	19.6	23 - 35	BBCH 18-19

Table 7: Application Summary (continued)

Trial No.	Plot ID	No. of Apps	Application Dates	Spray interval (days)	Test Substance	Rate (a) (lb ae/A or lb ai/A)	Additive & Rate (v/v) (b)	Spray Volume (GPA)	Crop Height (inches)	Crop Growth Stage
IA-17	T2	4	09 Jun 2009	N/A	2,4-D	0.97	NIS @ 0.25%	21.3	0	BBCH 05
			24 Jun 2009	15	2,4-D	1.04	NIS @ 0.25%	21.4	12	BBCH 14
			01 Jul 2009	7	Quizalofop-P-ethyl	0.084	COC @ 1.25%	20.3	19	BBCH 16
			06 Jul 2009	5	2,4-D	1.01	NIS @ 0.25%	20.1	29	BBCH 17
	T3	4	09 Jun 2009	N/A	2,4-D	0.98	NIS @ 0.25%	21.6	0	BBCH 05
			24 Jun 2009	15	2,4-D	1.04	NIS @ 0.25%	21.4	12	BBCH 14
			01 Jul 2009	7	Quizalofop-P-ethyl	0.081	COC @ 1.25%	19.6	19	BBCH 16
			06 Jul 2009	5	2,4-D	1.01	NIS @ 0.25%	20.2	29	BBCH 17
MO-18	T2	4	06 Jun 2009	N/A	2,4-D	1.02	NIS @ 0.25%	21.0	0	BBCH 00
			30 Jun 2009	24	2,4-D	1.00	NIS @ 0.25%	20.1	14 - 16	BBCH 15
			06 Jul 2009	6	Quizalofop-P-ethyl	0.084	COC @ 1.25%	20.0	27	BBCH 16.5
			12 Jul 2009	6	2,4-D	1.02	NIS @ 0.25%	21.06	36	BBCH 18
	T3	4	06 Jun 2009	N/A	2,4-D	1.02	NIS @ 0.25%	20.9	0	BBCH 00
			30 Jun 2009	24	2,4-D	1.00	NIS @ 0.25%	20.1	14 - 16	BBCH 15
			06 Jul 2009	6	Quizalofop-P-ethyl	0.083	COC @ 1.25%	19.9	30	BBCH 16.5
			12 Jul 2009	6	2,4-D	1.01	NIS @ 0.25%	20.73	46	BBCH 18

Table 7: Application Summary (continued)

Trial No.	Plot ID	No. of Apps	Application Dates	Spray interval (days)	Test Substance	Rate (a) (lb ae/A or lb ai/A)	Additive & Rate (v/v) (b)	Spray Volume (GPA)	Crop Height (inches)	Crop Growth Stage
IA-19	T2	4	05 Jun 2009	N/A	2,4-D	1.03	NIS @ 0.25%	23.9	0	Pre emerge
			11 Jul 2009	36	2,4-D	1.04	NIS @ 0.25%	27	24	V5
			18 Jul 2009	7	Quizalofop-P-ethyl	0.081	COC @ 1.25%	27.7	42	V7
			23 Jul 2009	5	2,4-D	0.98	NIS @ 0.25%	22.4	48	V8
	T3	4	05 Jun 2009	N/A	2,4-D	1.05	NIS @ 0.25%	24.4	0	Pre emerge
			11 Jul 2009	36	2,4-D	1.03	NIS @ 0.25%	26.7	24	V5
			18 Jul 2009	7	Quizalofop-P-ethyl	0.081	COC @ 1.25%	27.7	42	V7
			23 Jul 2009	5	2,4-D	0.98	NIS @ 0.25%	22.4	48	V8
MN-20	T2	4	05 Jun 2009	N/A	2,4-D	1.04	NIS @ 0.25%	23.7	0	Pre emerge
			11 Jul 2009	36	2,4-D	1.0	NIS @ 0.25%	26	20	V5
			18 Jul 2009	7	Quizalofop-P-ethyl	0.081	COC @ 1.25%	27.8	12 - 38	V7
			23 Jul 2009	5	2,4-D	1.03	NIS @ 0.25%	23.5	32	V8
	T3	4	05 Jun 2009	N/A	2,4-D	1.07	NIS @ 0.25%	24.4	0	Pre emerge
			11 Jul 2009	36	2,4-D	1.06	NIS @ 0.25%	27.5	27	V5
			18 Jul 2009	7	Quizalofop-P-ethyl	0.081	COC @ 1.25%	27.8	40	V7
			23 Jul 2009	5	2,4-D	1.03	NIS @ 0.25%	23.5	48	V8

Table 7: Application Summary (continued)

Trial No.	Plot ID	No. of Apps	Application Dates	Spray interval (days)	Test Substance	Rate (a) (lb ae/A or lb ai/A)	Additive & Rate (v/v) (b)	Spray Volume (GPA)	Crop Height (inches)	Crop Growth Stage
MO-21	T2	4	30 Jun 2009	N/A	2,4-D	1.008	NIS @ 0.25%	20.15	0	Pre Emerge
			23 Jul 2009	23	2,4-D	1.013	NIS @ 0.25%	20.25	10 - 12	BBCH 15
			30 Jul 2009	7	Quizalofop-P-ethyl	0.0822	COC @ 1.25%	20.10	20 - 22	BBCH 17
			04 Aug 2009	5	2,4-D	0.994	NIS @ 0.25%	19.88	36 - 38	BBCH 18
	T3	4	30 Jun 2009	N/A	2,4-D	0.996	NIS @ 0.25%	19.91	0	Pre-Emerge
			23 Jul 2009	23	2,4-D	1.011	NIS @ 0.25%	20.21	10 - 12	BBCH 15
			30 Jul 2009	7	Quizalofop-P-ethyl	0.0814	COC @ 1.25%	19.92	20 - 22	BBCH 17
			04 Aug 2009	5	2,4-D	0.993	NIS @ 0.25%	19.86	36 - 38	BBCH 18
OK-22	T2	4	27 Jun 2009	N/A	2,4-D	1.026	NIS @ 0.25%	23.9	0	BBCH 00
			13 Jul 2009	16	2,4-D	1.007	NIS @ 0.25%	23.3	18	BBCH 16
			18 Jul 2009	5	Quizalofop-P-ethyl	0.085	COC @ 1.25%	23.7	20	BBCH 17
			23 Jul 2009	5	2,4-D	1.013	NIS @ 0.25%	23.6	22	BBCH 18
	T3	4	27 Jun 2009	N/A	2,4-D	1.033	NIS @ 0.25%	24.1	0	BBCH 00
			13 Jul 2009	16	2,4-D	1.011	NIS @ 0.25%	23.4	18	BBCH 16
			18 Jul 2009	5	Quizalofop-P-ethyl	0.082	COC @ 1.25%	23.1	20	BBCH 17
			23 Jul 2009	5	2,4-D	1.02	NIS @ 0.25%	23.8	22	BBCH 18

Table 7: Application Summary (continued)

Trial No.	Plot ID	No. of Apps	Application Dates	Spray interval (days)	Test Substance	Rate (a) (lb ae/A or lb ai/A)	Additive & Rate (v/v) (b)	Spray Volume (GPA)	Crop Height (inches)	Crop Growth Stage
TX-23	T2	4	28 Jun 2009	N/A	2,4-D	1.0174	NIS @ 0.25%	23.66	0	BBCH 00
			14 Jul 2009	16	2,4-D	1.0101	NIS @ 0.25%	23.41	14	V3
			19 Jul 2009	5	Quizalofop-P-ethyl	0.0827	COC @ 1.25%	23.38	17	V6-V7
			24 Jul 2009	5	2,4-D	1.0035	NIS @ 0.25%	23.50	22	V8
	T3	4	28 Jun 2009	N/A	2,4-D	1.0039	NIS @ 0.25%	23.35	0	BBCH 00
			14 Jul 2009	16	2,4-D	1.0033	NIS @ 0.25%	23.25	14	V3
			19 Jul 2009	5	Quizalofop-P-ethyl	0.0827	COC @ 1.25%	23.38	17	V6-V7
			24 Jul 2009	5	2,4-D	1.0147	NIS @ 0.25%	23.77	22	V8
ON-24	T2	4	04 Jun 2009	N/A	2,4-D	1.034	NIS @ 0.25%	22.43	0	Pre Emerge
			01 Jul 2009	27	2,4-D	1.040	NIS @ 0.25%	22.52	14	V4-V5
			09 Jul 2009	8	Quizalofop-P-ethyl	0.082	COC @ 1.25%	21.63	24	V6-V7
			14 Jul 2009	5	2,4-D	1.034	NIS @ 0.25%	22.38	26 – 28	V7-V8
	T3	4	04 Jun 2009	N/A	2,4-D	1.020	NIS @ 0.25%	22.12	0	Pre Emerge
			01 Jul 2009	27	2,4-D	1.005	NIS @ 0.25%	21.75	14	V4-V5
			09 Jul 2009	8	Quizalofop-P-ethyl	0.083	COC @ 1.25%	21.96	24	V6-V7
			14 Jul 2009	5	2,4-D	1.045	NIS @ 0.25%	22.63	26 - 28	V7-V8

Table 7: Application Summary (continued)

Trial No.	Plot ID	No. of Apps	Application Dates	Spray interval (days)	Test Substance	Rate (a) (lb ae/A or lb ai/A)	Additive & Rate (v/v) (b)	Spray Volume (GPA)	Crop Height (inches)	Crop Growth Stage
ON-25	T2	4	10 Jun 2009	N/A	2,4-D	1.0041	NIS @ 0.25%	21.74	0	Pre Emerge
			06 Jul 2009	26	2,4-D	1.038	NIS @ 0.25%	22.47	16	V5
			13 Jul 2009	7	Quizalofop-P-ethyl	0.088	COC @ 1.25%	23.39	20 - 22	V6-V7
			18 Jul 2009	5	2,4-D	0.980	NIS @ 0.25%	21.22	32	V8
	T3	4	10 Jun 2009	N/A	2,4-D	1.002	NIS @ 0.25%	21.69	0	Pre Emerge
			06 Jul 2009	26	2,4-D	1.059	NIS @ 0.25%	22.92	16	V5
			13 Jul 2009	7	Quizalofop-P-ethyl	0.0844	COC @ 1.25%	22.36	20 - 22	V6-V7
			18 Jul 2009	5	2,4-D	0.978	NIS @ 0.25%	21.17	32	V8

(a) For Apps 1, 2 and 4 (2, 4-D) the rate is in units of “lbs ae/A”. For App 3 (quizalofop-P-ethyl) the rate is in units of “lbs ai/A”

(b) NIS = non-ionic surfactant, such as X-77; COC = crop oil concentrate, such as Agri-Dex

(c) Estimated visually

Table 8: Application Equipment and Pattern

Trial #	Equipment	Application Type	Swath Width	# of nozzles	Nozzle Spacing	Nozzle Size/Type
NJ-01	Boom Sprayer	Broadcast	5 ft (60 in)	3	20 in	11003 DG/ Flat Fan
PA-02	Boom Sprayer	Broadcast	5 ft (60 in)	3	20 in	10002 TT/Flat Fan
GA-03	Boom Sprayer	Broadcast	144 in	8	18 in	8002 VS Flat Fan
GA-04	Boom Sprayer	Broadcast	144 in	8	18 in	8002 VS Flat Fan, 50 Mesh
IL-05	Boom Sprayer	Broadcast	120 in	6	20 in	XR TeeJet Flat Fan 11004
IL-06	Boom Sprayer	Broadcast	15 ft (180 in)	6	30 in	11003 Flat Fan
IN-07	Boom Sprayer	Broadcast	10 ft (120 in)	6	20 in	8004 Flat Fan
MO-08	Boom Sprayer	Broadcast	60 in	4	15 in	Flat Fan XR 1100015VS
MO-09	Boom Sprayer	Broadcast	120 in	8	15 in	Flat Fan XR 8002 VS
	Boom Sprayer	Broadcast	60 in	8	15 in	Flat Fan XR 1100015VS

Table 8: Application Equipment and Pattern

Trial #	Equipment	Application Type	Swath Width	# of nozzles	Nozzle Spacing	Nozzle Size/Type
IL-10	Boom Sprayer	Broadcast	60 in	4	15 in	Flat Fan XR 1100015VS
MI-11	Boom Sprayer	Broadcast	60 in	4	15 in	Flat Fan Tee Jet 80015
OH-12	Boom Sprayer	Broadcast	60 in	4	15 in	Flat Fan Tee Jet 80015
IN-13	Boom Sprayer	Broadcast	60 in	4	15 in	Flat Fan Tee Jet 80015
NE-14	Boom Sprayer	Broadcast	240 in	12	20 in	8002 XR / Flat Fan
NE-15	Boom Sprayer	Broadcast	120 in	6	20 in	80015LP/ flat fan
	Boom Sprayer	Broadcast	60 in	2	30 in	11003/flat fan
KS-16	Boom Sprayer	Broadcast	120 in	6	20 in	80015LP/Flat Fan
	Boom Sprayer	Broadcast	60 in	2	30 in	TT11003/Flat Fan with Drop Nozzles
IA-17	Boom Sprayer	Broadcast	10 ft (120 in)	6	20 in	110-03 flat fan
MO-18	Boom Sprayer	Broadcast	10 ft (120 in)	6	20 in	110-02 flat fan
IA-19	Boom Sprayer	Broadcast	120 in	6	20 in	Flat Fan Airmix 110-04/06
MN-20	Boom Sprayer	Broadcast	120 in	6	20 in	Flat Fan Airmix 110-04
MO-21	Boom Sprayer	Broadcast	10 ft (120 in)	6	20 in	11002/Flat Fan
OK-22	Boom Sprayer	Broadcast	60 in	4	15 in	Flat Fan 110-02
TX-23	Boom Sprayer	Broadcast	60 in.	4	15 in	Flat Fan 110-02
ON-24	Boom Sprayer	Broadcast	118.1 in	6	19.7 in	8002 DG flat fan
ON-25	Boom Sprayer	Broadcast	118.1 in	6	19.7 in	8002 DG flat fan

5. SAMPLE COLLECTION AND HANDLING

5.1. Sample Collection

At each sampling event interval (defined in Tables 9 through 33) one sample was collected from the control plot and two independent samples were collected from each treated plot (only 1 sample was taken from the two treated plots (Trts. 4 and 5) for bulk grain samples). Three different sample commodities were collected for each trial in this study: field corn forage, field corn grain, and field corn stover. Care was taken to avoid plot edges where treatment may not have been uniform. Care was also taken to avoid the possibility of any contamination of the samples. Disposable gloves were worn during sampling and changed between collection of the untreated and treated samples. Samples from the untreated plot were collected prior to collecting samples from the treated plots. Samples from the untreated and treated plots were collected on the same day for each sample at each trial location.

Typically, each sample (except for the processing trials) weighed at least 1 kg and consisted of at least 12 different corn plants. Bulk field corn grain samples for RAC + processing trials weighed about 1100 pounds per sample. Finally, a top, middle, and bottom section of the plants (one each from the 3 different bunches of 4 plants) were combined into a single corn forage or stover sample. The composite sample contained a part (~1/3 of a stalk) of each of the 12 collected plants and included 4 bottom segments, 4 middle segments, and 4 top segments with each segment coming from a different corn plant.

Field Corn Forage
For RAC trials, field corn forage was collected at the two stages: 40 days (+/-2 days) (except for trials 04, 22 and 23) after final application , which is considered as an "early forage" sample timing and late-dough to early-dent growth stage, which is considered as "typical forage" sample timing. For decline + RAC trials, field corn forage was collected at 6 intervals after the final application: 40 days (+/-2 days) after final application, 14 and 7 days before late-dough to early-dent growth stage, at late-dough to early-dent growth stage, and 7 and 14 days (+/-2 days) after late-dough to early-dent growth stage. For RAC + processing trials, field corn forage was collected the same as for RAC trials from treatments 1, 2, and 3. Field corn forage is the whole aerial portion of the field corn plant with the leaves and ears attached (cut approximately 1 to 2 inches above the ground).

Field Corn Grain
For RAC trials, field corn grain was collected at typical mature harvest. For decline + RAC trials, field corn grain was collected at 5 intervals after application 4. These intervals were targeted to 14 and 7 days (+/-2 days) before corn is expected to reach typical mature harvest timing, at typical mature harvest timing (RAC sample), and 7 and 14 days (+/-2 days) after the RAC sample. For RAC + processing trials, field corn grain was collected the same as for RAC trials from treatments 1, 2, and 3. Also, bulk field corn grain samples were collected at typical mature harvest timing from treatments 1, 4, and 5 with a combine. Field corn grain is the seed removed from the cob and husk.

Field Corn Stover
For RAC trials, field corn stover was collected at typical mature harvest and on the same day as field corn grain. For decline + RAC trials, field corn stover was collected at 5 intervals after application 4. These intervals were targeted to 14 and 7 days (+/-2 days) before the corn was expected to reach typical maturity harvest timing, at typical maturity harvest timing (RAC sample), and 7 and 14 days (+/-2 days) after RAC sample. For RAC + processing trials, field corn stover was collected the same as for the RAC trials from treatments 1, 2, and 3. Stover samples were collected on the same day as grain samples. Field corn stover is the dried, mature plant stalk remaining after collection and removal of the grain or whole ear (cob and grain) and is cut approximately 1 to 2 inches above the ground.

5.2 Sample Processing

Field corn RAC, decline + RAC, and RAC + processing samples (except bulk grain) were placed either in a freezer within 4 hours of collection or chilled within 4 hours (except for trial 18) using a cooler with blue, dry, or wet ice and the samples placed in a freezer within 8 hours of collection. Bulk grain samples were shipped (may not have been frozen) within approximately 72 hours of collection and delivered within approximately 7 days of harvest. Times of collection and times when samples were placed in the freezer were recorded in field trial notebook. The temperature in the freezers storing samples were monitored and documented.

Table 9: Crop Samples (Trial 01)

Sample No.	Treatment	Sampling Event – Nominal PHI	Date of Collection	Crop Growth Stage
001-0001	1	Forage 40 DAT	17 Sep 09	early milk stage
001-0002	2	Forage 40 DAT	17 Sep 09	early milk stage
001-0003	2	Forage 40 DAT	17 Sep 09	early milk stage
001-0004	3	Forage 40 DAT	17 Sep 09	early milk stage
001-0005	3	Forage 40 DAT	17 Sep 09	early milk stage
002-0001	1	Forage 60 DAT	14 Oct 09	soft dough/early dent
002-0002	2	Forage 60 DAT	14 Oct 09	soft dough/early dent
002-0003	2	Forage 60 DAT	14 Oct 09	soft dough/early dent
002-0004	3	Forage 60 DAT	14 Oct 09	soft dough/early dent
002-0005	3	Forage 60 DAT	14 Oct 09	soft dough/early dent
003-0001	1	Grain 90 DAT	23 Nov 09	BBCH 97
003-0002	2	Grain 90 DAT	23 Nov 09	BBCH 97
003-0003	2	Grain 90 DAT	23 Nov 09	BBCH 97
003-0004	3	Grain 90 DAT	23 Nov 09	BBCH 97
003-0005	3	Grain 90 DAT	23 Nov 09	BBCH 97
004-0001	1	Stover 90 DAT	23 Nov 09	BBCH 97
004-0002	2	Stover 90 DAT	23 Nov 09	BBCH 97
004-0003	2	Stover 90 DAT	23 Nov 09	BBCH 97
004-0004	3	Stover 90 DAT	23 Nov 09	BBCH 97
004-0005	3	Stover 90 DAT	23 Nov 09	BBCH 97

Table 10: Crop Samples (Trial 02)

Sample No.	Treatment	Sampling Event – Nominal PHI	Date of Collection	Crop Growth Stage
005-0001	1	Forage 40 DAT	31 Aug 2009	Milk Stage
005-0002	2	Forage 40 DAT	31 Aug 2009	Milk Stage
005-0003	2	Forage 40 DAT	31 Aug 2009	Milk Stage
005-0004	3	Forage 40 DAT	31 Aug 2009	Milk Stage
005-0005	3	Forage 40 DAT	31 Aug 2009	Milk Stage
006-0001	1	Forage 60 DAT	14 Sep 2009	BBCH 85
006-0002	2	Forage 60 DAT	14 Sep 2009	BBCH 85
006-0003	2	Forage 60 DAT	14 Sep 2009	BBCH 85
006-0004	3	Forage 60 DAT	14 Sep 2009	BBCH 85
006-0005	3	Forage 60 DAT	14 Sep 2009	BBCH 85
007-0001	1	Grain 90 DAT	04 Nov 2009	BBCH 89
007-0002	2	Grain 90 DAT	04 Nov 2009	BBCH 89
007-0003	2	Grain 90 DAT	04 Nov 2009	BBCH 89
007-0004	3	Grain 90 DAT	04 Nov 2009	BBCH 89
007-0005	3	Grain 90 DAT	04 Nov 2009	BBCH 89
008-0001	1	Stover 90 DAT	04 Nov 2009	BBCH 89
008-0002	2	Stover 90 DAT	04 Nov 2009	BBCH 89
008-0003	2	Stover 90 DAT	04 Nov 2009	BBCH 89
008-0004	3	Stover 90 DAT	04 Nov 2009	BBCH 89
008-0005	3	Stover 90 DAT	04 Nov 2009	BBCH 89

Table 11: Crop Samples (Trial 03)

Sample No.	Treatment	Sampling Event – Nominal PHI	Date of Collection	Crop Growth Stage
009-0001	1	Forage 40 DAT	27 Aug 2009	BBCH 83
009-0002	2	Forage 40 DAT	27 Aug 2009	BBCH 83
009-0003	2	Forage 40 DAT	27 Aug 2009	BBCH 83
009-0004	3	Forage 40 DAT	27 Aug 2009	BBCH 83
009-0005	3	Forage 40 DAT	27 Aug 2009	BBCH 83
010-0001	1	Forage 60 DAT	01 Sep 2009	BBCH 85
010-0002	2	Forage 60 DAT	01 Sep 2009	BBCH 85
010-0003	2	Forage 60 DAT	01 Sep 2009	BBCH 85
010-0004	3	Forage 60 DAT	01 Sep 2009	BBCH 85
010-0005	3	Forage 60 DAT	01 Sep 2009	BBCH 85
011-0001	1	Grain 90 DAT	02 Oct 2009	BBCH 99
011-0002	2	Grain 90 DAT	02 Oct 2009	BBCH 99
011-0003	2	Grain 90 DAT	02 Oct 2009	BBCH 99
011-0004	3	Grain 90 DAT	02 Oct 2009	BBCH 99
011-0005	3	Grain 90 DAT	02 Oct 2009	BBCH 99
011-0006	4	Grain 90 DAT	02 Oct 2009	BBCH 99
011-0007	4	Grain 90 DAT	02 Oct 2009	BBCH 99
011-0008	5	Grain 90 DAT	02 Oct 2009	BBCH 99
011-0009	5	Grain 90 DAT	02 Oct 2009	BBCH 99
012-0001	1	Bulk Grain 90 DAT	02 Oct 2009	BBCH 99
012-0002	4	Bulk Grain 90 DAT	02 Oct 2009	BBCH 99
012-0003	5	Bulk Grain 90 DAT	02 Oct 2009	BBCH 99
013-0001	1	Stover 90 DAT	02 Oct 2009	BBCH 99
013-0002	2	Stover 90 DAT	02 Oct 2009	BBCH 99
013-0003	2	Stover 90 DAT	02 Oct 2009	BBCH 99
013-0004	3	Stover 90 DAT	02 Oct 2009	BBCH 99
013-0005	3	Stover 90 DAT	02 Oct 2009	BBCH 99

Table 12: Crop Samples (Trial 04)

Sample No.	Treatment	Sampling Event – Nominal PHI	Date of Collection	Crop Growth Stage
014-0001	1	Forage 40 DAT	30 Aug 2009	BBCH 83
014-0002	2	Forage 40 DAT	30 Aug 2009	BBCH 83
014-0003	2	Forage 40 DAT	30 Aug 2009	BBCH 83
014-0004	3	Forage 40 DAT	30 Aug 2009	BBCH 83
014-0005	3	Forage 40 DAT	30 Aug 2009	BBCH 83
015-0001	1	Forage 46 DAT	01 Sep 2009	BBCH 83
015-0002	2	Forage 46 DAT	01 Sep 2009	BBCH 83
015-0003	2	Forage 46 DAT	01 Sep 2009	BBCH 83
015-0004	3	Forage 46 DAT	01 Sep 2009	BBCH 83
015-0005	3	Forage 46 DAT	01 Sep 2009	BBCH 83
016-0001	1	Forage 53 DAT	08 Sep 2009	BBCH 85
016-0002	2	Forage 53 DAT	08 Sep 2009	BBCH 85
016-0003	2	Forage 53 DAT	08 Sep 2009	BBCH 85
016-0004	3	Forage 53 DAT	08 Sep 2009	BBCH 85
016-0005	3	Forage 53 DAT	08 Sep 2009	BBCH 85
017-0001	1	Forage 60 DAT	15 Sep 2009	BBCH 87
017-0002	2	Forage 60 DAT	15 Sep 2009	BBCH 87
017-0003	2	Forage 60 DAT	15 Sep 2009	BBCH 87
017-0004	3	Forage 60 DAT	15 Sep 2009	BBCH 87
017-0005	3	Forage 60 DAT	15 Sep 2009	BBCH 87
018-0001	1	Forage 67 DAT	22 Sep 2009	BBCH 89
018-0002	2	Forage 67 DAT	22 Sep 2009	BBCH 89
018-0003	2	Forage 67 DAT	22 Sep 2009	BBCH 89
018-0004	3	Forage 67 DAT	22 Sep 2009	BBCH 89
018-0005	3	Forage 67 DAT	22 Sep 2009	BBCH 89
019-0001	1	Forage 74 DAT	29 Sep 2009	BBCH 90
019-0002	2	Forage 74 DAT	29 Sep 2009	BBCH 90
019-0003	2	Forage 74 DAT	29 Sep 2009	BBCH 90
019-0004	3	Forage 74 DAT	29 Sep 2009	BBCH 90
019-0005	3	Forage 74 DAT	29 Sep 2009	BBCH 90
020-0001	1	Grain 76 DAT	01 Oct 2009	BBCH 90
020-0002	2	Grain 76 DAT	01 Oct 2009	BBCH 90
020-0003	2	Grain 76 DAT	01 Oct 2009	BBCH 90
020-0004	3	Grain 76 DAT	01 Oct 2009	BBCH 90
020-0005	3	Grain 76 DAT	01 Oct 2009	BBCH 90
021-0001	1	Stover 76 DAT	01 Oct 2009	BBCH 90
021-0002	2	Stover 76 DAT	01 Oct 2009	BBCH 90
021-0003	2	Stover 76 DAT	01 Oct 2009	BBCH 90
021-0004	3	Stover 76 DAT	01 Oct 2009	BBCH 90
021-0005	3	Stover 76 DAT	01 Oct 2009	BBCH 90
022-0001	1	Grain 83 DAT	08 Oct 2009	BBCH 90
022-0002	2	Grain 83 DAT	08 Oct 2009	BBCH 90

Table 12: Crop Samples (Trial 04) (continued)

Sample No.	Treatment	Sampling Event – Nominal PHI	Date of Collection	Crop Growth Stage
022-0003	2	Grain 83 DAT	08 Oct 2009	BBCH 90
022-0004	3	Grain 83 DAT	08 Oct 2009	BBCH 90
022-0005	3	Grain 83 DAT	08 Oct 2009	BBCH 90
023-0001	1	Stover 83 DAT	08 Oct 2009	BBCH 90
023-0002	2	Stover 83 DAT	08 Oct 2009	BBCH 90
023-0003	2	Stover 83 DAT	08 Oct 2009	BBCH 90
023-0004	3	Stover 83 DAT	08 Oct 2009	BBCH 90
023-0005	3	Stover 83 DAT	08 Oct 2009	BBCH 90
024-0001	1	Grain 90 DAT	15 Oct 2009	BBCH 92
024-0002	2	Grain 90 DAT	15 Oct 2009	BBCH 92
024-0003	2	Grain 90 DAT	15 Oct 2009	BBCH 92
024-0004	3	Grain 90 DAT	15 Oct 2009	BBCH 92
024-0005	3	Grain 90 DAT	15 Oct 2009	BBCH 92
025-0001	1	Stover 90 DAT	15 Oct 2009	BBCH 92
025-0002	2	Stover 90 DAT	15 Oct 2009	BBCH 92
025-0003	2	Stover 90 DAT	15 Oct 2009	BBCH 92
025-0004	3	Stover 90 DAT	15 Oct 2009	BBCH 92
025-0005	3	Stover 90 DAT	15 Oct 2009	BBCH 92
026-0001	1	Grain 97 DAT	22 Oct 2009	BBCH 99
026-0002	2	Grain 97 DAT	22 Oct 2009	BBCH 99
026-0003	2	Grain 97 DAT	22 Oct 2009	BBCH 99
026-0004	3	Grain 97 DAT	22 Oct 2009	BBCH 99
026-0005	3	Grain 97 DAT	22 Oct 2009	BBCH 99
027-0001	1	Stover 97 DAT	22 Oct 2009	BBCH 99
027-0002	2	Stover 97 DAT	22 Oct 2009	BBCH 99
027-0003	2	Stover 97 DAT	22 Oct 2009	BBCH 99
027-0004	3	Stover 97 DAT	22 Oct 2009	BBCH 99
027-0005	3	Stover 97 DAT	22 Oct 2009	BBCH 99
028-0001	1	Grain 104 DAT	29 Oct 2009	BBCH 99
028-0002	2	Grain 104 DAT	29 Oct 2009	BBCH 99
028-0003	2	Grain 104 DAT	29 Oct 2009	BBCH 99
028-0004	3	Grain 104 DAT	29 Oct 2009	BBCH 99
028-0005	3	Grain 104 DAT	29 Oct 2009	BBCH 99
029-0001	1	Stover 104 DAT	29 Oct 2009	BBCH 99
029-0002	2	Stover 104 DAT	29 Oct 2009	BBCH 99
029-0003	2	Stover 104 DAT	29 Oct 2009	BBCH 99
029-0004	3	Stover 104 DAT	29 Oct 2009	BBCH 99
029-0005	3	Stover 104 DAT	29 Oct 2009	BBCH 99

Table 13: Crop Samples (Trial 05)

Sample No.	Treatment	Sampling Event – Nominal PHI	Date of Collection	Crop Growth Stage
030-0001	1	Forage 40 DAT	18 Aug 2009	BBCH 73
030-0002	2	Forage 40 DAT	18 Aug 2009	BBCH 73
030-0003	2	Forage 40 DAT	18 Aug 2009	BBCH 73
030-0004	3	Forage 40 DAT	18 Aug 2009	BBCH 73
030-0005	3	Forage 40 DAT	18 Aug 2009	BBCH 73
031-0001	1	Forage - late dough/early dent (early dent)	14 Sep 2009	BBCH 85
031-0002	2		14 Sep 2009	BBCH 85
031-0003	2		14 Sep 2009	BBCH 85
031-0004	3		14 Sep 2009	BBCH 85
031-0005	3		14 Sep 2009	BBCH 85
032-0001	1	Grain - Mature	19 Nov 2009	BBCH 99
032-0002	2		19 Nov 2009	BBCH 99
032-0003	2		19 Nov 2009	BBCH 99
032-0004	3		19 Nov 2009	BBCH 99
032-0005	3		19 Nov 2009	BBCH 99
033-0001	1	Stover - Mature	19 Nov 2009	BBCH 99
033-0002	2		19 Nov 2009	BBCH 99
033-0003	2		19 Nov 2009	BBCH 99
033-0004	3		19 Nov 2009	BBCH 99
033-0005	3		19 Nov 2009	BBCH 99

Table 14: Crop Samples (Trial 06)

Sample No.	Treatment	Sampling Event – Nominal PHI	Date of Collection	Crop Growth Stage
034-0001	1	Forage 40 DAT	04 Sep 2009	BBCH 75
034-0002	2	Forage 40 DAT	04 Sep 2009	BBCH 75
034-0003	2	Forage 40 DAT	04 Sep 2009	BBCH 75
034-0004	3	Forage 40 DAT	04 Sep 2009	BBCH 75
034-0005	3	Forage 40 DAT	04 Sep 2009	BBCH 75
035-0001	1	Forage 60 DAT	29 Sep 2009	BBCH 85
035-0002	2	Forage 60 DAT	29 Sep 2009	BBCH 85
035-0003	2	Forage 60 DAT	29 Sep 2009	BBCH 85
035-0004	3	Forage 60 DAT	29 Sep 2009	BBCH 85
035-0005	3	Forage 60 DAT	29 Sep 2009	R6
036-0001	1	Grain 90 DAT	23 Nov 2009	R6
036-0002	2	Grain 90 DAT	23 Nov 2009	R6
036-0003	2	Grain 90 DAT	23 Nov 2009	R6
036-0004	3	Grain 90 DAT	23 Nov 2009	R6
036-0005	3	Grain 90 DAT	23 Nov 2009	R6
037-0001	1	Stover 90 DAT	23 Nov 2009	R6
037-0002	2	Stover 90 DAT	23 Nov 2009	R6
037-0003	2	Stover 90 DAT	23 Nov 2009	R6
037-0004	3	Stover 90 DAT	23 Nov 2009	R6
037-0005	3	Stover 90 DAT	23 Nov 2009	R6

Table 15: Crop Samples (Trial 07)

Sample No.	Treatment	Sampling Event – Nominal PHI	Date of Collection	Crop Growth Stage
038-0001	1	Forage 40 DAT	26 Aug 2009	BBCH 75-79
038-0002	2	Forage 40 DAT	26 Aug 2009	BBCH 75-79
038-0003	2	Forage 40 DAT	26 Aug 2009	BBCH 75-79
038-0004	3	Forage 40 DAT	26 Aug 2009	BBCH 75-79
038-0005	3	Forage 40 DAT	26 Aug 2009	BBCH 75-79
039-0001	1	Forage 60 DAT	14 Sep 2009	BBCH 85
039-0002	2	Forage 60 DAT	14 Sep 2009	BBCH 85
039-0003	2	Forage 60 DAT	14 Sep 2009	BBCH 85
039-0004	3	Forage 60 DAT	14 Sep 2009	BBCH 85
039-0005	3	Forage 60 DAT	14 Sep 2009	BBCH 85
040-0001	1	Grain 90 DAT	01 Dec 2009	BBCH 97-99
040-0002	2	Grain 90 DAT	01 Dec 2009	BBCH 97-99
040-0003	2	Grain 90 DAT	01 Dec 2009	BBCH 97-99
040-0004	3	Grain 90 DAT	01 Dec 2009	BBCH 97-99
040-0005	3	Grain 90 DAT	01 Dec 2009	BBCH 97-99
041-0001	1	Stover 90 DAT	01 Dec 2009	BBCH 97-99
041-0002	2	Stover 90 DAT	01 Dec 2009	BBCH 97-99
041-0003	2	Stover 90 DAT	01 Dec 2009	BBCH 97-99
041-0004	3	Stover 90 DAT	01 Dec 2009	BBCH 97-99
041-0005	3	Stover 90 DAT	01 Dec 2009	BBCH 97-99

Table 16: Crop Samples (Trial 08)

Sample No.	Treatment	Sampling Event – Nominal PHI	Date of Collection	Crop Growth Stage
042-0001	1	Forage 40 DAT	18 Sep 2009	BBCH 71
042-0002	2	Forage 40 DAT	18 Sep 2009	BBCH 71
042-0003	2	Forage 40 DAT	18 Sep 2009	BBCH 71
042-0004	3	Forage 40 DAT	18 Sep 2009	BBCH 71
042-0005	3	Forage 40 DAT	18 Sep 2009	BBCH 71
043-0001	1	Forage 60 DAT	25 Sep 2009	BBCH 75
043-0002	2	Forage 60 DAT	25 Sep 2009	BBCH 75
043-0003	2	Forage 60 DAT	25 Sep 2009	BBCH 75
043-0004	3	Forage 60 DAT	25 Sep 2009	BBCH 75
043-0005	3	Forage 60 DAT	25 Sep 2009	BBCH 75
044-0001	1	Grain 90 DAT	21 Nov 2009	BBCH 99
044-0002	2	Grain 90 DAT	21 Nov 2009	BBCH 99
044-0003	2	Grain 90 DAT	21 Nov 2009	BBCH 99
044-0004	3	Grain 90 DAT	21 Nov 2009	BBCH 99
044-0005	3	Grain 90 DAT	21 Nov 2009	BBCH 99
045-0001	1	Stover 90 DAT	21 Nov 2009	BBCH 99
045-0002	2	Stover 90 DAT	21 Nov 2009	BBCH 99
045-0003	2	Stover 90 DAT	21 Nov 2009	BBCH 99
045-0004	3	Stover 90 DAT	21 Nov 2009	BBCH 99
045-0005	3	Stover 90 DAT	21 Nov 2009	BBCH 99

Table 17: Crop Samples (Trial 09)

Sample No.	Treatment	Sampling Event – Nominal PHI	Date of Collection	Crop Growth Stage
046-0001	1	Forage 40 DAT	18 Sep 2009	BBCH 71
046-0002	2	Forage 40 DAT	18 Sep 2009	BBCH 71
046-0003	2	Forage 40 DAT	18 Sep 2009	BBCH 71
046-0004	3	Forage 40 DAT	18 Sep 2009	BBCH 71
046-0005	3	Forage 40 DAT	18 Sep 2009	BBCH 71
047-0001	1	Forage 60 DAT	25 Sep 2009	BBCH 75
047-0002	2	Forage 60 DAT	25 Sep 2009	BBCH 75
047-0003	2	Forage 60 DAT	25 Sep 2009	BBCH 75
047-0004	3	Forage 60 DAT	25 Sep 2009	BBCH 75
047-0005	3	Forage 60 DAT	25 Sep 2009	BBCH 75
048-0001	1	Grain 90 DAT	21 Nov 2009	BBCH 99
048-0002	2	Grain 90 DAT	21 Nov 2009	BBCH 99
048-0003	2	Grain 90 DAT	21 Nov 2009	BBCH 99
048-0004	3	Grain 90 DAT	21 Nov 2009	BBCH 99
048-0005	3	Grain 90 DAT	21 Nov 2009	BBCH 99
049-0001	1	Stover 90 DAT	21 Nov 2009	BBCH 99
049-0002	2	Stover 90 DAT	21 Nov 2009	BBCH 99
049-0003	2	Stover 90 DAT	21 Nov 2009	BBCH 99
049-0004	3	Stover 90 DAT	21 Nov 2009	BBCH 99
049-0005	3	Stover 90 DAT	21 Nov 2009	BBCH 99

Table 18: Crop Samples (Trial 10)

Sample No.	Treatment	Sampling Event – Nominal PHI	Date of Collection	Crop Growth Stage
050-0001	1	Forage 40 DAT	22 Sep 2009	BBCH 71
050-0002	2	Forage 40 DAT	22 Sep 2009	BBCH 71
050-0003	2	Forage 40 DAT	22 Sep 2009	BBCH 71
050-0004	3	Forage 40 DAT	22 Sep 2009	BBCH 71
050-0005	3	Forage 40 DAT	22 Sep 2009	BBCH 71
051-0001	1	Forage 60 DAT	25 Sep 2009	BBCH 75
051-0002	2	Forage 60 DAT	25 Sep 2009	BBCH 75
051-0003	2	Forage 60 DAT	25 Sep 2009	BBCH 75
051-0004	3	Forage 60 DAT	25 Sep 2009	BBCH 75
051-0005	3	Forage 60 DAT	25 Sep 2009	BBCH 75
052-0001	1	Grain 90 DAT	21 Nov 2009	BBCH 99
052-0002	2	Grain 90 DAT	21 Nov 2009	BBCH 99
052-0003	2	Grain 90 DAT	21 Nov 2009	BBCH 99
052-0004	3	Grain 90 DAT	21 Nov 2009	BBCH 99
052-0005	3	Grain 90 DAT	21 Nov 2009	BBCH 99
053-0001	1	Stover 90 DAT	21 Nov 2009	BBCH 99
053-0002	2	Stover 90 DAT	21 Nov 2009	BBCH 99
053-0003	2	Stover 90 DAT	21 Nov 2009	BBCH 99
053-0004	3	Stover 90 DAT	21 Nov 2009	BBCH 99
053-0005	3	Stover 90 DAT	21 Nov 2009	BBCH 99

Table 19: Crop Samples (Trial 11)

Sample No.	Treatment	Sampling Event – Nominal PHI	Date of Collection	Crop Growth Stage
054-0001	1	Forage 40 DAT	14 Sep 2009	BBCH 50
054-0002	2	Forage 40 DAT	14 Sep 2009	BBCH 50
054-0003	2	Forage 40 DAT	14 Sep 2009	BBCH 50
054-0004	3	Forage 40 DAT	14 Sep 2009	BBCH 50
054-0005	3	Forage 40 DAT	14 Sep 2009	BBCH 50
055-0001	1	Forage 46 DAT	20 Sep 2009	BBCH 55
055-0002	2	Forage 46 DAT	20 Sep 2009	BBCH 55
055-0003	2	Forage 46 DAT	20 Sep 2009	BBCH 55
055-0004	3	Forage 46 DAT	20 Sep 2009	BBCH 55
055-0005	3	Forage 46 DAT	20 Sep 2009	BBCH 55
056-0001	1	Forage 53 DAT	27 Sep 2009	BBCH 59
056-0002	2	Forage 53 DAT	27 Sep 2009	BBCH 59
056-0003	2	Forage 53 DAT	27 Sep 2009	BBCH 59
056-0004	3	Forage 53 DAT	27 Sep 2009	BBCH 59
056-0005	3	Forage 53 DAT	27 Sep 2009	BBCH 59
057-0001	1	Forage 60 DAT	03 Oct 2009	BBCH 67
057-0002	2	Forage 60 DAT	03 Oct 2009	BBCH 67
057-0003	2	Forage 60 DAT	03 Oct 2009	BBCH 67
057-0004	3	Forage 60 DAT	03 Oct 2009	BBCH 67
057-0005	3	Forage 60 DAT	03 Oct 2009	BBCH 67
058-0001	1	Forage 67 DAT	10 Oct 2009	BBCH 75
058-0002	2	Forage 67 DAT	10 Oct 2009	BBCH 75
058-0003	2	Forage 67 DAT	10 Oct 2009	BBCH 75
058-0004	3	Forage 67 DAT	10 Oct 2009	BBCH 75
058-0005	3	Forage 67 DAT	10 Oct 2009	BBCH 75
059-0001	1	Forage 74 DAT	17 Oct 2009	BBCH 85
059-0002	2	Forage 74 DAT	17 Oct 2009	BBCH 85
059-0003	2	Forage 74 DAT	17 Oct 2009	BBCH 85
059-0004	3	Forage 74 DAT	17 Oct 2009	BBCH 85
059-0005	3	Forage 74 DAT	17 Oct 2009	BBCH 85
060-0001	1	Grain 76 DAT	20 Oct 2009	BBCH 89
060-0002	2	Grain 76 DAT	20 Oct 2009	BBCH 89
060-0003	2	Grain 76 DAT	20 Oct 2009	BBCH 89
060-0004	3	Grain 76 DAT	20 Oct 2009	BBCH 89
060-0005	3	Grain 76 DAT	20 Oct 2009	BBCH 89
061-0001	1	Stover 76 DAT	20 Oct 2009	BBCH 89
061-0002	2	Stover 76 DAT	20 Oct 2009	BBCH 89
061-0003	2	Stover 76 DAT	20 Oct 2009	BBCH 89
061-0004	3	Stover 76 DAT	20 Oct 2009	BBCH 89
061-0005	3	Stover 76 DAT	20 Oct 2009	BBCH 89
062-0001	1	Grain 83 DAT	27 Oct 2009	BBCH 92
062-0002	2	Grain 83 DAT	27 Oct 2009	BBCH 92

Table 19: Crop Samples (Trial 11) (continued)

Sample No.	Treatment	Sampling Event – Nominal PHI	Date of Collection	Crop Growth Stage
062-0003	2	Grain 83 DAT	27 Oct 2009	BBCH 92
062-0004	3	Grain 83 DAT	27 Oct 2009	BBCH 92
062-0005	3	Grain 83 DAT	27 Oct 2009	BBCH 92
063-0001	1	Stover 83 DAT	27 Oct 2009	BBCH 92
063-0002	2	Stover 83 DAT	27 Oct 2009	BBCH 92
063-0003	2	Stover 83 DAT	27 Oct 2009	BBCH 92
063-0004	3	Stover 83 DAT	27 Oct 2009	BBCH 92
063-0005	3	Stover 83 DAT	27 Oct 2009	BBCH 92
064-0001	1	Grain 90 DAT	03 Nov 2009	BBCH 94
064-0002	2	Grain 90 DAT	03 Nov 2009	BBCH 94
064-0003	2	Grain 90 DAT	03 Nov 2009	BBCH 94
064-0004	3	Grain 90 DAT	03 Nov 2009	BBCH 94
064-0005	3	Grain 90 DAT	03 Nov 2009	BBCH 94
065-0001	1	Stover 90 DAT	03 Nov 2009	BBCH 94
065-0002	2	Stover 90 DAT	03 Nov 2009	BBCH 94
065-0003	2	Stover 90 DAT	03 Nov 2009	BBCH 94
065-0004	3	Stover 90 DAT	03 Nov 2009	BBCH 94
065-0005	3	Stover 90 DAT	03 Nov 2009	BBCH 94
066-0001	1	Grain 97 DAT	10 Nov 2009	BBCH 96
066-0002	2	Grain 97 DAT	10 Nov 2009	BBCH 96
066-0003	2	Grain 97 DAT	10 Nov 2009	BBCH 96
066-0004	3	Grain 97 DAT	10 Nov 2009	BBCH 96
066-0005	3	Grain 97 DAT	10 Nov 2009	BBCH 96
067-0001	1	Stover 97 DAT	10 Nov 2009	BBCH 96
067-0002	2	Stover 97 DAT	10 Nov 2009	BBCH 96
067-0003	2	Stover 97 DAT	10 Nov 2009	BBCH 96
067-0004	3	Stover 97 DAT	10 Nov 2009	BBCH 96
067-0005	3	Stover 97 DAT	10 Nov 2009	BBCH 96
068-0001	1	Grain 104 DAT	17 Nov 2009	BBCH 98
068-0002	2	Grain 104 DAT	17 Nov 2009	BBCH 98
068-0003	2	Grain 104 DAT	17 Nov 2009	BBCH 98
068-0004	3	Grain 104 DAT	17 Nov 2009	BBCH 98
068-0005	3	Grain 104 DAT	17 Nov 2009	BBCH 98
069-0001	1	Stover 104 DAT	17 Nov 2009	BBCH 98
069-0002	2	Stover 104 DAT	17 Nov 2009	BBCH 98
069-0003	2	Stover 104 DAT	17 Nov 2009	BBCH 98
069-0004	3	Stover 104 DAT	17 Nov 2009	BBCH 98
069-0005	3	Stover 104 DAT	17 Nov 2009	BBCH 98

Table 20: Crop Samples (Trial 12)

Sample No.	Treatment	Sampling Event – Nominal PHI	Date of Collection	Crop Growth Stage
070-0001	1	Forage 40 DAT	17 Sep 2009	BBCH 75-79
070-0002	2	Forage 40 DAT	17 Sep 2009	BBCH 75-79
070-0003	2	Forage 40 DAT	17 Sep 2009	BBCH 75-79
070-0004	3	Forage 40 DAT	17 Sep 2009	BBCH 75-79
070-0005	3	Forage 40 DAT	17 Sep 2009	BBCH 75-79
071-0001	1	Forage 60 DAT	02 Oct 2009	BBCH 80-85
071-0002	2	Forage 60 DAT	02 Oct 2009	BBCH 80-85
071-0003	2	Forage 60 DAT	02 Oct 2009	BBCH 80-85
071-0004	3	Forage 60 DAT	02 Oct 2009	BBCH 80-85
071-0005	3	Forage 60 DAT	02 Oct 2009	BBCH 80-85
072-0001	1	Grain 90 DAT	06 Nov 2009	BBCH 97
072-0002	2	Grain 90 DAT	06 Nov 2009	BBCH 97
072-0003	2	Grain 90 DAT	06 Nov 2009	BBCH 97
072-0004	3	Grain 90 DAT	06 Nov 2009	BBCH 97
072-0005	3	Grain 90 DAT	06 Nov 2009	BBCH 97
073-0001	1	Stover 90 DAT	06 Nov 2009	BBCH 97
073-0002	2	Stover 90 DAT	06 Nov 2009	BBCH 97
073-0003	2	Stover 90 DAT	06 Nov 2009	BBCH 97
073-0004	3	Stover 90 DAT	06 Nov 2009	BBCH 97
073-0005	3	Stover 90 DAT	06 Nov 2009	BBCH 97

Table 21: Crop Samples (Trial 13)

Sample No.	Treatment	Sampling Event – Nominal PHI	Date of Collection	Crop Growth Stage
074-0001	1	Forage 40 DAT	14 Sep 2009	BBCH 60
074-0002	2	Forage 40 DAT	14 Sep 2009	BBCH 60
074-0003	2	Forage 40 DAT	14 Sep 2009	BBCH 60
074-0004	3	Forage 40 DAT	14 Sep 2009	BBCH 60
074-0005	3	Forage 40 DAT	14 Sep 2009	BBCH 60
075-0001	1	Forage 60 DAT	04 Oct 2009	BBCH 85
075-0002	2	Forage 60 DAT	04 Oct 2009	BBCH 85
075-0003	2	Forage 60 DAT	04 Oct 2009	BBCH 85
075-0004	3	Forage 60 DAT	04 Oct 2009	BBCH 85
075-0005	3	Forage 60 DAT	04 Oct 2009	BBCH 85
076-0001	1	Grain 90 DAT	03 Nov 2009	BBCH 97
076-0002	2	Grain 90 DAT	03 Nov 2009	BBCH 97
076-0003	2	Grain 90 DAT	03 Nov 2009	BBCH 97
076-0004	3	Grain 90 DAT	03 Nov 2009	BBCH 97
076-0005	3	Grain 90 DAT	03 Nov 2009	BBCH 97
077-0001	1	Stover 90 DAT	03 Nov 2009	BBCH 97
077-0002	2	Stover 90 DAT	03 Nov 2009	BBCH 97
077-0003	2	Stover 90 DAT	03 Nov 2009	BBCH 97
077-0004	3	Stover 90 DAT	03 Nov 2009	BBCH 97
077-0005	3	Stover 90 DAT	03 Nov 2009	BBCH 97

Table 22: Crop Samples (Trial 14)

Sample No.	Treatment	Sampling Event – Nominal PHI	Date of Collection	Crop Growth Stage
079-0001	1	Forage 60 DAT (a)	01 Sep 2009	BBCH 85
079-0002	2	Forage 60 DAT	01 Sep 2009	BBCH 85
079-0003	2	Forage 60 DAT	01 Sep 2009	BBCH 85
079-0004	3	Forage 60 DAT	01 Sep 2009	BBCH 85
079-0005	3	Forage 60 DAT	01 Sep 2009	BBCH 85
080-0001	1	Grain 90 DAT	17 Nov 2009	BBCH 89
080-0002	2	Grain 90 DAT	17 Nov 2009	BBCH 89
080-0003	2	Grain 90 DAT	17 Nov 2009	BBCH 89
080-0004	3	Grain 90 DAT	17 Nov 2009	BBCH 89
080-0005	3	Grain 90 DAT	17 Nov 2009	BBCH 89
080-0006	4	Grain 90 DAT	17 Nov 2009	BBCH 89
080-0007	4	Grain 90 DAT	17 Nov 2009	BBCH 89
080-0008	5	Grain 90 DAT	17 Nov 2009	BBCH 89
080-0009	5	Grain 90 DAT	17 Nov 2009	BBCH 89
081-0001	1	Bulk Grain 90 DAT	17 Nov 2009	BBCH 89
081-0002	4	Bulk Grain 90 DAT	17 Nov 2009	BBCH 89
081-0003	5	Bulk Grain 90 DAT	17 Nov 2009	BBCH 89
082-0001	1	Stover 90 DAT	17 Nov 2009	BBCH 89
082-0002	2	Stover 90 DAT	17 Nov 2009	BBCH 89
082-0003	2	Stover 90 DAT	17 Nov 2009	BBCH 89
082-0004	3	Stover 90 DAT	17 Nov 2009	BBCH 89
082-0005	3	Stover 90 DAT	17 Nov 2009	BBCH 89

(a) 40 DAT forage not collected

Table 23: Crop Samples (Trial 15)

Sample No.	Treatment	Sampling Event – Nominal PHI	Date of Collection	Crop Growth Stage
083-0001	1	Forage 40 DAT	31 Aug 2009	BBCH 67
083-0002	2	Forage 40 DAT	31 Aug 2009	BBCH 67
083-0003	2	Forage 40 DAT	31 Aug 2009	BBCH 67
083-0004	3	Forage 40 DAT	31 Aug 2009	BBCH 67
083-0005	3	Forage 40 DAT	31 Aug 2009	BBCH 67
084-0001	1	Forage 60 DAT	24 Sep 2009	BBCH 85
084-0002	2	Forage 60 DAT	24 Sep 2009	BBCH 85
084-0003	2	Forage 60 DAT	24 Sep 2009	BBCH 85
084-0004	3	Forage 60 DAT	24 Sep 2009	BBCH 85
084-0005	3	Forage 60 DAT	24 Sep 2009	BBCH 85
085-0001	1	Grain 90 DAT	17 Oct 2009	BBCH 99
085-0002	2	Grain 90 DAT	17 Oct 2009	BBCH 99
085-0003	2	Grain 90 DAT	17 Oct 2009	BBCH 99
085-0004	3	Grain 90 DAT	17 Oct 2009	BBCH 99
085-0005	3	Grain 90 DAT	17 Oct 2009	BBCH 99
086-0001	1	Stover 90 DAT	17 Oct 2009	BBCH 99
086-0002	2	Stover 90 DAT	17 Oct 2009	BBCH 99
086-0003	2	Stover 90 DAT	17 Oct 2009	BBCH 99
086-0004	3	Stover 90 DAT	17 Oct 2009	BBCH 99
086-0005	3	Stover 90 DAT	17 Oct 2009	BBCH 99

Table 24: Crop Samples (Trial 16)

Sample No.	Treatment	Sampling Event – Nominal PHI	Date of Collection	Crop Growth Stage
087-0001	1	Forage 40 DAT	03 Sep 2009	BBCH 65-67
087-0002	2	Forage 40 DAT	03 Sep 2009	BBCH 65-67
087-0003	2	Forage 40 DAT	03 Sep 2009	BBCH 65-67
087-0004	3	Forage 40 DAT	03 Sep 2009	BBCH 65-67
087-0005	3	Forage 40 DAT	03 Sep 2009	BBCH 65-67
088-0001	1	Forage 60 DAT	29 Sep 2009	BBCH 85
088-0002	2	Forage 60 DAT	29 Sep 2009	BBCH 85
088-0003	2	Forage 60 DAT	29 Sep 2009	BBCH 85
088-0004	3	Forage 60 DAT	29 Sep 2009	BBCH 85
088-0005	3	Forage 60 DAT	29 Sep 2009	BBCH 85
089-0001	1	Grain 90 DAT	28 Oct 2009	BBCH 99
089-0002	2	Grain 90 DAT	28 Oct 2009	BBCH 99
089-0003	2	Grain 90 DAT	28 Oct 2009	BBCH 99
089-0004	3	Grain 90 DAT	28 Oct 2009	BBCH 99
089-0005	3	Grain 90 DAT	28 Oct 2009	BBCH 99
090-0001	1	Stover 90 DAT	28 Oct 2009	BBCH 99
090-0002	2	Stover 90 DAT	28 Oct 2009	BBCH 99
090-0003	2	Stover 90 DAT	28 Oct 2009	BBCH 99
090-0004	3	Stover 90 DAT	28 Oct 2009	BBCH 99
090-0005	3	Stover 90 DAT	28 Oct 2009	BBCH 99

Table 25: Crop Samples (Trial 17)

Sample No.	Treatment	Sampling Event – Nominal PHI	Date of Collection	Crop Growth Stage
091-0001	1	Forage 40 DAT	13 Aug 2009	BBCH 71
091-0002	2	Forage 40 DAT	13 Aug 2009	BBCH 71
091-0003	2	Forage 40 DAT	13 Aug 2009	BBCH 71
091-0004	3	Forage 40 DAT	13 Aug 2009	BBCH 71
091-0005	3	Forage 40 DAT	13 Aug 2009	BBCH 71
092-0001	1	Forage 60 DAT	04 Sep 2009	BBCH 85
092-0002	2	Forage 60 DAT	04 Sep 2009	BBCH 85
092-0003	2	Forage 60 DAT	04 Sep 2009	BBCH 85
092-0004	3	Forage 60 DAT	04 Sep 2009	BBCH 85
092-0005	3	Forage 60 DAT	04 Sep 2009	BBCH 85
093-0001	1	Grain 90 DAT	17 Oct 2009	BBCH 97
093-0002	2	Grain 90 DAT	17 Oct 2009	BBCH 97
093-0003	2	Grain 90 DAT	17 Oct 2009	BBCH 97
093-0004	3	Grain 90 DAT	17 Oct 2009	BBCH 97
093-0005	3	Grain 90 DAT	17 Oct 2009	BBCH 97
094-0001	1	Stover 90 DAT	17 Oct 2009	BBCH 97
094-0002	2	Stover 90 DAT	17 Oct 2009	BBCH 97
094-0003	2	Stover 90 DAT	17 Oct 2009	BBCH 97
094-0004	3	Stover 90 DAT	17 Oct 2009	BBCH 97
094-0005	3	Stover 90 DAT	17 Oct 2009	BBCH 97

Table 26: Crop Samples (Trial 18)

Sample No.	Treatment	Sampling Event – Nominal PHI	Date of Collection	Crop Growth Stage
095-0001	1	Forage 40 DAT	21 Aug 2009	BBCH 85
095-0002	2	Forage 40 DAT	21 Aug 2009	BBCH 85
095-0003	2	Forage 40 DAT	21 Aug 2009	BBCH 85
095-0004	3	Forage 40 DAT	21 Aug 2009	BBCH 85
095-0005	3	Forage 40 DAT	21 Aug 2009	BBCH 85
096-0001	1	Forage 60 DAT	03 Sep 2009	BBCH 87
096-0002	2	Forage 60 DAT	03 Sep 2009	BBCH 87
096-0003	2	Forage 60 DAT	03 Sep 2009	BBCH 87
096-0004	3	Forage 60 DAT	03 Sep 2009	BBCH 87
096-0005	3	Forage 60 DAT	03 Sep 2009	BBCH 87
097-0001	1	Grain 90 DAT	20 Oct 2009	BBCH 89
097-0002	2	Grain 90 DAT	20 Oct 2009	BBCH 89
097-0003	2	Grain 90 DAT	20 Oct 2009	BBCH 89
097-0004	3	Grain 90 DAT	20 Oct 2009	BBCH 89
097-0005	3	Grain 90 DAT	20 Oct 2009	BBCH 89
098-0001	1	Stover 90 DAT	20 Oct 2009	BBCH 89
098-0002	2	Stover 90 DAT	20 Oct 2009	BBCH 89
098-0003	2	Stover 90 DAT	20 Oct 2009	BBCH 89
098-0004	3	Stover 90 DAT	20 Oct 2009	BBCH 89
098-0005	3	Stover 90 DAT	20 Oct 2009	BBCH 89

Table 27: Crop Samples (Trial 19)

Sample No.	Treatment	Sampling Event – Nominal PHI	Date of Collection	Crop Growth Stage
099-0001	1	Forage 40 DAT	02 Sep 2009	Not recorded correctly
099-0002	2	Forage 40 DAT	02 Sep 2009	Not recorded correctly
099-0003	2	Forage 40 DAT	02 Sep 2009	Not recorded correctly
099-0004	3	Forage 40 DAT	02 Sep 2009	Not recorded correctly
099-0005	3	Forage 40 DAT	02 Sep 2009	Not recorded correctly
100-0001	1	Forage 60 DAT	29 Sep 2009	BBCH 85
100-0002	2	Forage 60 DAT	29 Sep 2009	BBCH 85
100-0003	2	Forage 60 DAT	29 Sep 2009	BBCH 85
100-0004	3	Forage 60 DAT	29 Sep 2009	BBCH 85
100-0005	3	Forage 60 DAT	29 Sep 2009	BBCH 85
101-0001	1	Grain 90 DAT	21 Nov 2009	BBCH 93
101-0002	2	Grain 90 DAT	21 Nov 2009	BBCH 93
101-0003	2	Grain 90 DAT	21 Nov 2009	BBCH 93
101-0004	3	Grain 90 DAT	21 Nov 2009	BBCH 93
101-0005	3	Grain 90 DAT	21 Nov 2009	BBCH 93
102-0001	1	Stover 90 DAT	21 Nov 2009	BBCH 93
102-0002	2	Stover 90 DAT	21 Nov 2009	BBCH 93
102-0003	2	Stover 90 DAT	21 Nov 2009	BBCH 93
102-0004	3	Stover 90 DAT	21 Nov 2009	BBCH 93
102-0005	3	Stover 90 DAT	21 Nov 2009	BBCH 93

Table 28: Crop Samples (Trial 20)

Sample No.	Treatment	Sampling Event – Nominal PHI	Date of Collection	Crop Growth Stage
103-0001	1	Forage 40 DAT	01 Sep 2009	BBCH 79
103-0002	2	Forage 40 DAT	01 Sep 2009	BBCH 79
103-0003	2	Forage 40 DAT	01 Sep 2009	BBCH 79
103-0004	3	Forage 40 DAT	01 Sep 2009	BBCH 79
103-0005	3	Forage 40 DAT	01 Sep 2009	BBCH 79
104-0001	1	Forage 60 DAT	29 Sep 2009	BBCH 85
104-0002	2	Forage 60 DAT	29 Sep 2009	BBCH 85
104-0003	2	Forage 60 DAT	29 Sep 2009	BBCH 85
104-0004	3	Forage 60 DAT	29 Sep 2009	BBCH 85
104-0005	3	Forage 60 DAT	29 Sep 2009	BBCH 85
105-0001	1	Grain 90 DAT	21 Nov 2009	BBCH 93
105-0002	2	Grain 90 DAT	21 Nov 2009	BBCH 93
105-0003	2	Grain 90 DAT	21 Nov 2009	BBCH 93
105-0004	3	Grain 90 DAT	21 Nov 2009	BBCH 93
105-0005	3	Grain 90 DAT	21 Nov 2009	BBCH 93
106-0001	1	Stover 90 DAT	21 Nov 2009	BBCH 93
106-0002	2	Stover 90 DAT	21 Nov 2009	BBCH 93
106-0003	2	Stover 90 DAT	21 Nov 2009	BBCH 93
106-0004	3	Stover 90 DAT	21 Nov 2009	BBCH 93
106-0005	3	Stover 90 DAT	21 Nov 2009	BBCH 93

Table 29: Crop Samples (Trial 21)

Sample No.	Treatment	Sampling Event – Nominal PHI	Date of Collection	Crop Growth Stage
107-0001	1	Forage 40 DAT	15 Sep 2009	BBCH 79
107-0002	2	Forage 40 DAT	15 Sep 2009	BBCH 79
107-0003	2	Forage 40 DAT	15 Sep 2009	BBCH 79
107-0004	3	Forage 40 DAT	15 Sep 2009	BBCH 79
107-0005	3	Forage 40 DAT	15 Sep 2009	BBCH 79
108-0001	1	Forage 60 DAT	30 Sep 2009	BBCH 85
108-0002	2	Forage 60 DAT	30 Sep 2009	BBCH 85
108-0003	2	Forage 60 DAT	30 Sep 2009	BBCH 85
108-0004	3	Forage 60 DAT	30 Sep 2009	BBCH 85
108-0005	3	Forage 60 DAT	30 Sep 2009	BBCH 85
109-0001	1	Grain 90 DAT	02 Nov 2009	BBCH 99
109-0002	2	Grain 90 DAT	02 Nov 2009	BBCH 99
109-0003	2	Grain 90 DAT	02 Nov 2009	BBCH 99
109-0004	3	Grain 90 DAT	02 Nov 2009	BBCH 99
109-0005	3	Grain 90 DAT	02 Nov 2009	BBCH 99
110-0001	1	Stover 90 DAT	02 Nov 2009	BBCH 99
110-0002	2	Stover 90 DAT	02 Nov 2009	BBCH 99
110-0003	2	Stover 90 DAT	02 Nov 2009	BBCH 99
110-0004	3	Stover 90 DAT	02 Nov 2009	BBCH 99
110-0005	3	Stover 90 DAT	02 Nov 2009	BBCH 99

Table 30: Crop Samples (Trial 22)

Sample No.	Treatment	Sampling Event – Nominal PHI	Date of Collection	Crop Growth Stage
111-0001	1	Forage 40 DAT	22 Aug 2009	BBCH 71
111-0002	2	Forage 40 DAT	22 Aug 2009	BBCH 71
111-0003	2	Forage 40 DAT	22 Aug 2009	BBCH 71
111-0004	3	Forage 40 DAT	22 Aug 2009	BBCH 71
111-0005	3	Forage 40 DAT	22 Aug 2009	BBCH 71
112-0001	1	Forage 46 DAT	29 Aug 2009	BBCH 73
112-0002	2	Forage 46 DAT	29 Aug 2009	BBCH 73
112-0003	2	Forage 46 DAT	29 Aug 2009	BBCH 73
112-0004	3	Forage 46 DAT	29 Aug 2009	BBCH 73
112-0005	3	Forage 46 DAT	29 Aug 2009	BBCH 73
113-0001	1	Forage 53 DAT	04 Sep 2009	BBCH 74
113-0002	2	Forage 53 DAT	04 Sep 2009	BBCH 74
113-0003	2	Forage 53 DAT	04 Sep 2009	BBCH 74
113-0004	3	Forage 53 DAT	04 Sep 2009	BBCH 74
113-0005	3	Forage 53 DAT	04 Sep 2009	BBCH 74
114-0001	1	Forage 60 DAT	10 Sep 2009	BBCH 83
114-0002	2	Forage 60 DAT	10 Sep 2009	BBCH 83
114-0003	2	Forage 60 DAT	10 Sep 2009	BBCH 83
114-0004	3	Forage 60 DAT	10 Sep 2009	BBCH 83

Table 30: Crop Samples (Trial 22) (continued)

Sample No.	Treatment	Sampling Event – Nominal PHI	Date of Collection	Crop Growth Stage
114-0005	3	Forage 60 DAT	10 Sep 2009	BBCH 83
115-0001	1	Forage 67 DAT	16 Sep 2009	BBCH 85
115-0002	2	Forage 67 DAT	16 Sep 2009	BBCH 85
115-0003	2	Forage 67 DAT	16 Sep 2009	BBCH 85
115-0004	3	Forage 67 DAT	16 Sep 2009	BBCH 85
115-0005	3	Forage 67 DAT	16 Sep 2009	BBCH 85
116-0001	1	Forage 74 DAT	22 Sep 2009	BBCH 87
116-0002	2	Forage 74 DAT	22 Sep 2009	BBCH 87
116-0003	2	Forage 74 DAT	22 Sep 2009	BBCH 87
116-0004	3	Forage 74 DAT	22 Sep 2009	BBCH 87
116-0005	3	Forage 74 DAT	22 Sep 2009	BBCH 87
117-0001	1	Grain 76 DAT	26 Sep 2009	BBCH 88
117-0002	2	Grain 76 DAT	26 Sep 2009	BBCH 88
117-0003	2	Grain 76 DAT	26 Sep 2009	BBCH 88
117-0004	3	Grain 76 DAT	26 Sep 2009	BBCH 88
117-0005	3	Grain 76 DAT	26 Sep 2009	BBCH 88
118-0001	1	Stover 76 DAT	26 Sep 2009	BBCH 88
118-0002	2	Stover 76 DAT	26 Sep 2009	BBCH 88
118-0003	2	Stover 76 DAT	26 Sep 2009	BBCH 88
118-0004	3	Stover 76 DAT	26 Sep 2009	BBCH 88
118-0005	3	Stover 76 DAT	26 Sep 2009	BBCH 88
119-0001	1	Grain 83 DAT	02 Oct 2009	BBCH 89
119-0002	2	Grain 83 DAT	02 Oct 2009	BBCH 89
119-0003	2	Grain 83 DAT	02 Oct 2009	BBCH 89
119-0004	3	Grain 83 DAT	02 Oct 2009	BBCH 89
119-0005	3	Grain 83 DAT	02 Oct 2009	BBCH 89
120-0001	1	Stover 83 DAT	02 Oct 2009	BBCH 89
120-0002	2	Stover 83 DAT	02 Oct 2009	BBCH 89
120-0003	2	Stover 83 DAT	02 Oct 2009	BBCH 89
120-0004	3	Stover 83 DAT	02 Oct 2009	BBCH 89
120-0005	3	Stover 83 DAT	02 Oct 2009	BBCH 89
121-0001	1	Grain 90 DAT	10 Oct 2009	BBCH 90
121-0002	2	Grain 90 DAT	10 Oct 2009	BBCH 90
121-0003	2	Grain 90 DAT	10 Oct 2009	BBCH 90
121-0004	3	Grain 90 DAT	10 Oct 2009	BBCH 90
121-0005	3	Grain 90 DAT	10 Oct 2009	BBCH 90
122-0001	1	Stover 90 DAT	10 Oct 2009	BBCH 90
122-0002	2	Stover 90 DAT	10 Oct 2009	BBCH 90
122-0003	2	Stover 90 DAT	10 Oct 2009	BBCH 90
122-0004	3	Stover 90 DAT	10 Oct 2009	BBCH 90
122-0005	3	Stover 90 DAT	10 Oct 2009	BBCH 90
123-0001	1	Grain 97 DAT	17 Oct 2009	BBCH 90
123-0002	2	Grain 97 DAT	17 Oct 2009	BBCH 90
123-0003	2	Grain 97 DAT	17 Oct 2009	BBCH 90
123-0004	3	Grain 97 DAT	17 Oct 2009	BBCH 90
123-0005	3	Grain 97 DAT	17 Oct 2009	BBCH 90
124-0001	1	Stover 97 DAT	17 Oct 2009	BBCH 90

Table 30: Crop Samples (Trial 22) (continued)

Sample No.	Treatment	Sampling Event – Nominal PHI	Date of Collection	Crop Growth Stage
124-0002	2	Stover 97 DAT	17 Oct 2009	BBCH 90
124-0003	2	Stover 97 DAT	17 Oct 2009	BBCH 90
124-0004	3	Stover 97 DAT	17 Oct 2009	BBCH 90
124-0005	3	Stover 97 DAT	17 Oct 2009	BBCH 90
125-0001	1	Grain 104 DAT	24 Oct 2009	BBCH 90
125-0002	2	Grain 104 DAT	24 Oct 2009	BBCH 90
125-0003	2	Grain 104 DAT	24 Oct 2009	BBCH 90
125-0004	3	Grain 104 DAT	24 Oct 2009	BBCH 90
125-0005	3	Grain 104 DAT	24 Oct 2009	BBCH 90
126-0001	1	Stover 104 DAT	24 Oct 2009	BBCH 90
126-0002	2	Stover 104 DAT	24 Oct 2009	BBCH 90
126-0003	2	Stover 104 DAT	24 Oct 2009	BBCH 90
126-0004	3	Stover 104 DAT	24 Oct 2009	BBCH 90
126-0005	3	Stover 104 DAT	24 Oct 2009	BBCH 90

Table 31: Crop Samples (Trial 23)

Sample No.	Treatment	Sampling Event – Nominal PHI	Date of Collection	Crop Growth Stage
127-0001	1	Forage 40 DAT	20 Aug 2009	BBCH 71
127-0002	2	Forage 40 DAT	20 Aug 2009	BBCH 71
127-0003	2	Forage 40 DAT	20 Aug 2009	BBCH 71
127-0004	3	Forage 40 DAT	20 Aug 2009	BBCH 71
127-0005	3	Forage 40 DAT	20 Aug 2009	BBCH 71
128-0001	1	Forage 60 DAT	01 Sep 2009	BBCH 76
128-0002	2	Forage 60 DAT	01 Sep 2009	BBCH 76
128-0003	2	Forage 60 DAT	01 Sep 2009	BBCH 76
128-0004	3	Forage 60 DAT	01 Sep 2009	BBCH 76
128-0005	3	Forage 60 DAT	01 Sep 2009	BBCH 76
129-0001	1	Grain 90 DAT	08 Nov 2009	BBCH 90
129-0002	2	Grain 90 DAT	08 Nov 2009	BBCH 90
129-0003	2	Grain 90 DAT	08 Nov 2009	BBCH 90
129-0004	3	Grain 90 DAT	08 Nov 2009	BBCH 90
129-0005	3	Grain 90 DAT	08 Nov 2009	BBCH 90
130-0001	1	Stover 90 DAT	08 Nov 2009	BBCH 90
130-0002	2	Stover 90 DAT	08 Nov 2009	BBCH 90
130-0003	2	Stover 90 DAT	08 Nov 2009	BBCH 90
130-0004	3	Stover 90 DAT	08 Nov 2009	BBCH 90
130-0005	3	Stover 90 DAT	08 Nov 2009	BBCH 90

Table 32: Crop Samples (Trial 24)

Sample No.	Treatment	Sampling Event – Nominal PHI	Date of Collection	Crop Growth Stage
131-0001	1	Forage 40 DAT	24 Aug 2009	R1
131-0002	2	Forage 40 DAT	24 Aug 2009	R1
131-0003	2	Forage 40 DAT	24 Aug 2009	R1
131-0004	3	Forage 40 DAT	24 Aug 2009	R1
131-0005	3	Forage 40 DAT	24 Aug 2009	R1
132-0001	1	Forage 60 DAT	30 Sep 2009	BBCH 85
132-0002	2	Forage 60 DAT	30 Sep 2009	BBCH 85
132-0003	2	Forage 60 DAT	30 Sep 2009	BBCH 85
132-0004	3	Forage 60 DAT	30 Sep 2009	BBCH 85
132-0005	3	Forage 60 DAT	30 Sep 2009	BBCH 85
133-0001	1	Grain 90 DAT	17 Nov 2009	BBCH 97
133-0002	2	Grain 90 DAT	17 Nov 2009	BBCH 97
133-0003	2	Grain 90 DAT	17 Nov 2009	BBCH 97
133-0004	3	Grain 90 DAT	17 Nov 2009	BBCH 97
133-0005	3	Grain 90 DAT	17 Nov 2009	BBCH 97
134-0001	1	Stover 90 DAT	17 Nov 2009	BBCH 97
134-0002	2	Stover 90 DAT	17 Nov 2009	BBCH 97
134-0003	2	Stover 90 DAT	17 Nov 2009	BBCH 97
134-0004	3	Stover 90 DAT	17 Nov 2009	BBCH 97
134-0005	3	Stover 90 DAT	17 Nov 2009	BBCH 97

Table 33: Crop Samples (Trial 25)

Sample No.	Treatment	Sampling Event – Nominal PHI	Date of Collection	Crop Growth Stage
135-0001	1	Forage 40 DAT	26 Aug 2009	R2
135-0002	2	Forage 40 DAT	26 Aug 2009	R2
135-0003	2	Forage 40 DAT	26 Aug 2009	R2
135-0004	3	Forage 40 DAT	26 Aug 2009	R2
135-0005	3	Forage 40 DAT	26 Aug 2009	R2
136-0001	1	Forage 60 DAT	24 Sep 2009	BBCH 85
136-0002	2	Forage 60 DAT	24 Sep 2009	BBCH 85
136-0003	2	Forage 60 DAT	24 Sep 2009	BBCH 85
136-0004	3	Forage 60 DAT	24 Sep 2009	BBCH 85
136-0005	3	Forage 60 DAT	24 Sep 2009	BBCH 85
137-0001	1	Grain 90 DAT	18 Nov 2009	BBCH 97
137-0002	2	Grain 90 DAT	18 Nov 2009	BBCH 97
137-0003	2	Grain 90 DAT	18 Nov 2009	BBCH 97
137-0004	3	Grain 90 DAT	18 Nov 2009	BBCH 97
137-0005	3	Grain 90 DAT	18 Nov 2009	BBCH 97
138-0001	1	Stover 90 DAT	18 Nov 2009	BBCH 97
138-0002	2	Stover 90 DAT	18 Nov 2009	BBCH 97
138-0003	2	Stover 90 DAT	18 Nov 2009	BBCH 97
138-0004	3	Stover 90 DAT	18 Nov 2009	BBCH 97
138-0005	3	Stover 90 DAT	18 Nov 2009	BBCH 97

5.3 Sample Field Storage and Shipping

All field samples were placed in frozen storage promptly after collection, and many samples were placed in temporary chilled storage containers (mobile freezers or coolers with blue or dry ice) immediately after collection and subsequently transferred to a facility freezer unit. The samples were maintained frozen at the field facility until shipment to the Dow AgroSciences. The samples were transported frozen and maintained frozen in transit to the Dow AgroSciences via freezer truck. Refer to Table 34 for details of storage and shipping.

Table 34: Sample Storage and Shipping

Trial No.	Samples Shipped	Collection to Frozen Storage (hrs:min)	Frozen Storage (days)	Storage Temp. Range	Date Shipped
NJ-01	001-0001 to 001-0005	00:05	28	-7 to 17°F	15 Oct 2009
	002-0001 to 004-0005	00:05-00:15	9-49	-6 to 16°F	02 Dec 2009
PA-02	005-0001 to 006-0005	0:10-1:00	31-45	-9 to 28°F	15 Oct 2009
	007-0001 to 008-0005	0:15-1:30	28	-6 to 34°F	02 Dec 2009
GA-03	All samples	00:16-1:28	9-46	-1 to 32°F	11 Oct 2009
GA-04	014-0001 to 021-0005	0:31-1:40	10-42	-1 to 32°F	11 Oct 2009
	022-0001 to 029-0005	0:20-1:55	32-53	-1 to 32°F	30 Nov 2009
IL-05	030-0001 to 030-0005	00:30-1:15	14	-26 to -13°C	01 Sep 2009
	031-0001 to 031-0005	00:45-1:10	33	-28 to -14°C	17 Oct 2009
	032-0001 to 033-0005	1:30-2:30	15	-28 to -18°C	04 Dec 2009
IL-06	034-0001 to 035-0005	0:18-1:40	24-49	-32 to 7°F	23 Oct 2009
	036-0001 to 037-0005	1:05-1:30	19	-28 to -6°F	12 Dec 2009
IN-07	038-0001 to 039-0005	00:20-1:15	4-23	-25 to -12°C	18 Sep 2009
	040-0001 to 041-0005	00:10-1:10	1	Packed in dry ice	02 Dec 2009
MO-08	All Samples	00:01-00:32[b]	27-91	-21.6 to 4.9°F	18 Dec 2009
MO-09	All Samples	00:02-00:30	27-91	-20 to +10°F	18 Dec 2009

Table 34: Sample Storage and Shipping (continued)

Trial No.	Samples Shipped	Collection to Frozen Storage (hrs:min)	Frozen Storage (days)	Storage Temp. Range	Date Shipped
IL-10	All Samples	<00:01-00:40	17-77	-21.6 to 4.9°F	18 Dec 2009
MI-11	054-0001 to 058-0005	~1:00	0-26	-42 to 21°F	10 Oct 2009
	059-0001 to 069-0005	~1:00	16-47	-47 to 4°F	03 Dec 2009
OH-12	070-0001 to 071-0005	~2:00	14-29	-18 to 21°F	16 Oct 2009
	072-0001 to 073-0005	~3:00	27	-41 to -1°F	03 Dec 2009
IN-13	074-0001 to 075-0005	~4:00	12-32	-18 to 21°F	16 Oct 2009
	076-0001 to 077-0005	~4:00	30	-41 to -1°F	03 Dec 2009
NE-14	079-0001 to 079-0005	00:33-1:14	50	-22 to 21°F	21 Oct 2009
	080-0001 to 082-0005	00:38-76:20(a)	21	-18 to 20°F	08 Dec 2009
NE-15	All Samples	0:20-2:00	5-52	-30 to 15°F	21 Oct 2009
KS-16	087-0001 to 088-0005	1:20-2:37	22-48	-30 to 8°F	21 Oct 2009
	089-0001 to 090-0005	1:43-2:32	44	-28 to -10°F	11 Dec 2009
IA-17	091-0001 to 091-0005	0:40-2:18	15	-24 to 2°F	28 Aug 2009
	092-0001 to 094-0005	0:21-1:45	2-45	-23 to -2°F	19 Oct 2009
MO-18	095-0001 to 095-0005	3:08-3:30	7	-19.7 to 6.8°F	28 Aug 2009
	096-0001 to 096-0005	5:50-6:10	46	-23 to -2°F	19 Oct 2009
	097-0001 to 098-0005	5:50-6:45	45	-25 to -3°F	04 Dec 2009
IA-19	099-0001 to 100-0005	0:42-1:54	21-48	-15 to 12°F[b]	20 Oct 2009
	101-0001 to 102-0005	1:39-1:45	16	-2 to 20°F	07 Dec 2009
MN-20	103-0001 to 104-0005	0:52-1:13	21-49	-15 to 12°F[b]	20 Oct 2009
	105-0001 to 106-0005	0:57-1:46	16	-2 to 20°F	07 Dec 2009

Table 34: Sample Storage and Shipping (continued)

Trial No.	Samples Shipped	Collection to Frozen Storage (hrs:min)	Frozen Storage (days)	Storage Temp. Range	Date Shipped
MO-21	107-0001 to 107-0005	1:15-2:00	6	-19 to -7°F	21 Sep 2009
	108-0001 to 108-0005	0:36-1:12	22	-20 to -6°F	22 Oct 2009
	109-0001 to 110-0005	1:24-2:00	40	-17 to -3°F	12 Dec 2009
OK-22	All Samples	~0:05	54-117	-24 to 26°F	17 Dec 2009
TX-23	All Samples	~0:05	39-119	-24 to 26°F	17 Dec 2009
ON-24	131-0001 to 131-0005	0:14-0:58	33	-17.0 to -14.7°C	25 Sep 2009
	132-0001 to 134-0005	0:07-1:59	4-52	-18.7 to -15.1°C	20 Nov 2009
ON-25	135-0001 to 136-0005	1:35-2:33	2-31	-17 to -15.8°C	25 Sep 2009
	137-0001 to 138-0005	1:53-2:37	3	-18.7 to -15.2°C	20 Nov 2009

a. Bulk grain for processing placed in freezer within 3 days of collection
b. Freezer temperatures of 02 Sep 2009 to 18 Nov 2009 for the untreated samples were lost while downloading due to a malfunction.

6. WEATHER SUMMARY DURING TEST PERIOD

6.1. Weather at Applications

Weather at the time of applications was favorable and typical, if not better than what would be encountered in a commercial situation. See Table 35 below for specific weather conditions at the time of applications at each trial.

Table 35: Weather Conditions at Applications

Trial #	Appl. No.	Air Temp (°F)	%Cloud Cover	Relative Air Humidity (%)	Wind Speed (mph) and Direction	Date
NJ-01	1	86.7	75	28	0-1 SE	07 Jul 2009
	2	75.0	50	65	2-5 NW	30 Jul 2009
	3	77.5	15	66	0-1 NW	04 Aug 2009
	4	90.0	50	45	3-5 SW	10 Aug 2009
PA-02	1	80.2	30	61.2	0-1 S	08 Jun 2009
	2	61.6	0	72.6	0-2 NW	08 Jul 2009
	3	67.4	0	60.8	0-1 NW	15 Jul 2009
	4	79.0	90	54	0, calm	20 Jul 2009
GA-03	1	88/87(a)	10	62/69(a)	2.5-3.5/2.0-3.6 NW(a)	12 Jun 2009
	2	9/90	50	64/63	2.0-2.5 NE	09 Jul 2009
	3	82/83	100	76/75	0-1.0 NE	15 Jul 2009
	4	77/79	10	64/62	1.0-2.0 NE	20 Jul 2009
GA-04	1	89.0	60	56	2.0-2.5 NW	12 Jun 2009
	2	80.0	20	60	2.0-2.5 NW	20 Jul 2009
	3	80.0	10	44	2.5-3.0 SW	27 Jul 2009
	4	78.0	80	87	1.0-1.5 SW	31 Jul 2009
IL-05	1	66.8	5	36.5	1.9 W	05 Jun 2009
	2	92.8	50	35.3	1.45 NNW	26 Jun 2009
	3	71	70	70	0-1 W	03 Jul 2009
	4	67	100	84	3-5 E	09 Jul 2009
IL-06	1	92.0	5	50	2-3 SW	26 Jun 2009
	2	88.0	10	46	2-3 W	16 Jul 2009
	3	73.0	95	68	3-4 N	22 Jul 2009
	4	88.0	20	44	2 S	28 Jul 2009
IN-07	1	71.0	0	76	3 WSW	09 Jun 2009
	2	71.0	10	58	4 WNW	02 Jul 2009
	3	75.0	10	72	2-3 SSE	10 Jul 2009
	4	78.0	20	67	3-4 NW	15 Jul 2009
MO-08	1	84.4	30	58.6	3.1 SSE	03 Jul 2009
	2	89.6	20	60	0.5 SW	28 Jul 2009
	3	90.6	0	45	1.0 W	04 Aug 2009
	4	78.5	10	60	0, calm	09 Aug 2009
MO-09	1	70.0	20	40	1.0 W	28 Jun 2009
	2	86.5	30	65	1.0 SW	27 Jul 2009
	3	93.6	30	60	1.4 SSW	03 Aug 2009
	4	94.0	30	50.5	1-3 SW	08 Aug 2009
IL-10	1	81.8	0	45	3.0 S	06 Jul 2009
	2	69.4	60	84	0, Calm	01 Aug 2009
	3	88.5	0	67	1.0 SSE	08 Aug 2009
	4	66.0	0	65.5	0, Calm	13 ug 2009

(a) First number listed is for Treatments 2 and 3 and the second number listed is for Treatments 4 and 5

Table 35: Weather Conditions at Applications (continued)

Trial #	Appl. No.	Air Temp (°F)	%Cloud Cover	Relative Air Humidity (%)	Wind Speed (mph) and Direction	Date
MI-11	1	74.0	95	75	3.0-4.0 SW	17 Jun 2009
	2	74.0	95	73	4 SW	25 Jul 2009
	3	77.0	50	60	1.0-2.5 W	31 Jul 2009
	4	85.0	35	38	2.0-3.0 SW	05 Aug 2009
OH-12	1	83.0	75	42	1-1.5 SE	14 Jun 2009
	2	78.0	95	67	2.5-4 SW	27 Jul 2009
	3	71.0	60	73	3.5-4.5 SW	03 Aug 2009
	4	65.0	100	83	1-3 SW	08 Aug 2009
IN-13	1	86.0	50	35	1-1.5 E	15 Jun 2009
	2	84.0	65	49	3.5-4 SW	25 Jul 2009
	3	78.0	45	50	1-3 W	31 Jul 2009
	4	67.0	75	70	2-3 SW	05 Aug 2009
NE-14	1	62	40	57	0-1 NW	04 Jun 2009
	2	72	30	64	2-5 SE	24 Jun 2009
	3	73-74	0	63-65	1-4 N	30 Jun 2009
	4	63	20	42	1-4 SW	06 Jul 2009
NE-15	1	83	40	77	0-3 SE	23 Jun 2009
	2	87	0	57	1-6 SE	08 Jul 2009
	3	80	0	44	3-9 NE	15 Jul 2009
	4	71	100	77	0-4 SE	20 Jul 2009
KS-16	1	94.0	0	23	0-3 W	29 Jun 2009
	2	75.0	0	50	2-6 N	15 Jul 2009
	3	81.0	0	50	2-6 NW	22 Jul 2009
	4	90.0	70	37	2-9 NE	27 Jul 2009
IA-17	1	69.5	50	59.9	1.6 NE	09 Jun 2009
	2	89.9	10	73.4	2.1 W	24 Jun 2009
	3	65.6	100	85.1	3.6 NW	01 Jul 2009
	4	83.3	45	54.5	2.6 W	06 Jul 2009
MO-18	1	72.5	95	85	0.6 N	06 Jun 2009
	2	77.8	10	43	4.2 NNW	30 Jun 2009
	3	81.3	10	44	0.7 SW	06 Jul 2009
	4	82.0	20	36	2.7 SW	12 Jul 2009
IA-19	1	59.3	0	33.6	0.8 NW	05 Jun 2009
	2	78.4	0	41.0	4.7 W	11 Jul 2009
	3	68.6	100	54.8	4 NW	18 Jul 2009
	4	83.4	70	32.9	3.2 N	23 Jul 2009
MN-20	1	65.2	10	24.9	2.7 NW	05 Jun 2009
	2	75.8	0	38.8	2.4 W	11 Jul 2009
	3	64.7	100	46.3	3.6 NW	18 Jul 2009
	4	85.0	40	35.5	1.1 NW	23 Jul 2009
MO-21	1	84.0	20	54	2-3 NW	30 Jun 2009
	2	83.0	40	56	1-2 NW	23 Jul 2009
	3	79.0	100	78	3-4 W	30 Jul 2009
	4	90.0	50	64	3-4 W	04 Aug 2009

Table 35: Weather Conditions at Applications (continued)

Trial #	Appl. No.	Air Temp (°F)	%Cloud Cover	Relative Air Humidity (%)	Wind Speed (mph) and Direction	Date
OK-22	1	110.7	30	35.8	2-3 S	27 Jun 2009
	2	105.1	0	42.3	1-2 S	13 Jul 2009
	3	93.0	10	28	1-3 S	18 Jul 2009
	4	100.6	10	48.0	1-2 SE	23 Jul 2009
TX-23	1	87.0	10	68.9	2-3 SSE	28 Jun 2009
	2	99.6	10	34	1-4 SSE	14 Jul 2009
	3	68.9	10	65	3-4 SE	19 Jul 2009
	4	59.7	10	40	2-3 SE	24 Jul 2009
ON-24	1	13.5[a]	10	67	3.1[b] N	04 Jun 2009
	2	16.8[a]	100	80	0	01 Jul 2009
	3	25.5[a]	50	48	2.2[b] W	09 Jul 2009
	4	15.7[a]	0	52	5.2[b] NW	14 Jul 2009
ON-25	1	16.2[a]	60	77.6	4.0[b] E	10 Jun 2009
	2	17.0[a]	50	90.0	0	06 Jul 2009
	3	17.6[a]	0	56	2.0[b] W	13 Jul 2009
	4	14.4[a]	40	83	8[b] W	18 Jul 2009

(a) Temperatures were recorded using units of °C instead of °F.
(b) Wind speed was recorded using units of km/h instead of mph.

6.2. Weather During Test Period

Weather during the trial period was considered by PFIs to be normal in most trial locations, warmer in some, and cooler in others. Rainfall was normal for some months, wetter for some months, and drier for some months in different trial locations. Weather was determined to be normal or not by comparing to historical averages for the region. Refer to Table 36 and 37 for detailed information on temperature and precipitation.

Table 36: Temperatures during Test Period (typically planting to final sampling)

Trial	State	Weather Source (distance from site)	Month, 2009[a]	Trial Temp Min °F	Trial Temp Max °F	Historical Temps Min °F	Historical Temps Max °F	Historical Temps Difference from Historical
NJ-01 Trial Period: 05 Jul 2009 to 23 Nov 2009	NJ	Trial: Weather station (on-site) Historical: NOAA Station 3029 (~8 miles)	Jul	59.9	79.9	62.8	85.2	Similar
			Aug	63.7	82.4	62.1	84.4	Similar
			Sep	54	72.9	54.3	77.6	Similar
			Oct	43.5	59	41.9	64.5	Similar
			Nov	41.2	54	34.6	54.3	Similar
PA-02 Trial Period: 08 Jun 2009 to 04 Nov 2009	PA	Trial:- CMS Inc. Weather Station (on site) Historical: NOAA 0106 Allentown, PA (~18 miles)	Jun	41	98	59	80.5	Similar
			Jul	52	97	62.8	84.7	Similar
			Aug	55	100	62.2	83.5	Similar
			Sep	41	86	53.9	76.3	Similar
			Oct	NA(a)	NA	42.2	64	Similar
			Nov	NA	NA	NA	NA	NA
GA-03 Trial Period: 09 Jun 2009 to 02 Oct 2009	GA	Trial :ARA Weather Station WS01 Sycamore, GA (on site) Historical: Weather Data UGA- Tifton Station Tifton, GA (~13 miles)	Jun	70	98	68.1	87.9	Similar
			Jul	63	97	70.9	90.3	Similar
			Aug	64	97	70.2	89.9	Similar
			Sep	53	92	65.9	86.4	Similar
			Oct	53	82	54.8	78.3	Similar

[a] NA = Not available

Table 36: Temperatures during Test Period (typically planting to final sampling) (continued)

Trial	State	Weather Source (distance from site)	Month, 2009[a]	Trial Temp Min °F	Trial Temp Max °F	Historical Temps Min °F	Historical Temps Max °F	Historical Temps Difference from Historical
GA-04 Trial Period: 10 Jun 2009 to 29 Oct 2009	GA	Trial and Historical: UGA Tifton Station Tifton, GA (~10 miles)	Jun	70.0	96.0	68.1	87.9	Warmer
			Jul	62.0	95.0	70.9	90.3	Warmer
			Aug	64.0	94.0	70.2	89.9	Warmer
			Sep	51.0	92.0	65.9	86.4	Warmer
			Oct	40.0	91.0	54.8	78.3	Warmer
IL-05 Trial Period: 04 Jun 2009 to 19 Nov 2009	IL	Trial :SGS AAR Wyoming ~ 10 miles Historical: Lacon NOAA Station – ~ 15 miles	Jun	60.3	81.6	60.5	83.5	Cooler
			Jul	58.1	78.6	65.2	87.4	Cooler
			Aug	57.7	79.5	63.2	85.6	Cooler
			Sep	52.3	75.7	54.6	80.3	Cooler
			Oct	39.5	55.5	44.5	67	Cooler
			Nov	34.2	53.6	33.8	53.4	Cooler
IL-06 Trial Period: 26 Jun 2009 to 23 Nov 2009	IL	Trial: AAR Weather Nettles (~ ½ mile) Historical: US Army Corp of Engineers Carlyle, IL (~4 miles)	Jun	62	96	64	83	Similar
			Jul	60	91	68	87	Similar
			Aug	50	92	66	86	Similar
			Sep	42	85	58	81	Similar
			Oct	29	73	47	68	Similar
			Nov	43	61	37	55	Similar
IN-07 Trial Period: 08 Jun 2009 to 01 Dec 2009	IN	Trial: SGS Spectrum Watchdog (on-site) Historical: Rockville NOAA (~8 miles)	Jun	63.1	82.8	61.3	83.4	Similar
			Jul	63.8	83.3	64.9	86.4	Cooler
			Aug	59.5	82.1	64.4	85.8	Much cooler
			Sep	56.1	79.4	55.3	80.2	Similar
			Oct	42	58.9	45	67.4	Much cooler
			Nov	37.2	55.6	36.3	55.1	Similar
			Dec	33.8	53.1	24.6	40.2	Similar
MO-08 Trial period: 03 Jul 2009 to 21 Nov 2009	MO	Trial: Tong (on site) Historical: NWS 231356 Carthage, MO (~8 miles)	Jul	56	99	69.2	91.2	Similar
			Aug	50	98	68.3	90.7	Similar
			Sep	43	88	58.6	83.5	Similar
			Oct	37	82	47.4	72.1	Similar
			Nov	34	79	38.1	60.6	Similar
MO-09 Trial Period: 28 Jun 2009 to 21 Nov 2009	MO	Trial: (onsite Historical: NWS 231711 Clinton, MO (~12.5 miles)	Jun	63	92	61.9	83.0	Similar
			Jul	57	92	66.4	89.2	Similar
			Aug	53	96	66.1	90.2	Similar
			Sep	41	86	55.1	80.0	Similar
			Oct	32	76	43.5	68.7	Similar
			Nov	29	76	32.3	55.4	Similar

Table 36: Temperatures during Test Period (typically planting to final sampling) (continued)

Trial	State	Weather Source (distance from site)	Month, 2009[a]	Trial Temp Min °F	Trial Temp Max °F	Historical Temps Min °F	Historical Temps Max °F	Historical Temps Difference from Historical
IL-10 Trial Period: 06 Jul 2009 to 21 Nov 2009	IL	Trial: Ross (on site) Historical: NWS 117551 Rushville, IL (~5.8 miles)	Jul	54	87	64.9	85.8	Similar
			Aug	48	92	64.6	86.1	Similar
			Sep	43	83	56.1	80.7	Similar
			Oct	32	72	44.0	67.2	Similar
			Nov	26	75	34.2	53.6	Similar
MI-11 Trial Period: 13 Jun 2009 to 17 Nov 2009	MI	Historical: COOP Adrian, MI (~15 miles)	Jun	47.0	95.9	57.0	80.5	Similar
			Jul	42.5	86.4	60.5	84.1	Similar
			Aug	39.9	92.0	59.1	82.1	Similar
			Sep	31.7	83.9	51.0	76.2	Similar
			Oct	22.5	74.6	40.5	62.2	Similar
			Nov	21.3	70.0	32.4	50.2	Similar
OH-12 Trial Period: 10 Jun 2009 to 06 Nov 2009	OH	Trial: Hudson Michigan State University (~20 miles) Historical: COOP Adrian, MI (~20 miles)	Jun	51.8	94.9	57.0	80.5	Similar
			Jul	46.6	84.6	60.5	84.1	Similar
			Aug	43.0	92.6	59.1	82.1	Similar
			Sep	36.4	83.7	51.0	76.2	Similar
			Oct	25.7	70.4	40.5	62.2	Similar
			Nov	24.3	55.8	32.4	50.2	Similar
IN-13 Trial Period: 15 Jun 2009 to 03 Nov 2009	IN	Trial: Constantine Michigan State University (~22 miles) Historical: COOP Angola, IN (~30 miles)	Jun	53.0	94.6	57.2	78.1	Similar
			Jul	46.7	84.1	60.6	81.8	Similar
			Aug	42.8	91.8	58.8	80.2	Similar
			Sep	37.1	86.1	50.4	74.7	Similar
			Oct	26.5	69.8	39.4	60.6	Similar
			Nov	27.6	58.3	31.3	48.9	Similar

Table 36: Temperatures during Test Period (typically planting to final sampling) (continued)

Trial	State	Weather Source (distance from site)	Month, 2009[a]	Trial Temp Min °F	Trial Temp Max °F	Historical Temps Min °F	Historical Temps Max °F	Historical Temps Difference from Historical
NE-14 Trial Period: 01 Jun 2009 to 17 Nov 2009	NE	Trial: NOAA 259513 12 miles NE of trial & ARA NE Weather Station (on site) Historical: NOAA 259510 York NE (~12 miles)	Jun	58	81	61	84	Similar
			Jul	58	84	67	89	Similar
			Aug	56	81	64	87	Similar
			Sep	41	66	53	80	Similar
			Oct	35	56	41	67	Similar
			Nov	35	57	30	54	Similar
NE-15 Trial Period: 22 Jun 2009 to 17 Oct 2009	NE	Trial and Historical: Osceola, NE NOAA c256375 (~6 miles)	Jun	56	93	60	81.6	Cooler
			Jul	50	91	65.7	86.8	Cooler
			Aug	46	92	63.6	84.7	Cooler
			Sep	34	80	53.6	78.5	Cooler
			Oct	23	71	41.6	65.4	Cooler
KS-16 Trial Period: 29 Jun 2009 to 28 Oct 2009	KS	Trial and Historical: NOAA c140682 Belleville, KS ~(8 miles)	Jun	60	91	61.7	85.3	Cooler
			Jul	52	97	68.4	92.4	Cooler
			Aug	49.2	101	67.1	91.2	Cooler
			Sep	36	84	56.2	81.5	Cooler
			Oct	57	78	44.4	68.2	Cooler
IA-17 Trial Period: 05 Jun 2009 to 17 Oct 2009	IA	Trial: Watchdog Model 900ET Bennett Agricultural Research Corp. (~0.7 miles) Historical: NOAA 132789 Fairfield, IA (~9 miles)	Jun	51.0	94.1	61.0	81.8	Cooler
			Jul	51.0	87.4	65.8	87.0	Cooler
			Aug	43.7	90.3	63.5	85.2	Cooler
			Sep	35.4	80.9	52.7	78.5	Cooler
			Oct	28.1	71.8	42.7	64.9	Cooler
MO-18 Trial Period: 05 Jun 2009 to 20 Oct 2009	MO	Trial: Specware 8 Pro Adair County Missouri (on site) Historical: NOAA 234544 Kirksville, Missouri (~15 miles)	Jun	52.0	98.7	60.3	80.8	Cooler
			Jul	53.2	89.7	65.7	86.6	Cooler
			Aug	46.5	97.3	63.6	85.4	Cooler
			Sep	39.4	85.3	53.6	77.9	Cooler
			Oct	27.9	72.7	43.0	65.1	Cooler

Table 36: Temperatures during Test Period (typically planting to final sampling) (continued)

Trial	State	Weather Source (distance from site)	Month, 2009[a]	Trial Temp Min °F	Trial Temp Max °F	Historical Temps Min °F	Historical Temps Max °F	Historical Temps Difference from Historical
IA-19 Trial Period: 02 Jun 2009 to 21 Nov 2009	IA	Trial: Weather Station IA-1 Lime Springs IA (~21 miles) Historical: NOAA 131954 Cresco, IA (~13.5 miles)	Jun	41	91.4	55.4	77.7	Cooler
			Jul	44.6	80.6	59.9	83.1	Cooler
			Aug	35.6	84.2	56.9	80.1	Cooler
			Sep	37.4	80.6	48.1	73.5	Cooler
			Oct	19.4	66.2	35.9	59.8	Cooler
			Nov	19.4	68	25.5	45.2	Cooler
MN-20 Trial Period: 01 Jun 2009 to 21 Nov 2009	MN	Trial: Weather Station IA-1 Lime Springs, IA (~6.5 miles) Historical: NOAA 131954 Cresco, IA (~15 miles)	Jun	41	91.4	55.4	77.7	Cooler
			Jul	44.6	80.6	59.9	83.1	Cooler
			Aug	35.6	84.2	56.9	80.1	Cooler
			Sep	37.4	80.6	48.1	73.5	Cooler
			Oct	19.4	66.2	35.9	59.8	Cooler
			Nov	19.4	68	25.5	45.2	Cooler
MO-21 Trial Period: 30 Jun 2009 to 02 Nov 2009	MO	Trial: Harty Farms Watch Dog Model 425 (~300 ft) Historical: NOAA COOP # 236791 Poplar Bluff, Missouri (~13 miles)	Jun	59	94	65.0	86.0	Similar
			Jul	62	89	69.0	90.0	Similar
			Aug	61	92	69.0	90.0	Similar
			Sep	59	85	59.0	82.0	Similar
			Oct	45	65	48.0	71.0	Cooler
			Nov	41	63	38.0	58.0	Similar

Table 36: Temperatures during Test Period (typically planting to final sampling) (continued)

Trial	State	Weather Source (distance from site)	Month, 2009[a]	Trial Temp		Historical Temps		
				Min °F	Max °F	Min °F	Max °F	Difference from Historical
OK-22 Trial Period: 27 Jun 2009 to 24 Oct 2009	OK	Trial and Historical: Madill NOAA 345468 Madill, OK (~10 miles)	Jun	72	103	67.9	89.0	Similar
			Jul	65	105	72.3	95.4	Similar
			Aug	59.0	103.0	72.2	96.0	Similar
			Sep	49.0	95.0	62.4	87.0	Similar
			Oct	39.0	88.0	52.7	76.7	Similar
TX-23 Trial Period: 28 Jun 2009 to 08 Nov 2009	TX	Trial: NOAA 419346 Decatur, TX (~19 miles) Rainfall (on site) Historical: NOAA 412334 Decatur, TX (~19 miles)	Jun	72	100	67.8	90.0	Similar
			Jul	63	104	70.4	92.6	Similar
			Aug	59	102	70.4	93.3	Similar
			Sep	47	95	61.6	87.6	Similar
			Oct	38	87	51.8	79.1	Similar
			Nov	38	78	42.8	67.9	Similar
ON-24 Trial Period: 03 Jun 2009 to 17 Nov 2009	Ontario	Trial: VARS WS1 Watchdog 900 ET VARS Facility (~150 meters) Historical:– Environment Canada Waterloo-Wellington Station A Waterloo Intl Airport (~30 kilometers)	Jun	52.88	73.94	52.16	74.12	Similar
			Jul	55.04	75.02	56.66	78.62	Similar
			Aug	56.84	77.36	54.86	76.46	Similar
			Sep	49.82	72.14	47.12	68.36	Similar
			Oct	37.58	54.5	37.22	56.12	Similar
			Nov	34.34	49.82	29.3	42.98	Similar
ON-25 Trial Period: 05 Jun 2009 to 18 Nov 2009	Ontario	Trial and Historical : Ridgetown RCS Ridgetown, Ontario Ridgetown College WMO ID 71307 (~33 kilometers)	Jun	52.52	73.58	56.3	75.02	Cooler
			Jul	54.5	75.92	61.16	80.24	Cooler
			Aug	57.02	77.36	60.08	77.9	Cooler
			Sep	50	71.78	53.6	70.88	Cooler
			Oct	38.66	56.12	43.34	58.64	Cooler
			Nov	36.14	51.26	33.8	46.4	Warmer

(a) NA = Not available or not applicable.

Table 37: Precipitation during Test Period (typically planting to last sampling)

Trial	State	Historical Weather Source (distance from site)	Month 2009	Rainfall: Trial Monthly Total inches	Rainfall: Historical Monthly Total inches	Rainfall: Difference from Historical	Irrigation Type/ inches
NJ-01 Trial Period: 05 Jul 2009 to 23 Nov 2009	NJ	Trial: Weather station (on site) Historical:NOAA Station 3029 (~8 miles)	Jul	5.87	4.91	Wetter	None
			Aug	10.07	4.06	Wetter	None
			Sep	4.41	5.78	Normal	None
			Oct	4.78	5.19	Normal	None
			Nov	1.23	3.69	Drier	None
PA-02 Oct and Nov data02 Trial Period: 08 Jun 2009 to 04 Nov 2009	PA	Trial: CMS Inc Weather Station (on site) Historical:- NOAA 0106 Allentown, PA (~18 miles)	Jun	9.67	4.86	Wetter	None
			Jul	5.79	4.68	Wetter	None
			Aug	8.48	3.27	Wetter	None
			Sep	1.62	5.78	Normal	None
			Oct	NA(a)	NA	NA	NA
			Nov	NA	NA	NA	NA
GA-03 Trial Period: 09 Jun 2009 to 02 Oct 2009	GA	Trial :ARA Weather Station WS01 Sycamore, GA (on site) Historical: Weather Data UGA- Tifton Station Tifton, GA (~13 miles)	Jun	1.2	4.11	Normal	Center Pivot 2.13
			Jul	1.96	4.54	Normal	Center Pivot 2.50
			Aug	6.60	4.09	Normal	Center Pivot 1.56
			Sep	1.91	3.47	Normal	None
			Oct	0	2.58	Normal	None

Table 37: Precipitation during Test Period (typically planting to last sampling) (continued)

Trial	State	Historical Weather Source (distance from site)	Month 2009	Rainfall: Trial Monthly Total inches	Rainfall: Historical Monthly Total inches	Rainfall: Difference from Historical	Irrigation Type/ inches
GA-04 Trial Period: 10 Jun 2009 to 29 Oct 2009	GA	Trial and Historical:- UGA Tifton Station Tifton, GA (~10 miles)	Jun	0.16	4.11	Drier	None
			Jul	7.78	4.54	Wetter	None
			Aug	7.02	4.09	Wetter	None
			Sep	1.70	3.47	Drier	None
			Oct	3.83	2.58	Wetter	None
IL-05 Trial Period: 04 Jun 2009 to 19 Nov 2009	IL	Trial :SGS AAR Wyoming (~ 10 miles) Historical: Lacon NOAA Station –(~15 miles)	Jun	4.37	3.69	Wetter	None
			Jul	5.7	3.27	Wetter	None
			Aug	3.91	3.21	Wetter	None
			Sep	2.88	2.78	Wetter	None
			Oct	5.91	2.51	Wetter	None
			Nov	2.21	2.19	Wetter	None
IL-06 Trial Period: 26 Jun 2009 to 23 Nov 2009	IL	Trial: AAR Weather Nettles (~1/2 mile) Historical: US Army Corp of Engineers Carlyle, IL (~4 miles)	Jun	0	4.43	Normal	None
			Jul	3.2	3.9	Normal	None
			Aug	2.6	3.61	Normal	None
			Sep	4	3.07	Normal	None
			Oct	11.3	3.56	Wetter	None
			Nov	2.1	3.58	Normal	None
IN-07 Trial Period: 08 Jun 2009 to 01 Dec 2009	IN	Trial : SGS Spectrum Watchdog (on-site) Historical: Rockville NOAA (~8 miles)	Jun	6.57	5.58	Wetter	None
			Jul	3.47	5.58	Drier	None
			Aug	3.34	3.66	Normal	None
			Sep	0.57	3.53	Drier	None
			Oct	7.92	4.29	Wetter	None
			Nov	1.99	3.12	Drier	None
MO-08 Trial period: 03 Jul 2009 to 21 Nov 2009	MO	Trial: Tong – (on site) Historical: NWS 231356 Carthage, MO (~7.9 miles)	Jul	2.99	3.46	Normal	None
			Aug	12.01	2.50	Normal	None
			Sep	5.81	4.19	Normal	None
			Oct	7.47	3.47	Normal	None
			Nov	0.54	2.59	Normal	None
MO-09 Trial Period: 28 Jun 2009 to 21 Nov 2009	MO	Trial:- (on site) Historical:- NWS 231711 Clinton, MO (~12.5 miles)	Jun	0.41	6.19	Normal	None
			Jul	6.86	3.09	Normal	None
			Aug	3.34	3.29	Normal	None
			Sep	2.98	3.11	Normal	None
			Oct	10.47	2.87	Normal	None
			Nov	0.32	1.70	Normal	None

Table 37: **Precipitation during Test Period (typically planting to last sampling) (continued)**

Trial	State	Historical Weather Source (distance from site)	Month 2009	Rainfall: Trial Monthly Total inches	Rainfall: Historical Monthly Total inches	Rainfall: Difference from Historical	Irrigation Type/ inches
IL-10 Trial Period: 06 Jul 2009 to 21 Nov 2009	IL	Trial: Ross – (on site) Historical: NWS 117551 Rushville, IL (~5.8 miles)	Jul	2.70	2.52	Normal	None
			Aug	4.83	3.07	Normal	None
			Sep	1.28	1.65	Normal	None
			Oct	7.27	2.42	Normal	None
			Nov	4.67	1.42	Normal	None
MI-11 Trial Period: 13 Jun 2009 to 17 Nov 2009	MI	Trial: Petersburg Michigan State University Dundee, MI (~8 miles) Historical: COOP Adrian, MI (~15 miles)	Jun	2.42	4.07	Normal	None
			Jul	0.51	3.81	Normal	None
			Aug	3.46	3.56	Normal	None
			Sep	1.31	3.72	Normal	None
			Oct	3.35	2.43	Normal	None
			Nov	0.20	2.47	Normal	None
OH-12 Trial Period: 10 Jun 2009 to 06 Nov 2009	OH	Trial: Hudson Michigan State University (~20 miles) Historical: COOP Adrian, MI (~20 miles)	Jun	5.71	4.07	Normal	None
			Jul	1.26	3.81	Normal	None
			Aug	4.07	3.56	Normal	None
			Sep	2.10	3.72	Normal	None
			Oct	3.49	2.43	Normal	None
			Nov	0	2.47	Normal	None
IN-13 Trial Period: 15 Jun 2009 to 03 Nov 2009	IN	Trial: Constantine Michigan State University (~22 miles) Historical: COOP Angola, IN (~30 miles)	Jun	2.76	3.18	Normal	None
			Jul	3.62	4.59	Normal	None
			Aug	4.75	3.89	Normal	None
			Sep	1.16	2.85	Normal	None
			Oct	4.97	2.53	Normal	None
			Nov	0	2.54	Normal	None

Table 37: Precipitation during Test Period (typically planting to last sampling) (continued)

				Rainfall			
Trial	**State**	**Historical Weather Source** (distance from site)	**Month 2009**	**Trial Monthly Total** inches	**Historical Monthly Total** inches	**Difference from Historical**	**Irrigation** Type/ inches
NE-14 Trial Period: 01 Jun 2009 to 17 Nov 2009	NE	Trial: NOAA 259513 12 miles NE & ARA NE Weather Station (on site) Historical : NOAA 259510 York NE (~12 miles)NE	Jun	5.96	3.55	Wetter	None
			Jul	1.34	2.92	Drier	6.0 in
			Aug	6.17	2.88	Wetter	1.0 in
			Sep	1.64	2.67	Drier	1.0 in
			Oct	3.66	2.42	Wetter	None
			Nov	0.27	1.03	Drier	None
NE-15 Trial Period: 22 Jun 2009 to 17 Oct 2009	NE	Trial and Historical: Osceola, NE NOAA c256375 (~6 miles)	Jun	1.58	3.67	Drier	None
			Jul	1.03	2.49	Normal	2.0 in
			Aug	3.75	3.23	Normal	2.0 in
			Sep	0.59	2.39	Drier	None
			Oct	1.06	1.76	Drier	None
KS-16 Trial Period: 29 Jun 2009 to 28 Oct 2009	KS	Trial and Historical: NOAA c140682 Belleville, KS (~8 miles)	Jun	0.05	6.54	Drier	None
			Jul	4.35	5.49	Drier	None
			Aug	4.28	5.36	Drier	None
			Sep	3	5.23	Drier	None
			Oct	2.16	3.49	Drier	None
IA-17 Trial Period: 05 Jun 2009 to 17 Oct 2009	IA	Trial: Watchdog Model 900ET Bennett Agricultural Research Corp. (~0.7 miles) Historical: NOAA 132789 Fairfield, IA (~9 miles)	Jun	9.92	5.12	Wetter	None
			Jul	3.8	3.94	Wetter	None
			Aug	11.39	4.40	Wetter	None
			Sep	2.96	2.92	Wetter	None
			Oct	7.45	3.37	Wetter	None

Table 37: Precipitation during Test Period (typically planting to last sampling) (continued)

Trial	State	Historical Weather Source (distance from site)	Month 2009	Rainfall: Trial Monthly Total inches	Rainfall: Historical Monthly Total inches	Rainfall: Difference from Historical	Irrigation Type/ inches
MO-18 Trial Period: 05 Jun 2009 to 20 Oct 2009	MO	Trial: Specware 8 Pro Adair County Missouri (on site) Historical: NOAA Kirksville, Missouri (~15 miles)	Jun	7.2	5.29	Wetter	None
			Jul	4.16	3.63	Wetter	None
			Aug	6.24	4.43	Wetter	None
			Sep	4.07	3.88	Wetter	None
			Oct	10.11	3.15	Wetter	None
IA-19 Trial Period: 02 Jun 2009 to 21 Nov 2009	IA	Trial: Weather Station IA-1 Lime Springs Iowa (~21 miles) Historical: Cresco 1 NE Cresco, IA (~13.5 miles)	Jun	3.50	5.99	Normal	None
			Jul	1.83	5.27	Hail 7/24/2009	None
			Aug	3.49	4.86	Normal	None
			Sep	1.19	3.00	Normal	None
			Oct	6.54	2.03	Normal	None
			Nov	0.45	1.44	Normal	None
MN-20 Trial Period: 01 Jun 2009 to 21 Nov 2009	MN	Trial: Weather Station IA-1 Lime Springs, IA (~6.5 miles) Historical: NOAA 131954 Cresco, IA (~15 miles)	Jun	3.50	5.99	Hail 6/12/2009	None
			Jul	1.83	5.27	Normal	None
			Aug	3.49	4.86	Normal	None
			Sep	1.19	3.00	Normal	None
			Oct	6.54	2.03	Normal	None
			Nov	0.45	1.44	Normal	None

Table 37: Precipitation during Test Period (typically planting to last sampling) (continued)

Trial	**State**	**Historical Weather Source** (distance from site)	**Month 2009**	**Rainfall**			**Irrigation** Type/ inches
				Trial Monthly Total inches	**Historical Monthly Total** inches	**Difference from Historical**	
MO-21 Trial Period: 30 Jun 2009 to 02 Nov 2009	MO	Trial: Harty Farms Watch Dog Model 425 (~300 ft) Historical: NOAA COOP # 236791 Poplar Bluff, Missouri (~13 miles)	Jun	5.12	3.59	Normal	NA
			Jul	2.43	4.87	Normal	NA
			Aug	3.12	3.63	Normal	NA
			Sep	1.56	2.87	Normal	NA
			Oct	13.71	3.53	Wetter	NA
			Nov	2.12	4.16	Normal	NA
OK-22 Trial Period: 27 Jun 2009 to 24 Oct 2009	OK	Trial and Historical:- Madill NOAA 345468 Madill, OK (~10 miles) Rainfall (on site)	Jun	0	4.86	Normal	1.75 in
			Jul	3.90	2.3	Normal	3.75 in
			Aug	3.30	2.4	Normal	4.75 in
			Sep	4.10	3.32	Normal	5.0 in
			Oct	8.90	3.67	Normal	1.75 in
TX-23 Trial Period: 28 Jun 2009 to 08 Nov 2009	TX	Trial:- NOAA 419346 Decatur, TX (~19 miles) Rainfall (on sit)e Historical: NOAA 412334 Decatur, TX (~19 miles)	Jun	0.06	5.04	Normal	None
			Jul	4.14	1.85	Normal	None
			Aug	1.46	1.86	Normal	None
			Sep	4.97	2.80	Normal	None
			Oct	10.65	3.18	Normal	None
			Nov	NA	2.41	Normal	None

Table 37: Precipitation during Test Period (typically planting to last sampling) (continued)

Trial	State	Historical Weather Source (distance from site)	Month 2009	Rainfall: Trial Monthly Total inches	Rainfall: Historical Monthly Total inches	Rainfall: Difference from Historical	Irrigation Type/ inches
ON-24 Trial Period: 03 Jun 2009 to 17 Nov 2009	Ontario	Trial: VARS WS1 Watchdog 900 ET VARS Facility (~150 meters) Historical : Environment Canada Waterloo-Wellington Station A Waterloo Intl Airport (~30 kilometers)	Jun	3.48	3.2	Normal	None
			Jul	4.2	3.61	Normal	None
			Aug	5.93	3.4	Wetter	None
			Sep	1.65	3.38	Drier	None
			Oct	3.53	2.58	Wetter	None
			Nov	1.66	3.26	Drier	None
ON-25 Trial Period: 05 Jun 2009 to 18 Nov 2009	Ontario	Trial and Historical:– Ridgetown RCS Ridgetown, Ontario Ridgetown College WMO ID 71307 (~33 kilometers)	Jun	2.57	3.23	Drier	None
			Jul	1.2	3.65	Drier	None
			Aug	3.64	4.13	Drier	None
			Sep	1.42	3.66	Drier	None
			Oct	2.76	2.18	Wetter	None
			Nov	1.19	3.67	Drier	None

(a) NA = not available or not applicable

7. REFERENCES

1. Residue Chemistry Test Guidelines, OPPTS 860.1500, Crop Field Trials, EPA 712-C-96-183, August 1996.

2. Residue Chemistry Test Guidelines, OPPTS 860.1520, Processed Food/Feed, EPA 712-C-96-184, August 1996.

3. Pest Management Regulatory Agency, Health Canada, Residue Chemistry Guidelines, Section 9: Crop Field Trials, Regulatory Directive Dir98-02, June 1998.

4. U.S. Environmental Protection Agency. 1989. Pesticide Programs; Good Laboratory Practice Standards; Final Rule (40 CFR, Part 160). *Federal Register*, Vol. 54, No. 158: 34052-34074.

5. U.S. Environmental Protection Agency. 1986. PR Notice 86-5. Standard Format for Data Submitted under the Federal Herbicide, Fungicide and Rodenticide Act (FIFRA) and Certain Provisions of the Federal Food, Drug and Cosmetic Act (FFDCA). 17 pages.

Appendix B – Processing Phase Summary

STUDY TITLE:

Magnitude of the Residue of 2,4-D and Quizalofop-P-ethyl in/on Herbicide Tolerant Field Corn Containing the Aryloxyalkanoate Dioxygenase-1 (AAD-1) Gene

PROCESSING REPORT:

Corn: Generation of Aspirated Grain Fractions and Batch Processing by Wet and Dry Milling Methods

PROCESSING FACILITY:

GLP Technologies
22273 State Highway 6 South
Navasota, Texas 77868

AUTHOR:

Dick L. Dusek
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Navasota, Texas 77868

SPONSOR:

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STUDY DIRECTOR:

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1730 Denham Road
Sycamore, Georgia 31790

STUDY IDENTIFICATION:

Study Number: ARA-09-15-10

Dick Dusek — **Author Signature**   June 3, 2010 — **Date**

GLP COMPLIANCE STATEMENT

STUDY TITLE: Magnitude of the Residue of 2,4-D and Quizalofop-P-ethyl in/on Herbicide Tolerant Field Corn Containing the Aryloxyalkanoate Dioxygenase-1 (AAD-1) Gene

This processing study was conducted and reported in accordance with the Environmental Protection Agency's Good Laboratory Practice Standards, 40 CFR 160, Federal Register, effective date October 16, 1989.

Dick L. Dusek
Processing Principal Investigator

June 3, 2010
Date

QUALITY ASSURANCE STATEMENT

STUDY TITLE: Magnitude of the Residue of 2,4-D and Quizalofop-P-ethyl in/on Herbicide Tolerant Field Corn Containing the Aryloxyalkanoate Dioxygenase-1 (AAD-1) Gene

In compliance with the Good Laboratory Practice regulations an inspector with the Quality Assurance Unit has inspected at least one phase of this study. Inspection findings were reported to GLP Technologies management, the study director and the study director's management. The Quality Assurance Unit has reviewed the processing report and certifies that it accurately describes the methods and standard operating procedures used, and the reported results accurately reflect the raw data generated during this processing phase.

Signed: ______________________ Date: 03 Jun 2010

Doyle L. Borchgardt
Quality Assurance Coordinator
GLP Technologies

INSPECTION		DATES REPORTED TO:	
TYPE	DATE	GLP TECHNOLOGIES MANAGER	STUDY DIRECTOR & STUDY DIRECTOR'S MANAGEMENT
1) Process Phase Trial No.: GA03 SOP G.3 R03 Sec. B.4: "Starch and Gluten Recovery"	02 thru 04 Feb 2010	09 Feb 2010	24 Feb 2010
2) Process Report	22 May 2010	22 May 2010	03 Jun 2010

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STUDY TITLE: Magnitude of the Residue of 2,4-D and Quizalofop-P-ethyl in/on Herbicide Tolerant Field Corn Containing the Aryloxyalkanoate Dioxygenase-1 (AAD-1) Gene

SPONSOR: Dow AgroSciences, LLC
Indianapolis, Indiana 46268

STUDY DIRECTOR: John "Jack" F. Culligan
Ag Research Associates, LLC (ARA)
Sycamore, Georgia 31790

PRINCIPAL PROCESSING INVESTIGATOR: Dick L. Dusek
GLP Technologies
Navasota, Texas 77868

PROCESSING, DATA RECORDING
& SHIPPING TECHNICIANS: Timothy R. Adams, Dick L. Dusek, Theodore F. Dusek Jr., K. Todd Hausman, Patrick W. Simecek, Joseph M. Gibson, C. Lee Jordan, Charles L. Schaefer and Luke S. Wagner

SAMPLE RECEIPT DATE: October 28, 2009 (Trial No. GA-03)
December 16, 2009 (Trial No. NE-14)

PROCESSING START DATE: January 25, 2010 (Trial No. GA-03)
February 8, 2010 (Trial No. NE-14)

PROCESSING TERMINATION DATE: February 15, 2010 (Trial No. GA-03)
February 23, 2010 (Trial No. NE-14)

FRACTION SHIPMENT DATE: February 16, 2010 (Trial No. GA-03)
March 2, 2010 (Trial No. NE-14)

INTRODUCTION:

Field corn samples received from two field trials were used to produce and collect commercially representative aspirated grain fractions and processed food/feed fractions. These fractions were sent to Dow AgroSciences, LLC in Indianapolis, Indiana.

TEST SUBSTANCES: [From protocol]

1.	Product Name:	Weedar 64®
	Active Ingredient:	2,4-Dichlorophenoxyacetic acid, dimethylamine salt
	CAS Number:	94-75-7
	Sponsor TSN Number:	TSN026491-0010
2.	Product Name:	Assure II Herbicide
	Active Ingredient:	Ethyl(R)-2-[4-(6-chloroquinoxalin-2-yl oxy)-phenoxy] propionate
	CAS Number:	76578-14-8
	Sponsor TSN Number:	TSN020252-0004

OBJECTIVE:

GLP Technologies' objective was to produce and collect commercially representative aspirated grain and processed food/feed fractions from the field corn samples grown in two field trials.

METHODS & MATERIALS:

Sample Receipt:

Trial No. GA-03

Three frozen field corn samples (RAC [Raw Agricultural Commodity]) were received from Chris Cromer with ARA in Sycamore, Georgia. Samples were delivered by ARA freezer truck on October 28, 2009.

Trial No. NE-14

Three frozen field corn samples (RAC [Raw Agricultural Commodity]) were received from Matt Krause with ARA in York, Nebraska. Samples were delivered by ARA freezer truck on December 16, 2009.

After receipt and inventory, all samples were stored in freezer "F1."

Storage Conditions:

GLP Technologies SOP D.2 "Storage of Samples in Freezers" requires that freezer temperatures be maintained at or below 10 degrees Fahrenheit with the exception of the defrost cycle and removal and placement of samples in the freezers.

Recorded in the data are the times and dates for removal or placement of samples/fractions in freezers or coolers. Processed fractions requested by the protocol were stored in freezer "F1." Included with this report are copies of temperature charts for applicable units.

Sample/Fraction Handling:

Samples were handled in a manner that minimizes the possibility of contamination. It is this facility's policy to use only containers and utensils washed with detergent and rinsed with water.

Processing Methods:

Samples from trial number GA-03 were identified and processed in the following order: 012-0001 (Control, T1), 012-0002 (Treated, T4), and 012-0003 (Treated, T5). Samples from trial number NE-14 were identified and processed in the following order: 081-0001 (Control, T1), 081-0002 (Treated, T4), and 081-0003 (Treated, T5). The two trials were processed separately.

Drying (For Generation of Aspirated Grain Fractions)

Moisture content of the incoming corn samples was determined. If the moisture content was greater than 13.0%, samples were dried in a Steelman Industries oven at 110-135°F to a moisture content of 10-13.0%. All samples required drying.

Generation of Aspirated Grain Fraction

To generate aspirated grain fractions, the samples were placed in a dust generation room containing a holding bin, two bucket conveyors, and a screw conveyor. As the samples were moved in the system, aspiration was used to remove light impurities (grain dust). Samples were moved for 120 minutes. Light impurities were classified using the following sieves: 2360 micron (8 mesh); 2000 micron (10mesh); 1180 micron (16 mesh); 850 micron (20 mesh); and 425 micron (40 mesh). After classification of each sample, the material through the 2360 micron sieve was recombined according to Study Director instructions to produce one aspirated grain fraction (AGF). A representative sample of the AGF was removed and the ash content was determined according to The American Oil Chemists' Society (AOCS) Method Ba 5a-49.

This generation of aspirated grain fractions procedure is outlined in form H.217 "Material Balance for Generation, Classification, and Ash Content Determination of Aspirated Grain Fractions" and is described in detail in SOP G.17 Revision 02, "Generation, Classification, and Ash Content Determination of Aspirated Grain Fractions."

Cleaning

Following dust generation, samples were cleaned by aspiration and screening. Light impurities were separated from the sample using a Kice aspirator. After aspiration, the sample was screened in a Hance Corporation cleaner to separate large and small foreign particles (screenings) from the whole corn sample.

Dry Milling Process

Representative samples of cleaned whole corn were moisture conditioned to 21.0% and tempered for approximately 2 hours. After tempering, the samples were fed into a C. S. Bell disc mill to crack the kernel. Corn stock from the mill was dried in a Steelman Industries oven for 30 minutes at 130-160°F. Dried cornstock was screened on the Hance cleaner fitted with a 1/8" screen to separate bran, germ, and large grits from grits, meal and flour.

Material on top of the 1/8" screen (bran, germ, and large grits) was aspirated with a Kice aspirator to remove bran from the germ, germ with attached hull and endosperm, and large grits. This material was again screened with the Hance cleaner equipped with a 10/64" and 14/64" (or similar size) screens to separate large grits and germ from germ with attached hull and endosperm. Germ with attached hull and endosperm was passed through the disc mill and screened with the Hance cleaner fitted with the 1/8" screen. Material on top of the screen was aspirated to remove detached bran. Material through the 1/8" screen was collected and weighed as large grits. These steps were repeated to reduce the amount of endosperm and bran in the germ. Germ and large grits passing through the 13/64" (or similar size) screen were separated using the Forsberg gravity separator. The two germ fractions were combined, weighed, and dried in the Steelman Industries oven at 130-160°F until the moisture content was 14-16%.

Material passing through the 1/8" screen was separated using a Great Western sifter equipped with a 14 mesh (0.0540") and 62 mesh (0.0098") screens. Material on top of the 14 mesh screen was collected and weighed as grits. Material through the 14 mesh and on top of the 62 mesh screen was collected and weighed as meal. Material through the 62 mesh screen was collected and weighed as flour. Grit fractions requested by protocol were a combination of large grits from the Forsberg gravity separator and grits from the Great Western. Grits, meal and flour fractions were collected and placed into freezer storage.

Germ material was heated to 175-195°F in a Marion mixer and held for 10 minutes. Heated material was flaked in an A. T. Ferrell flaking roll with a gap setting of 0.007"-0.010". Flaked kernel material was placed in stainless steel batch extractors and submerged in 120-140°F solvent (hexane). After 30 minutes, the miscella (crude oil and hexane) was drained and fresh hexane added to repeat the cycle two more times. Final two washings were for 15-20 minutes each. Following the final draining, the spent flakes were desolventized with ambient air or in a Reliance steam heated mixer to remove residual hexane. The resulting fractions from solvent extraction were miscella and solvent extracted germ meal.

Miscella was passed through a laboratory vacuum evaporator to separate the crude oil and hexane. Crude oil was then heated to 195-205°F for hexane removal. Crude oil was filtered before refining. For alkali refining, a weighed amount of crude oil and 16 degree Baumé sodium hydroxide was placed in a water bath and mixed for 15 minutes at high RPM at 68-75°F, and then for 12 minutes at low RPM at 145-153°F. Neutralized oil was centrifuged. Refined oil was decanted and filtered. Resulting fractions were alkali refined oil and soapstock. Refined oil was collected and placed into frozen storage. Soapstock was discarded.

Wet Milling Process

A representative sample of cleaned whole corn was steeped in 120-130°F water containing 0.1-0.2% sulfur dioxide (sulfurous acid) for 22-48 hours. At the end of the steeping period, the whole corn was passed through a C. S. Bell disc mill and a majority of the germ and hull was removed using a Dorr-Oliver hydroclone (water centrifuge).

Germ and hull were dried at 165-195°F to obtain a final moisture between 5-10%. After drying, the germ and hull were separated using aspiration and screening.

Cornstock (without germ and hull) was ground in the C. S. Bell disc mill. Ground material was passed over a Dynascreen equipped with a 325 mesh (50 micron) screen. In commercial industry, only bran (hull material) remains on top of the screen. Material on top of the screen was discarded. Process water (with starch and gluten) passing through the screen was separated into starch and gluten by batch centrifugation. Starch was dried at 130-160°F until the moisture content was 15.0% or less. Requested starch fractions were collected and placed into frozen storage.

Germ samples were moisture conditioned to 12%, heated to 190-220°F in a Marion mixer, flaked in a A. T. Ferrell flaking roll with a gap setting of 0.008"-0.012", and pressed in a Komet expeller to liberate part of the crude oil. Resulting fractions are expelled crude oil and presscake with residual crude oil.

Presscake was placed in stainless steel batch extractors and submerged in 120-140°F solvent (hexane). After 30 minutes, the miscella was drained and fresh hexane was added to repeat the cycle two more times. Final two washings were for 15 minutes each. Following the final draining, the spent presscake was desolventized with ambient air to remove residual hexane. The resulting fractions from solvent extraction were miscella and solvent extracted presscake (germ cake).

Miscella was passed through a laboratory vacuum evaporator to separate the crude oil and hexane. Crude oil was heated to 195-205°F for hexane removal.

Crude oil from expelling and solvent extraction was filtered and combined for refining. Crude oil samples from the wet milling process were refined using the same methods as in the dry milling procedure.

This processing procedure is outlined in form H.203 "Corn Processing Material Balance" and is described in detail in SOP G.4 Revision 03, "Corn: Batch Processing Method (Wet and Dry Milling)."

Remaining unprocessed whole corn samples from both trials were devitalized by grinding and discarded per facility SOP. No whole corn was observed in the ground material.

Comparison to Industrial Practice:

Generation of aspirated grain fractions simulates industrial practice used in terminal elevators to remove grain dust. Prevention of grain dust explosions is the reason for its removal. Because of compliance monitoring requirements and sample size, the samples were processed by batch rather than continuous, as in commercial operation.

Typically, softer varieties of corn are wet milled. Varieties with medium to hard endosperm are dry milled. In GLP Technologies' particular situation, we had only one variety of corn to wet and dry mill. Consequently, either the wet or dry mill procedure will provide better processing results.

Dry milling closely simulates commercial dry milling practices. Slight variations in industrial milling practices are designed to suit the buyer's needs. In comparison, the goal is to produce the same component parts for each sample within a study to be used in residue determination.

Wet milling simulates industrial practice as closely as possible. Due to equipment limitations and batch processing the material balance values for wet milling products were estimated using percentages from the CRC Handbook of Processing and Utilization in Agriculture. Fraction yields obtained by industry are not made public. Yields from commercial wet milling plants will vary between plants depending on type and quality of the corn and differences in milling practice and fiber and germ washing operations. The following table is for approximate yields.

Solubles from steeping	-	7.5%
Starch	-	67.5%
Gluten	-	5.8%
Germ	-	7.5%
Hull	-	11.5%

Processing Results:

Field corn samples were processed to collect aspirated grain fractions. Duplicated fractions of grits, meal, flour, dry milled alkali refined oil, starch, and alkali refined oil were collected during processing. Triplicated fractions of unprocessed whole corn (RAC) and whole corn (after drying and dust generation) were also collected.

Other Circumstances Pertaining to Study:

The following protocol deviations were reported to the Study Director via email:

1. Protocol requested approximately 1 liter of refined oil be collected. A liter of oil weighs approximately 800 grams. Refined oil samples from the dry milled portion for both trials failed to meet this requirement. Samples from NE-14 ranged from 370-409 grams and samples from GA-03 ranged from 303-361 grams.

2. Protocol requested report and raw data be completed and sent to the Study Director within approximately 6 weeks after all processed fractions are shipped

to Dow. Last fraction shipment was March 2, 2010. Report and data were not completed by April 13, 2010.

The following facility SOP deviations were reported to the Study Director via email:

1. (SOP G.3) Starch is dried at 130-160°F until the moisture content is less than 15.0%. Maximum recorded temperatures for samples 02-0001 (T1) and 012-0002 (T4) from trial GA-03 were 165.5°F and 161.9°F respectively.

2. (SOP D.11) "Pre-Process Verification will be completed prior to processing samples. This ensures that a visual inspection of the machine was performed and that it is clean and operational. The processing personnel will record the date, unique identifying number (test or protocol number pertinent to study and sample number [if applicable]), and commodity to be processed, and will initial the entries. Cleaning will be performed and documented prior to the next processing study." No pre-process verification or cleaning is documented for the Marion mixer, A.T. Ferrell flaking roll, Komet expeller and Recovery Unit #1. All machines were used on February 5, 2010 during expelling and solvent extraction of wet-milled corn germ from trial GA-03.

3. (SOP E.8) "During dispensing, technician will log in date, study number, commodity and initials." This information was not recorded into the dispense logbook for the sulfurous acid used to steep wet milled corn from sample 081-0003 (T5) from trial NE-14.

4. (SOP G.3) "Washed germ and fiber are dried in a Steelman Industries oven at 165-195°F for a final moisture of 5 to 10%" Samples 081-0001 (T1) and 081-0003 (T5) from trial NE-14 were dried to 3.2% and 4.7%, respectively.

5. (SOP G.3) "Miscella is separated in a laboratory vacuum evaporator. During this procedure, crude oil reaches a temperature of 195-205°F." Maximum temperature is not recorded for sample 081-0002 (T4) from the dry milled portion.

These deviations did not have a negative impact on processing.

Fraction Shipment:

Frozen fractions packed in dry ice were shipped to Sample Management at Dow AgroSciences, LLC in Indianapolis, Indiana via Federal Express priority overnight delivery. Fractions were shipped on February 16, 2010 (GA-03) and March 2, 2010 (NE-14). "Shipment of Fractions (Chain of Custody)" and "Fraction Shipment and Packing List" forms accompanied each shipment.

CONCLUSIONS:

Commercially representative corn aspirated grain and processed food/feed fractions were produced and collected from the field corn samples received from the two trial locations.

DATA ARCHIVAL:

Record Transfer and Retention:

This processing report and raw data have been transferred to the Study Director, John "Jack" F. Culligan at Ag Research Associates, LLC in Sycamore, Georgia.

GLP Technologies will archive the following study specific data:

-copy of the sponsor processing protocol
-exact copy of the processing report (main body)
-exact copy of the compliance statement
-exact copy of the sample material balance
-exact copy of the original raw processing data (includes communication logs, calculations, and deviation forms, when applicable)
-exact copy of personnel records (names and initials of personnel with processing study duties)
-exact copy of receiving records
-exact copy of shipping records
-exact copy of shipping bills of lading

GLP Technologies will archive the following non-study specific data indefinitely:

-original freezer temperature records
-original equipment logs (includes scales, temperature recording devices, and processing equipment records)
-CVs of personnel and training records

FORM H.217 Revision 00

MATERIAL BALANCE for GENERATION, CLASSIFICATION, AND ASH CONTENT DETERMINATION OF ASPIRATED GRAIN FRACTIONS

GA-03

Sample # **1 (Control, T1)** Code # **012-0001**

COMMODITY 1295.1 lbs.

Drying 1215.7 lbs. (after drying)

700.0 lbs. used for generation

Aspiration 0.9 lbs.

Classification

- **ASPIRATED GRAIN FRACTION > 2360 micron** 30.5 g
- **ASPIRATED GRAIN FRACTION > 2000 micron** 22.9 g
- **ASPIRATED GRAIN FRACTION > 1180 micron** 53.7 g
- **ASPIRATED GRAIN FRACTION > 850 micron** 19.1 g
- **ASPIRATED GRAIN FRACTION > 425 micron** 17.1 g
- **ASPIRATED GRAIN FRACTION < 425 micron** 277.4 g

Amounts recombined for AGF fraction:

22.9 g < 2360 micron and > 2000 micron
53.6 g > 1180 micron
19.0 g > 850 micron
17.1 g > 425 micron
277.3 g < 425 micron

ASH CONTENT: 1.6 %

FORM H.203 Revision 00
CORN PROCESSING MATERIAL BALANCE

GA-03

Sample # **1 (Control, T1)** Code # **012–0001**

WHOLE CORN 685.9 lbs.

Aspiration 4.9 lbs. **LIGHT IMPURITIES**

Screening 46.2 lbs. **SMALL SCREENINGS**
1.3 lbs. **LARGE SCREENINGS**

CLEANED CORN 632.9 lbs.

Cleaned Corn Dry Milled 225.0 lbs.

Steeping 26.5 lbs water added

Degermination, Drying, Screening, Aspiration, and Separation

27.5 lbs. **GRITS***
27.0 lbs. **MEAL***
7.1 lbs. **FLOUR***
8.1 lbs. **BRAN**
137.6 lbs. **LARGE GRITS**

GERM 35.5 lbs. (Dried to 33.3 lbs.)

Conditioning, Flaking, & Solvent Extraction 33.3 lbs. Germ Conditioned

CRUDE OIL 727 g
S. EXT. GERM FLAKES 27.9 lbs.

Refining
680 g Refined 32.6 g NaOH added

REFINED OIL 617 g **SOAPSTOCK** 81 g

* Calculated amounts.

Cleaned Corn Wet Milled 175.0 lbs

Steeping 34.0 gal water added

Draining **STEEPWATER** 19 gal

Steeped Corn 278.4 lbs.
Solubles from steeping** 13.1 lbs.

Degermination, Separation, Screening, and Water Washing

20.1 lbs. **FIBER**** 118.1 lbs. **STARCH****
10.2 lbs. **GLUTEN**** 13.1 lbs. **GERM****

Flaking, Conditioning, & Expelling
Germ pressed 10.6 lbs.
Water added 377.3 g

CRUDE OIL 889 g
PRESSCAKE 8.5 lbs.

Solvent Extraction

CRUDE OIL 739 g
S. EXT. GERM CAKE 6.0 lbs.

Refining
1550 g Refined 66.7 g NaOH added

REFINED OIL 1464 g **SOAPSTOCK** 124 g

** Calculated amounts based on commercial recovery percentages and starting weight of corn used for wet milling.

FORM H.217 Revision 00

MATERIAL BALANCE for GENERATION, CLASSIFICATION, AND ASH CONTENT DETERMINATION OF ASPIRATED GRAIN FRACTIONS

GA-03

Sample # **2 (Treated, T4)** Code # **012–0002**

COMMODITY 1090.4 lbs.

Drying 1038.0 lbs. (after drying)

700.0 lbs. used for generation

Aspiration 0.8 lbs.

Classification

- **ASPIRATED GRAIN FRACTION > 2360 micron** 44.2 g
- **ASPIRATED GRAIN FRACTION > 2000 micron** 26.5 g
- **ASPIRATED GRAIN FRACTION > 1180 micron** 51.8 g
- **ASPIRATED GRAIN FRACTION > 850 micron** 20.0 g
- **ASPIRATED GRAIN FRACTION > 425 micron** 18.5 g
- **ASPIRATED GRAIN FRACTION < 425 micron** 179.4 g

Amounts recombined for AGF fraction:

26.5 g < 2360 micron and > 2000 micron
51.7 g > 1180 micron
20.0 g > 850 micron
18.4 g > 425 micron
179.3 g < 425 micron

ASH CONTENT: 1.5 %

FORM H.203 Revision 00
CORN PROCESSING MATERIAL BALANCE

GA-03

Sample # **2 (Treated, T4)** Code # **012–0002**

WHOLE CORN 687.6 lbs.

Aspiration 5.2 lbs. **LIGHT IMPURITIES**

Screening 52.6 lbs. **SMALL SCREENINGS**
7.2 lbs. **LARGE SCREENINGS**

CLEANED CORN 622.6 lbs.

Cleaned Corn Dry Milled 225.0 lbs.

Steeping 26.5 lbs water added

Degermination, Drying, Screening, Aspiration, and Separation

24.3 lbs. **GRITS***
20.3 lbs. **MEAL***
4.0 lbs. **FLOUR***
8.1 lbs. **BRAN**
154.9 lbs. **LARGE GRITS**

GERM 31.1 lbs. (Dried to 30.2 lbs.)

Conditioning, Flaking, & Solvent Extraction 30.2 lbs. Germ Conditioned

CRUDE OIL 868 g
S. EXT. GERM FLAKES 24.9 lbs.

Refining
811 g Refined 40.6 g NaOH added

REFINED OIL 731 g
SOAPSTOCK 104 g

* Calculated amounts.

Cleaned Corn Wet Milled 175.0 lbs

Steeping 30.0 gal water added

Draining — **STEEPWATER** 17 gal

Steeped Corn 272.3 lbs.
Solubles from steeping** 13.1 lbs.

Degermination, Separation, Screening, and Water Washing

20.1 lbs. **FIBER**** 118.1 lbs. **STARCH****
10.2 lbs. **GLUTEN**** 13.1 lbs. **GERM****

Flaking, Conditioning, & Expelling
Germ pressed 11.0 lbs.
Water added 374.6 g

CRUDE OIL 938 g
PRESSCAKE 8.7 lbs.

Solvent Extraction

CRUDE OIL 926 g
S. EXT. GERM CAKE 6.4 lbs.

Refining
1749 g Refined 73.5 g NaOH added

REFINED OIL 1640 g
SOAPSTOCK 153 g

** Calculated amounts based on commercial recovery percentages and starting weight of corn used for wet milling.

FORM H.217 Revision 00

MATERIAL BALANCE for GENERATION, CLASSIFICATION, AND ASH CONTENT DETERMINATION OF ASPIRATED GRAIN FRACTIONS

GA-03

Sample # **3 (Treated, T5)** Code # **012–0003**

COMMODITY 873.1 lbs.

Drying 811.2 lbs. (after drying)

700.0 lbs. used for generation

Aspiration 1.5 lbs.

Classification

- **ASPIRATED GRAIN FRACTION > 2360 micron** 58.1 g
- **ASPIRATED GRAIN FRACTION > 2000 micron** 42.8 g
- **ASPIRATED GRAIN FRACTION > 1180 micron** 56.6 g
- **ASPIRATED GRAIN FRACTION > 850 micron** 23.6 g
- **ASPIRATED GRAIN FRACTION > 425 micron** 25.3 g
- **ASPIRATED GRAIN FRACTION < 425 micron** 457.7 g

Amounts recombined for AGF fraction:

42.1 g < 2360 micron and > 2000 micron
56.5 g > 1180 micron
23.5 g > 850 micron
25.3 g > 425 micron
457.5 g < 425 micron

ASH CONTENT: 1.5 %

FORM H.203 Revision 00
CORN PROCESSING MATERIAL BALANCE

GA-03

Sample # **3 (Treated, T5)** Code # **012–0003**

WHOLE CORN 686.1 lbs.
Aspiration 11.2 lbs. **LIGHT IMPURITIES**
Screening 49.1 lbs. **SMALL SCREENINGS**
10.8 lbs. **LARGE SCREENINGS**
CLEANED CORN 614.9 lbs.

Cleaned Corn Dry Milled 225.0 lbs.
Steeping 24.8 lbs water added
Degermination, Drying, Screening, Aspiration, and Separation
28.5 lbs. **GRITS***
25.7 lbs. **MEAL***
7.0 lbs. **FLOUR***
12.1 lbs. **BRAN**
139.8 lbs. **LARGE GRITS**
GERM 26.8 lbs. (Dried to 26.2 lbs.)
Conditioning, Flaking, & Solvent Extraction 26.2 lbs. Germ Conditioned
CRUDE OIL 866 g
S. EXT. GERM FLAKES 21.7 lbs.
Refining
806 g Refined 41.1 g NaOH added
REFINED OIL 729 g **SOAPSTOCK** 109 g

* Calculated amounts.

Cleaned Corn Wet Milled 175.0 lbs
Steeping 30.0 gal water added
Draining —— **STEEPWATER** 17.5 gal
Steeped Corn 354.0 lbs.
Solubles from steeping** 13.1 lbs.
(Possible weighing/recording error)
Degermination, Separation, Screening, and Water Washing
20.1 lbs. **FIBER**** 118.1 lbs. **STARCH****
10.2 lbs. **GLUTEN**** 13.1 lbs. **GERM****
Flaking, Conditioning, & Expelling
Germ pressed 10.6 lbs.
Water added 300.8 g
CRUDE OIL failed to record g
PRESSCAKE 8.2 lbs.
Solvent Extraction
CRUDE OIL 793 g **S. EXT. GERM CAKE** 6.2 lbs.
Refining
1715 g Refined 72.0 g NaOH added
REFINED OIL 1632 g **SOAPSTOCK** 124 g

** Calculated amounts based on commercial recovery percentages and starting weight of corn used for wet milling.

FORM H.217 Revision 00

MATERIAL BALANCE for GENERATION, CLASSIFICATION, AND ASH CONTENT DETERMINATION OF ASPIRATED GRAIN FRACTIONS

NE-14

Sample # **1 (Control, T1)** Code # **081-0001**

COMMODITY 1191.1 lbs.

Drying 1032.9 lbs. (after drying)

700.0 lbs. used for generation

Aspiration 1.0 lbs.

Classification

- **ASPIRATED GRAIN FRACTION > 2360 micron** 60.2 g
- **ASPIRATED GRAIN FRACTION > 2000 micron** 27.9 g
- **ASPIRATED GRAIN FRACTION > 1180 micron** 66.0 g
- **ASPIRATED GRAIN FRACTION > 850 micron** 35.2 g
- **ASPIRATED GRAIN FRACTION > 425 micron** 45.1 g
- **ASPIRATED GRAIN FRACTION < 425 micron** 219.2 g

Amounts recombined for AGF fraction:

27.4 g < 2360 micron and > 2000 micron
65.3 g > 1180 micron
35.0 g > 850 micron
45.0 g > 425 micron
218.9 g < 425 micron

ASH CONTENT: 2.9 %

FORM H.203 Revision 00
CORN PROCESSING MATERIAL BALANCE

NE-14

Sample # **1 (Control, T1)** Code # **081–0001**

WHOLE CORN 683.3 lbs.

Aspiration 7.2 lbs. **LIGHT IMPURITIES**

Screening 34.4 lbs. **SMALL SCREENINGS**
16.0 lbs. **LARGE SCREENINGS**

CLEANED CORN 624.8 lbs.

Cleaned Corn Dry Milled 225.0 lbs.

Steeping 25.1 lbs water added

Degermination, Drying, Screening, Aspiration, and Separation

19.6 lbs. **GRITS***
21.2 lbs. **MEAL***
4.8 lbs. **FLOUR***
13.1 lbs. **BRAN**
144.2 lbs. **LARGE GRITS**

GERM 36.0 lbs. (Dried to 35.8 lbs.)

Conditioning, Flaking, & Solvent Extraction 35.8 lbs. Germ Conditioned

CRUDE OIL 970 g
S. EXT. GERM FLAKES*** 34.9 lbs.

Refining
888 g Refined 45.3 g NaOH added

REFINED OIL 785 g **SOAPSTOCK** 118 g

* Calculated amounts.

*** Weight gain due to residual hexane in flakes.

Cleaned Corn Wet Milled 175.0 lbs

Steeping 33.0 gal water added

Draining —— **STEEPWATER** 18 gal

Steeped Corn 282.7 lbs.
Solubles from steeping** 13.1 lbs.

Degermination, Separation, Screening, and Water Washing

20.1 lbs. **FIBER**** 118.1 lbs. **STARCH****
10.2 lbs. **GLUTEN**** 13.1 lbs. **GERM****

Flaking, Conditioning, & Expelling
Germ pressed 9.4 lbs.
Water added 426.8 g

CRUDE OIL 1108 g
PRESSCAKE 6.7 lbs.

Solvent Extraction

CRUDE OIL 685 g
S. EXT. GERM CAKE 5.0 lbs.

Refining
1675 g Refined 68.7 g NaOH added

REFINED OIL 1579 g **SOAPSTOCK** 110 g

** Calculated amounts based on commercial recovery percentages and starting weight of corn used for wet milling.

FORM H.217 Revision 00

MATERIAL BALANCE for GENERATION, CLASSIFICATION, AND ASH CONTENT DETERMINATION OF ASPIRATED GRAIN FRACTIONS

NE-14

Sample # **2 (Treated, T4)** Code # **081–0002**

COMMODITY 1131.2 lbs.

Drying 986.7 lbs. (after drying)

700.0 lbs. used for generation

Aspiration 0.9 lbs.

Classification

- **ASPIRATED GRAIN FRACTION > 2360 micron** 60.7 g
- **ASPIRATED GRAIN FRACTION > 2000 micron** 28.7 g
- **ASPIRATED GRAIN FRACTION > 1180 micron** 67.2 g
- **ASPIRATED GRAIN FRACTION > 850 micron** 42.2 g
- **ASPIRATED GRAIN FRACTION > 425 micron** 56.1 g
- **ASPIRATED GRAIN FRACTION < 425 micron** 199.1 g

Amounts recombined for AGF fraction:

28.5 g < 2360 micron and > 2000 micron
66.9 g > 1180 micron
41.9 g > 850 micron
55.9 g > 425 micron
198.8 g < 425 micron

ASH CONTENT: 2.4 %

FORM H.203 Revision 00
CORN PROCESSING MATERIAL BALANCE

NE-14

Sample # **2 (Treated, T4)** Code # **081–0002**

WHOLE CORN 685.7 lbs.

Aspiration 6.5 lbs. **LIGHT IMPURITIES**

Screening 34.4 lbs. **SMALL SCREENINGS**
34.2 lbs. **LARGE SCREENINGS**

CLEANED CORN 609.8 lbs.

Cleaned Corn Dry Milled 225.0 lbs.

Steeping 26.5 lbs water added

Degermination, Drying, Screening, Aspiration, and Separation

20.0 lbs. **GRITS***
22.9 lbs. **MEAL***
4.9 lbs. **FLOUR***
14.5 lbs. **BRAN**
144.9 lbs. **LARGE GRITS**

GERM 36.5 lbs. (Dried to 35.2 lbs.)

Conditioning, Flaking, & Solvent Extraction 35.2 lbs. Germ Conditioned

CRUDE OIL 926 g

S. EXT. GERM FLAKES 34.5 lbs.***

Refining
834 g Refined 42.5 g NaOH added

REFINED OIL 746 g **SOAPSTOCK** 113 g

* Calculated amounts.

*** Weight gain due to residual hexane in flakes.

Cleaned Corn Wet Milled 175.0 lbs

Steeping 33.0 gal water added

Draining — **STEEPWATER** 18 gal

Steeped Corn 279.9 lbs.

Solubles from steeping** 13.1 lbs.

Degermination, Separation, Screening, and Water Washing

20.1 lbs. **FIBER**** 118.1 lbs. **STARCH****
10.2 lbs. **GLUTEN**** 13.1 lbs. **GERM****

Flaking, Conditioning, & Expelling Germ pressed 12.1 lbs.
Water added 405.8 g

CRUDE OIL 1114 g

PRESSCAKE 9.4 lbs.

Solvent Extraction

CRUDE OIL 1170 g **S. EXT. GERM CAKE** 6.7 lbs.

Refining
2122 g Refined 87.0 g NaOH added

REFINED OIL 2000 g **SOAPSTOCK** 182 g

** Calculated amounts based on commercial recovery percentages and starting weight of corn used for wet milling.

FORM H.217 Revision 00

MATERIAL BALANCE for GENERATION, CLASSIFICATION, AND ASH CONTENT DETERMINATION OF ASPIRATED GRAIN FRACTIONS

NE-14

Sample # **3 (Treated, T5)** Code # **081–0003**

COMMODITY 1145.0 lbs.

Drying 1013.0 lbs. (after drying)

700.0 lbs. used for generation

Aspiration 0.9 lbs.

Classification

- **ASPIRATED GRAIN FRACTION > 2360 micron** 52.3 g
- **ASPIRATED GRAIN FRACTION > 2000 micron** 20.9 g
- **ASPIRATED GRAIN FRACTION > 1180 micron** 53.4 g
- **ASPIRATED GRAIN FRACTION > 850 micron** 37.1 g
- **ASPIRATED GRAIN FRACTION > 425 micron** 54.3 g
- **ASPIRATED GRAIN FRACTION < 425 micron** 218.8 g

Amounts recombined for AGF fraction:

20.8 g < 2360 micron and > 2000 micron
53.2 g > 1180 micron
36.8 g > 850 micron
54.3 g > 425 micron
218.6 g < 425 micron

ASH CONTENT: 2.3 %

FORM H.203 Revision 00
CORNER PROCESSING MATERIAL BALANCE

NE-14

Sample # **3 (Treated, T5)** Code # **081–0003**

WHOLE CORN 685.7 lbs.
Aspiration 6.8 lbs. **LIGHT IMPURITIES**
Screening 29.5 lbs. **SMALL SCREENINGS**
29.0 lbs. **LARGE SCREENINGS**
CLEANED CORN 620.0 lbs.

Cleaned Corn Dry Milled 225.0 lbs.
Steeping 25.6 lbs water added
Degermination, Drying, Screening, Aspiration, and Separation
17.2 lbs. **GRITS***
19.8 lbs. **MEAL***
4.3 lbs. **FLOUR***
17.9 lbs. **BRAN**
149.8 lbs. **LARGE GRITS**
GERM 29.8 lbs. (Dried to 28.4 lbs.)
Conditioning, Flaking, & Solvent Extraction 28.4 lbs. Germ Conditioned
CRUDE OIL 993 g
S. EXT. GERM FLAKES 28.4 lbs.***
Refining
912 g Refined 45.6 g NaOH added
REFINED OIL 822 g
SOAPSTOCK 108 g

* Calculated amounts.

***Weight gain due to residual hexane in flakes.

Cleaned Corn Wet Milled 175.0 lbs
Steeping 33.0 gal water added
Draining — **STEEPWATER** 18 gal
Steeped Corn 280.0 lbs.
Solubles from steeping** 13.1 lbs.
Degermination, Separation, Screening, and Water Washing
20.1 lbs. **FIBER**** 118.1 lbs. **STARCH****
10.2 lbs. **GLUTEN**** 13.1 lbs. **GERM****
Flaking, Conditioning, & Expelling Germ pressed 11.9 lbs. Water added 448.2 g
CRUDE OIL 1083 g
PRESSCAKE 9.5 lbs.
Solvent Extraction
CRUDE OIL 1235 g
S. EXT. GERM CAKE 6.6 lbs.
Refining
2175 g Refined 89.2 g NaOH added
REFINED OIL 2033 g
SOAPSTOCK 199 g

** Calculated amounts based on commercial recovery percentages and starting weight of corn used for wet milling.

Appendix C – Analytical Phase Summary

ANALYTICAL SUMMARY TITLE

Analytical Summary for Magnitude of the Residue of 2,4-D and Quizalofop-P-ethyl in/on Herbicide Tolerant Field Corn Containing the Aryloxyalkanoate Dioxygenase-1 (AAD-1) Gene

PRINCIPAL ANALYST

B. M. Wendelburg

STUDY DIRECTOR

J. F. Culligan

ANALYTICAL SUMMARY COMPLETION DATE

04-Oct-2010

PERFORMING LABORATORY

Regulatory Sciences and Government Affairs—Indianapolis Lab
Dow AgroSciences LLC
9330 Zionsville Road
Indianapolis, Indiana 46268-1054

STUDY IDENTIFICATION

Protocol Number ARA-09-15-10

STATEMENT OF COMPLIANCE WITH GOOD LABORATORY PRACTICE STANDARDS

Title: Analytical Summary for Magnitude of the Residue of 2,4-D and Quizalofop-P-ethyl in/on Herbicide Tolerant Field Corn Containing the Aryloxyalkanoate Dioxygenase-1 (AAD-1) Gene

Study Initiation Date: 19-May-2009

This report represents data generated after the effective date of the EPA FIFRA Good Laboratory Practice Standards.

United States Environmental Protection Agency
Title 40 Code of Federal Regulations Part 160
FEDERAL REGISTER, August 17, 1989

Organisation for Economic Co-Operation and Development
ENV/MC/CHEM(98)17, Paris January 26, 1998

All aspects of this study were conducted in accordance with the requirements for Good Laboratory Practice Standards, 40 CFR 160, with the following exception.

B. M. Wendelburg Principal Analyst/Author Dow AgroSciences LLC	04Oct2010 Analytical Summary Completion Date
A. S. McGibbon Global Leader - Analytical Sciences, Fate and Metabolism Dow AgroSciences LLC	20 Sep 2010 Date

Dow AgroSciences Quality Assurance Unit

Good Laboratory Practice Statement Page

Study ID: ARA-09-15-10

Title: Analytical Summary for Magnitude of Residue of 2,4-D and Quizalofop-P-ethyl in/on Herbicide Tolerant Field Corn containing the Aryloxyalkanoate Dioxygenase-1 (AAD-1) Gene

Study Initiation Date: 19-May-2010 **Analytical Summary Completion Date:** 4-Oct-2010

GLP Quality Assurance Inspections

Date of GLP Inspection(s)	Date Reported to the Study Director and to Management	Phases that have received a GLP Inspection by the Quality Assurance Unit
11-Jun-2010	11-Jun-2010	Protocol Review
26-May-2010	15-Jun-2010	Dosing and Sample Extraction
8-Jun-2010	16-Jun-2010	Dosing (Repeat)
12-Aug-2010	20-Aug-2010	Dosing and Extraction (Repeat)
20, 21, 22, 23, 24, 27, 28-Sept-2010	30-Sept-2010	Analytical Summary, Raw Data, Reference Substance Verification

QUALITY ASSURANCE STATEMENT:

The Quality Assurance Unit has reviewed the analytical summary and has determined that the summary reflects the raw data generated during the analytical portion of this study.

T Froggatt

Tracey Froggatt
Dow AgroSciences, Quality Assurance

4-Oct-2010

Date

SIGNATURE PAGE

B. M. Wendelburg Author Dow AgroSciences LLC	20Sept 2010 Date
G. E. Dial Jr. Reviewer Dow AgroSciences LLC	20 Sept 2010 Date
J. T. Gesell Reviewer Dow AgroSciences LLC	20 SEP 2010 Date
A. S. McGibbon Manager Dow AgroSciences LLC	20 Sep 2010 Date

STUDY PERSONNEL

Title:	Analytical Summary for Magnitude of the Residue of 2,4-D and Quizalofop-P-ethyl in/on Herbicide Tolerant Field Corn Containing the Aryloxyalkanoate Dioxygenase-1 (AAD-1) Gene
Principal Investigator:	B. M. Wendelburg (current)
	G. E. Schelle (through 08-Jun-10)
Analysts:	D. D. Shackelford
	G. E. Dial Jr.
	L. J. Singletary, Kelly Scientific Resources
	J. A. Taylor, Kelly Scientific Resources
	A. C. Lehman, Kelly Scientific Resources
	M. J. Walter, Kelly Scientific Resources
	V. Dixit, Kelly Scientific Resources

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INTRODUCTION

Samples of corn commodities and processed products were collected according to Study Plan ARA-09-15-10 after treatment. The samples were analyzed for residues of 2,4-dichlorophenoxyacetic acid (2,4-D), 2,4-dichlorophenol (2,4-DCP), quizalofop-ethyl, and quizalofop acid at Dow AgroSciences LLC in Indianapolis, Indiana. This summary describes the analytical phase of the study and presents the analytical results obtained.

Sample Handling

Samples were assigned unique Dow AgroSciences sample numbers that were used to track the samples throughout receipt, preparation, storage, analysis, and reporting. Samples were received frozen and logged into the Regulatory Labs Information Management System (RLIMS). Each sample (with the exception of oil which needed no preparation) was blended with dry ice and ground using an RSI Robo Coupe bowl grinder with a 45-, 30-, or 3-L capacity bowl. After appropriate mixing, each sample was transferred to HDPE freezer containers and stored in temperature-monitored freezers at approximately -20 °C, except when removed to aliquot for analysis. All movements of samples at the Dow AgroSciences facility were documented within the RLIMS system.

Field sample analyses were conducted 18-May-2010 through 30-Aug-2010, as defined by the addition of extraction solution to samples. Table 1 summarizes the dates of sampling, shipment from the field, receipt at Dow AgroSciences, and analysis for each sample group.

ANALYTICAL METHODOLOGY

Reference Materials/Analytical Standards/Internal Standards

The following analytical standards were used as the reference material for calibration and fortification standards: 2,4-D, AGR275828, 99.5% pure (1); 2,4-DCP, AGR182992, 99% pure (2); quizalofop-ester, TSN106317, 99% pure (3); quizalofop acid, TSN106172, 96% pure (4). Certificates of analysis for these reference materials are illustrated in Figures 1-4. A 1000-µg/mL stock solution of each reference standard was prepared in 100 mL of methanol from a 0.1000-g weighed reference standard. A range of 2,4-D and 2,4-DCP mixed stock solutions was prepared in methanol by serial dilutions. A range of quizalofop-ethyl and quizalofop acid mixed stock solutions was prepared in methanol by serial dilutions The reagents 2,4-dichlorophenoxyacetic acid ($^{13}C_6$) , lot #SCHF-006, and 2,4-dichlorophenol ($^{13}C_6$) , lot #SCCJ-003, were used as stable isotope internal standards in the study. The stable isotopes used in this study required evaporative removal of the solvent they were delivered in to allow stock preparation. A stock solution of each stable isotope was prepared in a methanol/water (50:50) solution, and a further dilution containing both internal standards was prepared in a methanol/2N HCl (50:50) solution. A range of mixed calibration standards of 2,4-D and 2,4-DCP, containing 10 ng/mL of the 2,4-dichlorophenoxyacetic acid ($^{13}C_6$) and 2,4-dichlorophenol ($^{13}C_6$) internal standards, was prepared in a methanol/2N HCl (50:50) solution. A range of mixed calibration standards containing quizalofop-ethyl and quizalofop acid was prepared in a methanol/2N HCl (50:50) solution Common and chemical names, molecular formulas, and the nominal masses for the analytes and related compounds are given Table 2.

Analytical Methods

Two analytical methods were used for the determination of 2,4-dichlorophenoxyacetic acid (2,4-D), 2,4-dichlorophenol (2,4-DCP), quizalofop-ethyl, and quizalofop acid in corn commodities. The two methods are outlined separately below:

Analytical Method for 2,4-D and 2,4-DCP Determination in Corn

The analytical method followed for determination of 2,4-D and 2,4-DCP in corn commodities and processed products is documented in the study file. Chromatographic data was collected and integrated using MDS/Sciex Analyst software. Typical chromatograms for these analyses can be found in Figures 5-79.

Samples (with the exception of oil) were weighed into 8-oz HDPE containers in preparation for analysis; the sample weights were all 5.00±0.05 g. Oil samples were weighed into 50-mL HDPE centrifuge tubes in preparation for analysis; the sample weights were all 1.00±0.02 g. Residues of 2,4-D and 2,4-DCP were extracted from the sample matrices by homogenizing and shaking with a methanol /1.0 N sodium hydroxide (90:10 v/v) solution. The sample was centrifuged and an aliquot of the extraction solution was taken and combined with mixed stable isotope internal standard. The sample was mixed and evaporated to remove all organic content from the solution. Next, hydrochloric acid was added to the sample and was heated at 90 °C for 60 minutes. After cooling, the sample was diluted with methanol, sonicated, filtered (if required), and analyzed by online solid phase extraction (SPE) and liquid chromatography with negative ion APCI tandem mass spectrometry (LC-MS/MS). Field samples were reanalyzed according to the procedure below if concentrations were out of the calibration range.

Typical Online SPE Conditions (2,4-D and 2,4-DCP)

SPE Cartridge:	Hysphere C18HD, 2x10 mm
SPE Solvation:	Acetonitrile, 1 mL at 5 mL/min (SSM A[a])
SPE Equilibration:	0.1% formic acid in water, 1 mL at 5 mL/min (SSM D)
Sample Extraction:	70:30 water:methanol containing 0.1% formic acid, 750 µL at 2 mL/min (SSM E)
SPE Wash 1:	70:30 water:methanol containing 0.1% formic acid, 750 µL at 2 mL/min (SSM E)
SPE Elution:	Gradient elution mode using for 3.0 minutes
Clamp Flush 1:	Methanol containing 0.5% NH_4OH, 1 mL at 5 mL/min (HPD2, port 4)
Clamp Flush 2:	Water, 1 mL at 5 mL/min (HPD2, port 2)

[a] Denotes solvent line position on the Spark Holland Symbiosis system

Typical HPLC Operating Conditions (2,4-D and 2,4-DCP)

Instrumentation:	Symbiosis Pharma
Column:	Synergi Hydro-RP (75x4.6 mm id)
Column Temperature:	Ambient
Injection Volume:	30 µL
Injection Wash Program	1) 2 x 700 µL methanol containing 0.5% NH_4OH 2) 3 x 700 µL water
Run Time:	6.0 minutes
Mobile Phase:	A –Acetonitrile:methanol (80:20) with 0.1% acetic acid B –Water with 0.1% acetic acid
Flow Rate:	1.0 mL/min. Flow diverted for first 1.5 minutes

Isocratic:

Time, min	A, %	B, %
0:00	55	45
6:00	55	45

Flow Diverter Program: 1) 0.0→1.5 min: flow to waste
2) 1.5→4 min: flow to source
3) 4→end of run: flow to waste

Typical Mass Spectrometry Operating Conditions (2,4-D and 2,4-DCP)

Instrumentation:	MDS SCIEX API 5000 LC/MS/MS System MDS SCIEX Analyst 1.5.1 data system
Interface:	APCI
Polarity:	Negative
Scan Type:	MRM
Resolution:	Q1 – unit, Q3 – unit
Curtain Gas (CUR):	30
Collision Gas (CAD):	4.0 (High)
Temperature (TEM):	450 °C
Ion Source Gas 1 (GS1):	40
Ion Source Gas 2 (GS2):	N/A

Period 1

Time:	2.5 minutes
Acquisition Delay	1.5 minutes
APCI current	-4
Smart Settling:	Off
Settling Time:	0 ms
MR Pause:	5 ms

Compound:

	Ion			
	Q1	Q3	Time (ms)	DP/CE
2,4-D quant	219.0	161.0	50	-40/-22
2,4-D conf	221.0	163.0	50	-40/-22
2,4-DCP quant	161.0	125.0	50	-40/-24
2,4-DCP conf	163.0	127.0	50	-40/-24
2,4-D M+6 IS	225.0	167.0	50	-40/-22
2,4-DCP M+6 IS	167.0	131.0	50	-40/-24

Sample Analysis (2,4-D and 2,4-DCP)

1. Weigh 5.0 ± 0.05 g of the sample into an 8-oz HDPE bottle (1.0 ± 0.02 g in a 50-mL HDPE centrifuge tube for oil). For recovery samples, add appropriate aliquots of the spiking solutions to obtain concentrations ranging from 0.01 to 0.10 µg/g for grains and oil commodities and 0.01 to 10.0 µg/g for rest of plant samples.

2. Add 100 mL (20 mL for oil) of the methanol /1.0 N sodium hydroxide (90:10 v/v) extraction solution to the sample bottle.

3. Blend the sample for approximately 60 seconds at 10000 rpm using an Omni-Mixer homogenizer fitted with a 20-mm probe (10 mm for oil).

4. Shake the sample for a minimum of 60 minutes on a reciprocating shaker at approximately 180 excursions/minute.

5. Centrifuge the sample bottle for approximately 5 minutes at 2000 rpm.

6. Pipette a 1-mL aliquot (less if dilution is required, see Step 3.14) into a 11-dram vial and add 100 µL of the 0.1 µg/mL mixed stable isotope internal standard. Mix the sample by vortex, roll the vials by hand, and vortex again.

7. Evaporate the organic solvent to leave approximately 100 µL with a TurboVap set to 30ºC and 7 psi of nitrogen.

8. Add 500-µL of 2 N Hydrochloric acid to the sample, firmly cap, mix, and place in an oven set at 90ºC for 60 minutes.

9. Remove the sample from the oven and allow to cool. Allow approximately 20 minutes cooling at ambient conditions and an additional 20 minutes in a refrigerator.

10. Add 500-µL of methanol to the vial. Mix the sample by vortex, roll the vials by hand, and vortex again. Sonicate the sample for 30 seconds.

11. Filter the samples if necessary through a 0.45 µm PTFE filter device. Analyze the samples, calibration standards by On-Line SPE LC/MS with negative ion APCI tandem mass spectrometry.

12. Due to the wide concentration range of possible 2,4-D and 2,4-DCP metabolite residues in the corn and soybean samples, the following dilution scheme is suggested to result in samples concentrations that fall within the range of calibration standards. If dilutions are required, adjust the aliquot size taken in Step 6 according to the table below:

Expected Residue	Aliquot of Extracted Sample	Final Sample Concentration Range	Effective Dilution Factor
µg/g	µL	ng/mL	
0.01 – 1.0	1000	0.5 - 50	1
1.0 – 10	100	5.0 - 50	10
10.0 - 100	10	5.0 - 50	100[a]

[a] Step 3.7 may be skipped if there is only a 10 µL aliquot taken. The final volume is already approximately 100 µL

Analytical Method for Quizalofop-Ethyl and Quizalofop Acid Determination in Corn

The analytical method followed for determination of quizalofop-ethyl and quizalofop acid in corn commodities and processed products is documented in the study file. Chromatographic data was collected and integrated using MDS/Sciex Analyst software. Typical chromatograms for these analyses can be found in Figures 80-159.

Samples (with the exception of oil) were weighed into 8-oz HDPE containers in preparation for analysis; the sample weights were all 5.00±0.05 g. Oil samples were weighed into 50-mL HDPE centrifuge tubes in preparation for analysis; the sample weights were all 1.00±0.02 g. Residues of quizalofop-ethyl and quizalofop acid were extracted from the sample matrices by homogenizing and shaking with a methanol/water (90:10) solution. The sample was centrifuged and an aliquot of the extraction solution was taken and combined with a water/formic acid (99.9:0.1) solution. The sample was mixed, filtered (if required), and analyzed by online solid phase extraction (SPE) and liquid chromatography with APCI tandem mass spectrometry (LC-MS/MS).

Typical Online SPE Conditions (Quizalofop-Ethyl and Quizalofop Acid)

SPE Cartridge:	Hysphere C18HD, 2x10 mm
SPE Solvation:	Acetonitrile, 1 mL at 5 mL/min (SSM A[a])
SPE Equilibration:	0.1% formic acid in water, 1 mL at 5 mL/min (SSM D)
Sample Extraction:	Water, 1 mL at 2 mL/min (HPD2, port 2)
SPE Wash 1:	Water, 1 mL at 2 mL/min (HPD2, port 2)
SPE Elution:	Gradient elution mode using HPLC mobile phase for 2 minutes
Clamp Flush 1:	Methanol containing 0.5% NH_4OH, 1 mL at 5 mL/min (HPD2, port 4)
Clamp Flush 2:	Water, 1 mL at 5 mL/min (HPD2, port 2)

[a] Denotes solvent line position on the Spark Holland Symbiosis system

Typical HPLC Operating Conditions (Quizalofop-Ethyl and Quizalofop Acid)

Instrumentation:	Symbiosis Pharma
Column:	Synergi hydro (75x4.6 mm id)
Column Temperature:	Ambient
Injection Volume:	10 µL
Injection Wash Program	1) 2 x 700 µL methanol containing 0.5% NH_4OH 2) 3 x 700 µL water
Run Time:	12.0 minutes
Mobile Phase:	A –Acetonitrile:methanol (80:20) with 0.1% acetic acid B –Water with 0.1% acetic acid
Flow Rate:	1.0 mL/min. Flow diverted for first 3 minutes

Isocratic:

Time, min	A, %	B, %
0:00	40	60
5:00	100	0
7:00	100	0
8:00	40	60
12:00	40	60

Flow Diverter Program:
1) 0→3 min: flow to waste
2) 3→6 min: flow to source
3) 6→end of run: flow to waste

Typical Mass Spectrometry Operating Conditions (Quizalofop-Ethyl and Quizalofop Acid)

Instrumentation:	MDS SCIEX API 5000 LC/MS/MS System MDS SCIEX Analyst 1.4.2 data system
Interface:	APCI
Polarity:	Period 1: Negative Period 2: Positive
Scan Type:	MRM
Resolution:	Q1 – unit, Q3 – unit
Curtain Gas (CUR):	30
Collision Gas (CAD):	6.0
Temperature (TEM):	450 °C
Ion Source Gas 1 (GS1):	40
Ion Source Gas 2 (GS2):	N/A

Period 1

Time:	1.5 minutes
Acquisition Delay	3.0 minutes
APCI current	-1
Smart Settling:	Off
Settling Time:	0 ms
MR Pause:	5 ms
Compound:	

	Ion Q1	Q3	Time (ms)	DP/CE
Quizalofop acid quant	343.1	270.9	150	-40/-22
Quizalofop acid conf	343.1	108.1	150	-40/-60

Period 2

Time:	3.5 minutes
Acquisition Delay	0 minutes
APCI current	+1
Smart Settling:	Off
Settling Time:	700 ms
MR Pause:	5 ms
Compound:	

	Ion Q1	Q3	Time (ms)	DP/CE
Quizalofop ethyl quant	373.2	299.0	150	91/27
Quizalofop ethyl conf	373.2	91.0	150	91/41

Sample Analysis (Quizalofop-Ethyl and Quizalofop Acid)

1. Weigh 5.0 ± 0.05 g of the sample into an 8-oz HDPE bottle (1.0 ± 0.02 g in a 50-mL HDPE centrifuge tube for oil). For recovery samples, add appropriate aliquots of the spiking solutions to obtain concentrations ranging from 0.01 to 1.0 µg/g.

2. Add 100 mL (20 mL for oil) of methanol/water (90:10) extraction solution.

3. Homogenize the samples with a 20 mm homogenizer probe (10 mm for oil) for 1 minute. Cap the sample and shake for 60 minutes on a flat bed shaker at approximately 180 excursions per minute.

4. Centrifuge the sample for approximately 3 minutes at 2000 rpm.

5. Pipette a 0.5-mL aliquot into an HPLC vial and add 0.5 mL of a water/formic acid (99.9:0.1) solution and mix (filter through a 0.45-µm PTFE filter if necessary).

6. Analyze the samples, calibration standards by online SPE and liquid chromatography with APCI tandem mass spectrometry (LC-MS/MS).

Limits of Detection and Quantitation

The limit of detection and limit of quantitation were established at the initiation of the analytical phase of the study at 0.003 µg/g and 0.01 µg/g, respectively.

Confirmation of Residue Identity

The methods are specific for the determination of 2,4-dichlorophenoxyacetic acid (2,4-D), 2,4-dichlorophenol (2,4-DCP), quizalofop-ethyl, and quizalofop acid by virtue of the chromatographic separation and selective detection system used. The criteria used for qualitative confirmation of the presence of the analyte are similar to published guidelines (5). Confirmation is by comparison of the retention time (liquid chromatography) as well as the peak area ratios resulting from multiple reaction monitoring (MRM) of two separate ion transitions following LC-MS/MS analysis. The confirmation peak area ratios for recovery and field samples with reported residue values above LOQ were all within the range of ±20% of the average found for the standards within each set. Example chromatograms in Figures 5-159 include both quantitation and confirmation transitions.

Frozen Storage Stability in Field Samples

The longest frozen storage interval between the times the corn samples were collected in the field and sample analysis at Dow AgroSciences was 376 days.

CALCULATIONS

Determination of 2,4-D and 2,4-DCP Concentrations

Inject the series of calibration standards from 0.15 to 50.0 ng/mL (extending at least 20% above the concentration of all analyzed field samples – following any dilution) and determine the peak areas for the analytes and internal standards. For each sample and standard calculate the analyte quantitation ratios (analyte peak area/internal standard peak area).

Prepare a standard curve using linear regression analysis with 1/x weighting by plotting the analyte concentration on the abscissa (x-axis) and the respective quantitation ratio on the ordinate (y-axis) as shown in Figures 160-161.

Determine the concentration (µg/g) and/or recovery (%) from the sample as described in the example calculation outlined in Figure 164.

All calculations used a nominal sample mass of 5.0; oil samples used a nominal mass of 1.0 g. The actual weights of samples were 5.0±0.05 g, oil samples were 1.0±0.02 g and were individually recorded in the raw data.

Determination of Quizalofop-Ethyl and Quizalofop Acid Concentrations

Inject the series of calibration standards from 0.10 to 50.0 ng/mL (extending at least 20% above the concentration of all analyzed field samples – following any dilution) and determine the peak areas for the analytes.

Prepare a standard curve using linear regression analysis with 1/x weighting by plotting the analyte concentration on the abscissa (x-axis) and the respective peak area on the ordinate (y-axis) as shown in Figures 162-163.

Determine the concentration (µg/g) and/or recovery (%) from the sample as described in the example calculation outlined in Figure 165.

All calculations used a nominal sample mass of 5.0 g; oil samples used a nominal mass of 1.0 g. The actual weights of samples were 5.0±0.05 g, oil samples were 1.0±0.02 g and were individually recorded in the raw data.

Confirmation of Residue Identity

To demonstrate confirmation, two MS/MS ion transitions were monitored for each analyte. The following example is given for the analyte quizalofop acid

Inject the series of calibration standards described in the calibration standard preparation section using the conditions listed in the instrument section and determine the peak areas for the analyte as indicated below.

Quizalofop Acid	Q1/Q3 *m/z* 343/271 (quantitation)
	Q1/Q3 *m/z* 343/108 (confirmation)

For each standard, calculate the respective confirmation ratios.

$$\text{Confirmation Ratio} = \frac{\text{peak area of confirmation ion transition}}{\text{peak area of quantitation ion transition}}$$

$$\text{Confirmation Ratio} = \frac{\text{Quizalofop acid peak area at } m/z \text{ 343/108}}{\text{Quizalofop acid peak area at } m/z \text{ 343/271}}$$

For example, using the data for quizalofop acid from the 0.250-ng/mL standard, from set ARA-09-15-10 Q02, found in Figure 81:

$$\text{Confirmation Ratio} = \frac{323}{5520} = 0.0585$$

Confirmation of the presence of the analyte is indicated when the retention time of the samples matches that of the standards and the confirmation ratio is in the range of ±20% of the average found for the standards.

STATISTICS

The statistics used for this study were linear with 1/x weighting regression analysis, means, standard deviations, relative standard deviations, and coefficients of determination.

ANALYTICAL DATA

A procedural change involving a different extraction solution was required to ensure extraction efficiency. This change necessitated reanalysis of all samples for 2,4-D and 2,4-DCP. Initial analysis of samples for quizalofop acid and quizalofop ethyl were successful. Several individual and averaged concurrent recovery values, while outside a commonly accepted range of 70-120%, were deemed acceptable due to the consistency across all recovery samples of a given matrix.

Analysis results for all samples are summarized in the raw data in the form of spreadsheets. Each spreadsheet provides data from the analysis of a given set of samples, including the associated control, fortified recoveries, and the calibration standards. The residue concentration was reported as not detected (ND) if the calculated concentration was below the LOD. Residue concentrations reported between the LOD and LOQ have a higher degree of uncertainty than values above the LOQ (6).

The results of concurrent recovery samples demonstrating method performance are listed individually in Table 3 and summarized in Table 4. A summary of analytical set parameters are provided in Table 5. Analytical results for all analyzed field-treated samples are provided in Table 6. Residue results and associated field sample information is summarized in Table 7.

CONCLUSIONS

Residue values for field samples ranged from ND (not detected) to 8.92 µg/g for 2,4-D, ND (not detected) to 6.12 µg/g for 2,4-DCP, ND (not detected) to 0.20 µg/g for quizalofop-ethyl, and ND (not detected) to 0.17 µg/g for quizalofop acid. Method performance was demonstrated, with concurrent recoveries ranging from 77-109% for 2,4-D, 61-112% for 2,4-DCP, 72-116% for quizalofop-ethyl, and 75-143% for quizalofop acid.

ARCHIVING

The analytical summary and all of the associated raw data are all filed in the Dow AgroSciences LLC archives at 9330 Zionsville Road in Indianapolis, Indiana 46268-1054.

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Table 1. Summary of Sampling, Shipping, and Analysis Dates for Corn Samples

SGN[a]	Trial	TAT[b]	Matrix	Qualifier 1	Qualifier 2	Sampling Date[c]	Shipping Date[c]	Arrival Date[c]	Last Analysis[d]	Days of Storage[e]
001	NJ-01	40	Corn	Forage		17-Sep-2009	15-Oct-2009	26-Oct-2009	24-Aug-2010	341
002	NJ-01	60	Corn	Forage		14-Oct-2009	2-Dec-2009	21-Dec-2009	24-Aug-2010	314
003	NJ-01	90	Corn	Grain		23-Nov-2009	2-Dec-2009	21-Dec-2009	11-Aug-2010	261
004	NJ-01	90	Corn	Stover		23-Nov-2009	2-Dec-2009	21-Dec-2009	30-Aug-2010	280
005	PA-02	40	Corn	Forage		31-Aug-2009	15-Oct-2009	26-Oct-2009	24-Aug-2010	358
006	PA-02	60	Corn	Forage		14-Sep-2009	15-Oct-2009	26-Oct-2009	24-Aug-2010	344
007	PA-02	90	Corn	Grain		4-Nov-2009	2-Dec-2009	21-Dec-2009	11-Aug-2010	280
008	PA-02	90	Corn	Stover		4-Nov-2009	2-Dec-2009	21-Dec-2009	30-Aug-2010	299
009	GA-03	40	Corn	Forage		27-Aug-2009	11-Oct-2009	26-Oct-2009	24-Aug-2010	362
010	GA-03	60	Corn	Forage		1-Sep-2009	11-Oct-2009	26-Oct-2009	24-Aug-2010	357
011	GA-03	90	Corn	Grain		2-Oct-2009	11-Oct-2009	26-Oct-2009	12-Aug-2010	314
013	GA-03	90	Corn	Stover		2-Oct-2009	11-Oct-2009	26-Oct-2009	27-Aug-2010	329
014	GA-04	40	Corn	Forage		30-Aug-2009	11-Oct-2009	26-Oct-2009	24-Aug-2010	359
015	GA-04	46	Corn	Forage		1-Sep-2009	11-Oct-2009	26-Oct-2009	24-Aug-2010	357
016	GA-04	53	Corn	Forage		8-Sep-2009	11-Oct-2009	26-Oct-2009	24-Aug-2010	350
017	GA-04	60	Corn	Forage		15-Sep-2009	11-Oct-2009	26-Oct-2009	20-Aug-2010	339
018	GA-04	67	Corn	Forage		22-Sep-2009	11-Oct-2009	26-Oct-2009	20-Aug-2010	332
019	GA-04	74	Corn	Forage		29-Sep-2009	11-Oct-2009	26-Oct-2009	20-Aug-2010	325
020	GA-04	76	Corn	Grain		1-Oct-2009	11-Oct-2009	26-Oct-2009	11-Aug-2010	314
021	GA-04	76	Corn	Stover		1-Oct-2009	11-Oct-2009	26-Oct-2009	30-Aug-2010	333
022	GA-04	83	Corn	Grain		8-Oct-2009	30-Nov-2009	21-Dec-2009	11-Aug-2010	307
023	GA-04	83	Corn	Stover		8-Oct-2009	30-Nov-2009	21-Dec-2009	30-Aug-2010	326
024	GA-04	90	Corn	Grain		15-Oct-2009	30-Nov-2009	21-Dec-2009	11-Aug-2010	300
025	GA-04	90	Corn	Stover		15-Oct-2009	30-Nov-2009	21-Dec-2009	30-Aug-2010	319
026	GA-04	97	Corn	Grain		22-Oct-2009	30-Nov-2009	21-Dec-2009	11-Aug-2010	293
027	GA-04	97	Corn	Stover		22-Oct-2009	30-Nov-2009	21-Dec-2009	30-Aug-2010	312
028	GA-04	104	Corn	Grain		29-Oct-2009	30-Nov-2009	21-Dec-2009	11-Aug-2010	286
029	GA-04	104	Corn	Stover		29-Oct-2009	30-Nov-2009	21-Dec-2009	30-Aug-2010	305
030	IL-05	40	Corn	Forage		18-Aug-2009	1-Sep-2009	3-Sep-2009	24-Aug-2010	371
031	IL-05	60	Corn	Forage		14-Sep-2009	17-Oct-2009	26-Oct-2009	24-Aug-2010	344
032	IL-05	90	Corn	Grain		19-Nov-2009	4-Dec-2009	21-Dec-2009	12-Aug-2010	266
033	IL-05	90	Corn	Stover		19-Nov-2009	4-Dec-2009	21-Dec-2009	27-Aug-2010	281
034	IL-06	40	Corn	Forage		4-Sep-2009	23-Oct-2009	26-Oct-2009	24-Aug-2010	354
035	IL-06	60	Corn	Forage		29-Sep-2009	23-Oct-2009	26-Oct-2009	23-Aug-2010	328
036	IL-06	90	Corn	Grain		23-Nov-2009	12-Dec-2009	21-Dec-2009	11-Aug-2010	261
037	IL-06	90	Corn	Stover		23-Nov-2009	12-Dec-2009	21-Dec-2009	27-Aug-2010	277
038	IN-07	40	Corn	Forage		18-Sep-2009	18-Sep-2009	18-Sep-2009	24-Aug-2010	340
039	IN-07	60	Corn	Forage		14-Aug-2009	18-Sep-2009	18-Sep-2009	24-Aug-2010	375
040	IN-07	90	Corn	Grain		1-Dec-2009	2-Dec-2009	2-Dec-2009	11-Aug-2010	253
041	IN-07	90	Corn	Stover		1-Dec-2009	2-Dec-2009	2-Dec-2009	30-Aug-2010	272
042	MO-08	40	Corn	Forage		18-Sep-2009	18-Dec-2009	21-Dec-2009	20-Aug-2010	336
043	MO-08	60	Corn	Forage		25-Sep-2009	18-Dec-2009	21-Dec-2009	23-Aug-2010	332

Table 1 (Cont.). Summary of Sampling, Shipping, and Analysis Dates for Corn Samples

SGN[a]	Trial	TAT[b]	Matrix	Qualifier 1	Qualifier 2	Sampling Date[c]	Shipping Date[c]	Arrival Date[c]	Last Analysis[d]	Days of Storage[e]
044	MO-08	90	Corn	Grain		21-Nov-2009	18-Dec-2009	21-Dec-2009	11-Aug-2010	263
045	MO-08	90	Corn	Stover		21-Nov-2009	18-Dec-2009	21-Dec-2009	30-Aug-2010	282
046	MO-09	40	Corn	Forage		18-Sep-2009	18-Dec-2009	21-Dec-2009	24-Aug-2010	340
047	MO-09	60	Corn	Forage		25-Sep-2009	18-Dec-2009	21-Dec-2009	24-Aug-2010	333
048	MO-09	90	Corn	Grain		21-Nov-2009	18-Dec-2009	21-Dec-2009	11-Aug-2010	263
049	MO-09	90	Corn	Stover		21-Nov-2009	18-Dec-2009	21-Dec-2009	30-Aug-2010	282
050	IL-10	40	Corn	Forage		22-Sep-2009	18-Dec-2009	21-Dec-2009	23-Aug-2010	335
051	IL-10	60	Corn	Forage		25-Sep-2009	18-Dec-2009	21-Dec-2009	20-Aug-2010	329
052	IL-10	90	Corn	Grain		22-Nov-2009	18-Dec-2009	21-Dec-2009	11-Aug-2010	262
053	IL-10	90	Corn	Stover		22-Nov-2009	18-Dec-2009	21-Dec-2009	30-Aug-2010	281
054	MI-11	40	Corn	Forage		14-Sep-2009	16-Oct-2009	26-Oct-2009	24-Aug-2010	344
055	MI-11	46	Corn	Forage		20-Sep-2009	16-Oct-2009	26-Oct-2009	24-Aug-2010	338
056	MI-11	53	Corn	Forage		27-Sep-2009	16-Oct-2009	26-Oct-2009	24-Aug-2010	331
057	MI-11	60	Corn	Forage		3-Oct-2009	16-Oct-2009	26-Oct-2009	24-Aug-2010	325
058	MI-11	67	Corn	Forage		10-Oct-2009	16-Oct-2009	26-Oct-2009	24-Aug-2010	318
059	MI-11	74	Corn	Forage		17-Oct-2009	3-Dec-2009	21-Dec-2009	24-Aug-2010	311
060	MI-11	76	Corn	Grain		20-Oct-2009	3-Dec-2009	21-Dec-2009	11-Aug-2010	295
061	MI-11	76	Corn	Stover		20-Oct-2009	3-Dec-2009	21-Dec-2009	30-Aug-2010	314
062	MI-11	83	Corn	Grain		27-Oct-2009	3-Dec-2009	21-Dec-2009	11-Aug-2010	288
063	MI-11	83	Corn	Stover		27-Oct-2009	3-Dec-2009	21-Dec-2009	30-Aug-2010	307
064	MI-11	90	Corn	Grain		3-Nov-2009	3-Dec-2009	21-Dec-2009	11-Aug-2010	281
065	MI-11	90	Corn	Stover		3-Nov-2009	3-Dec-2009	21-Dec-2009	30-Aug-2010	300
066	MI-11	97	Corn	Grain		10-Nov-2009	3-Dec-2009	21-Dec-2009	11-Aug-2010	274
067	MI-11	97	Corn	Stover		10-Nov-2009	3-Dec-2009	21-Dec-2009	30-Aug-2010	293
068	MI-11	104	Corn	Grain		17-Nov-2009	3-Dec-2009	21-Dec-2009	11-Aug-2010	267
069	MI-11	104	Corn	Stover		17-Nov-2009	3-Dec-2009	21-Dec-2009	30-Aug-2010	286
070	OH-12	40	Corn	Forage		17-Sep-2009	16-Oct-2009	26-Oct-2009	24-Aug-2010	341
071	OH-12	60	Corn	Forage		2-Oct-2009	16-Oct-2009	26-Oct-2009	24-Aug-2010	326
072	OH-12	90	Corn	Grain		6-Nov-2009	3-Dec-2009	21-Dec-2009	11-Aug-2010	278
073	OH-12	90	Corn	Stover		6-Nov-2009	3-Dec-2009	21-Dec-2009	30-Aug-2010	297
074	IN-13	40	Corn	Forage		14-Sep-2009	16-Oct-2009	26-Oct-2009	24-Aug-2010	344
075	IN-13	60	Corn	Forage		4-Oct-2009	16-Oct-2009	26-Oct-2009	24-Aug-2010	324
076	IN-13	90	Corn	Grain		3-Nov-2009	3-Dec-2009	21-Dec-2009	11-Aug-2010	281
077	IN-13	90	Corn	Stover		3-Nov-2009	3-Dec-2009	21-Dec-2009	30-Aug-2010	300
079	NE-14	60	Corn	Forage		1-Sep-2009	21-Oct-2009	26-Oct-2009	23-Aug-2010	356
080	NE-14	90	Corn	Grain		17-Nov-2009	8-Dec-2009	21-Dec-2009	12-Aug-2010	268
082	NE-14	90	Corn	Stover		17-Nov-2009	8-Dec-2009	21-Dec-2009	27-Aug-2010	283
083	NE-15	40	Corn	Forage		31-Aug-2009	21-Oct-2009	26-Oct-2009	23-Aug-2010	357
084	NE-15	60	Corn	Forage		24-Sep-2009	21-Oct-2009	26-Oct-2009	23-Aug-2010	333
085	NE-15	90	Corn	Grain		17-Oct-2009	21-Oct-2009	26-Oct-2009	11-Aug-2010	298

Table 1 (Cont.). Summary of Sampling, Shipping, and Analysis Dates for Corn Samples

SGN[a]	Trial	TAT[b]	Matrix	Qualifier 1	Qualifier 2	Sampling Date[c]	Shipping Date[c]	Arrival Date[c]	Last Analysis[d]	Days of Storage[e]
086	NE-15	90	Corn	Stover		17-Oct-2009	21-Oct-2009	26-Oct-2009	27-Aug-2010	314
087	KS-16	40	Corn	Forage		3-Sep-2009	21-Oct-2009	26-Oct-2009	23-Aug-2010	354
088	KS-16	60	Corn	Forage		29-Sep-2009	21-Oct-2009	26-Oct-2009	23-Aug-2010	328
089	KS-16	90	Corn	Grain		28-Oct-2009	11-Dec-2009	21-Dec-2009	12-Aug-2010	288
090	KS-16	90	Corn	Stover		28-Oct-2009	11-Dec-2009	21-Dec-2009	27-Aug-2010	303
091	IA-17	40	Corn	Forage		13-Aug-2009	28-Aug-2009	3-Sep-2009	24-Aug-2010	376
092	IA-17	60	Corn	Forage		4-Sep-2009	19-Oct-2009	26-Oct-2009	24-Aug-2010	354
093	IA-17	90	Corn	Grain		17-Oct-2009	19-Oct-2009	26-Oct-2009	12-Aug-2010	299
094	IA-17	90	Corn	Stover		17-Oct-2009	19-Oct-2009	26-Oct-2009	27-Aug-2010	314
095	MO-18	40	Corn	Forage		21-Aug-2009	28-Aug-2009	3-Sep-2009	23-Aug-2010	367
096	MO-18	60	Corn	Forage		3-Sep-2009	19-Oct-2009	26-Oct-2009	23-Aug-2010	354
097	MO-18	90	Corn	Grain		20-Oct-2009	4-Dec-2009	21-Dec-2009	12-Aug-2010	296
098	MO-18	90	Corn	Stover		20-Oct-2009	4-Dec-2009	21-Dec-2009	27-Aug-2010	311
099	IA-19	40	Corn	Forage		2-Sep-2009	20-Oct-2009	26-Oct-2009	24-Aug-2010	356
100	IA-19	60	Corn	Forage		29-Sep-2009	20-Oct-2009	26-Oct-2009	24-Aug-2010	329
101	IA-19	90	Corn	Grain		21-Nov-2009	7-Dec-2009	21-Dec-2009	12-Aug-2010	264
102	IA-19	90	Corn	Stover		21-Nov-2009	7-Dec-2009	21-Dec-2009	30-Aug-2010	282
103	MN-20	40	Corn	Forage		1-Sep-2009	20-Oct-2009	26-Oct-2009	24-Aug-2010	357
104	MN-20	60	Corn	Forage		29-Sep-2009	20-Oct-2009	26-Oct-2009	24-Aug-2010	329
105	MN-20	90	Corn	Grain		21-Nov-2009	7-Dec-2009	21-Dec-2009	12-Aug-2010	264
106	MN-20	90	Corn	Stover		21-Nov-2009	7-Dec-2009	21-Dec-2009	30-Aug-2010	282
107	MO-21	40	Corn	Forage		15-Sep-2009	21-Sep-2009	26-Oct-2009	24-Aug-2010	343
108	MO-21	60	Corn	Forage		30-Sep-2009	22-Oct-2009	26-Oct-2009	24-Aug-2010	328
109	MO-21	90	Corn	Grain		2-Nov-2009	12-Dec-2009	21-Dec-2009	12-Aug-2010	283
110	MO-21	90	Corn	Stover		2-Nov-2009	12-Dec-2009	21-Dec-2009	27-Aug-2010	298
111	OK-22	40	Corn	Forage		22-Aug-2009	17-Dec-2009	21-Dec-2009	23-Aug-2010	366
112	OK-22	46	Corn	Forage		29-Aug-2009	17-Dec-2009	21-Dec-2009	24-Aug-2010	360
113	OK-22	53	Corn	Forage		4-Sep-2009	17-Dec-2009	21-Dec-2009	24-Aug-2010	354
114	OK-22	60	Corn	Forage		10-Sep-2009	17-Dec-2009	21-Dec-2009	24-Aug-2010	348
115	OK-22	67	Corn	Forage		16-Sep-2009	17-Dec-2009	21-Dec-2009	24-Aug-2010	342
116	OK-22	74	Corn	Forage		22-Sep-2009	17-Dec-2009	21-Dec-2009	23-Aug-2010	335
117	OK-22	76	Corn	Grain		26-Sep-2009	17-Dec-2009	21-Dec-2009	12-Aug-2010	320
118	OK-22	76	Corn	Stover		26-Sep-2009	17-Dec-2009	21-Dec-2009	27-Aug-2010	335
119	OK-22	83	Corn	Grain		2-Oct-2009	17-Dec-2009	21-Dec-2009	12-Aug-2010	314
120	OK-22	83	Corn	Stover		2-Oct-2009	17-Dec-2009	21-Dec-2009	27-Aug-2010	329
121	OK-22	90	Corn	Grain		10-Oct-2009	17-Dec-2009	21-Dec-2009	12-Aug-2010	306
122	OK-22	90	Corn	Stover		10-Oct-2009	17-Dec-2009	21-Dec-2009	27-Aug-2010	321
123	OK-22	97	Corn	Grain		17-Oct-2009	17-Dec-2009	21-Dec-2009	12-Aug-2010	299
124	OK-22	97	Corn	Stover		17-Oct-2009	17-Dec-2009	21-Dec-2009	27-Aug-2010	314
125	OK-22	104	Corn	Grain		24-Oct-2009	17-Dec-2009	21-Dec-2009	12-Aug-2010	292

Table 1 (Cont.). Summary of Sampling, Shipping, and Analysis Dates for Corn Samples

SGN[a]	Trial	TAT[b]	Matrix	Qualifier 1	Qualifier 2	Sampling Date[c]	Shipping Date[c]	Arrival Date[c]	Last Analysis[d]	Days of Storage[e]
126	OK-22	104	Corn	Stover		24-Oct-2009	17-Dec-2009	21-Dec-2009	27-Aug-2010	307
127	TX-23	40	Corn	Forage		20-Aug-2009	17-Dec-2009	21-Dec-2009	23-Aug-2010	368
128	TX-23	60	Corn	Forage		1-Sep-2009	17-Dec-2009	21-Dec-2009	24-Aug-2010	357
129	TX-23	90	Corn	Grain		8-Nov-2009	17-Dec-2009	21-Dec-2009	12-Aug-2010	277
130	TX-23	90	Corn	Stover		8-Nov-2009	17-Dec-2009	21-Dec-2009	30-Aug-2010	295
131	ON-24	40	Corn	Forage		24-Aug-2009	25-Sep-2009	29-Sep-2009	24-Aug-2010	365
132	ON-24	60	Corn	Forage		30-Sep-2009	20-Nov-2009	3-Dec-2009	23-Aug-2010	327
133	ON-24	90	Corn	Grain		17-Nov-2009	20-Nov-2009	3-Dec-2009	12-Aug-2010	268
134	ON-24	90	Corn	Stover		17-Nov-2009	20-Nov-2009	3-Dec-2009	27-Aug-2010	283
135	ON-25	40	Corn	Forage		26-Aug-2009	25-Sep-2009	29-Sep-2009	23-Aug-2010	362
136	ON-25	60	Corn	Forage		24-Sept-2009	25-Sep-2009	29-Sep-2009	23-Aug-2010	333
137	ON-25	90	Corn	Grain		18-Nov-2009	20-Nov-2009	3-Dec-2009	12-Aug-2010	267
138	ON-25	90	Corn	Stover		18-Nov-2009	20-Nov-2009	3-Dec-2009	27-Aug-2010	282
139	GA-03	90	Corn	Grain		25-Jan-2010	16-Feb-2010	17-Feb-2010	12-Aug-2010	199
140	GA-03	90	Corn	Grain		26-Jan-2010	16-Feb-2010	16-Feb-2010	12-Aug-2010	198
141	GA-03	90	Corn	Grain	Aspirated Grain Fractions	15-Feb-2010	16-Feb-2010	17-Feb-2010	16-Aug-2010	182
142	GA-03	90	Corn	Grain	Starch	3-Feb-2010	16-Feb-2010	17-Feb-2010	16-Aug-2010	194
143	GA-03	90	Corn	Grain	Oil, refined	8-Feb-2010	16-Feb-2010	17-Feb-2010	13-Aug-2010	186
144	GA-03	90	Corn	Grain	Meal	29-Jan-2010	16-Feb-2010	17-Feb-2010	16-Aug-2010	199
145	GA-03	90	Corn	Grain	Grits	29-Jan-2010	16-Feb-2010	17-Feb-2010	16-Aug-2010	199
146	GA-03	90	Corn	Grain	Flour	29-Jan-2010	16-Feb-2010	17-Feb-2010	16-Aug-2010	199
147	GA-03	90	Corn	Grain	Oil, refined	3-Feb-2010	16-Feb-2010	17-Feb-2010	13-Aug-2010	191
148	NE-14	90	Corn	Grain		8-Feb-2010	2-Mar-2010	3-Mar-2010	12-Aug-2010	185
149	NE-14	90	Corn	Grain		9-Feb-2010	2-Mar-2010	3-Mar-2010	12-Aug-2010	184
150	NE-14	90	Corn	Grain	Aspirated Grain Fractions	22-Feb-2010	2-Mar-2010	3-Mar-2010	16-Aug-2010	175
151	NE-14	90	Corn	Grain	Starch	17-Feb-2010	2-Mar-2010	3-Mar-2010	16-Aug-2010	180
152	NE-14	90	Corn	Grain	Oil, refined	23-Feb-2010	2-Mar-2010	3-Mar-2010	13-Aug-2010	171
153	NE-14	90	Corn	Grain	Meal	12-Feb-2010	2-Mar-2010	3-Mar-2010	16-Aug-2010	185
154	NE-14	90	Corn	Grain	Grits	12-Feb-2010	2-Mar-2010	3-Mar-2010	16-Aug-2010	185
155	NE-14	90	Corn	Grain	Flour	12-Feb-2010	2-Mar-2010	3-Mar-2010	16-Aug-2010	185
156	NE-14	90	Corn	Grain	Oil, refined	23-Feb-2010	2-Mar-2010	3-Mar-2010	13-Aug-2010	171
157	Control	1	Corn	Forage		24-May-2010	N/A	24-May-2010	26-May-2010	2
158	Control	1	Corn	Stover		24-May-2010	N/A	24-May-2010	28-May-2010	4

[a] SGN = Sample Group Number
[b] TAT = Time After Treatment
[c] First date of occurance.
[d] Date when analysis was last initiated by the addition of extraction solution.
[e] Maximum days of storage for a given sample group.

Table 2. Identity and Structures of Analytes and Related Compounds

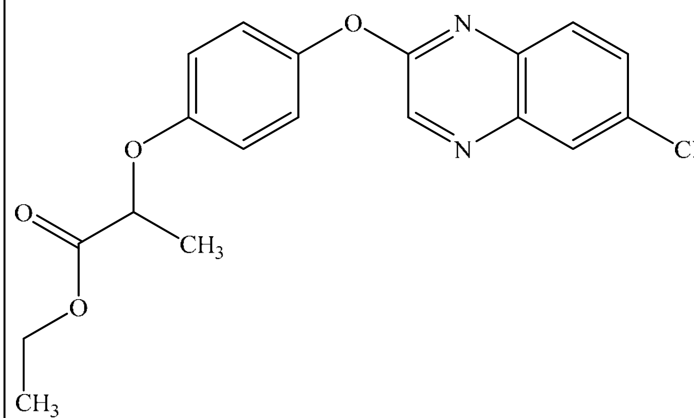
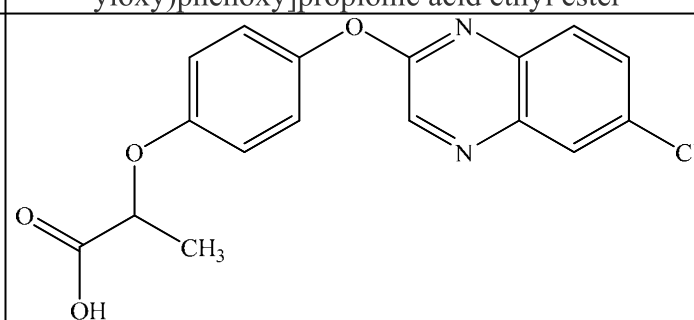
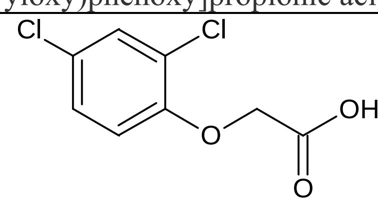
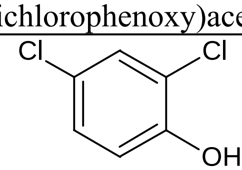
Common Name of Compound	Structural Formula and Chemical Name
Quizalofop-Ester Molecular Formula: $C_{19}H_{17}ClN_2O_4$ Formula Weight: 372.81 Nominal Mass: 372	 (R)-2-[4-(6-chloroquinoxalin-2-yloxy)phenoxy]propionic acid ethyl ester
Quizalofop Acid Molecular Formula: $C_{17}H_{13}ClN_2O_4$ Formula Weight: 344.76 Nominal Mass: 344	 (R)-2-[4-(6-chloroquinoxalin-2-yloxy)phenoxy]propionic acid
2,4-D Molecular Formula: $C_8H_6Cl_2O_3$ Formula Weight: 221.04 Nominal Mass: 220	 (2,4-dichlorophenoxy)acetic acid
2,4-DCP Molecular Formula: $C_6H_4Cl_2O$ Formula Weight: 163.00 Nominal Mass: 162	 2,4-dichlorophenol

Table 2 (Cont.). Identity and Structures of Analytes and Related Compounds

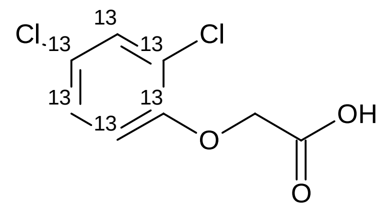
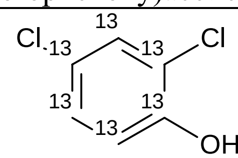
Common Name of Compound	Structural Formula and Chemical Name
2,4-D Stable Isotope Molecular Formula: $^{13}C_6C_2H_6Cl_2O_3$ Formula Weight: 226.99 Nominal Mass: 226	 (2,4-dichlorophenoxy)acetic acid ($^{13}C_6$)
2,4-DCP Stable Isotope Molecular Formula: $^{13}C_6H_4Cl_2O$ Formula Weight: 168.96 Nominal Mass: 168	 2,4-dichlorophenol ($^{13}C_6$)

Table 3. Procedural Recovery Raw Data for Corn Samples

Analyte	Matrix	Control Sample ID	Inj. No	Analyte Peak Area	ISTD Peak Area	Dilution Factor	Fortified Conc. (µg/g)	Found (µg/g)[a]	% Rec	Set ID
2,4-D	Grain	ARA-09-15-10-011-0001A2	3	0	775839	1	NA[b]	ND[c]	NA	D16
2,4-D	Grain	ARA-09-15-10-011-0001A3 + 0.003 µg/g	57	7107	351669	1	0.003	0.0025	NA	D16
2,4-D	Grain	ARA-09-15-10-011-0001A4 + 0.01 µg/g	58	19594	344637	1	0.01	0.0093	93	D16
2,4-D	Grain	ARA-09-15-10-011-0001A5 + 0.01 µg/g	59	19493	337652	1	0.01	0.0094	94	D16
2,4-D	Grain	ARA-09-15-10-011-0001A6 + 0.10 µg/g	60	170938	348600	1	0.1	0.0897	90	D16
2,4-D	Grain	ARA-09-15-10-011-0001A7 + 0.10 µg/g	61	157625	326457	1	0.1	0.0883	88	D16
2,4-D	Grain	ARA-09-15-10-011-0001A8	3	0	627634	1	NA	ND	NA	D17
2,4-D	Grain	ARA-09-15-10-011-0001A9 + 0.003 µg/g	58	6614	315633	1	0.003	0.0027	NA	D17
2,4-D	Grain	ARA-09-15-10-011-0001A10 + 0.01 µg/g	59	17899	320234	1	0.01	0.0091	91	D17
2,4-D	Grain	ARA-09-15-10-011-0001A11 + 0.01 µg/g	60	16597	311223	1	0.01	0.0086	86	D17
2,4-D	Grain	ARA-09-15-10-011-0001A12 + 0.10 µg/g	62	167644	319657	1	0.1	0.0946	95	D17
2,4-D	Grain	ARA-09-15-10-011-0001A13 + 0.10 µg/g	63	159858	303753	1	0.1	0.0949	95	D17
2,4-D	Grain	ARA-09-15-10-011-0001A14	3	0	548705	1	NA	ND	NA	D18
2,4-D	Grain	ARA-09-15-10-011-0001A15 + 0.003 µg/g	53	5361	299676	1	0.003	0.0024	NA	D18
2,4-D	Grain	ARA-09-15-10-011-0001A16 + 0.01 µg/g	54	16097	300882	1	0.01	0.0089	89	D18
2,4-D	Grain	ARA-09-15-10-011-0001A17 + 0.01 µg/g	56	15266	298873	1	0.01	0.0085	85	D18
2,4-D	Grain	ARA-09-15-10-011-0001A18 + 0.10 µg/g	57	145296	293958	1	0.1	0.0902	90	D18
2,4-D	Grain	ARA-09-15-10-011-0001A19 + 0.10 µg/g	58	144865	288493	1	0.1	0.0917	92	D18
2,4-D	Aspirated grain fractions	ARA-09-15-10-141-0001A2	3	0	279413	1	NA	ND	NA	D19
2,4-D	Aspirated grain fractions	ARA-09-15-10-141-0001A3 + 0.003 µg/g	43	5129	243926	1	0.003	0.0025	NA	D19
2,4-D	Aspirated grain fractions	ARA-09-15-10-141-0001A4 + 0.01 µg/g	48	12467	237781	1	0.01	0.0082	82	D19
2,4-D	Aspirated grain fractions	ARA-09-15-10-141-0001A5 + 0.01 µg/g	49	12557	242006	1	0.01	0.0081	81	D19
2,4-D	Aspirated grain fractions	ARA-09-15-10-141-0001A6 + 0.10 µg/g	59	101218	225996	1	0.1	0.0803	80	D19
2,4-D	Aspirated grain fractions	ARA-09-15-10-141-0001A7 + 0.10 µg/g	60	105121	226830	1	0.1	0.0831	83	D19
2,4-D	Starch	ARA-09-15-10-142-0001A30	34	0	235788	1	NA	ND	NA	D19

Table 3 (Cont.). Procedural Recovery Raw Data for Corn Samples

Analyte	Matrix	Control Sample ID	Inj. No	Analyte Peak Area	ISTD Peak Area	Dilution Factor	Fortified Conc. (µg/g)	Found (µg/g)[a]	% Rec	Set ID
2,4-D	Starch	ARA-09-15-10-142-0001A31 + 0.003 µg/g	47	4906	228238	1	0.003	0.0026	NA	D19
2,4-D	Starch	ARA-09-15-10-142-0001A32 + 0.01 µg/g	57	11275	213529	1	0.01	0.0083	83	D19
2,4-D	Starch	ARA-09-15-10-142-0001A33 + 0.01 µg/g	58	11766	214223	1	0.01	0.0087	87	D19
2,4-D	Starch	ARA-09-15-10-142-0001A34 + 0.10 µg/g	68	103187	225189	1	0.1	0.0821	82	D19
2,4-D	Starch	ARA-09-15-10-142-0001A35 + 0.10 µg/g	69	103996	214061	1	0.1	0.0872	87	D19
2,4-D	Meal	ARA-09-15-10-144-0001A22	25	0	221073	1	NA	ND	NA	D19
2,4-D	Meal	ARA-09-15-10-144-0001A23 + 0.003 µg/g	46	4957	227393	1	0.003	0.0026	NA	D19
2,4-D	Meal	ARA-09-15-10-144-0001A24 + 0.01 µg/g	55	12333	220343	1	0.01	0.0088	88	D19
2,4-D	Meal	ARA-09-15-10-144-0001A25 + 0.01 µg/g	56	13379	223536	1	0.01	0.0096	96	D19
2,4-D	Meal	ARA-09-15-10-144-0001A26 + 0.10 µg/g	66	98534	200959	1	0.1	0.0880	88	D19
2,4-D	Meal	ARA-09-15-10-144-0001A27 + 0.10 µg/g	67	107726	214517	1	0.1	0.0902	90	D19
2,4-D	Grits	ARA-09-15-10-145-0001A22	16	0	233741	1	NA	ND	NA	D19
2,4-D	Grits	ARA-09-15-10-145-0001A23 + 0.003 µg/g	45	4397	223224	1	0.003	0.0022	NA	D19
2,4-D	Grits	ARA-09-15-10-145-0001A24 + 0.01 µg/g	53	12482	230486	1	0.01	0.0085	85	D19
2,4-D	Grits	ARA-09-15-10-145-0001A25 + 0.01 µg/g	54	12603	219091	1	0.01	0.0091	91	D19
2,4-D	Grits	ARA-09-15-10-145-0001A26 + 0.10 µg/g	64	107416	215863	1	0.1	0.0893	89	D19
2,4-D	Grits	ARA-09-15-10-145-0001A27 + 0.10 µg/g	65	103988	211591	1	0.1	0.0882	88	D19
2,4-D	Flour	ARA-09-15-10-146-0001A3	7	0	231570	1	NA	ND	NA	D19
2,4-D	Flour	ARA-09-15-10-146-0001A4 + 0.003 µg/g	44	5259	241301	1	0.003	0.0026	NA	D19
2,4-D	Flour	ARA-09-15-10-146-0001A5 + 0.01 µg/g	50	12783	228026	1	0.01	0.0089	89	D19
2,4-D	Flour	ARA-09-15-10-146-0001A6 + 0.01 µg/g	52	13046	233421	1	0.01	0.0088	88	D19
2,4-D	Flour	ARA-09-15-10-146-0001A7 + 0.10 µg/g	62	112413	230348	1	0.1	0.0876	88	D19
2,4-D	Flour	ARA-09-15-10-146-0001A8 + 0.10 µg/g	63	107520	222553	1	0.1	0.0867	87	D19
2,4-D	Oil, refined	ARA-09-15-10-143-0001A30	3	7301	591836	1	NA	0.0008	NA	D20
2,4-D	Oil, refined	ARA-09-15-10-143-0001A31 + 0.003 µg/g	23	8057	407032	1	0.003	0.0014	NA	D20
2,4-D	Oil, refined	ARA-09-15-10-143-0001A32 + 0.01 µg/g	25	20554	377955	1	0.01	0.0077	77	D20
2,4-D	Oil, refined	ARA-09-15-10-143-0001A33 + 0.01 µg/g	27	23310	383130	1	0.01	0.0089	89	D20
2,4-D	Oil, refined	ARA-09-15-10-143-0001A34 + 0.10 µg/g	30	186415	355512	1	0.1	0.0937	94	D20

Table 3 (Cont.). Procedural Recovery Raw Data for Corn Samples

Analyte	Matrix	Control Sample ID	Inj. No	Analyte Peak Area	ISTD Peak Area	Dilution Factor	Fortified Conc. (µg/g)	Found (µg/g)[a]	% Rec	Set ID
2,4-D	Oil, refined	ARA-09-15-10-143-0001A35 + 0.10 µg/g	32	194629	359192	1	0.1	0.0969	97	D20
2,4-D	Oil, refined	ARA-09-15-10-152-0001A2	13	0	510397	1	NA	ND	NA	D20
2,4-D	Oil, refined	ARA-09-15-10-152-0001A3 + 0.003 µg/g	24	8297	401402	1	0.003	0.0024	NA	D20
2,4-D	Oil, refined	ARA-09-15-10-152-0001A4 + 0.01 µg/g	28	22652	384183	1	0.01	0.0094	94	D20
2,4-D	Oil, refined	ARA-09-15-10-152-0001A5 + 0.01 µg/g	29	22351	365333	1	0.01	0.0098	98	D20
2,4-D	Oil, refined	ARA-09-15-10-152-0001A6 + 0.10 µg/g	33	189609	351324	1	0.1	0.0973	97	D20
2,4-D	Oil, refined	ARA-09-15-10-152-0001A7 + 0.10 µg/g	34	185997	343575	1	0.1	0.0976	98	D20
2,4-D	Forage	ARA-09-15-10-046-0001A2	26	0	297466	1	NA	ND	NA	D21
2,4-D	Forage	ARA-09-15-10-046-0001A4 + 0.003 µg/g	46	6821	247811	1	0.003	0.0029	NA	D21
2,4-D	Forage	ARA-09-15-10-046-0001A5 + 0.01 µg/g	47	16956	249962	1	0.01	0.0103	103	D21
2,4-D	Forage	ARA-09-15-10-046-0001A6 + 0.01 µg/g	48	16108	246722	1	0.01	0.0098	98	D21
2,4-D	Forage	ARA-09-15-10-046-0001A7 + 0.10 µg/g	51	130190	244919	1	0.1	0.0955	95	D21
2,4-D	Forage	ARA-09-15-10-046-0001A8 + 0.10 µg/g	52	132347	244816	1	0.1	0.0971	97	D21
2,4-D	Forage	ARA-09-15-10-046-0001A9	3	3254	572889	1	NA	ND	NA	D22
2,4-D	Forage	ARA-09-15-10-046-0001A10 + 0.003 µg/g	46	6265	274937	1	0.003	0.0031	NA	D22
2,4-D	Forage	ARA-09-15-10-046-0001A11 + 0.01 µg/g	47	16105	276054	1	0.01	0.0095	95	D22
2,4-D	Forage	ARA-09-15-10-046-0001A12 + 0.01 µg/g	48	16080	271990	1	0.01	0.0097	97	D22
2,4-D	Forage	ARA-09-15-10-046-0001A13 + 0.10 µg/g	51	148798	271005	1	0.1	0.0984	98	D22
2,4-D	Forage	ARA-09-15-10-046-0001A14 + 0.10 µg/g	52	146283	268578	1	0.1	0.0976	98	D22
2,4-D	Forage	ARA-09-15-10-046-0001A15	3	3249	600794	1	NA	0.0009	NA	D23
2,4-D	Forage	ARA-09-15-10-046-0001A16 + 0.003 µg/g	4	13901	583589	1	0.003	0.0033	NA	D23
2,4-D	Forage	ARA-09-15-10-046-0001A17 + 0.01 µg/g	5	33555	583077	1	0.01	0.0095	95	D23
2,4-D	Forage	ARA-09-15-10-046-0001A18 + 0.01 µg/g	6	33128	566226	1	0.01	0.0096	96	D23
2,4-D	Forage	ARA-09-15-10-046-0001A19 + 2.00 µg/g	51	308487	292648	10	2	1.9095	95	D23
2,4-D	Forage	ARA-09-15-10-046-0001A20 + 2.00 µg/g	52	306405	282067	10	2	1.9678	98	D23
2,4-D	Forage	ARA-09-15-10-046-0001A21	3	4323	521329	1	NA	ND	NA	D24
2,4-D	Forage	ARA-09-15-10-046-0001A22 + 0.003 µg/g	4	11512	519716	1	0.003	0.0025	NA	D24
2,4-D	Forage	ARA-09-15-10-046-0001A23 + 0.01 µg/g	5	33059	500274	1	0.01	0.0104	104	D24

Table 3 (Cont.). Procedural Recovery Raw Data for Corn Samples

Analyte	Matrix	Control Sample ID	Inj. No	Analyte Peak Area	ISTD Peak Area	Dilution Factor	Fortified Conc. (µg/g)	Found (µg/g)[a]	% Rec	Set ID
2,4-D	Forage	ARA-09-15-10-046-0001A24 + 0.01 µg/g	6	31934	489880	1	0.01	0.0102	102	D24
2,4-D	Forage	ARA-09-15-10-046-0001A25 + 4.00 µg/g	51	627729	258598	10	4	4.3672	109	D24
2,4-D	Forage	ARA-09-15-10-046-0001A26 + 4.00 µg/g	52	571039	251341	10	4	4.0866	102	D24
2,4-D	Forage	ARA-09-15-10-046-0001A3	3	0	287532	1	NA	ND	NA	D25
2,4-D	Forage	ARA-09-15-10-046-0001A27 + 0.003 µg/g	4	5394	283855	1	0.003	0.0026	NA	D25
2,4-D	Forage	ARA-09-15-10-046-0001A28 + 0.01 µg/g	5	16955	285749	1	0.01	0.0098	98	D25
2,4-D	Forage	ARA-09-15-10-046-0001A29 + 0.01 µg/g	6	16821	269840	1	0.01	0.0103	103	D25
2,4-D	Forage	ARA-09-15-10-046-0001A30 + 10.0 µg/g	47	1276262	237807	10	10	9.5007	95	D25
2,4-D	Forage	ARA-09-15-10-046-0001A31 + 10.0 µg/g	48	1255771	236499	10	10	9.3998	94	D25
2,4-D	Forage	ARA-09-15-10-046-0001A32 + 10.0 µg/g	50	1190848	242740	10	10	8.6841	87	D25
2,4-D	Forage	ARA-09-15-10-046-0001A33 + 10.0 µg/g	51	1201698	235378	10	10	9.0376	90	D25
2,4-D	Forage	ARA-09-15-10-046-0001A34 + 10.0 µg/g	52	1213897	224906	10	10	9.5548	96	D25
2,4-D	Forage	ARA-09-15-10-046-0001A35 + 10.0 µg/g	53	1239965	233465	10	10	9.4021	94	D25
2,4-D	Stover	ARA-09-15-10-021-0001A10	5	4259	245162	1	NA	0.0018	NA	D26
2,4-D	Stover	ARA-09-15-10-021-0001A11 + 0.003 µg/g	46	7504	233378	1	0.003	0.0027	NA	D26
2,4-D	Stover	ARA-09-15-10-021-0001A12 + 0.01 µg/g	47	15587	231003	1	0.01	0.0092	92	D26
2,4-D	Stover	ARA-09-15-10-021-0001A13 + 0.01 µg/g	48	16962	247143	1	0.01	0.0095	95	D26
2,4-D	Stover	ARA-09-15-10-021-0001A14 + 0.10 µg/g	51	124423	227446	1	0.1	0.0977	98	D26
2,4-D	Stover	ARA-09-15-10-021-0001A15 + 0.10 µg/g	52	123165	232687	1	0.1	0.0944	94	D26
2,4-D	Stover	ARA-09-15-10-021-0001A3	3	8024	446540	1	NA	0.0020	NA	D27
2,4-D	Stover	ARA-09-15-10-021-0001A4 + 0.003 µg/g	12	11852	368153	1	0.003	0.0026	NA	D27
2,4-D	Stover	ARA-09-15-10-021-0001A5 + 0.01 µg/g	13	26492	374864	1	0.01	0.0097	97	D27
2,4-D	Stover	ARA-09-15-10-021-0001A6 + 0.01 µg/g	14	26498	371678	1	0.01	0.0099	99	D27
2,4-D	Stover	ARA-09-15-10-021-0001A7 + 1.00 µg/g	48	1311372	246288	1	1	0.9804	98	D27
2,4-D	Stover	ARA-09-15-10-021-0001A8 + 1.00 µg/g	49	1276225	247461	1	1	0.9495	95	D27
2,4-D	Stover	ARA-09-15-10-021-0001A9	3	4246	303349	1	NA	0.0015	NA	D28
2,4-D	Stover	ARA-09-15-10-021-0001A16 + 0.003 µg/g	4	8536	288953	1	0.003	0.0028	NA	D28
2,4-D	Stover	ARA-09-15-10-021-0001A17 + 0.01 µg/g	5	19468	280837	1	0.01	0.0100	100	D28

Table 3 (Cont.). Procedural Recovery Raw Data for Corn Samples

Analyte	Matrix	Control Sample ID	Inj. No	Analyte Peak Area	ISTD Peak Area	Dilution Factor	Fortified Conc. (µg/g)	Found (µg/g)[a]	% Rec	Set ID
2,4-D	Stover	ARA-09-15-10-021-0001A18 + 0.01 µg/g	6	19505	281992	1	0.01	0.0100	100	D28
2,4-D	Stover	ARA-09-15-10-021-0001A19 + 20.0 µg/g	10*	376499	364200	100	20	18.6370	93	D28
2,4-D	Stover	ARA-09-15-10-021-0001A2 + 20.0 µg/g	15*	390707	371355	100	20	18.9694	95	D28
2,4-D	Stover	ARA-09-15-10-021-0001A20 + 20.0 µg/g	11*	415055	393961	100	20	18.9954	95	D28
2,4-D	Stover	ARA-09-15-10-021-0001A21 + 20.0 µg/g	12*	405376	388237	100	20	18.8250	94	D28
2,4-D	Stover	ARA-09-15-10-021-0001A22 + 20.0 µg/g	13*	335499	373670	100	20	16.1730	81	D28
2,4-D	Stover	ARA-09-15-10-021-0001A23 + 20.0 µg/g	14*	397225	399058	100	20	17.9415	90	D28
2,4-DCP	Grain	ARA-09-15-10-011-0001A2	3	0	28149	1	NA	ND	NA	D16
2,4-DCP	Grain	ARA-09-15-10-011-0001A3 + 0.003 µg/g	57	3087	20577	1	0.003	0.0030	NA	D16
2,4-DCP	Grain	ARA-09-15-10-011-0001A4 + 0.01 µg/g	58	8911	22417	1	0.01	0.0099	99	D16
2,4-DCP	Grain	ARA-09-15-10-011-0001A5 + 0.01 µg/g	59	9027	22682	1	0.01	0.0099	99	D16
2,4-DCP	Grain	ARA-09-15-10-011-0001A6 + 0.10 µg/g	60	79842	22757	1	0.1	0.0976	98	D16
2,4-DCP	Grain	ARA-09-15-10-011-0001A7 + 0.10 µg/g	61	69074	19978	1	0.1	0.0961	96	D16
2,4-DCP	Grain	ARA-09-15-10-011-0001A8	3	0	26445	1	NA	ND	NA	D17
2,4-DCP	Grain	ARA-09-15-10-011-0001A9 + 0.003 µg/g	58	3428	22841	1	0.003	0.0030	NA	D17
2,4-DCP	Grain	ARA-09-15-10-011-0001A10 + 0.01 µg/g	59	8349	19954	1	0.01	0.0106	106	D17
2,4-DCP	Grain	ARA-09-15-10-011-0001A11 + 0.01 µg/g	60	9936	25400	1	0.01	0.0098	98	D17
2,4-DCP	Grain	ARA-09-15-10-011-0001A12 + 0.10 µg/g	62	87352	25219	1	0.1	0.0968	97	D17
2,4-DCP	Grain	ARA-09-15-10-011-0001A13 + 0.10 µg/g	63	72218	20194	1	0.1	0.1000	100	D17
2,4-DCP	Grain	ARA-09-15-10-011-0001A14	3	1424	22537	1	NA	0.0001	NA	D18
2,4-DCP	Grain	ARA-09-15-10-011-0001A15 + 0.003 µg/g	53	3684	22774	1	0.003	0.0028	NA	D18
2,4-DCP	Grain	ARA-09-15-10-011-0001A16 + 0.01 µg/g	54	9457	22780	1	0.01	0.0100	100	D18
2,4-DCP	Grain	ARA-09-15-10-011-0001A17 + 0.01 µg/g	56	7599	17148	1	0.01	0.0108	108	D18
2,4-DCP	Grain	ARA-09-15-10-011-0001A18 + 0.10 µg/g	57	55382	16415	1	0.1	0.0938	94	D18
2,4-DCP	Grain	ARA-09-15-10-011-0001A19 + 0.10 µg/g	58	52693	14901	1	0.1	0.0984	98	D18
2,4-DCP	Aspirated grain fractions	ARA-09-15-10-141-0001A2	3	1393	16399	1	NA	0.0012	NA	D19
2,4-DCP	Aspirated grain fractions	ARA-09-15-10-141-0001A3 + 0.003 µg/g	43	4123	22834	1	0.003	0.0027	NA	D19

Table 3 (Cont.). Procedural Recovery Raw Data for Corn Samples

Analyte	Matrix	Control Sample ID	Inj. No	Analyte Peak Area	ISTD Peak Area	Dilution Factor	Fortified Conc. (µg/g)	Found (µg/g)[a]	% Rec	Set ID
2,4-DCP	Aspirated grain fractions	ARA-09-15-10-141-0001A4 + 0.01 µg/g	48	7184	17902	1	0.01	0.0090	90	D19
2,4-DCP	Aspirated grain fractions	ARA-09-15-10-141-0001A5 + 0.01 µg/g	49	7671	18837	1	0.01	0.0091	91	D19
2,4-DCP	Aspirated grain fractions	ARA-09-15-10-141-0001A6 + 0.10 µg/g	59	59990	18540	1	0.1	0.0894	89	D19
2,4-DCP	Aspirated grain fractions	ARA-09-15-10-141-0001A7 + 0.10 µg/g	60	65667	18686	1	0.1	0.0973	97	D19
2,4-DCP	Starch	ARA-09-15-10-142-0001A30	34	1562	17827	1	NA	0.0013	NA	D19
2,4-DCP	Starch	ARA-09-15-10-142-0001A31 + 0.003 µg/g	47	3894	20771	1	0.003	0.0028	NA	D19
2,4-DCP	Starch	ARA-09-15-10-142-0001A32 + 0.01 µg/g	57	7108	20297	1	0.01	0.0074	74	D19
2,4-DCP	Starch	ARA-09-15-10-142-0001A33 + 0.01 µg/g	58	7262	21160	1	0.01	0.0073	73	D19
2,4-DCP	Starch	ARA-09-15-10-142-0001A34 + 0.10 µg/g	68	47076	21162	1	0.1	0.0606	61	D19
2,4-DCP	Starch	ARA-09-15-10-142-0001A35 + 0.10 µg/g	69	39147	15515	1	0.1	0.0691	69	D19
2,4-DCP	Meal	ARA-09-15-10-144-0001A22	25	1292	15916	1	NA	0.0011	NA	D19
2,4-DCP	Meal	ARA-09-15-10-144-0001A23 + 0.003 µg/g	46	4220	23073	1	0.003	0.0029	NA	D19
2,4-DCP	Meal	ARA-09-15-10-144-0001A24 + 0.01 µg/g	55	8765	21665	1	0.01	0.0092	92	D19
2,4-DCP	Meal	ARA-09-15-10-144-0001A25 + 0.01 µg/g	56	8942	20939	1	0.01	0.0098	98	D19
2,4-DCP	Meal	ARA-09-15-10-144-0001A26 + 0.10 µg/g	66	73435	20739	1	0.1	0.0982	98	D19
2,4-DCP	Meal	ARA-09-15-10-144-0001A27 + 0.10 µg/g	67	69889	19777	1	0.1	0.0980	98	D19
2,4-DCP	Grits	ARA-09-15-10-145-0001A22	16	1723	23418	1	NA	0.0009	NA	D19
2,4-DCP	Grits	ARA-09-15-10-145-0001A23 + 0.003 µg/g	45	3946	21022	1	0.003	0.0032	NA	D19
2,4-DCP	Grits	ARA-09-15-10-145-0001A24 + 0.01 µg/g	53	8381	20063	1	0.01	0.0098	98	D19
2,4-DCP	Grits	ARA-09-15-10-145-0001A25 + 0.01 µg/g	54	7305	16009	1	0.01	0.0109	109	D19
2,4-DCP	Grits	ARA-09-15-10-145-0001A26 + 0.10 µg/g	64	70618	20264	1	0.1	0.0968	97	D19
2,4-DCP	Grits	ARA-09-15-10-145-0001A27 + 0.10 µg/g	65	67753	19363	1	0.1	0.0972	97	D19

Table 3 (Cont.). Procedural Recovery Raw Data for Corn Samples

Analyte	Matrix	Control Sample ID	Inj. No	Analyte Peak Area	ISTD Peak Area	Dilution Factor	Fortified Conc. (µg/g)	Found (µg/g)[a]	% Rec	Set ID
2,4-DCP	Flour	ARA-09-15-10-146-0001A3	7	0	20826	1	NA	ND	NA	D19
2,4-DCP	Flour	ARA-09-15-10-146-0001A4 + 0.003 µg/g	44	3202	22835	1	0.003	0.0028	NA	D19
2,4-DCP	Flour	ARA-09-15-10-146-0001A5 + 0.01 µg/g	50	8189	20456	1	0.01	0.0102	102	D19
2,4-DCP	Flour	ARA-09-15-10-146-0001A6 + 0.01 µg/g	52	7079	16782	1	0.01	0.0108	108	D19
2,4-DCP	Flour	ARA-09-15-10-146-0001A7 + 0.10 µg/g	62	59090	17697	1	0.1	0.0936	94	D19
2,4-DCP	Flour	ARA-09-15-10-146-0001A8 + 0.10 µg/g	63	72463	21086	1	0.1	0.0963	96	D19
2,4-DCP	Oil, refined	ARA-09-15-10-143-0001A30	3	0	25816	1	NA	ND	NA	D20
2,4-DCP	Oil, refined	ARA-09-15-10-143-0001A31 + 0.003 µg/g	23	3934	30358	1	0.003	0.0028	NA	D20
2,4-DCP	Oil, refined	ARA-09-15-10-143-0001A32 + 0.01 µg/g	25	10582	34121	1	0.01	0.0080	80	D20
2,4-DCP	Oil, refined	ARA-09-15-10-143-0001A33 + 0.01 µg/g	27	11848	30434	1	0.01	0.0102	102	D20
2,4-DCP	Oil, refined	ARA-09-15-10-143-0001A34 + 0.10 µg/g	30	64882	22992	1	0.1	0.0797	80	D20
2,4-DCP	Oil, refined	ARA-09-15-10-143-0001A35 + 0.10 µg/g	32	96144	31433	1	0.1	0.0864	86	D20
2,4-DCP	Oil, refined	ARA-09-15-10-152-0001A2	13	1355	32576	1	NA	0.0003	NA	D20
2,4-DCP	Oil, refined	ARA-09-15-10-152-0001A3 + 0.003 µg/g	24	3160	24703	1	0.003	0.0025	NA	D20
2,4-DCP	Oil, refined	ARA-09-15-10-152-0001A4 + 0.01 µg/g	28	10809	29719	1	0.01	0.0092	92	D20
2,4-DCP	Oil, refined	ARA-09-15-10-152-0001A5 + 0.01 µg/g	29	9544	29843	1	0.01	0.0079	79	D20
2,4-DCP	Oil, refined	ARA-09-15-10-152-0001A6 + 0.10 µg/g	33	71703	23426	1	0.1	0.0862	86	D20
2,4-DCP	Oil, refined	ARA-09-15-10-152-0001A7 + 0.10 µg/g	34	70384	22425	1	0.1	0.0884	88	D20
2,4-DCP	Forage	ARA-09-15-10-046-0001A2	26	2909	15380	1	NA	0.0020	NA	D21
2,4-DCP	Forage	ARA-09-15-10-046-0001A4 + 0.003 µg/g	46	5502	19997	1	0.003	0.0025	NA	D21
2,4-DCP	Forage	ARA-09-15-10-046-0001A5 + 0.01 µg/g	47	9611	20258	1	0.01	0.0082	82	D21
2,4-DCP	Forage	ARA-09-15-10-046-0001A6 + 0.01 µg/g	48	8490	17666	1	0.01	0.0084	84	D21
2,4-DCP	Forage	ARA-09-15-10-046-0001A7 + 0.10 µg/g	51	68806	18788	1	0.1	0.1002	100	D21

Table 3 (Cont.). Procedural Recovery Raw Data for Corn Samples

Analyte	Matrix	Control Sample ID	Inj. No	Analyte Peak Area	ISTD Peak Area	Dilution Factor	Fortified Conc. (µg/g)	Found (µg/g)[a]	% Rec	Set ID
2,4-DCP	Forage	ARA-09-15-10-046-0001A8 + 0.10 µg/g	52	65663	19966	1	0.1	0.0894	89	D21
2,4-DCP	Forage	ARA-09-15-10-046-0001A9	3	1291	20991	1	NA	0.0002	NA	D22
2,4-DCP	Forage	ARA-09-15-10-046-0001A10 + 0.003 µg/g	46	3219	16263	1	0.003	0.0038	NA	D22
2,4-DCP	Forage	ARA-09-15-10-046-0001A11 + 0.01 µg/g	47	8234	18101	1	0.01	0.0111	111	D22
2,4-DCP	Forage	ARA-09-15-10-046-0001A12 + 0.01 µg/g	48	6585	15740	1	0.01	0.0101	101	D22
2,4-DCP	Forage	ARA-09-15-10-046-0001A13 + 0.10 µg/g	51	77449	20351	1	0.1	0.1057	106	D22
2,4-DCP	Forage	ARA-09-15-10-046-0001A14 + 0.10 µg/g	52	73659	21202	1	0.1	0.0963	96	D22
2,4-DCP	Forage	ARA-09-15-10-046-0001A15	3	2286	21022	1	NA	0.0014	NA	D23
2,4-DCP	Forage	ARA-09-15-10-046-0001A16 + 0.003 µg/g	4	3890	18234	1	0.003	0.0030	NA	D23
2,4-DCP	Forage	ARA-09-15-10-046-0001A17 + 0.01 µg/g	5	6923	14216	1	0.01	0.0109	109	D23
2,4-DCP	Forage	ARA-09-15-10-046-0001A18 + 0.01 µg/g	6	7888	17534	1	0.01	0.0098	98	D23
2,4-DCP	Forage	ARA-09-15-10-046-0001A19 + 2.00 µg/g	51	157709	25072	10	2	1.7972	90	D23
2,4-DCP	Forage	ARA-09-15-10-046-0001A20 + 2.00 µg/g	52	152688	23832	10	2	1.8308	92	D23
2,4-DCP	Forage	ARA-09-15-10-046-0001A21	3	2409	16177	1	NA	0.0027	NA	D24
2,4-DCP	Forage	ARA-09-15-10-046-0001A22 + 0.003 µg/g	4	4986	22142	1	0.003	0.0021	NA	D24
2,4-DCP	Forage	ARA-09-15-10-046-0001A23 + 0.01 µg/g	5	8389	16959	1	0.01	0.0097	97	D24
2,4-DCP	Forage	ARA-09-15-10-046-0001A24 + 0.01 µg/g	6	10789	21455	1	0.01	0.0099	99	D24
2,4-DCP	Forage	ARA-09-15-10-046-0001A25 + 4.00 µg/g	51	204568	14205	10	4	4.0133	100	D24
2,4-DCP	Forage	ARA-09-15-10-046-0001A26 + 4.00 µg/g	52	249968	18526	10	4	3.7591	94	D24
2,4-DCP	Forage	ARA-09-15-10-046-0001A3	3	2196	20467	1	NA	0.0017	NA	D25
2,4-DCP	Forage	ARA-09-15-10-046-0001A27 + 0.003 µg/g	4	4308	18096	1	0.003	0.0036	NA	D25
2,4-DCP	Forage	ARA-09-15-10-046-0001A28 + 0.01 µg/g	5	9940	20872	1	0.01	0.0102	102	D25
2,4-DCP	Forage	ARA-09-15-10-046-0001A29 + 0.01 µg/g	6	7807	16042	1	0.01	0.0104	104	D25

Table 3 (Cont.). Procedural Recovery Raw Data for Corn Samples

Analyte	Matrix	Control Sample ID	Inj. No	Analyte Peak Area	ISTD Peak Area	Dilution Factor	Fortified Conc. (µg/g)	Found (µg/g)[a]	% Rec	Set ID
2,4-DCP	Forage	ARA-09-15-10-046-0001A30 + 10.0 µg/g	47	659880	19137	10	10	9.4745	95	D25
2,4-DCP	Forage	ARA-09-15-10-046-0001A31 + 10.0 µg/g	48	700042	21175	10	10	9.0832	91	D25
2,4-DCP	Forage	ARA-09-15-10-046-0001A32 + 10.0 µg/g	50	562462	16932	10	10	9.1270	91	D25
2,4-DCP	Forage	ARA-09-15-10-046-0001A33 + 10.0 µg/g	51	681497	22822	10	10	8.2031	82	D25
2,4-DCP	Forage	ARA-09-15-10-046-0001A34 + 10.0 µg/g	52	586176	17340	10	10	9.2882	93	D25
2,4-DCP	Forage	ARA-09-15-10-046-0001A35 + 10.0 µg/g	53	697631	22274	10	10	8.6046	86	D25
2,4-DCP	Stover	ARA-09-15-10-021-0001A10	5	3323	24075	1	NA	0.0025	NA	D26
2,4-DCP	Stover	ARA-09-15-10-021-0001A11 + 0.003 µg/g	46	5470	20996	1	0.003	0.0034	NA	D26
2,4-DCP	Stover	ARA-09-15-10-021-0001A12 + 0.01 µg/g	47	11021	21266	1	0.01	0.0105	105	D26
2,4-DCP	Stover	ARA-09-15-10-021-0001A13 + 0.01 µg/g	48	10714	19657	1	0.01	0.0112	112	D26
2,4-DCP	Stover	ARA-09-15-10-021-0001A14 + 0.10 µg/g	51	75136	19971	1	0.1	0.0999	100	D26
2,4-DCP	Stover	ARA-09-15-10-021-0001A15 + 0.10 µg/g	52	76977	19344	1	0.1	0.1059	106	D26
2,4-DCP	Stover	ARA-09-15-10-021-0001A3	3	3316	23640	1	NA	0.0026	NA	D27
2,4-DCP	Stover	ARA-09-15-10-021-0001A4 + 0.003 µg/g	12	5182	21738	1	0.003	0.0028	NA	D27
2,4-DCP	Stover	ARA-09-15-10-021-0001A5 + 0.01 µg/g	13	10369	21224	1	0.01	0.0099	99	D27
2,4-DCP	Stover	ARA-09-15-10-021-0001A6 + 0.01 µg/g	14	11303	22725	1	0.01	0.0102	102	D27
2,4-DCP	Stover	ARA-09-15-10-021-0001A7 + 1.00 µg/g	48	780612	21329	1	1	1.0366	104	D27
2,4-DCP	Stover	ARA-09-15-10-021-0001A8 + 1.00 µg/g	49	801361	21830	1	1	1.0397	104	D27
2,4-DCP	Stover	ARA-09-15-10-021-0001A9	3	3411	27257	1	NA	0.0019	NA	D28
2,4-DCP	Stover	ARA-09-15-10-021-0001A16 + 0.003 µg/g	4	5748	24652	1	0.003	0.0031	NA	D28
2,4-DCP	Stover	ARA-09-15-10-021-0001A17 + 0.01 µg/g	5	12196	25154	1	0.01	0.0103	103	D28
2,4-DCP	Stover	ARA-09-15-10-021-0001A18 + 0.01 µg/g	6	11426	22216	1	0.01	0.0111	111	D28
2,4-DCP	Stover	ARA-09-15-10-021-0001A19 + 20.0 µg/g	10*	223042	39241	100	20	16.0603	80	D28
2,4-DCP	Stover	ARA-09-15-10-021-0001A2 + 20.0 µg/g	15*	205002	33154	100	20	17.4863	87	D28

Table 3 (Cont.). Procedural Recovery Raw Data for Corn Samples

Analyte	Matrix	Control Sample ID	Inj. No	Analyte Peak Area	ISTD Peak Area	Dilution Factor	Fortified Conc. (µg/g)	Found (µg/g)[a]	% Rec	Set ID
2,4-DCP	Stover	ARA-09-15-10-021-0001A20 + 20.0 µg/g	11*	169236	27720	100	20	17.2633	86	D28
2,4-DCP	Stover	ARA-09-15-10-021-0001A21 + 20.0 µg/g	12*	222168	35464	100	20	17.7184	89	D28
2,4-DCP	Stover	ARA-09-15-10-021-0001A22 + 20.0 µg/g	13*	183807	34172	100	20	15.1894	76	D28
2,4-DCP	Stover	ARA-09-15-10-021-0001A23 + 20.0 µg/g	14*	194996	34301	100	20	16.0631	80	D28
QUIZALOFOP ACID	Grain	ARA-09-15-10-020-0001A2	3	0	--	1	NA	ND	NA	Q01
QUIZALOFOP ACID	Grain	ARA-09-15-10-020-0001A3 + 0.003 µg/g	4	1397	--	1	0.003	0.0033	NA	Q01
QUIZALOFOP ACID	Grain	ARA-09-15-10-020-0001A4 + 0.01 µg/g	5	5095	--	1	0.01	0.0104	104	Q01
QUIZALOFOP ACID	Grain	ARA-09-15-10-020-0001A5 + 0.01 µg/g	6	5182	--	1	0.01	0.0105	105	Q01
QUIZALOFOP ACID	Grain	ARA-09-15-10-020-0001A6 + 1.0 µg/g	7	529698	--	1	1	1.0107	101	Q01
QUIZALOFOP ACID	Grain	ARA-09-15-10-020-0001A7 + 1.0 µg/g	8	508181	--	1	1	0.9697	97	Q01
QUIZALOFOP ETHYL	Grain	ARA-09-15-10-020-0001A2	3	0	--	1	NA	ND	NA	Q01
QUIZALOFOP ETHYL	Grain	ARA-09-15-10-020-0001A3 + 0.003 µg/g	4	1104	--	1	0.003	0.0032	NA	Q01
QUIZALOFOP ETHYL	Grain	ARA-09-15-10-020-0001A4 + 0.01 µg/g	5	3968	--	1	0.01	0.0108	108	Q01
QUIZALOFOP ETHYL	Grain	ARA-09-15-10-020-0001A5 + 0.01 µg/g	6	3940	--	1	0.01	0.0107	107	Q01
QUIZALOFOP ETHYL	Grain	ARA-09-15-10-020-0001A6 + 1.0 µg/g	7	426810	--	1	1	1.1228	112	Q01
QUIZALOFOP ETHYL	Grain	ARA-09-15-10-020-0001A7 + 1.0 µg/g	8	406139	--	1	1	1.0685	107	Q01
QUIZALOFOP ACID	Grain	ARA-09-15-10-020-0001A8	3	0	--	1	NA	ND	NA	Q02
QUIZALOFOP ACID	Grain	ARA-09-15-10-020-0001A9 + 0.003 µg/g	4	1455	--	1	0.003	0.0028	NA	Q02
QUIZALOFOP ACID	Grain	ARA-09-15-10-020-0001A10 + 0.01 µg/g	5	5172	--	1	0.01	0.0100	100	Q02
QUIZALOFOP ACID	Grain	ARA-09-15-10-020-0001A11 + 0.01 µg/g	6	5258	--	1	0.01	0.0102	102	Q02
QUIZALOFOP ACID	Grain	ARA-09-15-10-020-0001A12 + 1.0 µg/g	7	508658	--	1	1	0.9896	99	Q02
QUIZALOFOP ACID	Grain	ARA-09-15-10-020-0001A13 + 1.0 µg/g	8	508119	--	1	1	0.9886	99	Q02
QUIZALOFOP ETHYL	Grain	ARA-09-15-10-020-0001A8	3	0	--	1	NA	ND	NA	Q02
QUIZALOFOP ETHYL	Grain	ARA-09-15-10-020-0001A9 + 0.003 µg/g	4	1396	--	1	0.003	0.0030	NA	Q02
QUIZALOFOP ETHYL	Grain	ARA-09-15-10-020-0001A10 + 0.01 µg/g	5	3964	--	1	0.01	0.0098	98	Q02
QUIZALOFOP ETHYL	Grain	ARA-09-15-10-020-0001A11 + 0.01 µg/g	6	4179	--	1	0.01	0.0103	103	Q02

Table 3 (Cont.). Procedural Recovery Raw Data for Corn Samples

Analyte	Matrix	Control Sample ID	Inj. No	Analyte Peak Area	ISTD Peak Area	Dilution Factor	Fortified Conc. (µg/g)	Found (µg/g)[a]	% Rec	Set ID
QUIZALOFOP ETHYL	Grain	ARA-09-15-10-020-0001A12 + 1.0 µg/g	7	406851	--	1	1	1.0729	107	Q02
QUIZALOFOP ETHYL	Grain	ARA-09-15-10-020-0001A13 + 1.0 µg/g	8	415269	--	1	1	1.0951	110	Q02
QUIZALOFOP ACID	Grain	ARA-09-15-10-020-0001A14	3	0	--	1	NA	ND	NA	Q03
QUIZALOFOP ACID	Grain	ARA-09-15-10-020-0001A15 + 0.003 µg/g	4	1308	--	1	0.003	0.0040	NA	Q03
QUIZALOFOP ACID	Grain	ARA-09-15-10-020-0001A16 + 0.01 µg/g	5	4407	--	1	0.01	0.0116	116	Q03
QUIZALOFOP ACID	Grain	ARA-09-15-10-020-0001A17 + 0.01 µg/g	6	4383	--	1	0.01	0.0115	115	Q03
QUIZALOFOP ACID	Grain	ARA-09-15-10-020-0001A18 + 1.0 µg/g	7	427224	--	1	1	1.0509	105	Q03
QUIZALOFOP ACID	Grain	ARA-09-15-10-020-0001A19 + 1.0 µg/g	8	421267	--	1	1	1.0363	104	Q03
QUIZALOFOP ETHYL	Grain	ARA-09-15-10-020-0001A14	3	0	--	1	NA	ND	NA	Q03
QUIZALOFOP ETHYL	Grain	ARA-09-15-10-020-0001A15 + 0.003 µg/g	4	1020	--	1	0.003	0.0035	NA	Q03
QUIZALOFOP ETHYL	Grain	ARA-09-15-10-020-0001A16 + 0.01 µg/g	5	3511	--	1	0.01	0.0103	103	Q03
QUIZALOFOP ETHYL	Grain	ARA-09-15-10-020-0001A17 + 0.01 µg/g	6	3534	--	1	0.01	0.0104	104	Q03
QUIZALOFOP ETHYL	Grain	ARA-09-15-10-020-0001A18 + 1.0 µg/g	7	356917	--	1	1	0.9677	97	Q03
QUIZALOFOP ETHYL	Grain	ARA-09-15-10-020-0001A19 + 1.0 µg/g	8	382296	--	1	1	1.0365	104	Q03
QUIZALOFOP ACID	Forage	ARA-09-15-10-157-0001A1	3	0	--	1	NA	ND	NA	Q04
QUIZALOFOP ACID	Forage	ARA-09-15-10-157-0001A2 + 0.003 µg/g	4	1185	--	1	0.003	0.0026	NA	Q04
QUIZALOFOP ACID	Forage	ARA-09-15-10-157-0001A3 + 0.01 µg/g	5	3615	--	1	0.01	0.0102	102	Q04
QUIZALOFOP ACID	Forage	ARA-09-15-10-157-0001A4 + 0.01 µg/g	6	3604	--	1	0.01	0.0102	102	Q04
QUIZALOFOP ACID	Forage	ARA-09-15-10-157-0001A5 + 1.0 µg/g	7	353079	--	1	1	1.1001	110	Q04
QUIZALOFOP ACID	Forage	ARA-09-15-10-157-0001A6 + 1.0 µg/g	8	340255	--	1	1	1.0601	106	Q04
QUIZALOFOP ETHYL	Forage	ARA-09-15-10-157-0001A1	3	0	--	1	NA	ND	NA	Q04
QUIZALOFOP ETHYL	Forage	ARA-09-15-10-157-0001A2 + 0.003 µg/g	4	1426	--	1	0.003	0.0025	NA	Q04
QUIZALOFOP ETHYL	Forage	ARA-09-15-10-157-0001A3 + 0.01 µg/g	5	4650	--	1	0.01	0.0080	80	Q04
QUIZALOFOP ETHYL	Forage	ARA-09-15-10-157-0001A4 + 0.01 µg/g	6	4643	--	1	0.01	0.0080	80	Q04
QUIZALOFOP ETHYL	Forage	ARA-09-15-10-157-0001A5 + 1.0 µg/g	7	466354	--	1	1	0.8052	81	Q04

Table 3 (Cont.). Procedural Recovery Raw Data for Corn Samples

Analyte	Matrix	Control Sample ID	Inj. No	Analyte Peak Area	ISTD Peak Area	Dilution Factor	Fortified Conc. (µg/g)	Found (µg/g)[a]	% Rec	Set ID
QUIZALOFOP ETHYL	Forage	ARA-09-15-10-157-0001A6 + 1.0 µg/g	8	451229	--	1	1	0.7791	78	Q04
QUIZALOFOP ACID	Forage	ARA-09-15-10-157-0001A7	3	0	--	1	NA	ND	NA	Q05
QUIZALOFOP ACID	Forage	ARA-09-15-10-157-0001A8 + 0.003 µg/g	4	827	--	1	0.003	0.0034	NA	Q05
QUIZALOFOP ACID	Forage	ARA-09-15-10-157-0001A9 + 0.01 µg/g	5	3092	--	1	0.01	0.0115	115	Q05
QUIZALOFOP ACID	Forage	ARA-09-15-10-157-0001A10 + 0.01 µg/g	6	3071	--	1	0.01	0.0114	114	Q05
QUIZALOFOP ACID	Forage	ARA-09-15-10-157-0001A11 + 1.0 µg/g	7	344059	--	1	1	1.2279	123	Q05
QUIZALOFOP ACID	Forage	ARA-09-15-10-157-0001A12 + 1.0 µg/g	8	319902	--	1	1	1.1417	114	Q05
QUIZALOFOP ETHYL	Forage	ARA-09-15-10-157-0001A7	3	0	--	1	NA	ND	NA	Q05
QUIZALOFOP ETHYL	Forage	ARA-09-15-10-157-0001A8 + 0.003 µg/g	4	1389	--	1	0.003	0.0035	NA	Q05
QUIZALOFOP ETHYL	Forage	ARA-09-15-10-157-0001A9 + 0.01 µg/g	5	4646	--	1	0.01	0.0100	100	Q05
QUIZALOFOP ETHYL	Forage	ARA-09-15-10-157-0001A10 + 0.01 µg/g	6	4758	--	1	0.01	0.0102	102	Q05
QUIZALOFOP ETHYL	Forage	ARA-09-15-10-157-0001A11 + 1.0 µg/g	7	489698	--	1	1	0.9801	98	Q05
QUIZALOFOP ETHYL	Forage	ARA-09-15-10-157-0001A12 + 1.0 µg/g	8	485655	--	1	1	0.9720	97	Q05
QUIZALOFOP ACID	Forage	ARA-09-15-10-157-0001A13	3	0	--	1	NA	ND	NA	Q06
QUIZALOFOP ACID	Forage	ARA-09-15-10-157-0001A14 + 0.003 µg/g	4	704	--	1	0.003	0.0025	NA	Q06
QUIZALOFOP ACID	Forage	ARA-09-15-10-157-0001A15 + 0.01 µg/g	5	3110	--	1	0.01	0.0105	105	Q06
QUIZALOFOP ACID	Forage	ARA-09-15-10-157-0001A16 + 0.01 µg/g	6	2782	--	1	0.01	0.0094	94	Q06
QUIZALOFOP ACID	Forage	ARA-09-15-10-157-0001A17 + 1.0 µg/g	7	284917	--	1	1	0.9460	95	Q06
QUIZALOFOP ACID	Forage	ARA-09-15-10-157-0001A18 + 1.0 µg/g	8	324730	--	1	1	1.0781	108	Q06
QUIZALOFOP ETHYL	Forage	ARA-09-15-10-157-0001A13	3	0	--	1	NA	ND	NA	Q06
QUIZALOFOP ETHYL	Forage	ARA-09-15-10-157-0001A14 + 0.003 µg/g	4	1707	--	1	0.003	0.0025	NA	Q06
QUIZALOFOP ETHYL	Forage	ARA-09-15-10-157-0001A15 + 0.01 µg/g	5	4705	--	1	0.01	0.0084	84	Q06
QUIZALOFOP ETHYL	Forage	ARA-09-15-10-157-0001A16 + 0.01 µg/g	6	5384	--	1	0.01	0.0097	97	Q06
QUIZALOFOP ETHYL	Forage	ARA-09-15-10-157-0001A17 + 1.0 µg/g	7	527430	--	1	1	1.0287	103	Q06
QUIZALOFOP ETHYL	Forage	ARA-09-15-10-157-0001A18 + 1.0 µg/g	8	555169	--	1	1	1.0829	108	Q06

Table 3 (Cont.). Procedural Recovery Raw Data for Corn Samples

Analyte	Matrix	Control Sample ID	Inj. No	Analyte Peak Area	ISTD Peak Area	Dilution Factor	Fortified Conc. (µg/g)	Found (µg/g)[a]	% Rec	Set ID
QUIZALOFOP ACID	Forage	ARA-09-15-10-157-0001A19	3	0	--	1	NA	ND	NA	Q07
QUIZALOFOP ACID	Forage	ARA-09-15-10-157-0001A20 + 0.003 µg/g	4	916	--	1	0.003	0.0013	NA	Q07
QUIZALOFOP ACID	Forage	ARA-09-15-10-157-0001A21 + 0.01 µg/g	5	3076	--	1	0.01	0.0091	91	Q07
QUIZALOFOP ACID	Forage	ARA-09-15-10-157-0001A22 + 0.01 µg/g	6	3061	--	1	0.01	0.0090	90	Q07
QUIZALOFOP ACID	Forage	ARA-09-15-10-157-0001A23 + 1.0 µg/g	7	301962	--	1	1	1.0867	109	Q07
QUIZALOFOP ACID	Forage	ARA-09-15-10-157-0001A24 + 1.0 µg/g	8	318686	--	1	1	1.1470	115	Q07
QUIZALOFOP ETHYL	Forage	ARA-09-15-10-157-0001A19	3	0	--	1	NA	ND	NA	Q07
QUIZALOFOP ETHYL	Forage	ARA-09-15-10-157-0001A20 + 0.003 µg/g	4	1634	--	1	0.003	0.0013	NA	Q07
QUIZALOFOP ETHYL	Forage	ARA-09-15-10-157-0001A21 + 0.01 µg/g	5	5205	--	1	0.01	0.0087	87	Q07
QUIZALOFOP ETHYL	Forage	ARA-09-15-10-157-0001A22 + 0.01 µg/g	6	5133	--	1	0.01	0.0085	85	Q07
QUIZALOFOP ETHYL	Forage	ARA-09-15-10-157-0001A23 + 1.0 µg/g	7	503556	--	1	1	1.0397	104	Q07
QUIZALOFOP ETHYL	Forage	ARA-09-15-10-157-0001A24 + 1.0 µg/g	8	508661	--	1	1	1.0502	105	Q07
QUIZALOFOP ACID	Forage	ARA-09-15-10-157-0001A25	3	0	--	1	NA	ND	NA	Q08
QUIZALOFOP ACID	Forage	ARA-09-15-10-157-0001A26 + 0.003 µg/g	4	826	--	1	0.003	0.0030	NA	Q08
QUIZALOFOP ACID	Forage	ARA-09-15-10-157-0001A27 + 0.01 µg/g	5	2717	--	1	0.01	0.0093	93	Q08
QUIZALOFOP ACID	Forage	ARA-09-15-10-157-0001A28 + 0.01 µg/g	6	3127	--	1	0.01	0.0107	107	Q08
QUIZALOFOP ACID	Forage	ARA-09-15-10-157-0001A29 + 1.0 µg/g	8	299252	--	1	1	1.0018	100	Q08
QUIZALOFOP ACID	Forage	ARA-09-15-10-157-0001A30 + 1.0 µg/g	9	295568	--	1	1	0.9895	99	Q08
QUIZALOFOP ETHYL	Forage	ARA-09-15-10-157-0001A25	3	0	--	1	NA	ND	NA	Q08
QUIZALOFOP ETHYL	Forage	ARA-09-15-10-157-0001A26 + 0.003 µg/g	4	1948	--	1	0.003	0.0021	NA	Q08
QUIZALOFOP ETHYL	Forage	ARA-09-15-10-157-0001A27 + 0.01 µg/g	5	4691	--	1	0.01	0.0075	75	Q08
QUIZALOFOP ETHYL	Forage	ARA-09-15-10-157-0001A28 + 0.01 µg/g	6	5312	--	1	0.01	0.0087	87	Q08
QUIZALOFOP ETHYL	Forage	ARA-09-15-10-157-0001A29 + 1.0 µg/g	8	523587	--	1	1	1.0230	102	Q08
QUIZALOFOP ETHYL	Forage	ARA-09-15-10-157-0001A30 + 1.0 µg/g	9	516272	--	1	1	1.0087	101	Q08
QUIZALOFOP ACID	Stover	ARA-09-15-10-158-0001A1	3	0	--	1	NA	ND	NA	Q09

Table 3 (Cont.). Procedural Recovery Raw Data for Corn Samples

Analyte	Matrix	Control Sample ID	Inj. No	Analyte Peak Area	ISTD Peak Area	Dilution Factor	Fortified Conc. (µg/g)	Found (µg/g)[a]	% Rec	Set ID
QUIZALOFOP ACID	Stover	ARA-09-15-10-158-0001A2 + 0.003 µg/g	4	1205	--	1	0.003	0.0027	NA	Q09
QUIZALOFOP ACID	Stover	ARA-09-15-10-158-0001A3 + 0.01 µg/g	5	3488	--	1	0.01	0.0109	109	Q09
QUIZALOFOP ACID	Stover	ARA-09-15-10-158-0001A4 + 0.01 µg/g	6	3452	--	1	0.01	0.0107	107	Q09
QUIZALOFOP ACID	Stover	ARA-09-15-10-158-0001A5 + 1.0 µg/g	7	387200	--	1	1	1.3778	138	Q09
QUIZALOFOP ACID	Stover	ARA-09-15-10-158-0001A6 + 1.0 µg/g	8	386515	--	1	1	1.3754	138	Q09
QUIZALOFOP ETHYL	Stover	ARA-09-15-10-158-0001A1	3	0	--	1	NA	ND	NA	Q09
QUIZALOFOP ETHYL	Stover	ARA-09-15-10-158-0001A2 + 0.003 µg/g	4	2164	--	1	0.003	0.0022	NA	Q09
QUIZALOFOP ETHYL	Stover	ARA-09-15-10-158-0001A3 + 0.01 µg/g	5	5667	--	1	0.01	0.0091	91	Q09
QUIZALOFOP ETHYL	Stover	ARA-09-15-10-158-0001A4 + 0.01 µg/g	6	5591	--	1	0.01	0.0089	89	Q09
QUIZALOFOP ETHYL	Stover	ARA-09-15-10-158-0001A5 + 1.0 µg/g	7	554123	--	1	1	1.0935	109	Q09
QUIZALOFOP ETHYL	Stover	ARA-09-15-10-158-0001A6 + 1.0 µg/g	8	553889	--	1	1	1.0930	109	Q09
QUIZALOFOP ACID	Stover	ARA-09-15-10-158-0001A7	3	0	--	1	NA	ND	NA	Q10
QUIZALOFOP ACID	Stover	ARA-09-15-10-158-0001A8 + 0.003 µg/g	4	1113	--	1	0.003	0.0031	NA	Q10
QUIZALOFOP ACID	Stover	ARA-09-15-10-158-0001A9 + 0.01 µg/g	5	3383	--	1	0.01	0.0110	110	Q10
QUIZALOFOP ACID	Stover	ARA-09-15-10-158-0001A10 + 0.01 µg/g	6	3343	--	1	0.01	0.0109	109	Q10
QUIZALOFOP ACID	Stover	ARA-09-15-10-158-0001A11 + 1.0 µg/g	7	361856	--	1	1	1.2606	126	Q10
QUIZALOFOP ACID	Stover	ARA-09-15-10-158-0001A12 + 1.0 µg/g	8	371377	--	1	1	1.2938	129	Q10
QUIZALOFOP ETHYL	Stover	ARA-09-15-10-158-0001A7	3	0	--	1	NA	ND	NA	Q10
QUIZALOFOP ETHYL	Stover	ARA-09-15-10-158-0001A8 + 0.003 µg/g	4	1916	--	1	0.003	0.0023	NA	Q10
QUIZALOFOP ETHYL	Stover	ARA-09-15-10-158-0001A9 + 0.01 µg/g	5	5146	--	1	0.01	0.0087	87	Q10
QUIZALOFOP ETHYL	Stover	ARA-09-15-10-158-0001A10 + 0.01 µg/g	6	4827	--	1	0.01	0.0081	81	Q10
QUIZALOFOP ETHYL	Stover	ARA-09-15-10-158-0001A11 + 1.0 µg/g	7	530552	--	1	1	1.0541	105	Q10
QUIZALOFOP ETHYL	Stover	ARA-09-15-10-158-0001A12 + 1.0 µg/g	8	505027	--	1	1	1.0033	100	Q10
QUIZALOFOP ACID	Stover	ARA-09-15-10-158-0001A13	3	0	--	1	NA	ND	NA	Q11
QUIZALOFOP ACID	Stover	ARA-09-15-10-158-0001A14 + 0.003 µg/g	4	1081	--	1	0.003	0.0031	NA	Q11

Table 3 (Cont.). Procedural Recovery Raw Data for Corn Samples

Analyte	Matrix	Control Sample ID	Inj. No	Analyte Peak Area	ISTD Peak Area	Dilution Factor	Fortified Conc. (µg/g)	Found (µg/g)[a]	% Rec	Set ID
QUIZALOFOP ACID	Stover	ARA-09-15-10-158-0001A15 + 0.01 µg/g	5	3483	--	1	0.01	0.0118	118	Q11
QUIZALOFOP ACID	Stover	ARA-09-15-10-158-0001A16 + 0.01 µg/g	6	3256	--	1	0.01	0.0110	110	Q11
QUIZALOFOP ACID	Stover	ARA-09-15-10-158-0001A17 + 1.0 µg/g	7	320931	--	1	1	1.1725	117	Q11
QUIZALOFOP ACID	Stover	ARA-09-15-10-158-0001A18 + 1.0 µg/g	9	319385	--	1	1	1.1669	117	Q11
QUIZALOFOP ETHYL	Stover	ARA-09-15-10-158-0001A13	3	0	--	1	NA	ND	NA	Q11
QUIZALOFOP ETHYL	Stover	ARA-09-15-10-158-0001A14 + 0.003 µg/g	4	2020	--	1	0.003	0.0028	NA	Q11
QUIZALOFOP ETHYL	Stover	ARA-09-15-10-158-0001A15 + 0.01 µg/g	5	6349	--	1	0.01	0.0104	104	Q11
QUIZALOFOP ETHYL	Stover	ARA-09-15-10-158-0001A16 + 0.01 µg/g	6	5997	--	1	0.01	0.0098	98	Q11
QUIZALOFOP ETHYL	Stover	ARA-09-15-10-158-0001A17 + 1.0 µg/g	7	614628	--	1	1	1.0853	109	Q11
QUIZALOFOP ETHYL	Stover	ARA-09-15-10-158-0001A18 + 1.0 µg/g	9	608445	--	1	1	1.0743	107	Q11
QUIZALOFOP ACID	Aspirated grain fractions	ARA-09-15-10-150-0001A1	3	0	--	1	NA	ND	NA	Q12
QUIZALOFOP ACID	Flour	ARA-09-15-10-155-0001A1	4	0	--	1	NA	ND	NA	Q12
QUIZALOFOP ACID	Grits	ARA-09-15-10-145-0001A1	5	0	--	1	NA	ND	NA	Q12
QUIZALOFOP ACID	Meal	ARA-09-15-10-144-0001A1	6	0	--	1	NA	ND	NA	Q12
QUIZALOFOP ACID	Aspirated grain fractions	ARA-09-15-10-150-0001A2 + 0.003 µg/g	7	1337	--	1	0.003	0.0035	NA	Q12
QUIZALOFOP ACID	Flour	ARA-09-15-10-155-0001A2 + 0.003 µg/g	8	1377	--	1	0.003	0.0036	NA	Q12
QUIZALOFOP ACID	Grits	ARA-09-15-10-145-0001A2 + 0.003 µg/g	10	1394	--	1	0.003	0.0036	NA	Q12
QUIZALOFOP ACID	Meal	ARA-09-15-10-144-0001A2 + 0.003 µg/g	11	1339	--	1	0.003	0.0035	NA	Q12
QUIZALOFOP ACID	Aspirated grain fractions	ARA-09-15-10-150-0001A3 + 0.01 µg/g	12	4875	--	1	0.01	0.0107	107	Q12
QUIZALOFOP ACID	Aspirated grain fractions	ARA-09-15-10-150-0001A4 + 0.01 µg/g	13	5075	--	1	0.01	0.0111	111	Q12
QUIZALOFOP ACID	Aspirated grain fractions	ARA-09-15-10-150-0001A5 + 0.01 µg/g	14	4930	--	1	0.01	0.0108	108	Q12
QUIZALOFOP ACID	Flour	ARA-09-15-10-155-0001A3 + 0.01 µg/g	15	4634	--	1	0.01	0.0102	102	Q12

Table 3 (Cont.). Procedural Recovery Raw Data for Corn Samples

Analyte	Matrix	Control Sample ID	Inj. No	Analyte Peak Area	ISTD Peak Area	Dilution Factor	Fortified Conc. (µg/g)	Found (µg/g)[a]	% Rec	Set ID
QUIZALOFOP ACID	Flour	ARA-09-15-10-155-0001A4 + 0.01 µg/g	16	4903	--	1	0.01	0.0107	107	Q12
QUIZALOFOP ACID	Flour	ARA-09-15-10-155-0001A5 + 0.01 µg/g	18	4990	--	1	0.01	0.0109	109	Q12
QUIZALOFOP ACID	Grits	ARA-09-15-10-145-0001A3 + 0.01 µg/g	19	5203	--	1	0.01	0.0113	113	Q12
QUIZALOFOP ACID	Grits	ARA-09-15-10-145-0001A4 + 0.01 µg/g	20	5182	--	1	0.01	0.0113	113	Q12
QUIZALOFOP ACID	Grits	ARA-09-15-10-145-0001A5 + 0.01 µg/g	21	5172	--	1	0.01	0.0113	113	Q12
QUIZALOFOP ACID	Meal	ARA-09-15-10-144-0001A3 + 0.01 µg/g	22	5271	--	1	0.01	0.0115	115	Q12
QUIZALOFOP ACID	Meal	ARA-09-15-10-144-0001A4 + 0.01 µg/g	23	5306	--	1	0.01	0.0115	115	Q12
QUIZALOFOP ACID	Meal	ARA-09-15-10-144-0001A5 + 0.01 µg/g	24	4935	--	1	0.01	0.0108	108	Q12
QUIZALOFOP ACID	Aspirated grain fractions	ARA-09-15-10-150-0001A17 + 1.0 µg/g	26	486347	--	1	1	0.9843	98	Q12
QUIZALOFOP ACID	Aspirated grain fractions	ARA-09-15-10-150-0001A18 + 1.0 µg/g	27	499603	--	1	1	1.0111	101	Q12
QUIZALOFOP ACID	Aspirated grain fractions	ARA-09-15-10-150-0001A19 + 1.0 µg/g	28	497950	--	1	1	1.0078	101	Q12
QUIZALOFOP ACID	Flour	ARA-09-15-10-155-0001A17 + 1.0 µg/g	29	470333	--	1	1	0.9519	95	Q12
QUIZALOFOP ACID	Flour	ARA-09-15-10-155-0001A18 + 1.0 µg/g	30	498757	--	1	1	1.0094	101	Q12
QUIZALOFOP ACID	Flour	ARA-09-15-10-155-0001A19 + 1.0 µg/g	31	513508	--	1	1	1.0392	104	Q12
QUIZALOFOP ACID	Grits	ARA-09-15-10-145-0001A17 + 1.0 µg/g	32	547723	--	1	1	1.1084	111	Q12
QUIZALOFOP ACID	Grits	ARA-09-15-10-145-0001A18 + 1.0 µg/g	34	541236	--	1	1	1.0953	110	Q12
QUIZALOFOP ACID	Grits	ARA-09-15-10-145-0001A19 + 1.0 µg/g	35	558014	--	1	1	1.1292	113	Q12
QUIZALOFOP ACID	Meal	ARA-09-15-10-144-0001A17 + 1.0 µg/g	36	547716	--	1	1	1.1084	111	Q12
QUIZALOFOP ACID	Meal	ARA-09-15-10-144-0001A18 + 1.0 µg/g	37	548905	--	1	1	1.1108	111	Q12
QUIZALOFOP ACID	Meal	ARA-09-15-10-144-0001A19 + 1.0 µg/g	38	524121	--	1	1	1.0607	106	Q12
QUIZALOFOP ETHYL	Aspirated grain fractions	ARA-09-15-10-150-0001A1	3	0	--	1	NA	ND	NA	Q12
QUIZALOFOP ETHYL	Flour	ARA-09-15-10-155-0001A1	4	0	--	1	NA	ND	NA	Q12
QUIZALOFOP ETHYL	Grits	ARA-09-15-10-145-0001A1	5	0	--	1	NA	ND	NA	Q12

Table 3 (Cont.). Procedural Recovery Raw Data for Corn Samples

Analyte	Matrix	Control Sample ID	Inj. No	Analyte Peak Area	ISTD Peak Area	Dilution Factor	Fortified Conc. (µg/g)	Found (µg/g)[a]	% Rec	Set ID
QUIZALOFOP ETHYL	Meal	ARA-09-15-10-144-0001A1	6	0	--	1	NA	ND	NA	Q12
QUIZALOFOP ETHYL	Aspirated grain fractions	ARA-09-15-10-150-0001A2 + 0.003 µg/g	7	1772	--	1	0.003	0.0032	NA	Q12
QUIZALOFOP ETHYL	Flour	ARA-09-15-10-155-0001A2 + 0.003 µg/g	8	1804	--	1	0.003	0.0032	NA	Q12
QUIZALOFOP ETHYL	Grits	ARA-09-15-10-145-0001A2 + 0.003 µg/g	10	1711	--	1	0.003	0.0030	NA	Q12
QUIZALOFOP ETHYL	Meal	ARA-09-15-10-144-0001A2 + 0.003 µg/g	11	1648	--	1	0.003	0.0029	NA	Q12
QUIZALOFOP ETHYL	Aspirated grain fractions	ARA-09-15-10-150-0001A3 + 0.01 µg/g	12	5130	--	1	0.01	0.0098	98	Q12
QUIZALOFOP ETHYL	Aspirated grain fractions	ARA-09-15-10-150-0001A4 + 0.01 µg/g	13	5517	--	1	0.01	0.0106	106	Q12
QUIZALOFOP ETHYL	Aspirated grain fractions	ARA-09-15-10-150-0001A5 + 0.01 µg/g	14	4822	--	1	0.01	0.0092	92	Q12
QUIZALOFOP ETHYL	Flour	ARA-09-15-10-155-0001A3 + 0.01 µg/g	15	5524	--	1	0.01	0.0106	106	Q12
QUIZALOFOP ETHYL	Flour	ARA-09-15-10-155-0001A4 + 0.01 µg/g	16	5706	--	1	0.01	0.0110	110	Q12
QUIZALOFOP ETHYL	Flour	ARA-09-15-10-155-0001A5 + 0.01 µg/g	18	5675	--	1	0.01	0.0109	109	Q12
QUIZALOFOP ETHYL	Grits	ARA-09-15-10-145-0001A3 + 0.01 µg/g	19	5526	--	1	0.01	0.0106	106	Q12
QUIZALOFOP ETHYL	Grits	ARA-09-15-10-145-0001A4 + 0.01 µg/g	20	5389	--	1	0.01	0.0103	103	Q12
QUIZALOFOP ETHYL	Grits	ARA-09-15-10-145-0001A5 + 0.01 µg/g	21	5540	--	1	0.01	0.0106	106	Q12
QUIZALOFOP ETHYL	Meal	ARA-09-15-10-144-0001A3 + 0.01 µg/g	22	4923	--	1	0.01	0.0094	94	Q12
QUIZALOFOP ETHYL	Meal	ARA-09-15-10-144-0001A4 + 0.01 µg/g	23	5133	--	1	0.01	0.0098	98	Q12
QUIZALOFOP ETHYL	Meal	ARA-09-15-10-144-0001A5 + 0.01 µg/g	24	4935	--	1	0.01	0.0094	94	Q12
QUIZALOFOP ETHYL	Aspirated grain fractions	ARA-09-15-10-150-0001A17 + 1.0 µg/g	26	545848	--	1	1	1.0834	108	Q12
QUIZALOFOP ETHYL	Aspirated grain fractions	ARA-09-15-10-150-0001A18 + 1.0 µg/g	27	548670	--	1	1	1.0890	109	Q12
QUIZALOFOP ETHYL	Aspirated grain fractions	ARA-09-15-10-150-0001A19 + 1.0 µg/g	28	536778	--	1	1	1.0654	107	Q12
QUIZALOFOP ETHYL	Flour	ARA-09-15-10-155-0001A17 + 1.0 µg/g	29	555633	--	1	1	1.1028	110	Q12

Table 3 (Cont.). Procedural Recovery Raw Data for Corn Samples

Analyte	Matrix	Control Sample ID	Inj. No	Analyte Peak Area	ISTD Peak Area	Dilution Factor	Fortified Conc. (µg/g)	Found (µg/g)[a]	% Rec	Set ID
QUIZALOFOP ETHYL	Flour	ARA-09-15-10-155-0001A18 + 1.0 µg/g	30	567635	--	1	1	1.1267	113	Q12
QUIZALOFOP ETHYL	Flour	ARA-09-15-10-155-0001A19 + 1.0 µg/g	31	569085	--	1	1	1.1295	113	Q12
QUIZALOFOP ETHYL	Grits	ARA-09-15-10-145-0001A17 + 1.0 µg/g	32	552848	--	1	1	1.0973	110	Q12
QUIZALOFOP ETHYL	Grits	ARA-09-15-10-145-0001A18 + 1.0 µg/g	34	568857	--	1	1	1.1291	113	Q12
QUIZALOFOP ETHYL	Grits	ARA-09-15-10-145-0001A19 + 1.0 µg/g	35	543443	--	1	1	1.0786	108	Q12
QUIZALOFOP ETHYL	Meal	ARA-09-15-10-144-0001A17 + 1.0 µg/g	36	515479	--	1	1	1.0231	102	Q12
QUIZALOFOP ETHYL	Meal	ARA-09-15-10-144-0001A18 + 1.0 µg/g	37	527023	--	1	1	1.0460	105	Q12
QUIZALOFOP ETHYL	Meal	ARA-09-15-10-144-0001A19 + 1.0 µg/g	38	517416	--	1	1	1.0270	103	Q12
QUIZALOFOP ACID	Aspirated grain fractions	ARA-09-15-10-150-0001A9	3	0	--	1	NA	ND	NA	Q13
QUIZALOFOP ACID	Flour	ARA-09-15-10-155-0001A9	4	0	--	1	NA	ND	NA	Q13
QUIZALOFOP ACID	Grits	ARA-09-15-10-145-0001A9	5	0	--	1	NA	ND	NA	Q13
QUIZALOFOP ACID	Meal	ARA-09-15-10-144-0001A9	6	0	--	1	NA	ND	NA	Q13
QUIZALOFOP ACID	Aspirated grain fractions	ARA-09-15-10-150-0001A10 + 0.003 µg/g	7	1388	--	1	0.003	0.0033	NA	Q13
QUIZALOFOP ACID	Flour	ARA-09-15-10-155-0001A10 + 0.003 µg/g	8	1447	--	1	0.003	0.0034	NA	Q13
QUIZALOFOP ACID	Grits	ARA-09-15-10-145-0001A10 + 0.003 µg/g	10	1601	--	1	0.003	0.0037	NA	Q13
QUIZALOFOP ACID	Meal	ARA-09-15-10-144-0001A10 + 0.003 µg/g	11	1551	--	1	0.003	0.0036	NA	Q13
QUIZALOFOP ACID	Aspirated grain fractions	ARA-09-15-10-150-0001A11 + 0.01 µg/g	12	5205	--	1	0.01	0.0105	105	Q13
QUIZALOFOP ACID	Aspirated grain fractions	ARA-09-15-10-150-0001A12 + 0.01 µg/g	13	5258	--	1	0.01	0.0106	106	Q13
QUIZALOFOP ACID	Aspirated grain fractions	ARA-09-15-10-150-0001A13 + 0.01 µg/g	14	5294	--	1	0.01	0.0107	107	Q13
QUIZALOFOP ACID	Flour	ARA-09-15-10-155-0001A11 + 0.01 µg/g	15	4842	--	1	0.01	0.0099	99	Q13
QUIZALOFOP ACID	Flour	ARA-09-15-10-155-0001A12 + 0.01 µg/g	16	4551	--	1	0.01	0.0093	93	Q13
QUIZALOFOP ACID	Flour	ARA-09-15-10-155-0001A13 + 0.01 µg/g	18	4730	--	1	0.01	0.0096	96	Q13

Table 3 (Cont.). Procedural Recovery Raw Data for Corn Samples

Analyte	Matrix	Control Sample ID	Inj. No	Analyte Peak Area	ISTD Peak Area	Dilution Factor	Fortified Conc. (µg/g)	Found (µg/g)[a]	% Rec	Set ID
QUIZALOFOP ACID	Grits	ARA-09-15-10-145-0001A11 + 0.01 µg/g	19	5013	--	1	0.01	0.0102	102	Q13
QUIZALOFOP ACID	Grits	ARA-09-15-10-145-0001A12 + 0.01 µg/g	20	5403	--	1	0.01	0.0109	109	Q13
QUIZALOFOP ACID	Grits	ARA-09-15-10-145-0001A13 + 0.01 µg/g	21	5497	--	1	0.01	0.0111	111	Q13
QUIZALOFOP ACID	Meal	ARA-09-15-10-144-0001A11 + 0.01 µg/g	22	5503	--	1	0.01	0.0111	111	Q13
QUIZALOFOP ACID	Meal	ARA-09-15-10-144-0001A12 + 0.01 µg/g	23	6027	--	1	0.01	0.0121	121	Q13
QUIZALOFOP ACID	Meal	ARA-09-15-10-144-0001A13 + 0.01 µg/g	24	6089	--	1	0.01	0.0122	122	Q13
QUIZALOFOP ACID	Aspirated grain fractions	ARA-09-15-10-150-0001A14 + 1.0 µg/g	26	539808	--	1	1	1.0254	103	Q13
QUIZALOFOP ACID	Aspirated grain fractions	ARA-09-15-10-150-0001A15 + 1.0 µg/g	27	519314	--	1	1	0.9865	99	Q13
QUIZALOFOP ACID	Aspirated grain fractions	ARA-09-15-10-150-0001A16 + 1.0 µg/g	28	553367	--	1	1	1.0511	105	Q13
QUIZALOFOP ACID	Flour	ARA-09-15-10-155-0001A14 + 1.0 µg/g	29	470101	--	1	1	0.8931	89	Q13
QUIZALOFOP ACID	Flour	ARA-09-15-10-155-0001A15 + 1.0 µg/g	30	493068	--	1	1	0.9367	94	Q13
QUIZALOFOP ACID	Flour	ARA-09-15-10-155-0001A16 + 1.0 µg/g	31	474703	--	1	1	0.9018	90	Q13
QUIZALOFOP ACID	Grits	ARA-09-15-10-145-0001A14 + 1.0 µg/g	32	542883	--	1	1	1.0312	103	Q13
QUIZALOFOP ACID	Grits	ARA-09-15-10-145-0001A15 + 1.0 µg/g	34	537787	--	1	1	1.0216	102	Q13
QUIZALOFOP ACID	Grits	ARA-09-15-10-145-0001A16 + 1.0 µg/g	35	572007	--	1	1	1.0865	109	Q13
QUIZALOFOP ACID	Meal	ARA-09-15-10-144-0001A21 + 1.0 µg/g	36	535257	--	1	1	1.0168	102	Q13
QUIZALOFOP ACID	Meal	ARA-09-15-10-144-0001A15 + 1.0 µg/g	37	543649	--	1	1	1.0327	103	Q13
QUIZALOFOP ACID	Meal	ARA-09-15-10-144-0001A16 + 1.0 µg/g	38	555212	--	1	1	1.0546	105	Q13
QUIZALOFOP ETHYL	Aspirated grain fractions	ARA-09-15-10-150-0001A9	3	0	--	1	NA	ND	NA	Q13
QUIZALOFOP ETHYL	Flour	ARA-09-15-10-155-0001A9	4	0	--	1	NA	ND	NA	Q13
QUIZALOFOP ETHYL	Grits	ARA-09-15-10-145-0001A9	5	0	--	1	NA	ND	NA	Q13
QUIZALOFOP ETHYL	Meal	ARA-09-15-10-144-0001A9	6	0	--	1	NA	ND	NA	Q13

Table 3 (Cont.). Procedural Recovery Raw Data for Corn Samples

Analyte	Matrix	Control Sample ID	Inj. No	Analyte Peak Area	ISTD Peak Area	Dilution Factor	Fortified Conc. (µg/g)	Found (µg/g)[a]	% Rec	Set ID
QUIZALOFOP ETHYL	Aspirated grain fractions	ARA-09-15-10-150-0001A10 + 0.003 µg/g	7	2162	--	1	0.003	0.0031	NA	Q13
QUIZALOFOP ETHYL	Flour	ARA-09-15-10-155-0001A10 + 0.003 µg/g	8	2127	--	1	0.003	0.0030	NA	Q13
QUIZALOFOP ETHYL	Grits	ARA-09-15-10-145-0001A10 + 0.003 µg/g	10	1869	--	1	0.003	0.0025	NA	Q13
QUIZALOFOP ETHYL	Meal	ARA-09-15-10-144-0001A10 + 0.003 µg/g	11	1686	--	1	0.003	0.0021	NA	Q13
QUIZALOFOP ETHYL	Aspirated grain fractions	ARA-09-15-10-150-0001A11 + 0.01 µg/g	12	5281	--	1	0.01	0.0094	94	Q13
QUIZALOFOP ETHYL	Aspirated grain fractions	ARA-09-15-10-150-0001A12 + 0.01 µg/g	13	5694	--	1	0.01	0.0102	102	Q13
QUIZALOFOP ETHYL	Aspirated grain fractions	ARA-09-15-10-150-0001A13 + 0.01 µg/g	14	5671	--	1	0.01	0.0101	101	Q13
QUIZALOFOP ETHYL	Flour	ARA-09-15-10-155-0001A11 + 0.01 µg/g	15	5654	--	1	0.01	0.0101	101	Q13
QUIZALOFOP ETHYL	Flour	ARA-09-15-10-155-0001A12 + 0.01 µg/g	16	5469	--	1	0.01	0.0097	97	Q13
QUIZALOFOP ETHYL	Flour	ARA-09-15-10-155-0001A13 + 0.01 µg/g	18	5303	--	1	0.01	0.0094	94	Q13
QUIZALOFOP ETHYL	Grits	ARA-09-15-10-145-0001A11 + 0.01 µg/g	19	5328	--	1	0.01	0.0095	95	Q13
QUIZALOFOP ETHYL	Grits	ARA-09-15-10-145-0001A12 + 0.01 µg/g	20	5616	--	1	0.01	0.0100	100	Q13
QUIZALOFOP ETHYL	Grits	ARA-09-15-10-145-0001A13 + 0.01 µg/g	21	5403	--	1	0.01	0.0096	96	Q13
QUIZALOFOP ETHYL	Meal	ARA-09-15-10-144-0001A11 + 0.01 µg/g	22	5310	--	1	0.01	0.0094	94	Q13
QUIZALOFOP ETHYL	Meal	ARA-09-15-10-144-0001A12 + 0.01 µg/g	23	5615	--	1	0.01	0.0100	100	Q13
QUIZALOFOP ETHYL	Meal	ARA-09-15-10-144-0001A13 + 0.01 µg/g	24	5841	--	1	0.01	0.0105	105	Q13
QUIZALOFOP ETHYL	Aspirated grain fractions	ARA-09-15-10-150-0001A14 + 1.0 µg/g	26	527958	--	1	1	1.0631	106	Q13
QUIZALOFOP ETHYL	Aspirated grain fractions	ARA-09-15-10-150-0001A15 + 1.0 µg/g	27	543549	--	1	1	1.0945	109	Q13
QUIZALOFOP ETHYL	Aspirated grain fractions	ARA-09-15-10-150-0001A16 + 1.0 µg/g	28	546711	--	1	1	1.1009	110	Q13
QUIZALOFOP ETHYL	Flour	ARA-09-15-10-155-0001A14 + 1.0 µg/g	29	548018	--	1	1	1.1035	110	Q13
QUIZALOFOP ETHYL	Flour	ARA-09-15-10-155-0001A15 + 1.0 µg/g	30	554141	--	1	1	1.1159	112	Q13

Table 3 (Cont.). Procedural Recovery Raw Data for Corn Samples

Analyte	Matrix	Control Sample ID	Inj. No	Analyte Peak Area	ISTD Peak Area	Dilution Factor	Fortified Conc. (µg/g)	Found (µg/g)[a]	% Rec	Set ID
QUIZALOFOP ETHYL	Flour	ARA-09-15-10-155-0001A16 + 1.0 µg/g	31	543655	--	1	1	1.0948	109	Q13
QUIZALOFOP ETHYL	Grits	ARA-09-15-10-145-0001A14 + 1.0 µg/g	32	541478	--	1	1	1.0904	109	Q13
QUIZALOFOP ETHYL	Grits	ARA-09-15-10-145-0001A15 + 1.0 µg/g	34	541164	--	1	1	1.0897	109	Q13
QUIZALOFOP ETHYL	Grits	ARA-09-15-10-145-0001A16 + 1.0 µg/g	35	548341	--	1	1	1.1042	110	Q13
QUIZALOFOP ETHYL	Meal	ARA-09-15-10-144-0001A21 + 1.0 µg/g	36	516191	--	1	1	1.0394	104	Q13
QUIZALOFOP ETHYL	Meal	ARA-09-15-10-144-0001A15 + 1.0 µg/g	37	536567	--	1	1	1.0805	108	Q13
QUIZALOFOP ETHYL	Meal	ARA-09-15-10-144-0001A16 + 1.0 µg/g	38	550303	--	1	1	1.1082	111	Q13
QUIZALOFOP ACID	Oil, refined	ARA-09-15-10-143-0001A1	3	0	--	1	NA	ND	NA	Q14
QUIZALOFOP ACID	Oil, refined	ARA-09-15-10-147-0001A1	4	0	--	1	NA	ND	NA	Q14
QUIZALOFOP ACID	Starch	ARA-09-15-10-142-0001A1	5	0	--	1	NA	ND	NA	Q14
QUIZALOFOP ACID	Oil, refined	ARA-09-15-10-143-0001A2 + 0.003 µg/g	6	1119	--	1	0.003	0.0013	NA	Q14
QUIZALOFOP ACID	Oil, refined	ARA-09-15-10-147-0001A2 + 0.003 µg/g	8	1251	--	1	0.003	0.0016	NA	Q14
QUIZALOFOP ACID	Starch	ARA-09-15-10-142-0001A2 + 0.003 µg/g	9	1633	--	1	0.003	0.0027	NA	Q14
QUIZALOFOP ACID	Oil, refined	ARA-09-15-10-143-0001A3 + 0.01 µg/g	10	5179	--	1	0.01	0.0124	124	Q14
QUIZALOFOP ACID	Oil, refined	ARA-09-15-10-143-0001A4 + 0.01 µg/g	11	5183	--	1	0.01	0.0124	124	Q14
QUIZALOFOP ACID	Oil, refined	ARA-09-15-10-143-0001A5 + 0.01 µg/g	12	5232	--	1	0.01	0.0126	126	Q14
QUIZALOFOP ACID	Oil, refined	ARA-09-15-10-147-0001A3 + 0.01 µg/g	14	4885	--	1	0.01	0.0116	116	Q14
QUIZALOFOP ACID	Oil, refined	ARA-09-15-10-147-0001A4 + 0.01 µg/g	15	5202	--	1	0.01	0.0125	125	Q14
QUIZALOFOP ACID	Oil, refined	ARA-09-15-10-147-0001A5 + 0.01 µg/g	16	5103	--	1	0.01	0.0122	122	Q14
QUIZALOFOP ACID	Starch	ARA-09-15-10-142-0001A3 + 0.01 µg/g	17	4187	--	1	0.01	0.0097	97	Q14
QUIZALOFOP ACID	Starch	ARA-09-15-10-142-0001A4 + 0.01 µg/g	18	4268	--	1	0.01	0.0099	99	Q14
QUIZALOFOP ACID	Starch	ARA-09-15-10-142-0001A5 + 0.01 µg/g	20	4400	--	1	0.01	0.0103	103	Q14
QUIZALOFOP ACID	Oil, refined	ARA-09-15-10-143-0001A6 + 1.0 µg/g	21	464654	--	1	1	1.2746	127	Q14
QUIZALOFOP ACID	Oil, refined	ARA-09-15-10-143-0001A7 + 1.0 µg/g	22	477443	--	1	1	1.3097	131	Q14
QUIZALOFOP ACID	Oil, refined	ARA-09-15-10-143-0001A8 + 1.0 µg/g	23	506502	--	1	1	1.3896	139	Q14

Table 3 (Cont.). Procedural Recovery Raw Data for Corn Samples

Analyte	Matrix	Control Sample ID	Inj. No	Analyte Peak Area	ISTD Peak Area	Dilution Factor	Fortified Conc. (µg/g)	Found (µg/g)[a]	% Rec	Set ID
QUIZALOFOP ACID	Oil, refined	ARA-09-15-10-147-0001A7 + 1.0 µg/g	26	493187	--	1	1	1.353	135	Q14
QUIZALOFOP ACID	Oil, refined	ARA-09-15-10-147-0001A8 + 1.0 µg/g	27	520495	--	1	1	1.428	143	Q14
QUIZALOFOP ACID	Starch	ARA-09-15-10-142-0001A6 + 1.0 µg/g	28	372313	--	1	1	1.0209	102	Q14
QUIZALOFOP ACID	Starch	ARA-09-15-10-142-0001A7 + 1.0 µg/g	29	380194	--	1	1	1.0426	104	Q14
QUIZALOFOP ACID	Starch	ARA-09-15-10-142-0001A8 + 1.0 µg/g	30	360817	--	1	1	0.9894	99	Q14
QUIZALOFOP ETHYL	Oil, refined	ARA-09-15-10-143-0001A1	3	0	--	1	NA	ND	NA	Q14
QUIZALOFOP ETHYL	Oil, refined	ARA-09-15-10-147-0001A1	4	0	--	1	NA	ND	NA	Q14
QUIZALOFOP ETHYL	Starch	ARA-09-15-10-142-0001A1	5	0	--	1	NA	ND	NA	Q14
QUIZALOFOP ETHYL	Oil, refined	ARA-09-15-10-143-0001A2 + 0.003 µg/g	6	891	--	1	0.003	0.0021	NA	Q14
QUIZALOFOP ETHYL	Oil, refined	ARA-09-15-10-147-0001A2 + 0.003 µg/g	8	1144	--	1	0.003	0.0027	NA	Q14
QUIZALOFOP ETHYL	Starch	ARA-09-15-10-142-0001A2 + 0.003 µg/g	9	1848	--	1	0.003	0.0041	NA	Q14
QUIZALOFOP ETHYL	Oil, refined	ARA-09-15-10-143-0001A3 + 0.01 µg/g	10	4325	--	1	0.01	0.0092	92	Q14
QUIZALOFOP ETHYL	Oil, refined	ARA-09-15-10-143-0001A4 + 0.01 µg/g	11	4740	--	1	0.01	0.0100	100	Q14
QUIZALOFOP ETHYL	Oil, refined	ARA-09-15-10-143-0001A5 + 0.01 µg/g	12	4724	--	1	0.01	0.0100	100	Q14
QUIZALOFOP ETHYL	Oil, refined	ARA-09-15-10-147-0001A3 + 0.01 µg/g	14	4916	--	1	0.01	0.0104	104	Q14
QUIZALOFOP ETHYL	Oil, refined	ARA-09-15-10-147-0001A4 + 0.01 µg/g	15	5020	--	1	0.01	0.0106	106	Q14
QUIZALOFOP ETHYL	Oil, refined	ARA-09-15-10-147-0001A5 + 0.01 µg/g	16	5338	--	1	0.01	0.0112	112	Q14
QUIZALOFOP ETHYL	Starch	ARA-09-15-10-142-0001A3 + 0.01 µg/g	17	5507	--	1	0.01	0.0116	116	Q14
QUIZALOFOP ETHYL	Starch	ARA-09-15-10-142-0001A4 + 0.01 µg/g	18	5140	--	1	0.01	0.0108	108	Q14
QUIZALOFOP ETHYL	Starch	ARA-09-15-10-142-0001A5 + 0.01 µg/g	20	5240	--	1	0.01	0.0110	110	Q14
QUIZALOFOP ETHYL	Oil, refined	ARA-09-15-10-143-0001A6 + 1.0 µg/g	21	474718	--	1	1	0.9723	97	Q14
QUIZALOFOP ETHYL	Oil, refined	ARA-09-15-10-143-0001A7 + 1.0 µg/g	22	493965	--	1	1	1.0118	101	Q14
QUIZALOFOP ETHYL	Oil, refined	ARA-09-15-10-143-0001A8 + 1.0 µg/g	23	501755	--	1	1	1.0277	103	Q14
QUIZALOFOP ETHYL	Oil, refined	ARA-09-15-10-147-0001A7 + 1.0 µg/g	26	518347	--	1	1	1.0617	106	Q14
QUIZALOFOP ETHYL	Oil, refined	ARA-09-15-10-147-0001A8 + 1.0 µg/g	27	511343	--	1	1	1.0473	105	Q14

Table 3 (Cont.). Procedural Recovery Raw Data for Corn Samples

Analyte	Matrix	Control Sample ID	Inj. No	Analyte Peak Area	ISTD Peak Area	Dilution Factor	Fortified Conc. (µg/g)	Found (µg/g)[a]	% Rec	Set ID
QUIZALOFOP ETHYL	Starch	ARA-09-15-10-142-0001A6 + 1.0 µg/g	28	526024	--	1	1	1.0774	108	Q14
QUIZALOFOP ETHYL	Starch	ARA-09-15-10-142-0001A7 + 1.0 µg/g	29	497743	--	1	1	1.0195	102	Q14
QUIZALOFOP ETHYL	Starch	ARA-09-15-10-142-0001A8 + 1.0 µg/g	30	525546	--	1	1	1.0764	108	Q14
QUIZALOFOP ACID	Oil, refined	ARA-09-15-10-143-0001A9	3	0	--	1	NA	ND	NA	Q15
QUIZALOFOP ACID	Oil, refined	ARA-09-15-10-147-0001A9	4	0	--	1	NA	ND	NA	Q15
QUIZALOFOP ACID	Starch	ARA-09-15-10-142-0001A9	5	0	--	1	NA	ND	NA	Q15
QUIZALOFOP ACID	Oil, refined	ARA-09-15-10-143-0001A10 + 0.003 µg/g	6	1251	--	1	0.003	0.0025	NA	Q15
QUIZALOFOP ACID	Oil, refined	ARA-09-15-10-147-0001A10 + 0.003 µg/g	7	1284	--	1	0.003	0.0026	NA	Q15
QUIZALOFOP ACID	Starch	ARA-09-15-10-142-0001A10 + 0.003 µg/g	9	1542	--	1	0.003	0.0035	NA	Q15
QUIZALOFOP ACID	Oil, refined	ARA-09-15-10-143-0001A11 + 0.01 µg/g	10	3399	--	1	0.01	0.0095	95	Q15
QUIZALOFOP ACID	Oil, refined	ARA-09-15-10-143-0001A12 + 0.01 µg/g	11	3806	--	1	0.01	0.0108	108	Q15
QUIZALOFOP ACID	Oil, refined	ARA-09-15-10-143-0001A13 + 0.01 µg/g	12	3489	--	1	0.01	0.0098	98	Q15
QUIZALOFOP ACID	Oil, refined	ARA-09-15-10-147-0001A11 + 0.01 µg/g	13	3530	--	1	0.01	0.0099	99	Q15
QUIZALOFOP ACID	Oil, refined	ARA-09-15-10-147-0001A12 + 0.01 µg/g	14	3908	--	1	0.01	0.0112	112	Q15
QUIZALOFOP ACID	Oil, refined	ARA-09-15-10-147-0001A13 + 0.01 µg/g	16	3433	--	1	0.01	0.0096	96	Q15
QUIZALOFOP ACID	Starch	ARA-09-15-10-142-0001A11 + 0.01 µg/g	17	2773	--	1	0.01	0.0075	75	Q15
QUIZALOFOP ACID	Starch	ARA-09-15-10-142-0001A12 + 0.01 µg/g	18	2846	--	1	0.01	0.0077	77	Q15
QUIZALOFOP ACID	Starch	ARA-09-15-10-142-0001A13 + 0.01 µg/g	19	2937	--	1	0.01	0.0080	80	Q15
QUIZALOFOP ACID	Oil, refined	ARA-09-15-10-143-0001A14 + 1.00 µg/g	20	333192	--	1	1	1.0850	108	Q15
QUIZALOFOP ACID	Oil, refined	ARA-09-15-10-143-0001A15 + 1.00 µg/g	21	338294	--	1	1	1.1016	110	Q15
QUIZALOFOP ACID	Oil, refined	ARA-09-15-10-143-0001A16 + 1.00 µg/g	23	335140	--	1	1	1.0913	109	Q15
QUIZALOFOP ACID	Oil, refined	ARA-09-15-10-147-0001A14 + 1.00 µg/g	24	328673	--	1	1	1.0702	107	Q15
QUIZALOFOP ACID	Oil, refined	ARA-09-15-10-147-0001A15 + 1.00 µg/g	25	346832	--	1	1	1.1295	113	Q15
QUIZALOFOP ACID	Oil, refined	ARA-09-15-10-147-0001A16 + 1.00 µg/g	26	332059	--	1	1	1.0813	108	Q15
QUIZALOFOP ACID	Starch	ARA-09-15-10-142-0001A14 + 1.00 µg/g	27	318791	--	1	1	1.0380	104	Q15

Table 3 (Cont.). Procedural Recovery Raw Data for Corn Samples

Analyte	Matrix	Control Sample ID	Inj. No	Analyte Peak Area	ISTD Peak Area	Dilution Factor	Fortified Conc. (µg/g)	Found (µg/g)[a]	% Rec	Set ID
QUIZALOFOP ACID	Starch	ARA-09-15-10-142-0001A15 + 1.00 µg/g	28	335974	--	1	1	1.0941	109	Q15
QUIZALOFOP ACID	Starch	ARA-09-15-10-142-0001A16 + 1.00 µg/g	30	341980	--	1	1	1.1136	111	Q15
QUIZALOFOP ETHYL	Oil, refined	ARA-09-15-10-143-0001A9	3	0	--	1	NA	ND	NA	Q15
QUIZALOFOP ETHYL	Oil, refined	ARA-09-15-10-147-0001A9	4	0	--	1	NA	ND	NA	Q15
QUIZALOFOP ETHYL	Starch	ARA-09-15-10-142-0001A9	5	0	--	1	NA	ND	NA	Q15
QUIZALOFOP ETHYL	Oil, refined	ARA-09-15-10-143-0001A10 + 0.003 µg/g	6	1535	--	1	0.003	0.0022	NA	Q15
QUIZALOFOP ETHYL	Oil, refined	ARA-09-15-10-147-0001A10 + 0.003 µg/g	7	1442	--	1	0.003	0.0020	NA	Q15
QUIZALOFOP ETHYL	Starch	ARA-09-15-10-142-0001A10 + 0.003 µg/g	9	2051	--	1	0.003	0.0032	NA	Q15
QUIZALOFOP ETHYL	Oil, refined	ARA-09-15-10-143-0001A11 + 0.01 µg/g	10	4044	--	1	0.01	0.0074	74	Q15
QUIZALOFOP ETHYL	Oil, refined	ARA-09-15-10-143-0001A12 + 0.01 µg/g	11	4160	--	1	0.01	0.0076	76	Q15
QUIZALOFOP ETHYL	Oil, refined	ARA-09-15-10-143-0001A13 + 0.01 µg/g	12	3975	--	1	0.01	0.0072	72	Q15
QUIZALOFOP ETHYL	Oil, refined	ARA-09-15-10-147-0001A11 + 0.01 µg/g	13	4275	--	1	0.01	0.0078	78	Q15
QUIZALOFOP ETHYL	Oil, refined	ARA-09-15-10-147-0001A12 + 0.01 µg/g	14	3980	--	1	0.01	0.0072	72	Q15
QUIZALOFOP ETHYL	Oil, refined	ARA-09-15-10-147-0001A13 + 0.01 µg/g	16	4165	--	1	0.01	0.0076	76	Q15
QUIZALOFOP ETHYL	Starch	ARA-09-15-10-142-0001A11 + 0.01 µg/g	17	5012	--	1	0.01	0.0093	93	Q15
QUIZALOFOP ETHYL	Starch	ARA-09-15-10-142-0001A12 + 0.01 µg/g	18	5190	--	1	0.01	0.0097	97	Q15
QUIZALOFOP ETHYL	Starch	ARA-09-15-10-142-0001A13 + 0.01 µg/g	19	5229	--	1	0.01	0.0098	98	Q15
QUIZALOFOP ETHYL	Oil, refined	ARA-09-15-10-143-0001A14 + 1.00 µg/g	20	408662	--	1	1	0.8419	84	Q15
QUIZALOFOP ETHYL	Oil, refined	ARA-09-15-10-143-0001A15 + 1.00 µg/g	21	415619	--	1	1	0.8563	86	Q15
QUIZALOFOP ETHYL	Oil, refined	ARA-09-15-10-143-0001A16 + 1.00 µg/g	23	409640	--	1	1	0.8440	84	Q15
QUIZALOFOP ETHYL	Oil, refined	ARA-09-15-10-147-0001A14 + 1.00 µg/g	24	400536	--	1	1	0.8252	83	Q15
QUIZALOFOP ETHYL	Oil, refined	ARA-09-15-10-147-0001A15 + 1.00 µg/g	25	415309	--	1	1	0.8557	86	Q15

Table 3 (Cont.). Procedural Recovery Raw Data for Corn Samples

Analyte	Matrix	Control Sample ID	Inj. No	Analyte Peak Area	ISTD Peak Area	Dilution Factor	Fortified Conc. (µg/g)	Found (µg/g)[a]	% Rec	Set ID
QUIZALOFOP ETHYL	Oil, refined	ARA-09-15-10-147-0001A16 + 1.00 µg/g	26	401286	--	1	1	0.8267	83	Q15
QUIZALOFOP ETHYL	Starch	ARA-09-15-10-142-0001A14 + 1.00 µg/g	27	524708	--	1	1	1.0813	108	Q15
QUIZALOFOP ETHYL	Starch	ARA-09-15-10-142-0001A15 + 1.00 µg/g	28	517375	--	1	1	1.0662	107	Q15
QUIZALOFOP ETHYL	Starch	ARA-09-15-10-142-0001A16 + 1.00 µg/g	30	551046	--	1	1	1.1356	114	Q15

[a] All calculations were performed using Microsoft Excel 2003 with full precision.
[b] NA = Not Applicable
[c] ND = Not Detected

Table 4. Summary of Concurrent Recoveries from Fortified Corn Samples

Matrix	Fort. Level (µg/g)	Number of Samples	Recoveries, %	Mean Recovery, %	Std. Dev.
2,4-D					
Aspirated grain fractions	0.01	2	81, 82	82	1
Aspirated grain fractions	0.1	2	80, 83	82	2
Flour	0.01	2	88, 89	89	0
Flour	0.1	2	87, 88	87	1
Forage	0.01	10	95, 95, 96, 97, 98, 98, 102, 103, 103, 104	99	4
Forage	0.1	4	95, 97, 98, 98	97	1
Forage	2	2	95, 98	97	2
Forage	4	2	102, 109	106	5
Forage	10	6	87, 90, 94, 94, 95, 96	93	3
Grain	0.01	6	85, 86, 89, 91, 93, 94	90	4
Grain	0.1	6	88, 90, 90, 92, 95, 95	92	3
Grits	0.01	2	85, 91	88	4
Grits	0.1	2	88, 89	89	1
Meal	0.01	2	88, 96	92	5
Meal	0.1	2	88, 90	89	2
Oil, refined	0.01	4	77, 89, 94, 98	89	9
Oil, refined	0.1	4	94, 97, 97, 98	96	2
Starch	0.01	2	83, 87	85	3
Starch	0.1	2	82, 87	85	4
Stover	0.01	6	92, 95, 97, 99, 100, 100	97	3
Stover	0.1	2	94, 98	96	2
Stover	1	2	95, 98	96	2
Stover	20	6	81, 90, 93, 94, 95, 95	91	5
2,4-DCP					
Aspirated grain fractions	0.01	2	90, 91	91	1
Aspirated grain fractions	0.1	2	89, 97	93	6
Flour	0.01	2	102, 108	105	4
Flour	0.1	2	94, 96	95	2
Forage	0.01	10	82, 84, 97, 98, 99, 101, 102, 104, 109, 111	99	9
Forage	0.1	4	89, 96, 100, 106	98	7
Forage	2	2	90, 92	91	1
Forage	4	2	94, 100	97	4
Forage	10	6	82, 86, 91, 91, 93, 95	90	5
Grain	0.01	6	98, 99, 99, 100, 106, 108	102	4
Grain	0.1	6	94, 96, 97, 98, 98, 100	97	2
Grits	0.01	2	98, 109	103	8
Grits	0.1	2	97, 97	97	0
Meal	0.01	2	92, 98	95	5
Meal	0.1	2	98, 98	98	0
Oil, refined	0.01	4	79, 80, 92, 102	88	11
Oil, refined	0.1	4	80, 86, 86, 88	85	4
Starch	0.01	2	73, 74	74	1
Starch	0.1	2	61, 69	65	6
Stover	0.01	6	99, 102, 103, 105, 111, 112	105	5
Stover	0.1	2	100, 106	103	4
Stover	1	2	104, 104	104	0
Stover	20	6	76, 80, 80, 86, 87, 89	83	5

Table 4 (Cont.). Summary of Concurrent Recoveries from Fortified Corn Samples

Matrix	Fort. Level (µg/g)	Number of Samples	Recoveries, %	Mean Recovery, %	Std. Dev.
QUIZALOFOP ACID					
Aspirated grain fractions	0.01	6	105, 106, 107, 107, 108, 111	107	2
Aspirated grain fractions	1	6	98, 99, 101, 101, 103, 105	101	3
Flour	0.01	6	93, 96, 99, 102, 107, 109	101	6
Flour	1	6	89, 90, 94, 95, 101, 104	96	6
Forage	0.01	10	90, 91, 93, 94, 102, 102, 105, 107, 114, 115	101	9
Forage	1	10	95, 99, 100, 106, 108, 109, 110, 114, 115, 123	108	8
Grain	0.01	6	100, 102, 104, 105, 115, 116	107	7
Grain	1	6	97, 99, 99, 101, 104, 105	101	3
Grits	0.01	6	102, 109, 111, 113, 113, 113	110	4
Grits	1	6	102, 103, 109, 110, 111, 113	108	4
Meal	0.01	6	108, 111, 115, 115, 121, 122	115	6
Meal	1	6	102, 103, 105, 106, 111, 111	106	4
Oil, refined	0.01	12	95, 96, 98, 99, 108, 112, 116, 122, 124, 124, 125, 126	112	12
Oil, refined	1	11	107, 108, 108, 109, 110, 113, 127, 131, 135, 139, 143	121	14
Starch	0.01	6	75, 77, 80, 97, 99, 103	88	12
Starch	1	6	99, 102, 104, 104, 109, 111	105	5
Stover	0.01	6	107, 109, 109, 110, 110, 118	111	4
Stover	1	6	117, 117, 126, 129, 138, 138	127	9
QUIZALOFOP ETHYL					
Aspirated grain fractions	0.01	6	92, 94, 98, 101, 102, 106	99	5
Aspirated grain fractions	1	6	106, 107, 108, 109, 109, 110	108	2
Flour	0.01	6	94, 97, 101, 106, 109, 110	103	6
Flour	1	6	109, 110, 110, 112, 113, 113	111	1
Forage	0.01	10	75, 80, 80, 84, 85, 87, 87, 97, 100, 102	88	9
Forage	1	10	78, 81, 97, 98, 101, 102, 103, 104, 105, 108	98	10
Grain	0.01	6	98, 103, 103, 104, 107, 108	104	4
Grain	1	6	97, 104, 107, 107, 110, 112	106	5
Grits	0.01	6	95, 96, 100, 103, 106, 106	101	5
Grits	1	6	108, 109, 109, 110, 110, 113	110	2
Meal	0.01	6	94, 94, 94, 98, 100, 105	98	4
Meal	1	6	102, 103, 104, 105, 108, 111	105	3
Oil, refined	0.01	12	72, 72, 74, 76, 76, 78, 92, 100, 100, 104, 106, 112	88	15
Oil, refined	1	11	83, 83, 84, 84, 86, 86, 97, 101, 103, 105, 106	92	10
Starch	0.01	6	93, 97, 98, 108, 110, 116	104	9
Starch	1	6	102, 107, 108, 108, 108, 114	108	4
Stover	0.01	6	81, 87, 89, 91, 98, 104	92	8
Stover	1	6	100, 105, 107, 109, 109, 109	107	3

Table 5. Analytical Set Parameters for the Analysis of Corn Samples

Set ID	Analysis Date[a]	Analyte	Average Set Recovery, (%)	Std. Dev., %	% Rel. Std. Dev.	Slope	Intercept	r Squared	LOD (µg/g)	LOQ (µg/g)
D16	10-Aug-2010	2,4-D	91	3	3	0.1079	0.0068	0.9995	0.003	0.010
D16	10-Aug-2010	2,4-DCP	98	2	2	0.7101	0.0450	0.9994	0.003	0.010
D17	11-Aug-2010	2,4-D	92	4	5	0.1095	0.0062	0.9998	0.003	0.010
D17	11-Aug-2010	2,4-DCP	100	4	4	0.7064	0.0456	0.9995	0.003	0.010
D18	12-Aug-2010	2,4-D	89	3	3	0.1084	0.0051	0.9996	0.003	0.010
D18	12-Aug-2010	2,4-DCP	100	6	6	0.7057	0.0582	0.9993	0.003	0.010
D19	16-Aug-2010	2,4-D	87	4	4	0.1098	0.0074	0.9998	0.003	0.010
D19	16-Aug-2010	2,4-DCP	92	13	14	0.7049	0.0412	0.9991	0.003	0.010
D20	13-Aug-2010	2,4-D	93	7	8	0.1093	0.0078	0.9998	0.003	0.010
D20	13-Aug-2010	2,4-DCP	87	8	9	0.7006	0.0315	0.9990	0.003	0.010
D21	19-Aug-2010	2,4-D	98	3	3	0.1089	0.0118	0.9994	0.003	0.010
D21	19-Aug-2010	2,4-DCP	89	8	9	0.6931	0.1189	0.9991	0.003	0.010
D22	20-Aug-2010	2,4-D	97	1	1	0.1104	0.0057	0.9994	0.003	0.010
D22	20-Aug-2010	2,4-DCP	103	6	6	0.7088	0.0543	1.0000	0.003	0.010
D23	23-Aug-2010	2,4-D	96	2	2	0.1103	0.0004	0.9998	0.003	0.010
D23	23-Aug-2010	2,4-DCP	97	9	9	0.6929	0.0587	0.9984	0.003	0.010
D24	24-Aug-2010	2,4-D	104	3	3	0.1108	0.0084	0.9993	0.003	0.010
D24	24-Aug-2010	2,4-DCP	98	3	3	0.7145	0.0540	0.9995	0.003	0.010
D25	24-Aug-2010	2,4-D	95	5	5	0.1129	0.0042	0.9963	0.003	0.010
D25	24-Aug-2010	2,4-DCP	93	7	8	0.7268	0.0447	0.9987	0.003	0.010
D26	26-Aug-2010	2,4-D	95	2	2	0.1085	0.0077	0.9993	0.003	0.010
D26	26-Aug-2010	2,4-DCP	106	5	5	0.7253	0.0468	0.9998	0.003	0.010
D27	27-Aug-2010	2,4-D	97	2	2	0.1082	0.0072	0.9991	0.003	0.010
D27	27-Aug-2010	2,4-DCP	102	2	2	0.7034	0.0475	0.9987	0.003	0.010
D28	30-Aug-2010	2,4-D	94	6	7	0.1103	0.0056	0.9999	0.003	0.010
D28	30-Aug-2010	2,4-DCP	89	12	13	0.7005	0.0584	0.9990	0.003	0.010
Q01	18-May-2010	QUIZALOFOP ACID	102	4	4	20976.9768	-349.7667	0.9991	0.003	0.010
Q01	18-May-2010	QUIZALOFOP ETHYL	108	3	2	15208.8980	-122.0972	0.9992	0.003	0.010
Q02	19-May-2010	QUIZALOFOP ACID	100	1	1	20558.0513	27.6469	0.9990	0.003	0.010
Q02	19-May-2010	QUIZALOFOP ETHYL	104	5	5	15157.9224	264.4343	0.9997	0.003	0.010
Q03	20-May-2010	QUIZALOFOP ACID	110	7	6	16272.3212	-310.6477	0.9980	0.003	0.010
Q03	20-May-2010	QUIZALOFOP ETHYL	102	3	3	14764.8572	-287.4279	0.9999	0.003	0.010
Q04	24-May-2010	QUIZALOFOP ACID	105	4	4	12825.3667	345.8435	0.9988	0.003	0.010
Q04	24-May-2010	QUIZALOFOP ETHYL	80	1	2	23166.1290	-6.5948	0.9998	0.003	0.010
Q05	25-May-2010	QUIZALOFOP ACID	116	4	4	11211.9488	-127.7482	0.9982	0.003	0.010
Q05	25-May-2010	QUIZALOFOP ETHYL	99	2	2	19999.9734	-364.2213	0.9998	0.003	0.010

Table 5 (Cont.). Analytical Set Parameters for the Analysis of Corn Samples

Set ID	Analysis Date	Analyte	Average Set Recovery, (%)	Std. Dev., %	% Rel. Std. Dev.	Slope	Intercept	r Squared	LOD (µg/g)	LOQ (µg/g)
Q06	25-May-2010	QUIZALOFOP ACID	100	7	7	12049.7116	-46.9374	0.9994	0.003	0.010
Q06	25-May-2010	QUIZALOFOP ETHYL	98	10	11	20491.9674	408.2194	0.9995	0.003	0.010
Q07	26-May-2010	QUIZALOFOP ACID	101	12	12	11094.0317	551.6570	0.9979	0.003	0.010
Q07	26-May-2010	QUIZALOFOP ETHYL	95	11	11	19334.1134	1022.0416	0.9995	0.003	0.010
Q08	26-May-2010	QUIZALOFOP ACID	100	6	6	11951.2094	-74.5330	0.9972	0.003	0.010
Q08	26-May-2010	QUIZALOFOP ETHYL	91	13	14	20438.1049	871.2463	0.9997	0.003	0.010
Q09	27-May-2010	QUIZALOFOP ACID	123	17	14	11228.3055	434.6416	0.9998	0.003	0.010
Q09	27-May-2010	QUIZALOFOP ETHYL	100	11	11	20230.8802	1075.0856	0.9999	0.003	0.010
Q10	27-May-2010	QUIZALOFOP ACID	119	11	9	11475.2919	219.1655	0.9995	0.003	0.010
Q10	27-May-2010	QUIZALOFOP ETHYL	93	12	12	20102.9230	778.1610	0.9996	0.003	0.010
Q11	28-May-2010	QUIZALOFOP ACID	116	4	3	10940.0423	244.4839	0.9998	0.003	0.010
Q11	28-May-2010	QUIZALOFOP ETHYL	105	5	5	22636.5714	456.0076	0.9996	0.003	0.010
Q12	8-Jun-2010	QUIZALOFOP ACID	108	5	5	19780.4721	-399.7863	0.9962	0.003	0.010
Q12	8-Jun-2010	QUIZALOFOP ETHYL	105	6	6	20146.3379	183.4868	0.9999	0.003	0.010
Q13	9-Jun-2010	QUIZALOFOP ACID	104	8	8	21071.3347	-350.4326	0.9971	0.003	0.010
Q13	9-Jun-2010	QUIZALOFOP ETHYL	104	6	6	19840.7519	637.2191	0.9996	0.003	0.010
Q14	9-Jun-2010	QUIZALOFOP ACID	119	15	13	14561.3095	660.0110	0.9995	0.003	0.010
Q14	9-Jun-2010	QUIZALOFOP ETHYL	105	6	6	19535.1112	-151.7177	0.9998	0.003	0.010
Q15	10-Jun-2010	QUIZALOFOP ACID	101	12	12	12266.0657	481.4907	0.9992	0.003	0.010
Q15	10-Jun-2010	QUIZALOFOP ETHYL	87	13	15	19392.3552	479.2116	0.9996	0.003	0.010

[a] Date when analysis was initiated by the addition of extraction solution.

Table 6. Individual Residue Results from Corn Samples

Analyte	Sample ID[a]	Inj. No	Analyte Peak Area	ISTD Peak Area	Dilution Factor	Uncorrected (µg/g)	Reported (µg/g)[b]	Set ID
2,4-D	011-0001	3	0	775839	1	ND	ND[c]	D16
2,4-D	003-0001	4	0	765921	1	ND	ND	D16
2,4-D	007-0001	5	0	747516	1	ND	ND	D16
2,4-D	020-0001	6	0	741989	1	ND	ND	D16
2,4-D	022-0001	7	0	703412	1	ND	ND	D16
2,4-D	024-0001	8	0	698561	1	ND	ND	D16
2,4-D	026-0001	9	0	675646	1	ND	ND	D16
2,4-D	028-0001	11	0	651318	1	ND	ND	D16
2,4-D	032-0001	12	0	651021	1	ND	ND	D16
2,4-D	036-0001	13	0	639314	1	ND	ND	D16
2,4-D	040-0001	14	0	616849	1	ND	ND	D16
2,4-D	044-0001	15	0	591572	1	ND	ND	D16
2,4-D	048-0001	16	4624	574600	1	0.0002	ND	D16
2,4-D	052-0001	17	2140	572527	1	ND	ND	D16
2,4-D	060-0001	18	0	565757	1	ND	ND	D16
2,4-D	062-0001	20	5562	564575	1	0.0006	ND	D16
2,4-D	064-0001	21	0	548261	1	ND	ND	D16
2,4-D	066-0001	22	0	538155	1	ND	ND	D16
2,4-D	068-0001	23	0	530613	1	ND	ND	D16
2,4-D	072-0001	24	0	508514	1	ND	ND	D16
2,4-D	076-0001	25	2897	511865	1	ND	ND	D16
2,4-D	080-0001	26	0	483616	1	ND	ND	D16
2,4-D	085-0001	27	0	489786	1	ND	ND	D16
2,4-D	089-0001	29	0	485663	1	ND	ND	D16
2,4-D	093-0001	30	0	481061	1	ND	ND	D16
2,4-D	097-0001	31	0	457391	1	ND	ND	D16
2,4-D	101-0001	32	0	460374	1	ND	ND	D16
2,4-D	105-0001	33	0	452531	1	ND	ND	D16
2,4-D	109-0001	34	0	437909	1	ND	ND	D16
2,4-D	117-0001	35	0	427615	1	ND	ND	D16
2,4-D	119-0001	36	0	416811	1	ND	ND	D16
2,4-D	121-0001	38	0	427188	1	ND	ND	D16
2,4-D	123-0001	39	0	418876	1	ND	ND	D16
2,4-D	125-0001	40	0	411742	1	ND	ND	D16
2,4-D	129-0001	41	0	392491	1	ND	ND	D16
2,4-D	133-0001	42	0	389890	1	ND	ND	D16
2,4-D	137-0001	43	0	392961	1	ND	ND	D16
2,4-D	139-0001	44	0	370612	1	ND	ND	D16
2,4-D	139-0002	45	0	380088	1	ND	ND	D16
2,4-D	139-0003	47	0	364246	1	ND	ND	D16
2,4-D	140-0001	48	0	370980	1	ND	ND	D16
2,4-D	140-0002	49	0	369108	1	ND	ND	D16
2,4-D	140-0003	50	0	359680	1	ND	ND	D16
2,4-D	148-0001	51	0	359388	1	ND	ND	D16
2,4-D	148-0002	52	0	341475	1	ND	ND	D16
2,4-D	148-0003	53	0	352870	1	ND	ND	D16
2,4-D	149-0001	54	0	336608	1	ND	ND	D16
2,4-D	032-0002	56	0	366642	1	ND	ND	D16
2,4-DCP	011-0001	3	0	28149	1	ND	ND	D16
2,4-DCP	003-0001	4	2749	28849	1	0.0014	ND	D16

Table 6 (Cont.). Individual Residue Results from Corn Samples

Analyte	Sample ID[a]	Inj. No	Analyte Peak Area	ISTD Peak Area	Dilution Factor	Uncorrected (µg/g)	Reported (µg/g)[b]	Set ID
2,4-DCP	007-0001	5	2986	29806	1	0.0016	ND	D16
2,4-DCP	020-0001	6	1470	25355	1	0.0004	ND	D16
2,4-DCP	022-0001	7	2102	29408	1	0.0007	ND	D16
2,4-DCP	024-0001	8	1439	26258	1	0.0003	ND	D16
2,4-DCP	026-0001	9	2364	29519	1	0.0010	ND	D16
2,4-DCP	028-0001	11	1843	23789	1	0.0009	ND	D16
2,4-DCP	032-0001	12	2185	26841	1	0.0010	ND	D16
2,4-DCP	036-0001	13	1121	26727	1	ND	ND	D16
2,4-DCP	040-0001	14	0	27789	1	ND	ND	D16
2,4-DCP	044-0001	15	1778	26467	1	0.0006	ND	D16
2,4-DCP	048-0001	16	3192	28140	1	0.0019	ND	D16
2,4-DCP	052-0001	17	7223	25546	1	0.0067	[0.0067][d]	D16
2,4-DCP	060-0001	18	3474	25133	1	0.0026	ND	D16
2,4-DCP	062-0001	20	2421	29240	1	0.0011	ND	D16
2,4-DCP	064-0001	21	1736	25234	1	0.0007	ND	D16
2,4-DCP	066-0001	22	2415	24197	1	0.0015	ND	D16
2,4-DCP	068-0001	23	3729	24961	1	0.0029	ND	D16
2,4-DCP	072-0001	24	1162	22893	1	0.0002	ND	D16
2,4-DCP	076-0001	25	0	24552	1	ND	ND	D16
2,4-DCP	080-0001	26	2852	24809	1	0.0020	ND	D16
2,4-DCP	085-0001	27	0	23289	1	ND	ND	D16
2,4-DCP	089-0001	29	3837	23853	1	0.0033	[0.0033]	D16
2,4-DCP	093-0001	30	1333	23352	1	0.0003	ND	D16
2,4-DCP	097-0001	31	3163	26937	1	0.0020	ND	D16
2,4-DCP	101-0001	32	1831	24512	1	0.0008	ND	D16
2,4-DCP	105-0001	33	2124	25164	1	0.0011	ND	D16
2,4-DCP	109-0001	34	1478	22962	1	0.0005	ND	D16
2,4-DCP	117-0001	35	1171	25859	1	ND	ND	D16
2,4-DCP	119-0001	36	2085	23792	1	0.0012	ND	D16
2,4-DCP	121-0001	38	1876	22717	1	0.0011	ND	D16
2,4-DCP	123-0001	39	2304	22826	1	0.0016	ND	D16
2,4-DCP	125-0001	40	1504	21497	1	0.0007	ND	D16
2,4-DCP	129-0001	41	1468	24188	1	0.0004	ND	D16
2,4-DCP	133-0001	42	1607	22701	1	0.0007	ND	D16
2,4-DCP	137-0001	43	2037	26605	1	0.0009	ND	D16
2,4-DCP	139-0001	44	1498	22370	1	0.0006	ND	D16
2,4-DCP	139-0002	45	2118	22450	1	0.0014	ND	D16
2,4-DCP	139-0003	47	2103	19956	1	0.0017	ND	D16
2,4-DCP	140-0001	48	1681	24739	1	0.0006	ND	D16
2,4-DCP	140-0002	49	1578	25231	1	0.0005	ND	D16
2,4-DCP	140-0003	50	1340	24629	1	0.0003	ND	D16
2,4-DCP	148-0001	51	2898	21271	1	0.0026	ND	D16
2,4-DCP	148-0002	52	2370	22931	1	0.0016	ND	D16
2,4-DCP	148-0003	53	1146	20422	1	0.0003	ND	D16
2,4-DCP	149-0001	54	1403	25995	1	0.0003	ND	D16
2,4-DCP	032-0002	56	2047	21631	1	0.0014	ND	D16
2,4-D	011-0001	3	0	627634	1	ND	ND	D17
2,4-D	149-0002	4	0	598876	1	ND	ND	D17
2,4-D	149-0003	5	0	589610	1	ND	ND	D17
2,4-D	003-0002	6	0	585812	1	ND	ND	D17
2,4-D	003-0003	7	0	594237	1	ND	ND	D17

Table 6 (Cont.). Individual Residue Results from Corn Samples

Analyte	Sample ID[a]	Inj. No	Analyte Peak Area	ISTD Peak Area	Dilution Factor	Uncorrected (µg/g)	Reported (µg/g)[b]	Set ID
2,4-D	007-0002	8	0	564746	1	ND	ND	D17
2,4-D	007-0003	9	0	535256	1	ND	ND	D17
2,4-D	011-0002	10	0	530017	1	ND	ND	D17
2,4-D	011-0003	12	0	520064	1	ND	ND	D17
2,4-D	020-0002	13	0	519815	1	ND	ND	D17
2,4-D	020-0003	14	0	506196	1	ND	ND	D17
2,4-D	022-0002	15	0	479250	1	ND	ND	D17
2,4-D	022-0003	16	0	483446	1	ND	ND	D17
2,4-D	024-0002	17	0	493226	1	ND	ND	D17
2,4-D	024-0003	18	0	462631	1	ND	ND	D17
2,4-D	026-0002	19	0	461950	1	ND	ND	D17
2,4-D	026-0003	20	9037	465169	1	0.0024	ND	D17
2,4-D	028-0002	22	0	464084	1	ND	ND	D17
2,4-D	028-0003	23	0	449214	1	ND	ND	D17
2,4-D	032-0002	24	0	438382	1	ND	ND	D17
2,4-D	032-0003	25	0	432903	1	ND	ND	D17
2,4-D	036-0002	26	0	419500	1	ND	ND	D17
2,4-D	036-0003	27	0	412647	1	ND	ND	D17
2,4-D	040-0002	28	0	411610	1	ND	ND	D17
2,4-D	040-0003	29	0	395759	1	ND	ND	D17
2,4-D	044-0002	30	150596	403112	1	0.0671	0.0671	D17
2,4-D	044-0003	32	0	392774	1	ND	ND	D17
2,4-D	048-0002	33	0	392335	1	ND	ND	D17
2,4-D	048-0003	34	0	375978	1	ND	ND	D17
2,4-D	052-0002	35	0	365790	1	ND	ND	D17
2,4-D	052-0003	36	0	361672	1	ND	ND	D17
2,4-D	060-0002	37	0	368352	1	ND	ND	D17
2,4-D	060-0003	38	0	363983	1	ND	ND	D17
2,4-D	062-0002	39	3429	358986	1	0.0006	ND	D17
2,4-D	062-0003	40	0	366823	1	ND	ND	D17
2,4-D	064-0002	42	0	354285	1	ND	ND	D17
2,4-D	064-0003	43	0	346586	1	ND	ND	D17
2,4-D	066-0002	44	0	349082	1	ND	ND	D17
2,4-D	066-0003	45	0	339725	1	ND	ND	D17
2,4-D	068-0002	46	0	327742	1	ND	ND	D17
2,4-D	068-0003	47	0	338277	1	ND	ND	D17
2,4-D	072-0002	48	9110	341543	1	0.0037	[0.0037]	D17
2,4-D	072-0003	49	6722	324130	1	0.0026	ND	D17
2,4-D	076-0002	50	0	326585	1	ND	ND	D17
2,4-D	076-0003	52	0	333098	1	ND	ND	D17
2,4-D	080-0002	53	0	322728	1	ND	ND	D17
2,4-D	080-0003	54	0	313167	1	ND	ND	D17
2,4-D	085-0002	55	0	323863	1	ND	ND	D17
2,4-D	085-0003	56	0	325691	1	ND	ND	D17
2,4-D	089-0002	57	0	321522	1	ND	ND	D17
2,4-DCP	011-0001	3	0	26445	1	ND	ND	D17
2,4-DCP	149-0002	4	1880	22114	1	0.0011	ND	D17
2,4-DCP	149-0003	5	0	25040	1	ND	ND	D17
2,4-DCP	003-0002	6	1536	23083	1	0.0006	ND	D17
2,4-DCP	003-0003	7	0	21052	1	ND	ND	D17

Table 6 (Cont.). Individual Residue Results from Corn Samples

Analyte	Sample ID[a]	Inj. No	Analyte Peak Area	ISTD Peak Area	Dilution Factor	Uncorrected (µg/g)	Reported (µg/g)[b]	Set ID
2,4-DCP	007-0002	8	1511	25529	1	0.0004	ND	D17
2,4-DCP	007-0003	9	0	22242	1	ND	ND	D17
2,4-DCP	011-0002	10	1383	21386	1	0.0005	ND	D17
2,4-DCP	011-0003	12	1582	22783	1	0.0007	ND	D17
2,4-DCP	020-0002	13	1963	21207	1	0.0013	ND	D17
2,4-DCP	020-0003	14	2503	21450	1	0.0020	ND	D17
2,4-DCP	022-0002	15	1267	20335	1	0.0005	ND	D17
2,4-DCP	022-0003	16	1607	23019	1	0.0007	ND	D17
2,4-DCP	024-0002	17	3094	22828	1	0.0025	ND	D17
2,4-DCP	024-0003	18	1937	20420	1	0.0014	ND	D17
2,4-DCP	026-0002	19	2095	20136	1	0.0017	ND	D17
2,4-DCP	026-0003	20	2135	21127	1	0.0016	ND	D17
2,4-DCP	028-0002	22	2363	22940	1	0.0016	ND	D17
2,4-DCP	028-0003	23	2562	22623	1	0.0019	ND	D17
2,4-DCP	032-0002	24	1881	20177	1	0.0013	ND	D17
2,4-DCP	032-0003	25	0	20666	1	ND	ND	D17
2,4-DCP	036-0002	26	0	19821	1	ND	ND	D17
2,4-DCP	036-0003	27	1455	23607	1	0.0005	ND	D17
2,4-DCP	040-0002	28	1621	21451	1	0.0008	ND	D17
2,4-DCP	040-0003	29	1872	21752	1	0.0011	ND	D17
2,4-DCP	044-0002	30	1789	24727	1	0.0008	ND	D17
2,4-DCP	044-0003	32	1707	23803	1	0.0007	ND	D17
2,4-DCP	048-0002	33	2459	24131	1	0.0016	ND	D17
2,4-DCP	048-0003	34	1544	23418	1	0.0006	ND	D17
2,4-DCP	052-0002	35	2526	23360	1	0.0018	ND	D17
2,4-DCP	052-0003	36	0	24050	1	ND	ND	D17
2,4-DCP	060-0002	37	4624	23897	1	0.0042	[0.0042]	D17
2,4-DCP	060-0003	38	4045	23202	1	0.0036	[0.0036]	D17
2,4-DCP	062-0002	39	2850	18584	1	0.0031	[0.0031]	D17
2,4-DCP	062-0003	40	3652	22606	1	0.0033	[0.0033]	D17
2,4-DCP	064-0002	42	2460	24203	1	0.0016	ND	D17
2,4-DCP	064-0003	43	3264	20324	1	0.0033	[0.0033]	D17
2,4-DCP	066-0002	44	4634	22905	1	0.0044	[0.0044]	D17
2,4-DCP	066-0003	45	3901	22406	1	0.0036	[0.0036]	D17
2,4-DCP	068-0002	46	2330	17395	1	0.0025	ND	D17
2,4-DCP	068-0003	47	4521	22741	1	0.0043	[0.0043]	D17
2,4-DCP	072-0002	48	0	20773	1	ND	ND	D17
2,4-DCP	072-0003	49	0	17940	1	ND	ND	D17
2,4-DCP	076-0002	50	1345	22985	1	0.0004	ND	D17
2,4-DCP	076-0003	52	2449	22357	1	0.0018	ND	D17
2,4-DCP	080-0002	53	1575	19355	1	0.0010	ND	D17
2,4-DCP	080-0003	54	0	18726	1	ND	ND	D17
2,4-DCP	085-0002	55	1368	17699	1	0.0009	ND	D17
2,4-DCP	085-0003	56	1125	26127	1	ND	ND	D17
2,4-DCP	089-0002	57	0	24874	1	ND	ND	D17
2,4-D	011-0001	3	0	548705	1	ND	ND	D18
2,4-D	089-0003	4	0	545677	1	ND	ND	D18
2,4-D	093-0002	5	0	521525	1	ND	ND	D18
2,4-D	093-0003	6	0	516200	1	ND	ND	D18
2,4-D	097-0002	7	0	501511	1	ND	ND	D18
2,4-D	097-0003	8	0	488747	1	ND	ND	D18

Table 6 (Cont.). Individual Residue Results from Corn Samples

Analyte	Sample ID[a]	Inj. No	Analyte Peak Area	ISTD Peak Area	Dilution Factor	Uncorrected (µg/g)	Reported (µg/g)[b]	Set ID
2,4-D	101-0002	9	0	476345	1	ND	ND	D18
2,4-D	101-0003	11	0	473345	1	ND	ND	D18
2,4-D	105-0002	12	0	463803	1	ND	ND	D18
2,4-D	105-0003	13	0	461566	1	ND	ND	D18
2,4-D	109-0002	14	0	443749	1	ND	ND	D18
2,4-D	109-0003	15	0	439141	1	ND	ND	D18
2,4-D	117-0002	16	0	433503	1	ND	ND	D18
2,4-D	117-0003	17	0	404263	1	ND	ND	D18
2,4-D	119-0002	18	0	406715	1	ND	ND	D18
2,4-D	119-0003	20	6536	404727	1	0.0020	ND	D18
2,4-D	121-0002	21	9751	398996	1	0.0036	[0.0036]	D18
2,4-D	121-0003	22	10625	399819	1	0.0040	[0.0040]	D18
2,4-D	123-0002	23	3583	386292	1	0.0008	ND	D18
2,4-D	123-0003	24	5289	383989	1	0.0016	ND	D18
2,4-D	125-0002	25	0	371222	1	ND	ND	D18
2,4-D	125-0003	26	0	361209	1	ND	ND	D18
2,4-D	129-0002	27	0	362347	1	ND	ND	D18
2,4-D	129-0003	29	0	375519	1	ND	ND	D18
2,4-D	133-0002	30	0	367886	1	ND	ND	D18
2,4-D	133-0003	31	0	359917	1	ND	ND	D18
2,4-D	137-0002	32	0	353239	1	ND	ND	D18
2,4-D	137-0003	33	0	344272	1	ND	ND	D18
2,4-D	011-0006	34	0	346936	1	ND	ND	D18
2,4-D	011-0007	35	0	337929	1	ND	ND	D18
2,4-D	080-0006	36	0	338927	1	ND	ND	D18
2,4-D	080-0007	38	0	340988	1	ND	ND	D18
2,4-D	139-0004	39	0	333763	1	ND	ND	D18
2,4-D	139-0005	40	0	327701	1	ND	ND	D18
2,4-D	139-0006	41	0	338879	1	ND	ND	D18
2,4-D	140-0004	42	0	331998	1	ND	ND	D18
2,4-D	140-0005	43	0	316631	1	ND	ND	D18
2,4-D	140-0006	44	0	309916	1	ND	ND	D18
2,4-D	148-0004	45	0	315603	1	ND	ND	D18
2,4-D	148-0005	47	0	309290	1	ND	ND	D18
2,4-D	148-0006	48	0	310144	1	ND	ND	D18
2,4-D	149-0004	49	0	300321	1	ND	ND	D18
2,4-D	149-0005	50	0	296431	1	ND	ND	D18
2,4-D	149-0006	51	0	254572	1	ND	ND	D18
2,4-D	032-0002	52	0	298130	1	ND	ND	D18
2,4-DCP	011-0001	3	1424	22537	1	0.0001	ND	D18
2,4-DCP	089-0003	4	1956	24676	1	0.0006	ND	D18
2,4-DCP	093-0002	5	0	25267	1	ND	ND	D18
2,4-DCP	093-0003	6	1715	28692	1	ND	ND	D18
2,4-DCP	097-0002	7	1716	18723	1	0.0009	ND	D18
2,4-DCP	097-0003	8	1957	16824	1	0.0016	ND	D18
2,4-DCP	101-0002	9	2506	21641	1	0.0016	ND	D18
2,4-DCP	101-0003	11	2660	20856	1	0.0020	ND	D18
2,4-DCP	105-0002	12	2278	20361	1	0.0015	ND	D18
2,4-DCP	105-0003	13	1928	22703	1	0.0008	ND	D18
2,4-DCP	109-0002	14	1835	18343	1	0.0012	ND	D18
2,4-DCP	109-0003	15	1851	25495	1	0.0004	ND	D18

Table 6 (Cont.). Individual Residue Results from Corn Samples

Analyte	Sample ID[a]	Inj. No	Analyte Peak Area	ISTD Peak Area	Dilution Factor	Uncorrected (µg/g)	Reported (µg/g)[b]	Set ID
2,4-DCP	117-0002	16	1614	22825	1	0.0004	ND	D18
2,4-DCP	117-0003	17	2922	19961	1	0.0025	ND	D18
2,4-DCP	119-0002	18	0	18214	1	ND	ND	D18
2,4-DCP	119-0003	20	1971	21584	1	0.0009	ND	D18
2,4-DCP	121-0002	21	1866	22562	1	0.0007	ND	D18
2,4-DCP	121-0003	22	2262	24007	1	0.0010	ND	D18
2,4-DCP	123-0002	23	2141	21451	1	0.0012	ND	D18
2,4-DCP	123-0003	24	1858	19960	1	0.0010	ND	D18
2,4-DCP	125-0002	25	2860	17910	1	0.0029	ND	D18
2,4-DCP	125-0003	26	2111	22092	1	0.0011	ND	D18
2,4-DCP	129-0002	27	1849	17416	1	0.0014	ND	D18
2,4-DCP	129-0003	29	2361	19243	1	0.0018	ND	D18
2,4-DCP	133-0002	30	0	20771	1	ND	ND	D18
2,4-DCP	133-0003	31	0	23791	1	ND	ND	D18
2,4-DCP	137-0002	32	1497	23912	1	0.0001	ND	D18
2,4-DCP	137-0003	33	1417	21073	1	0.0003	ND	D18
2,4-DCP	011-0006	34	2504	19852	1	0.0019	ND	D18
2,4-DCP	011-0007	35	2209	25263	1	0.0008	ND	D18
2,4-DCP	080-0006	36	1796	24413	1	0.0004	ND	D18
2,4-DCP	080-0007	38	1825	25438	1	0.0004	ND	D18
2,4-DCP	139-0004	39	4016	24674	1	0.0030	[0.0030]	D18
2,4-DCP	139-0005	40	1881	24414	1	0.0005	ND	D18
2,4-DCP	139-0006	41	0	23979	1	ND	ND	D18
2,4-DCP	140-0004	42	2487	24928	1	0.0012	ND	D18
2,4-DCP	140-0005	43	2317	17471	1	0.0021	ND	D18
2,4-DCP	140-0006	44	2176	18413	1	0.0017	ND	D18
2,4-DCP	148-0004	45	12148	22314	1	0.0138	0.0138	D18
2,4-DCP	148-0005	47	6119	20125	1	0.0070	[0.0070]	D18
2,4-DCP	148-0006	48	3999	21187	1	0.0037	[0.0037]	D18
2,4-DCP	149-0004	49	2645	23869	1	0.0015	ND	D18
2,4-DCP	149-0005	50	2107	19856	1	0.0014	ND	D18
2,4-DCP	149-0006	51	1713	16020	1	0.0014	ND	D18
2,4-DCP	032-0002	52	0	21302	1	ND	ND	D18
2,4-D	141-0001	3	0	279413	1	ND	ND	D19
2,4-D	141-0002	4	32324	262099	1	0.0211	0.0211	D19
2,4-D	150-0001	5	0	277059	1	ND	ND	D19
2,4-D	150-0002	6	3404	261343	1	0.0010	ND	D19
2,4-D	146-0001	7	0	231570	1	ND	ND	D19
2,4-D	146-0002	8	0	237368	1	ND	ND	D19
2,4-D	146-0003	9	0	243740	1	ND	ND	D19
2,4-D	146-0004	10	0	221944	1	ND	ND	D19
2,4-D	155-0001	12	0	221006	1	ND	ND	D19
2,4-D	155-0002	13	0	224878	1	ND	ND	D19
2,4-D	155-0003	14	0	245812	1	ND	ND	D19
2,4-D	155-0004	15	0	231625	1	ND	ND	D19
2,4-D	145-0001	16	0	233741	1	ND	ND	D19
2,4-D	145-0002	17	0	232720	1	ND	ND	D19
2,4-D	145-0003	18	0	229432	1	ND	ND	D19
2,4-D	145-0004	19	0	229652	1	ND	ND	D19
2,4-D	154-0001	20	0	239530	1	ND	ND	D19
2,4-D	154-0002	22	0	238393	1	ND	ND	D19

Table 6 (Cont.). Individual Residue Results from Corn Samples

Analyte	Sample ID[a]	Inj. No	Analyte Peak Area	ISTD Peak Area	Dilution Factor	Uncorrected (µg/g)	Reported (µg/g)[b]	Set ID
2,4-D	154-0003	23	0	228984	1	ND	ND	D19
2,4-D	154-0004	24	0	220475	1	ND	ND	D19
2,4-D	144-0001	25	0	221073	1	ND	ND	D19
2,4-D	144-0002	26	0	233569	1	ND	ND	D19
2,4-D	144-0003	27	0	216864	1	ND	ND	D19
2,4-D	144-0004	28	0	226225	1	ND	ND	D19
2,4-D	153-0001	29	0	225070	1	ND	ND	D19
2,4-D	153-0002	30	0	226535	1	ND	ND	D19
2,4-D	153-0003	32	0	226376	1	ND	ND	D19
2,4-D	153-0004	33	0	230515	1	ND	ND	D19
2,4-D	142-0001	34	0	235788	1	ND	ND	D19
2,4-D	142-0002	35	0	227063	1	ND	ND	D19
2,4-D	142-0003	36	0	214185	1	ND	ND	D19
2,4-D	142-0004	37	0	218707	1	ND	ND	D19
2,4-D	151-0001	38	0	222618	1	ND	ND	D19
2,4-D	151-0002	39	0	220881	1	ND	ND	D19
2,4-D	151-0003	40	0	244850	1	ND	ND	D19
2,4-D	151-0004	42	0	225477	1	ND	ND	D19
2,4-DCP	141-0001	3	1393	16399	1	0.0012	ND	D19
2,4-DCP	141-0002	4	5890	23316	1	0.0060	[0.0060]	D19
2,4-DCP	150-0001	5	1385	21634	1	0.0006	ND	D19
2,4-DCP	150-0002	6	1783	16581	1	0.0019	ND	D19
2,4-DCP	146-0001	7	0	20826	1	ND	ND	D19
2,4-DCP	146-0002	8	1082	24062	1	0.0001	ND	D19
2,4-DCP	146-0003	9	1979	22971	1	0.0013	ND	D19
2,4-DCP	146-0004	10	1341	16015	1	0.0012	ND	D19
2,4-DCP	155-0001	12	0	17522	1	ND	ND	D19
2,4-DCP	155-0002	13	0	20720	1	ND	ND	D19
2,4-DCP	155-0003	14	0	19944	1	ND	ND	D19
2,4-DCP	155-0004	15	0	20469	1	ND	ND	D19
2,4-DCP	145-0001	16	1723	23418	1	0.0009	ND	D19
2,4-DCP	145-0002	17	1607	18855	1	0.0012	ND	D19
2,4-DCP	145-0003	18	2128	20103	1	0.0018	ND	D19
2,4-DCP	145-0004	19	2320	22037	1	0.0018	ND	D19
2,4-DCP	154-0001	20	1608	20372	1	0.0011	ND	D19
2,4-DCP	154-0002	22	1788	21224	1	0.0012	ND	D19
2,4-DCP	154-0003	23	1657	22422	1	0.0009	ND	D19
2,4-DCP	154-0004	24	1703	17754	1	0.0016	ND	D19
2,4-DCP	144-0001	25	1292	15916	1	0.0011	ND	D19
2,4-DCP	144-0002	26	0	19417	1	ND	ND	D19
2,4-DCP	144-0003	27	1813	17571	1	0.0018	ND	D19
2,4-DCP	144-0004	28	1894	22253	1	0.0012	ND	D19
2,4-DCP	153-0001	29	0	22589	1	ND	ND	D19
2,4-DCP	153-0002	30	1250	16754	1	0.0009	ND	D19
2,4-DCP	153-0003	32	1353	14614	1	0.0015	ND	D19
2,4-DCP	153-0004	33	1329	18377	1	0.0009	ND	D19
2,4-DCP	142-0001	34	1562	17827	1	0.0013	ND	D19
2,4-DCP	142-0002	35	2041	21372	1	0.0015	ND	D19
2,4-DCP	142-0003	36	1994	19865	1	0.0017	ND	D19
2,4-DCP	142-0004	37	1880	14382	1	0.0025	ND	D19
2,4-DCP	151-0001	38	0	21809	1	ND	ND	D19

Table 6 (Cont.). Individual Residue Results from Corn Samples

Analyte	Sample ID[a]	Inj. No	Analyte Peak Area	ISTD Peak Area	Dilution Factor	Uncorrected (µg/g)	Reported (µg/g)[b]	Set ID
2,4-DCP	151-0002	39	1215	20676	1	0.0005	ND	D19
2,4-DCP	151-0003	40	1210	23235	1	0.0003	ND	D19
2,4-DCP	151-0004	42	1667	20275	1	0.0012	ND	D19
2,4-D	143-0001	3	7301	591836	1	0.0008	ND	D20
2,4-D	143-0002	4	0	580093	1	ND	ND	D20
2,4-D	143-0003	5	0	559797	1	ND	ND	D20
2,4-D	143-0004	7	0	539523	1	ND	ND	D20
2,4-D	147-0001	8	0	523925	1	ND	ND	D20
2,4-D	147-0002	9	0	543384	1	ND	ND	D20
2,4-D	147-0003	10	0	505943	1	ND	ND	D20
2,4-D	147-0004	12	0	516416	1	ND	ND	D20
2,4-D	152-0001	13	0	510397	1	ND	ND	D20
2,4-D	152-0002	14	0	486878	1	ND	ND	D20
2,4-D	152-0003	15	0	488604	1	ND	ND	D20
2,4-D	152-0004	17	0	442277	1	ND	ND	D20
2,4-D	156-0001	18	0	443782	1	ND	ND	D20
2,4-D	156-0002	19	0	444951	1	ND	ND	D20
2,4-D	156-0003	20	0	414635	1	ND	ND	D20
2,4-D	156-0004	22	0	418194	1	ND	ND	D20
2,4-DCP	143-0001	3	0	25816	1	ND	ND	D20
2,4-DCP	143-0002	4	0	29949	1	ND	ND	D20
2,4-DCP	143-0003	5	0	27392	1	ND	ND	D20
2,4-DCP	143-0004	7	0	30726	1	ND	ND	D20
2,4-DCP	147-0001	8	0	30667	1	ND	ND	D20
2,4-DCP	147-0002	9	0	30317	1	ND	ND	D20
2,4-DCP	147-0003	10	0	27606	1	ND	ND	D20
2,4-DCP	147-0004	12	1490	30493	1	0.0005	ND	D20
2,4-DCP	152-0001	13	1355	32576	1	0.0003	ND	D20
2,4-DCP	152-0002	14	0	24710	1	ND	ND	D20
2,4-DCP	152-0003	15	0	24848	1	ND	ND	D20
2,4-DCP	152-0004	17	0	30750	1	ND	ND	D20
2,4-DCP	156-0001	18	0	22593	1	ND	ND	D20
2,4-DCP	156-0002	19	0	20845	1	ND	ND	D20
2,4-DCP	156-0003	20	0	26129	1	ND	ND	D20
2,4-DCP	156-0004	22	1504	30556	1	0.0005	ND	D20
2,4-D	001-0001	3	6858	389455	1	0.0011	ND	D21
2,4-D	002-0001	4	0	394483	1	ND	ND	D21
2,4-D	005-0001	5	0	384974	1	ND	ND	D21
2,4-D	006-0001	6	0	365742	1	ND	ND	D21
2,4-D	009-0001	7	0	360640	1	ND	ND	D21
2,4-D	010-0001	8	0	362299	1	ND	ND	D21
2,4-D	014-0001	10	0	355061	1	ND	ND	D21
2,4-D	015-0001	11	0	351122	1	ND	ND	D21
2,4-D	016-0001	12	0	354642	1	ND	ND	D21
2,4-D	017-0001	13	0	343548	1	ND	ND	D21
2,4-D	018-0001	14	0	343717	1	ND	ND	D21
2,4-D	019-0001	15	0	323960	1	ND	ND	D21
2,4-D	030-0001	16	0	321545	1	ND	ND	D21
2,4-D	031-0001	18	0	322069	1	ND	ND	D21
2,4-D	034-0001	19	0	307018	1	ND	ND	D21
2,4-D	035-0001	20	0	300683	1	ND	ND	D21

Table 6 (Cont.). Individual Residue Results from Corn Samples

Analyte	Sample ID[a]	Inj. No	Analyte Peak Area	ISTD Peak Area	Dilution Factor	Uncorrected (µg/g)	Reported (µg/g)[b]	Set ID
2,4-D	038-0001	21	4807	309734	1	0.0007	ND	D21
2,4-D	039-0001	22	0	299391	1	ND	ND	D21
2,4-D	042-0001	23	0	304656	1	ND	ND	D21
2,4-D	043-0001	24	0	293163	1	ND	ND	D21
2,4-D	046-0001	26	0	297466	1	ND	ND	D21
2,4-D	047-0001	27	0	284359	1	ND	ND	D21
2,4-D	050-0001	28	0	278827	1	ND	ND	D21
2,4-D	051-0001	29	0	282823	1	ND	ND	D21
2,4-D	054-0001	30	0	281076	1	ND	ND	D21
2,4-D	055-0001	31	0	272522	1	ND	ND	D21
2,4-D	056-0001	32	0	279792	1	ND	ND	D21
2,4-D	057-0001	34	0	274192	1	ND	ND	D21
2,4-D	058-0001	35	0	266870	1	ND	ND	D21
2,4-D	059-0001	36	0	263528	1	ND	ND	D21
2,4-D	070-0001	37	0	260909	1	ND	ND	D21
2,4-D	071-0001	38	3405	262380	1	0.0002	ND	D21
2,4-D	074-0001	39	0	259732	1	ND	ND	D21
2,4-D	075-0001	40	0	263346	1	ND	ND	D21
2,4-D	079-0001	42	0	260902	1	ND	ND	D21
2,4-D	083-0001	43	0	259059	1	ND	ND	D21
2,4-D	084-0001	44	0	256268	1	ND	ND	D21
2,4-D	087-0001	45	4545	244999	1	0.0012	ND	D21
2,4-D	115-0003	50	138094	247049	1	0.1005	0.1005	D21
2,4-DCP	001-0001	3	3482	20602	1	0.0014	ND	D21
2,4-DCP	002-0001	4	3530	16270	1	0.0028	ND	D21
2,4-DCP	005-0001	5	3585	16245	1	0.0029	ND	D21
2,4-DCP	006-0001	6	5444	14788	1	0.0072	[0.0072]	D21
2,4-DCP	009-0001	7	3972	17998	1	0.0029	ND	D21
2,4-DCP	010-0001	8	4085	15248	1	0.0043	[0.0043]	D21
2,4-DCP	014-0001	10	2607	16599	1	0.0011	ND	D21
2,4-DCP	015-0001	11	2886	14387	1	0.0024	ND	D21
2,4-DCP	016-0001	12	3967	18127	1	0.0029	ND	D21
2,4-DCP	017-0001	13	2594	16352	1	0.0011	ND	D21
2,4-DCP	018-0001	14	3494	16293	1	0.0028	ND	D21
2,4-DCP	019-0001	15	2570	19039	1	0.0005	ND	D21
2,4-DCP	030-0001	16	2689	13233	1	0.0024	ND	D21
2,4-DCP	031-0001	18	3094	14996	1	0.0025	ND	D21
2,4-DCP	034-0001	19	2084	16310	1	0.0003	ND	D21
2,4-DCP	035-0001	20	2644	15351	1	0.0015	ND	D21
2,4-DCP	038-0001	21	2646	15983	1	0.0013	ND	D21
2,4-DCP	039-0001	22	2360	13890	1	0.0015	ND	D21
2,4-DCP	042-0001	23	4595	16007	1	0.0049	[0.0049]	D21
2,4-DCP	043-0001	24	4467	14238	1	0.0056	[0.0056]	D21
2,4-DCP	046-0001	26	2909	15380	1	0.0020	ND	D21
2,4-DCP	047-0001	27	3240	13404	1	0.0035	[0.0035]	D21
2,4-DCP	050-0001	28	6067	13594	1	0.0094	[0.0094]	D21
2,4-DCP	051-0001	29	4904	16352	1	0.0052	[0.0052]	D21
2,4-DCP	054-0001	30	7253	17699	1	0.0084	[0.0084]	D21
2,4-DCP	055-0001	31	3818	15964	1	0.0035	[0.0035]	D21
2,4-DCP	056-0001	32	3265	18067	1	0.0018	ND	D21
2,4-DCP	057-0001	34	4016	18096	1	0.0030	[0.0030]	D21

Table 6 (Cont.). Individual Residue Results from Corn Samples

Analyte	Sample ID[a]	Inj. No	Analyte Peak Area	ISTD Peak Area	Dilution Factor	Uncorrected (µg/g)	Reported (µg/g)[b]	Set ID
2,4-DCP	058-0001	35	3649	16501	1	0.0030	[0.0030]	D21
2,4-DCP	059-0001	36	2700	16021	1	0.0014	ND	D21
2,4-DCP	070-0001	37	2591	14968	1	0.0016	ND	D21
2,4-DCP	071-0001	38	2624	17842	1	0.0008	ND	D21
2,4-DCP	074-0001	39	3540	15937	1	0.0030	[0.0030]	D21
2,4-DCP	075-0001	40	3953	17912	1	0.0029	ND	D21
2,4-DCP	079-0001	42	2819	13802	1	0.0025	ND	D21
2,4-DCP	083-0001	43	3713	17398	1	0.0027	ND	D21
2,4-DCP	084-0001	44	3103	17023	1	0.0018	ND	D21
2,4-DCP	087-0001	45	5475	14883	1	0.0072	[0.0072]	D21
2,4-DCP	115-0003	50	23807	17398	1	0.0361	0.0361	D21
2,4-D	046-0001	3	3254	572889	1	ND	ND	D22
2,4-D	088-0001	4	7004	549927	1	0.0013	ND	D22
2,4-D	091-0001	5	0	505454	1	ND	ND	D22
2,4-D	092-0001	6	0	500681	1	ND	ND	D22
2,4-D	095-0001	7	0	476215	1	ND	ND	D22
2,4-D	096-0001	8	0	484253	1	ND	ND	D22
2,4-D	099-0001	10	2380	464868	1	ND	ND	D22
2,4-D	103-0001	11	0	456336	1	ND	ND	D22
2,4-D	104-0001	12	0	455535	1	ND	ND	D22
2,4-D	107-0001	13	0	462052	1	ND	ND	D22
2,4-D	108-0001	14	0	432366	1	ND	ND	D22
2,4-D	100-0001	15	0	418594	1	ND	ND	D22
2,4-D	111-0001	16	0	411992	1	ND	ND	D22
2,4-D	112-0001	18	0	395548	1	ND	ND	D22
2,4-D	113-0001	19	0	389346	1	ND	ND	D22
2,4-D	114-0001	20	0	389606	1	ND	ND	D22
2,4-D	115-0001	21	0	376002	1	ND	ND	D22
2,4-D	116-0001	22	0	366601	1	ND	ND	D22
2,4-D	127-0001	23	0	352740	1	ND	ND	D22
2,4-D	128-0001	24	3235	358735	1	0.0006	ND	D22
2,4-D	131-0001	26	0	343248	1	ND	ND	D22
2,4-D	132-0001	27	0	325940	1	ND	ND	D22
2,4-D	135-0001	28	0	326524	1	ND	ND	D22
2,4-D	136-0001	29	0	335283	1	ND	ND	D22
2,4-D	042-0002	30	19775	329019	1	0.0099	[0.0099]	D22
2,4-D	083-0003	31	14736	330832	1	0.0070	[0.0070]	D22
2,4-D	087-0003	32	16892	326410	1	0.0083	[0.0083]	D22
2,4-D	096-0002	34	24120	325282	1	0.0124	0.0124	D22
2,4-D	051-0002	35	33981	318703	1	0.0183	0.0183	D22
2,4-D	088-0003	36	22872	302317	1	0.0127	0.0127	D22
2,4-D	051-0003	37	39718	293273	1	0.0235	0.0235	D22
2,4-D	017-0002	38	11551	300229	1	0.0059	[0.0059]	D22
2,4-D	017-0003	39	0	299733	1	ND	ND	D22
2,4-D	018-0002	40	0	299448	1	ND	ND	D22
2,4-D	018-0003	42	0	301761	1	ND	ND	D22
2,4-D	019-0002	43	0	292521	1	ND	ND	D22
2,4-D	019-0003	44	0	288755	1	ND	ND	D22
2,4-D	042-0003	45	3655	284727	1	0.0013	ND	D22
2,4-D	115-0003	50	173477	278290	1	0.1119	0.1119	D22
2,4-DCP	046-0001	3	1291	20991	1	0.0002	ND	D22

Table 6 (Cont.). Individual Residue Results from Corn Samples

Analyte	Sample ID[a]	Inj. No	Analyte Peak Area	ISTD Peak Area	Dilution Factor	Uncorrected (µg/g)	Reported (µg/g)[b]	Set ID
2,4-DCP	088-0001	4	4341	24307	1	0.0035	[0.0035]	D22
2,4-DCP	091-0001	5	3309	22340	1	0.0026	ND	D22
2,4-DCP	092-0001	6	2314	21613	1	0.0015	ND	D22
2,4-DCP	095-0001	7	7707	18959	1	0.0099	[0.0099]	D22
2,4-DCP	096-0001	8	1833	19303	1	0.0011	ND	D22
2,4-DCP	099-0001	10	6733	22273	1	0.0070	[0.0070]	D22
2,4-DCP	103-0001	11	3103	19678	1	0.0029	ND	D22
2,4-DCP	104-0001	12	2433	17793	1	0.0023	ND	D22
2,4-DCP	107-0001	13	3079	22158	1	0.0024	ND	D22
2,4-DCP	108-0001	14	1445	18092	1	0.0007	ND	D22
2,4-DCP	100-0001	15	1966	18550	1	0.0015	ND	D22
2,4-DCP	111-0001	16	3130	18704	1	0.0032	[0.0032]	D22
2,4-DCP	112-0001	18	2759	19778	1	0.0024	ND	D22
2,4-DCP	113-0001	19	4076	18572	1	0.0047	[0.0047]	D22
2,4-DCP	114-0001	20	2537	16473	1	0.0028	ND	D22
2,4-DCP	115-0001	21	3513	16675	1	0.0044	[0.0044]	D22
2,4-DCP	116-0001	22	3916	17028	1	0.0050	[0.0050]	D22
2,4-DCP	127-0001	23	2415	17943	1	0.0023	ND	D22
2,4-DCP	128-0001	24	2559	19892	1	0.0021	ND	D22
2,4-DCP	131-0001	26	2821	17014	1	0.0031	[0.0031]	D22
2,4-DCP	132-0001	27	2432	16625	1	0.0026	ND	D22
2,4-DCP	135-0001	28	2516	17167	1	0.0026	ND	D22
2,4-DCP	136-0001	29	3303	21309	1	0.0028	ND	D22
2,4-DCP	042-0002	30	9025	19602	1	0.0115	0.0115	D22
2,4-DCP	083-0003	31	33995	19803	1	0.0469	0.0469	D22
2,4-DCP	087-0003	32	22243	21934	1	0.0271	0.0271	D22
2,4-DCP	096-0002	34	27801	18983	1	0.0398	0.0398	D22
2,4-DCP	051-0002	35	20247	18768	1	0.0289	0.0289	D22
2,4-DCP	088-0003	36	15506	17604	1	0.0233	0.0233	D22
2,4-DCP	051-0003	37	17037	18144	1	0.0250	0.0250	D22
2,4-DCP	017-0002	38	5420	22432	1	0.0053	[0.0053]	D22
2,4-DCP	017-0003	39	3953	19316	1	0.0042	[0.0042]	D22
2,4-DCP	018-0002	40	3002	19603	1	0.0028	ND	D22
2,4-DCP	018-0003	42	3857	19577	1	0.0040	[0.0040]	D22
2,4-DCP	019-0002	43	3873	17807	1	0.0046	[0.0046]	D22
2,4-DCP	019-0003	44	4849	18692	1	0.0058	[0.0058]	D22
2,4-DCP	042-0003	45	3170	19655	1	0.0030	[0.0030]	D22
2,4-DCP	115-0003	50	23948	16748	1	0.0388	0.0388	D22
2,4-D	046-0001	3	3249	600794	1	0.0009	ND	D23
2,4-D	050-0002	7	3695	543657	1	0.0012	ND	D23
2,4-D	115-0003	8	294907	533606	1	0.1001	0.1001	D23
2,4-D	084-0002	11	12485	497416	1	0.0045	[0.0045]	D23
2,4-D	084-0003	12	5632	479124	1	0.0021	ND	D23
2,4-D	088-0002	13	5023	468754	1	0.0019	ND	D23
2,4-D	095-0002	14	12411	454550	1	0.0049	[0.0049]	D23
2,4-D	095-0003	15	0	453809	1	ND	ND	D23
2,4-D	096-0003	16	14649	435098	1	0.0060	[0.0060]	D23
2,4-D	127-0002	18	0	434447	1	ND	ND	D23
2,4-D	127-0003	19	0	420197	1	ND	ND	D23
2,4-D	043-0003	20	68161	418453	1	0.0295	0.0295	D23
2,4-D	083-0002	21	49309	405283	1	0.0220	0.0220	D23

Table 6 (Cont.). Individual Residue Results from Corn Samples

Analyte	Sample ID[a]	Inj. No	Analyte Peak Area	ISTD Peak Area	Dilution Factor	Uncorrected (µg/g)	Reported (µg/g)[b]	Set ID
2,4-D	132-0003	22	60745	402754	1	0.0273	0.0273	D23
2,4-D	136-0003	23	66938	405226	1	0.0299	0.0299	D23
2,4-D	087-0002	24	72702	378759	1	0.0347	0.0347	D23
2,4-D	132-0002	26	28030	391596	1	0.0129	0.0129	D23
2,4-D	043-0002	27	131859	370892	1	0.0644	0.0644	D23
2,4-D	136-0002	28	63656	378586	1	0.0304	0.0304	D23
2,4-D	079-0002	29	83032	370811	1	0.0405	0.0405	D23
2,4-D	116-0002	30	119505	370033	1	0.0585	0.0585	D23
2,4-D	111-0002	31	41057	360264	1	0.0206	0.0206	D23
2,4-D	115-0003	32	203877	344417	1	0.1072	0.1072	D23
2,4-D	050-0003	34	161068	345996	1	0.0843	0.0843	D23
2,4-D	079-0003	35	156797	347414	1	0.0818	0.0818	D23
2,4-D	116-0003	36	157635	340096	1	0.0840	0.0840	D23
2,4-D	135-0002	37	130535	321928	1	0.0734	0.0734	D23
2,4-D	115-0002	38	343971	325546	1	0.1915	0.1915	D23
2,4-D	016-0003	39	220047	320679	1	0.1243	0.1243	D23
2,4-D	135-0003	40	122491	335373	1	0.0661	0.0661	D23
2,4-D	111-0003	42	446057	319232	1	0.2533	0.2533	D23
2,4-D	092-0002	43	276821	313017	1	0.1603	0.1603	D23
2,4-D	035-0002	44	373457	303910	1	0.2227	0.2227	D23
2,4-D	035-0003	45	375374	310893	1	0.2188	0.2188	D23
2,4-D	108-0003	46	44404	285724	10	0.2811	0.2811	D23
2,4-D	010-0002	47	56368	303327	10	0.3362	0.3362	D23
2,4-D	113-0002	48	29071	280278	10	0.1873	0.1873	D23
2,4-D	131-0002	50	33355	298841	10	0.2017	0.2017	D23
2,4-DCP	046-0001	3	2286	21022	1	0.0014	ND	D23
2,4-DCP	050-0002	7	5095	23378	1	0.0046	[0.0046]	D23
2,4-DCP	115-0003	8	30008	23499	1	0.0352	0.0352	D23
2,4-DCP	084-0002	11	20747	15071	1	0.0380	0.0380	D23
2,4-DCP	084-0003	12	9191	15166	1	0.0158	0.0158	D23
2,4-DCP	088-0002	13	14128	23219	1	0.0159	0.0159	D23
2,4-DCP	095-0002	14	12664	23981	1	0.0135	0.0135	D23
2,4-DCP	095-0003	15	3610	22246	1	0.0030	[0.0030]	D23
2,4-DCP	096-0003	16	9705	15663	1	0.0162	0.0162	D23
2,4-DCP	127-0002	18	1570	17828	1	0.0008	ND	D23
2,4-DCP	127-0003	19	1234	13088	1	0.0010	ND	D23
2,4-DCP	043-0003	20	44669	24121	1	0.0518	0.0518	D23
2,4-DCP	083-0002	21	6529	13514	1	0.0123	0.0123	D23
2,4-DCP	132-0003	22	43792	14004	1	0.0886	0.0886	D23
2,4-DCP	136-0003	23	172216	18000	1	0.2745	0.2745	D23
2,4-DCP	087-0002	24	24902	9053	1	0.0777	0.0777	D23
2,4-DCP	132-0002	26	35806	24580	1	0.0404	0.0404	D23
2,4-DCP	043-0002	27	35045	11869	1	0.0835	0.0835	D23
2,4-DCP	136-0002	28	165472	23670	1	0.2001	0.2001	D23
2,4-DCP	079-0002	29	129943	22699	1	0.1635	0.1635	D23
2,4-DCP	116-0002	30	25680	19650	1	0.0360	0.0360	D23
2,4-DCP	111-0002	31	16099	13977	1	0.0316	0.0316	D23
2,4-DCP	115-0003	32	25604	20026	1	0.0352	0.0352	D23
2,4-DCP	050-0003	34	73302	18384	1	0.1134	0.1134	D23
2,4-DCP	079-0003	35	207609	20771	1	0.2868	0.2868	D23
2,4-DCP	116-0003	36	22375	20599	1	0.0297	0.0297	D23

Table 6 (Cont.). Individual Residue Results from Corn Samples

Analyte	Sample ID[a]	Inj. No	Analyte Peak Area	ISTD Peak Area	Dilution Factor	Uncorrected (µg/g)	Reported (µg/g)[b]	Set ID
2,4-DCP	135-0002	37	271194	12935	1	0.6035	0.6035	D23
2,4-DCP	115-0002	38	25535	21692	1	0.0323	0.0323	D23
2,4-DCP	016-0003	39	44934	12786	1	0.0997	0.0997	D23
2,4-DCP	135-0003	40	334431	16936	1	0.5683	0.5683	D23
2,4-DCP	111-0003	42	75791	16595	1	0.1301	0.1301	D23
2,4-DCP	092-0002	43	247275	15501	1	0.4587	0.4587	D23
2,4-DCP	035-0002	44	208795	13740	1	0.4369	0.4369	D23
2,4-DCP	035-0003	45	361885	19722	1	0.5279	0.5279	D23
2,4-DCP	108-0003	46	8922	12777	10	0.1846	0.1846	D23
2,4-DCP	010-0002	47	12776	19884	10	0.1685	0.1685	D23
2,4-DCP	113-0002	48	5752	17334	10	0.0788	0.0788	D23
2,4-DCP	131-0002	50	49545	20027	10	0.6971	0.6971	D23
2,4-D	046-0001	3	4323	521329	1	ND	ND	D24
2,4-D	115-0003	7	307478	472011	1	0.1161	0.1161	D24
2,4-D	071-0003	8	55573	422429	10	0.2223	0.2223	D24
2,4-D	100-0003	10	43771	394025	10	0.1853	0.1853	D24
2,4-D	092-0003	11	51746	412854	10	0.2110	0.2110	D24
2,4-D	131-0003	12	53898	397249	10	0.2297	0.2297	D24
2,4-D	112-0002	13	632478	451204	1	0.2515	0.2515	D24
2,4-D	010-0003	14	1152787	431732	1	0.4805	0.4805	D24
2,4-D	107-0003	15	861209	410787	1	0.3770	0.3770	D24
2,4-D	030-0003	16	672780	399081	1	0.3028	0.3028	D24
2,4-D	107-0002	18	707825	384159	1	0.3311	0.3311	D24
2,4-D	100-0002	19	60276	336299	10	0.3083	0.3083	D24
2,4-D	114-0003	20	530543	446771	1	0.2129	0.2129	D24
2,4-D	108-0002	21	1260960	368849	1	0.6157	0.6157	D24
2,4-D	114-0002	22	587688	358419	1	0.2945	0.2945	D24
2,4-D	104-0002	23	57074	334741	10	0.2926	0.2926	D24
2,4-D	112-0003	24	541896	355910	1	0.2734	0.2734	D24
2,4-D	104-0003	26	57864	338803	10	0.2931	0.2931	D24
2,4-D	091-0002	27	61226	301382	10	0.3515	0.3515	D24
2,4-D	071-0002	28	53324	266332	10	0.3462	0.3462	D24
2,4-D	099-0002	29	47547	297723	10	0.2731	0.2731	D24
2,4-D	034-0002	30	75477	301890	10	0.4361	0.4361	D24
2,4-D	030-0002	31	76773	298438	10	0.4492	0.4492	D24
2,4-D	015-0003	32	110093	296680	10	0.6547	0.6547	D24
2,4-D	016-0002	34	73313	286310	10	0.4470	0.4470	D24
2,4-D	009-0002	35	99476	284533	10	0.6159	0.6159	D24
2,4-D	015-0002	36	81714	275024	10	0.5212	0.5212	D24
2,4-D	034-0003	37	79356	295898	10	0.4689	0.4689	D24
2,4-D	091-0003	38	77192	274061	10	0.4933	0.4933	D24
2,4-D	047-0003	39	95891	269030	10	0.6283	0.6283	D24
2,4-D	009-0003	40	110645	283920	10	0.6883	0.6883	D24
2,4-D	103-0002	42	72218	263259	10	0.4800	0.4800	D24
2,4-D	099-0003	43	76858	257755	10	0.5231	0.5231	D24
2,4-D	031-0002	44	84917	261409	10	0.5712	0.5712	D24
2,4-D	039-0003	45	93839	277660	10	0.5949	0.5949	D24
2,4-D	103-0003	46	105849	255918	10	0.7315	0.7315	D24
2,4-D	014-0002	47	138860	277417	10	0.8884	0.8884	D24
2,4-D	014-0003	48	176845	269780	10	1.1682	1.1682	D24
2,4-D	046-0003	50	146529	269178	10	0.9675	0.9675	D24

Table 6 (Cont.). Individual Residue Results from Corn Samples

Analyte	Sample ID[a]	Inj. No	Analyte Peak Area	ISTD Peak Area	Dilution Factor	Uncorrected (µg/g)	Reported (µg/g)[b]	Set ID
2,4-DCP	046-0001	3	2409	16177	1	0.0027	ND	D24
2,4-DCP	115-0003	7	29838	20012	1	0.0402	0.0402	D24
2,4-DCP	071-0003	8	38421	14035	10	0.7512	0.7512	D24
2,4-DCP	100-0003	10	40357	13113	10	0.8464	0.8464	D24
2,4-DCP	092-0003	11	27481	11310	10	0.6650	0.6650	D24
2,4-DCP	131-0003	12	63200	18907	10	0.9205	0.9205	D24
2,4-DCP	112-0002	13	59786	17527	1	0.0940	0.0940	D24
2,4-DCP	010-0003	14	132515	21134	1	0.1740	0.1740	D24
2,4-DCP	107-0003	15	235271	18998	1	0.3451	0.3451	D24
2,4-DCP	030-0003	16	362396	21164	1	0.4778	0.4778	D24
2,4-DCP	107-0002	18	153632	13243	1	0.3232	0.3232	D24
2,4-DCP	100-0002	19	62806	13358	10	1.3010	1.3010	D24
2,4-DCP	114-0003	20	31098	15108	1	0.0561	0.0561	D24
2,4-DCP	108-0002	21	145734	10997	1	0.3694	0.3694	D24
2,4-DCP	114-0002	22	31637	14773	1	0.0584	0.0584	D24
2,4-DCP	104-0002	23	49926	13509	10	1.0194	1.0194	D24
2,4-DCP	112-0003	24	46312	17472	1	0.0727	0.0727	D24
2,4-DCP	104-0003	26	57344	19400	10	0.8123	0.8123	D24
2,4-DCP	091-0002	27	47984	14725	10	0.8970	0.8970	D24
2,4-DCP	071-0002	28	26042	12585	10	0.5641	0.5641	D24
2,4-DCP	099-0002	29	102791	18416	10	1.5473	1.5473	D24
2,4-DCP	034-0002	30	45691	14147	10	0.8889	0.8889	D24
2,4-DCP	030-0002	31	23407	14968	10	0.4226	0.4226	D24
2,4-DCP	015-0003	32	31076	16155	10	0.5233	0.5233	D24
2,4-DCP	016-0002	34	14454	9891	10	0.3939	0.3939	D24
2,4-DCP	009-0002	35	26366	18013	10	0.3946	0.3946	D24
2,4-DCP	015-0002	36	22384	12901	10	0.4705	0.4705	D24
2,4-DCP	034-0003	37	59188	19197	10	0.8479	0.8479	D24
2,4-DCP	091-0003	38	87925	20999	10	1.1569	1.1569	D24
2,4-DCP	047-0003	39	69622	19104	10	1.0050	1.0050	D24
2,4-DCP	009-0003	40	36064	17829	10	0.5511	0.5511	D24
2,4-DCP	103-0002	42	86335	15990	10	1.4962	1.4962	D24
2,4-DCP	099-0003	43	118582	15317	10	2.1519	2.1519	D24
2,4-DCP	031-0002	44	44401	19756	10	0.6140	0.6140	D24
2,4-DCP	039-0003	45	59949	16739	10	0.9874	0.9874	D24
2,4-DCP	103-0003	46	165707	20835	10	2.2111	2.2111	D24
2,4-DCP	014-0002	47	57819	22904	10	0.6915	0.6915	D24
2,4-DCP	014-0003	48	69024	20514	10	0.9267	0.9267	D24
2,4-DCP	046-0003	50	87305	16432	10	1.4721	1.4721	D24
2,4-D	046-0001	3	0	287532	1	ND	ND	D25
2,4-D	115-0003	7	154816	260190	1	0.1047	0.1047	D25
2,4-D	055-0002	8	101896	250968	10	0.7119	0.7119	D25
2,4-D	070-0002	10	115767	249599	10	0.8143	0.8143	D25
2,4-D	070-0003	11	123896	243634	10	0.8935	0.8935	D25
2,4-D	031-0003	12	128996	251020	10	0.9030	0.9030	D25
2,4-D	059-0003	13	126284	268231	10	0.8267	0.8267	D25
2,4-D	047-0002	14	177499	243315	10	1.2850	1.2850	D25
2,4-D	055-0003	15	125928	242419	10	0.9129	0.9129	D25
2,4-D	038-0002	16	144287	240740	10	1.0544	1.0544	D25
2,4-D	059-0002	18	126544	249551	10	0.8910	0.8910	D25
2,4-D	038-0003	19	176201	239359	10	1.2968	1.2968	D25

Table 6 (Cont.). Individual Residue Results from Corn Samples

Analyte	Sample ID[a]	Inj. No	Analyte Peak Area	ISTD Peak Area	Dilution Factor	Uncorrected (µg/g)	Reported (µg/g)[b]	Set ID
2,4-D	113-0003	20	151391	243250	10	1.0952	1.0952	D25
2,4-D	039-0002	21	220051	246017	10	1.5772	1.5772	D25
2,4-D	046-0002	22	220257	243211	10	1.5970	1.5970	D25
2,4-D	075-0002	23	187797	247148	10	1.3388	1.3388	D25
2,4-D	056-0002	24	195134	242334	10	1.4192	1.4192	D25
2,4-D	002-0003	26	298297	254551	10	2.0687	2.0687	D25
2,4-D	005-0003	27	201577	226971	10	1.5660	1.5660	D25
2,4-D	054-0002	28	196337	234602	10	1.4753	1.4753	D25
2,4-D	058-0003	29	283556	236656	10	2.1153	2.1153	D25
2,4-D	054-0003	30	214670	236805	10	1.5986	1.5986	D25
2,4-D	058-0002	31	265553	241937	10	1.9372	1.9372	D25
2,4-D	001-0002	32	292437	230906	10	2.2363	2.2363	D25
2,4-D	006-0002	34	247624	246096	10	1.7752	1.7752	D25
2,4-D	006-0003	35	245350	239899	10	1.8045	1.8045	D25
2,4-D	075-0003	36	308182	222679	10	2.4445	2.4445	D25
2,4-D	074-0002	37	232834	224400	10	1.8308	1.8308	D25
2,4-D	005-0002	38	278231	220641	10	2.2267	2.2267	D25
2,4-D	057-0003	39	264971	230920	10	2.0255	2.0255	D25
2,4-D	002-0002	40	455921	240362	10	3.3531	3.3531	D25
2,4-D	056-0003	42	322836	227211	10	2.5099	2.5099	D25
2,4-D	001-0003	43	330464	195433	10	2.9883	2.9883	D25
2,4-D	057-0002	44	410708	229196	10	3.1673	3.1673	D25
2,4-D	128-0003	45	250483	235672	10	1.8756	1.8756	D25
2,4-D	128-0002	46	553380	223797	10	4.3733	4.3733	D25
2,4-D	074-0003	54	80812	232396	10	0.6086	0.6086	D25
2,4-DCP	046-0001	3	2196	20467	1	0.0017	ND	D25
2,4-DCP	115-0003	7	19615	14072	1	0.0371	0.0371	D25
2,4-DCP	055-0002	8	140801	26125	10	1.4708	1.4708	D25
2,4-DCP	070-0002	10	84705	19572	10	1.1786	1.1786	D25
2,4-DCP	070-0003	11	113758	19840	10	1.5655	1.5655	D25
2,4-DCP	031-0003	12	96224	21864	10	1.1988	1.1988	D25
2,4-DCP	059-0003	13	129992	20389	10	1.7421	1.7421	D25
2,4-DCP	047-0002	14	122974	21539	10	1.5588	1.5588	D25
2,4-DCP	055-0003	15	163408	20854	10	2.1439	2.1439	D25
2,4-DCP	038-0002	16	146190	20887	10	1.9137	1.9137	D25
2,4-DCP	059-0002	18	123136	21182	10	1.5874	1.5874	D25
2,4-DCP	038-0003	19	165479	21408	10	2.1148	2.1148	D25
2,4-DCP	113-0003	20	19384	24361	10	0.2067	0.2067	D25
2,4-DCP	039-0002	21	186128	20967	10	2.4305	2.4305	D25
2,4-DCP	046-0002	22	131647	21343	10	1.6850	1.6850	D25
2,4-DCP	075-0002	23	216075	21206	10	2.7916	2.7916	D25
2,4-DCP	056-0002	24	171690	21982	10	2.1370	2.1370	D25
2,4-DCP	002-0003	26	106707	21307	10	1.3658	1.3658	D25
2,4-DCP	005-0003	27	184565	23873	10	2.1151	2.1151	D25
2,4-DCP	054-0002	28	227184	20717	10	3.0053	3.0053	D25
2,4-DCP	058-0003	29	254586	16278	10	4.2914	4.2914	D25
2,4-DCP	054-0003	30	202326	20737	10	2.6725	2.6725	D25
2,4-DCP	058-0002	31	226820	18998	10	3.2731	3.2731	D25
2,4-DCP	001-0002	32	135336	20801	10	1.7781	1.7781	D25
2,4-DCP	006-0002	34	254563	22496	10	3.1016	3.1016	D25
2,4-DCP	006-0003	35	180288	18366	10	2.6889	2.6889	D25

Table 6 (Cont.). Individual Residue Results from Corn Samples

Analyte	Sample ID[a]	Inj. No	Analyte Peak Area	ISTD Peak Area	Dilution Factor	Uncorrected (µg/g)	Reported (µg/g)[b]	Set ID
2,4-DCP	075-0003	36	311946	17101	10	5.0073	5.0073	D25
2,4-DCP	074-0002	37	227458	19288	10	3.2328	3.2328	D25
2,4-DCP	005-0002	38	194393	13342	10	3.9970	3.9970	D25
2,4-DCP	057-0003	39	241588	20667	10	3.2044	3.2044	D25
2,4-DCP	002-0002	40	149829	20669	10	1.9824	1.9824	D25
2,4-DCP	056-0003	42	297690	19998	10	4.0840	4.0840	D25
2,4-DCP	001-0003	43	153724	19209	10	2.1899	2.1899	D25
2,4-DCP	057-0002	44	337735	22344	10	4.1470	4.1470	D25
2,4-DCP	128-0003	45	61761	21523	10	0.7773	0.7773	D25
2,4-DCP	128-0002	46	111473	26102	10	1.1629	1.1629	D25
2,4-DCP	074-0003	54	120544	16420	10	2.0079	2.0079	D25
2,4-D	008-0001	3	0	259482	1	ND	ND	D26
2,4-D	013-0001	4	0	254676	1	ND	ND	D26
2,4-D	021-0001	5	4259	245162	1	0.0018	ND	D26
2,4-D	025-0001	6	3852	254746	1	0.0014	ND	D26
2,4-D	027-0001	7	6140	247002	1	0.0032	[0.0032]	D26
2,4-D	029-0001	8	0	252533	1	ND	ND	D26
2,4-D	033-0001	10	0	253850	1	ND	ND	D26
2,4-D	037-0001	11	4296	255823	1	0.0017	ND	D26
2,4-D	045-0001	12	0	251635	1	ND	ND	D26
2,4-D	061-0001	13	0	258591	1	ND	ND	D26
2,4-D	063-0001	14	0	255596	1	ND	ND	D26
2,4-D	065-0001	15	0	260900	1	ND	ND	D26
2,4-D	067-0001	16	0	253433	1	ND	ND	D26
2,4-D	069-0001	18	5651	257393	1	0.0026	ND	D26
2,4-D	073-0001	19	5010	257617	1	0.0022	ND	D26
2,4-D	077-0001	20	0	252662	1	ND	ND	D26
2,4-D	082-0001	21	3233	259921	1	0.0009	ND	D26
2,4-D	086-0001	22	3117	253364	1	0.0008	ND	D26
2,4-D	090-0001	23	4462	255838	1	0.0018	ND	D26
2,4-D	094-0001	24	0	254765	1	ND	ND	D26
2,4-D	098-0001	26	0	259122	1	ND	ND	D26
2,4-D	102-0001	27	0	253544	1	ND	ND	D26
2,4-D	106-0001	28	0	246709	1	ND	ND	D26
2,4-D	110-0001	29	0	249216	1	ND	ND	D26
2,4-D	118-0001	30	0	252067	1	ND	ND	D26
2,4-D	120-0001	31	0	252376	1	ND	ND	D26
2,4-D	122-0001	32	0	249943	1	ND	ND	D26
2,4-D	124-0001	34	0	259006	1	ND	ND	D26
2,4-D	126-0001	35	0	249201	1	ND	ND	D26
2,4-D	130-0001	36	3902	253054	1	0.0014	ND	D26
2,4-D	134-0001	37	0	240866	1	ND	ND	D26
2,4-D	138-0001	38	0	240813	1	ND	ND	D26
2,4-D	049-0001	39	12247	249597	1	0.0076	[0.0076]	D26
2,4-D	023-0001	40	5616	244639	1	0.0028	ND	D26
2,4-D	004-0001	42	7433	249698	1	0.0041	[0.0041]	D26
2,4-D	041-0001	43	7534	241089	1	0.0043	[0.0043]	D26
2,4-D	053-0001	44	9710	249399	1	0.0058	[0.0058]	D26
2,4-D	033-0002	45	208647	246987	1	0.1544	0.1544	D26
2,4-D	045-0002	50	24817	238741	1	0.0177	0.0177	D26
2,4-DCP	008-0001	3	2516	27231	1	0.0013	ND	D26

Table 6 (Cont.). Individual Residue Results from Corn Samples

Analyte	Sample ID[a]	Inj. No	Analyte Peak Area	ISTD Peak Area	Dilution Factor	Uncorrected (µg/g)	Reported (µg/g)[b]	Set ID
2,4-DCP	013-0001	4	3448	23835	1	0.0027	ND	D26
2,4-DCP	021-0001	5	3323	24075	1	0.0025	ND	D26
2,4-DCP	025-0001	6	2727	24828	1	0.0017	ND	D26
2,4-DCP	027-0001	7	3897	24565	1	0.0031	[0.0031]	D26
2,4-DCP	029-0001	8	2768	22911	1	0.0020	ND	D26
2,4-DCP	033-0001	10	3906	23258	1	0.0033	[0.0033]	D26
2,4-DCP	037-0001	11	4098	27854	1	0.0028	ND	D26
2,4-DCP	045-0001	12	3478	24374	1	0.0026	ND	D26
2,4-DCP	061-0001	13	5300	24190	1	0.0048	[0.0048]	D26
2,4-DCP	063-0001	14	7993	24409	1	0.0077	[0.0077]	D26
2,4-DCP	065-0001	15	5021	23745	1	0.0045	[0.0045]	D26
2,4-DCP	067-0001	16	6353	24308	1	0.0059	[0.0059]	D26
2,4-DCP	069-0001	18	14400	25008	1	0.0146	0.0146	D26
2,4-DCP	073-0001	19	5517	22453	1	0.0055	[0.0055]	D26
2,4-DCP	077-0001	20	6879	22243	1	0.0072	[0.0072]	D26
2,4-DCP	082-0001	21	5700	23413	1	0.0054	[0.0054]	D26
2,4-DCP	086-0001	22	5705	25683	1	0.0048	[0.0048]	D26
2,4-DCP	090-0001	23	6029	23857	1	0.0057	[0.0057]	D26
2,4-DCP	094-0001	24	3816	22901	1	0.0033	[0.0033]	D26
2,4-DCP	098-0001	26	5632	25030	1	0.0049	[0.0049]	D26
2,4-DCP	102-0001	27	2749	23093	1	0.0020	ND	D26
2,4-DCP	106-0001	28	3536	20931	1	0.0034	[0.0034]	D26
2,4-DCP	110-0001	29	6553	21750	1	0.0070	[0.0070]	D26
2,4-DCP	118-0001	30	4449	28734	1	0.0030	[0.0030]	D26
2,4-DCP	120-0001	31	9003	23294	1	0.0094	[0.0094]	D26
2,4-DCP	122-0001	32	2933	21728	1	0.0024	ND	D26
2,4-DCP	124-0001	34	3258	23146	1	0.0026	ND	D26
2,4-DCP	126-0001	35	3382	21874	1	0.0030	[0.0030]	D26
2,4-DCP	130-0001	36	5615	21192	1	0.0060	[0.0060]	D26
2,4-DCP	134-0001	37	6059	21763	1	0.0064	[0.0064]	D26
2,4-DCP	138-0001	38	2902	20493	1	0.0026	ND	D26
2,4-DCP	049-0001	39	3409	22982	1	0.0028	ND	D26
2,4-DCP	023-0001	40	3876	22043	1	0.0036	[0.0036]	D26
2,4-DCP	004-0001	42	3222	23981	1	0.0024	ND	D26
2,4-DCP	041-0001	43	9745	21024	1	0.0115	0.0115	D26
2,4-DCP	053-0001	44	9626	22251	1	0.0106	0.0106	D26
2,4-DCP	033-0002	45	112901	20920	1	0.1475	0.1475	D26
2,4-DCP	045-0002	50	7192	22191	1	0.0076	[0.0076]	D26
2,4-D	021-0001	3	8024	446540	1	0.0020	ND	D27
2,4-D	033-0003	4	249130	432657	1	0.1050	0.1050	D27
2,4-D	086-0002	5	11051	417913	1	0.0035	[0.0035]	D27
2,4-D	098-0002	6	0	411011	1	ND	ND	D27
2,4-D	098-0003	7	8069	376744	1	0.0026	ND	D27
2,4-D	090-0003	8	62450	374202	1	0.0295	0.0295	D27
2,4-D	090-0002	10	57383	371210	1	0.0272	0.0272	D27
2,4-D	045-0003	11	56143	387024	1	0.0255	0.0255	D27
2,4-D	086-0003	15	24022	355453	1	0.0111	0.0111	D27
2,4-D	126-0002	16	188093	356400	1	0.0962	0.0962	D27
2,4-D	126-0003	18	191966	355187	1	0.0985	0.0985	D27
2,4-D	122-0003	19	324337	337337	1	0.1763	0.1763	D27
2,4-D	122-0002	20	328830	340915	1	0.1769	0.1769	D27

Table 6 (Cont.). Individual Residue Results from Corn Samples

Analyte	Sample ID[a]	Inj. No	Analyte Peak Area	ISTD Peak Area	Dilution Factor	Uncorrected (µg/g)	Reported (µg/g)[b]	Set ID
2,4-D	124-0002	21	381334	334853	1	0.2091	0.2091	D27
2,4-D	094-0003	22	166883	330792	1	0.0919	0.0919	D27
2,4-D	120-0003	23	280460	312557	1	0.1644	0.1644	D27
2,4-D	124-0003	24	338956	330933	1	0.1879	0.1879	D27
2,4-D	138-0002	26	216596	311550	1	0.1271	0.1271	D27
2,4-D	037-0002	27	473729	314230	1	0.2772	0.2772	D27
2,4-D	120-0002	28	426985	304192	1	0.2580	0.2580	D27
2,4-D	082-0003	29	398647	292583	1	0.2504	0.2504	D27
2,4-D	094-0002	30	257669	297603	1	0.1586	0.1586	D27
2,4-D	134-0002	31	361799	288049	1	0.2307	0.2307	D27
2,4-D	118-0003	32	452672	300479	1	0.2770	0.2770	D27
2,4-D	110-0003	34	1227262	294548	1	0.7685	0.7685	D27
2,4-D	138-0003	35	266192	286321	1	0.1704	0.1704	D27
2,4-D	041-0003	36	930127	275196	1	0.6231	0.6231	D27
2,4-D	082-0002	37	585510	266199	1	0.4050	0.4050	D27
2,4-D	037-0003	38	704545	267228	1	0.4858	0.4858	D27
2,4-D	118-0002	39	435882	279748	1	0.2865	0.2865	D27
2,4-D	110-0002	40	1186996	275901	1	0.7935	0.7935	D27
2,4-D	013-0003	42	245160	277832	1	0.1617	0.1617	D27
2,4-D	134-0003	43	494020	255816	1	0.3555	0.3555	D27
2,4-D	130-0002	44	850892	260417	1	0.6024	0.6024	D27
2,4-D	013-0002	45	450845	256420	1	0.3235	0.3235	D27
2,4-D	045-0002	47	24316	253863	1	0.0164	0.0164	D27
2,4-DCP	021-0001	3	3316	23640	1	0.0026	ND	D27
2,4-DCP	033-0003	4	73338	21768	1	0.0944	0.0944	D27
2,4-DCP	086-0002	5	6573	22375	1	0.0070	[0.0070]	D27
2,4-DCP	098-0002	6	2724	21844	1	0.0022	ND	D27
2,4-DCP	098-0003	7	4375	24304	1	0.0038	[0.0038]	D27
2,4-DCP	090-0003	8	26849	20978	1	0.0350	0.0350	D27
2,4-DCP	090-0002	10	20798	23624	1	0.0237	0.0237	D27
2,4-DCP	045-0003	11	15353	21244	1	0.0192	0.0192	D27
2,4-DCP	086-0003	15	25952	22463	1	0.0315	0.0315	D27
2,4-DCP	126-0002	16	18542	22536	1	0.0220	0.0220	D27
2,4-DCP	126-0003	18	16054	23069	1	0.0184	0.0184	D27
2,4-DCP	122-0003	19	24757	20564	1	0.0329	0.0329	D27
2,4-DCP	122-0002	20	28336	22021	1	0.0352	0.0352	D27
2,4-DCP	124-0002	21	31540	21920	1	0.0396	0.0396	D27
2,4-DCP	094-0003	22	99325	22046	1	0.1267	0.1267	D27
2,4-DCP	120-0003	23	27170	22224	1	0.0334	0.0334	D27
2,4-DCP	124-0003	24	26543	23040	1	0.0314	0.0314	D27
2,4-DCP	138-0002	26	428484	19669	1	0.6180	0.6180	D27
2,4-DCP	037-0002	27	143397	24051	1	0.1682	0.1682	D27
2,4-DCP	120-0002	28	33202	21450	1	0.0427	0.0427	D27
2,4-DCP	082-0003	29	181568	20764	1	0.2473	0.2473	D27
2,4-DCP	094-0002	30	160800	23718	1	0.1914	0.1914	D27
2,4-DCP	134-0002	31	232263	20943	1	0.3140	0.3140	D27
2,4-DCP	118-0003	32	45651	23020	1	0.0550	0.0550	D27
2,4-DCP	110-0003	34	235750	21897	1	0.3048	0.3048	D27
2,4-DCP	138-0003	35	548491	23076	1	0.6744	0.6744	D27
2,4-DCP	041-0003	36	407703	20868	1	0.5541	0.5541	D27
2,4-DCP	082-0002	37	291152	20740	1	0.3978	0.3978	D27

Table 6 (Cont.). Individual Residue Results from Corn Samples

Analyte	Sample ID[a]	Inj. No	Analyte Peak Area	ISTD Peak Area	Dilution Factor	Uncorrected (µg/g)	Reported (µg/g)[b]	Set ID
2,4-DCP	037-0003	38	225587	20601	1	0.3100	0.3100	D27
2,4-DCP	118-0002	39	48411	23044	1	0.0584	0.0584	D27
2,4-DCP	110-0002	40	251718	26277	1	0.2710	0.2710	D27
2,4-DCP	013-0003	42	64832	25211	1	0.0718	0.0718	D27
2,4-DCP	134-0003	43	319992	21077	1	0.4303	0.4303	D27
2,4-DCP	130-0002	44	210977	26077	1	0.2287	0.2287	D27
2,4-DCP	013-0002	45	151624	20707	1	0.2068	0.2068	D27
2,4-DCP	045-0002	47	5823	21632	1	0.0063	[0.0063]	D27
2,4-D	021-0001	3	4246	303349	1	0.0015	ND	D28
2,4-D	045-0002	7	30404	276565	1	0.0189	0.0189	D28
2,4-D	102-0003	8	109356	259235	10	0.7546	0.7546	D28
2,4-D	130-0003	9	124903	256080	10	0.8740	0.8740	D28
2,4-D	049-0002	11	192686	292209	10	1.1852	1.1852	D28
2,4-D	106-0002	12	185587	279006	10	1.1957	1.1957	D28
2,4-D	102-0002	13	208874	286680	10	1.3106	1.3106	D28
2,4-D	023-0002	14	117911	268678	10	0.7854	0.7854	D28
2,4-D	008-0003	15	241469	267421	10	1.6267	1.6267	D28
2,4-D	073-0002	16	249462	268145	10	1.6763	1.6763	D28
2,4-D	029-0003	17	165221	271102	10	1.0946	1.0946	D28
2,4-D	023-0003	18	127443	264982	10	0.8617	0.8617	D28
2,4-D	029-0002	20	131465	268485	10	0.8775	0.8775	D28
2,4-D	025-0002	21	209448	274698	10	1.3720	1.3720	D28
2,4-D	027-0003	22	190729	265325	10	1.2930	1.2930	D28
2,4-D	008-0002	23	391119	262110	10	2.6949	2.6949	D28
2,4-D	027-0002	24	193218	262015	10	1.3267	1.3267	D28
2,4-D	049-0003	25	368664	270105	10	2.4641	2.4641	D28
2,4-D	025-0003	26	156877	260392	10	1.0820	1.0820	D28
2,4-D	053-0002	27	369619	262726	10	2.5402	2.5402	D28
2,4-D	073-0003	29	347672	259147	10	2.4219	2.4219	D28
2,4-D	077-0002	30	475847	277248	10	3.1011	3.1011	D28
2,4-D	077-0003	31	462671	266737	10	3.1342	3.1342	D28
2,4-D	004-0003	32	881675	271496	10	5.8767	5.8767	D28
2,4-D	061-0002	33	505850	257868	10	3.5459	3.5459	D28
2,4-D	004-0002	34	1193026	263721	10	8.1905	8.1905	D28
2,4-D	069-0003	35	445781	249575	10	3.2277	3.2277	D28
2,4-D	063-0002	36	597098	264625	10	4.0802	4.0802	D28
2,4-D	021-0002	38	420226	265221	10	2.8621	2.8621	D28
2,4-D	021-0003	39	384256	262980	10	2.6386	2.6386	D28
2,4-D	061-0003	40	911680	275951	10	5.9788	5.9788	D28
2,4-D	067-0002	41	707406	270241	10	4.7351	4.7351	D28
2,4-D	069-0002	42	823010	256881	10	5.7977	5.7977	D28
2,4-D	063-0003	43	940955	257429	10	6.6159	6.6159	D28
2,4-D	067-0003	44	1047128	268843	10	7.0505	7.0505	D28
2,4-D	065-0002	45	1070559	267942	10	7.2327	7.2327	D28
2,4-D	041-0002	54	209973	261940	10	1.4430	1.4430	D28
2,4-D	106-0003	56	126860	259919	10	0.8746	0.8746	D28
2,4-D	053-0003	57	133516	261939	10	0.9139	0.9139	D28
2,4-D	065-0003	9b	190795	383409	100	8.9195	8.9195	D28
2,4-DCP	021-0001	3	3411	27257	1	0.0019	ND	D28
2,4-DCP	045-0002	7	7853	24268	1	0.0076	[0.0076]	D28
2,4-DCP	102-0003	8	97534	17482	10	1.5763	1.5763	D28

Table 6 (Cont.). Individual Residue Results from Corn Samples

Analyte	Sample ID[a]	Inj. No	Analyte Peak Area	ISTD Peak Area	Dilution Factor	Uncorrected (µg/g)	Reported (µg/g)[b]	Set ID
2,4-DCP	130-0003	9	39532	21159	10	0.5168	0.5168	D28
2,4-DCP	049-0002	11	55440	20900	10	0.7407	0.7407	D28
2,4-DCP	106-0002	12	140570	23159	10	1.7164	1.7164	D28
2,4-DCP	102-0002	13	148840	17253	10	2.4465	2.4465	D28
2,4-DCP	023-0002	14	22260	16636	10	0.3654	0.3654	D28
2,4-DCP	008-0003	15	96714	22556	10	1.2076	1.2076	D28
2,4-DCP	073-0002	16	99428	23578	10	1.1874	1.1874	D28
2,4-DCP	029-0003	17	33546	15552	10	0.5992	0.5992	D28
2,4-DCP	023-0003	18	28677	21267	10	0.3683	0.3683	D28
2,4-DCP	029-0002	20	31279	19928	10	0.4315	0.4315	D28
2,4-DCP	025-0002	21	44602	23640	10	0.5220	0.5220	D28
2,4-DCP	027-0003	22	44620	23013	10	0.5369	0.5369	D28
2,4-DCP	008-0002	23	144420	24140	10	1.6915	1.6915	D28
2,4-DCP	027-0002	24	43762	21873	10	0.5546	0.5546	D28
2,4-DCP	049-0003	25	107614	26659	10	1.1359	1.1359	D28
2,4-DCP	025-0003	26	36931	21400	10	0.4761	0.4761	D28
2,4-DCP	053-0002	27	99271	21424	10	1.3063	1.3063	D28
2,4-DCP	073-0003	29	122187	19549	10	1.7679	1.7679	D28
2,4-DCP	077-0002	30	249972	22364	10	3.1748	3.1748	D28
2,4-DCP	077-0003	31	221475	20743	10	3.0319	3.0319	D28
2,4-DCP	004-0003	32	127597	23155	10	1.5567	1.5567	D28
2,4-DCP	061-0002	33	228498	20410	10	3.1799	3.1799	D28
2,4-DCP	004-0002	34	187291	23692	10	2.2405	2.2405	D28
2,4-DCP	069-0003	35	206928	22512	10	2.6079	2.6079	D28
2,4-DCP	063-0002	36	245562	21590	10	3.2309	3.2309	D28
2,4-DCP	021-0002	38	93336	23066	10	1.1387	1.1387	D28
2,4-DCP	021-0003	39	74935	26245	10	0.7986	0.7986	D28
2,4-DCP	061-0003	40	413809	21511	10	5.4760	5.4760	D28
2,4-DCP	067-0002	41	329607	24208	10	3.8709	3.8709	D28
2,4-DCP	069-0002	42	267772	20198	10	3.7686	3.7686	D28
2,4-DCP	063-0003	43	490253	22806	10	6.1212	6.1212	D28
2,4-DCP	067-0003	44	383851	24610	10	4.4368	4.4368	D28
2,4-DCP	065-0002	45	373127	21617	10	4.9117	4.9117	D28
2,4-DCP	041-0002	54	98615	21742	10	1.2784	1.2784	D28
2,4-DCP	106-0003	56	105751	25686	10	1.1589	1.1589	D28
2,4-DCP	053-0003	57	42037	23638	10	0.4911	0.4911	D28
2,4-DCP	065-0003	9b	75474	35866	100	5.8416	5.8416	D28
QUIZALOFOP ACID	020-0001	3	0	--	1	ND	ND	Q01
QUIZALOFOP ACID	003-0001	10	0	--	1	ND	ND	Q01
QUIZALOFOP ACID	007-0001	11	0	--	1	ND	ND	Q01
QUIZALOFOP ACID	011-0001	12	0	--	1	ND	ND	Q01
QUIZALOFOP ACID	022-0001	13	0	--	1	ND	ND	Q01
QUIZALOFOP ACID	024-0001	14	0	--	1	ND	ND	Q01
QUIZALOFOP ACID	026-0001	15	0	--	1	ND	ND	Q01
QUIZALOFOP ACID	028-0001	16	0	--	1	ND	ND	Q01
QUIZALOFOP ACID	032-0001	18	0	--	1	ND	ND	Q01
QUIZALOFOP ACID	036-0001	19	0	--	1	ND	ND	Q01
QUIZALOFOP ACID	040-0001	20	0	--	1	ND	ND	Q01
QUIZALOFOP ACID	044-0001	21	0	--	1	ND	ND	Q01
QUIZALOFOP ACID	048-0001	22	0	--	1	ND	ND	Q01
QUIZALOFOP ACID	052-0001	23	0	--	1	ND	ND	Q01
QUIZALOFOP ACID	060-0001	24	0	--	1	ND	ND	Q01

Table 6 (Cont.). Individual Residue Results from Corn Samples

Analyte	Sample ID[a]	Inj. No	Analyte Peak Area	ISTD Peak Area	Dilution Factor	Uncorrected (µg/g)	Reported (µg/g)[b]	Set ID
QUIZALOFOP ACID	062-0001	26	0	--	1	ND	ND	Q01
QUIZALOFOP ACID	064-0001	27	0	--	1	ND	ND	Q01
QUIZALOFOP ACID	066-0001	28	0	--	1	ND	ND	Q01
QUIZALOFOP ACID	068-0001	29	0	--	1	ND	ND	Q01
QUIZALOFOP ACID	072-0001	30	0	--	1	ND	ND	Q01
QUIZALOFOP ACID	076-0001	31	0	--	1	ND	ND	Q01
QUIZALOFOP ACID	080-0001	32	0	--	1	ND	ND	Q01
QUIZALOFOP ACID	085-0001	34	888	--	1	0.0024	ND	Q01
QUIZALOFOP ACID	089-0001	35	0	--	1	ND	ND	Q01
QUIZALOFOP ACID	093-0001	36	0	--	1	ND	ND	Q01
QUIZALOFOP ACID	097-0001	37	0	--	1	ND	ND	Q01
QUIZALOFOP ACID	101-0001	38	0	--	1	ND	ND	Q01
QUIZALOFOP ACID	105-0001	39	0	--	1	ND	ND	Q01
QUIZALOFOP ACID	109-0001	40	0	--	1	ND	ND	Q01
QUIZALOFOP ACID	117-0001	42	1053	--	1	0.0027	ND	Q01
QUIZALOFOP ACID	119-0001	43	0	--	1	ND	ND	Q01
QUIZALOFOP ACID	121-0001	44	0	--	1	ND	ND	Q01
QUIZALOFOP ACID	123-0001	45	0	--	1	ND	ND	Q01
QUIZALOFOP ACID	125-0001	46	0	--	1	ND	ND	Q01
QUIZALOFOP ACID	129-0001	47	0	--	1	ND	ND	Q01
QUIZALOFOP ACID	133-0001	48	0	--	1	ND	ND	Q01
QUIZALOFOP ACID	137-0001	50	2865	--	1	0.0061	[0.0061]	Q01
QUIZALOFOP ACID	139-0001	51	748	--	1	0.0021	ND	Q01
QUIZALOFOP ACID	139-0002	52	0	--	1	ND	ND	Q01
QUIZALOFOP ACID	139-0003	53	0	--	1	ND	ND	Q01
QUIZALOFOP ETHYL	020-0001	3	0	--	1	ND	ND	Q01
QUIZALOFOP ETHYL	003-0001	10	0	--	1	ND	ND	Q01
QUIZALOFOP ETHYL	007-0001	11	0	--	1	ND	ND	Q01
QUIZALOFOP ETHYL	011-0001	12	0	--	1	ND	ND	Q01
QUIZALOFOP ETHYL	022-0001	13	0	--	1	ND	ND	Q01
QUIZALOFOP ETHYL	024-0001	14	0	--	1	ND	ND	Q01
QUIZALOFOP ETHYL	026-0001	15	0	--	1	ND	ND	Q01
QUIZALOFOP ETHYL	028-0001	16	0	--	1	ND	ND	Q01
QUIZALOFOP ETHYL	032-0001	18	0	--	1	ND	ND	Q01
QUIZALOFOP ETHYL	036-0001	19	0	--	1	ND	ND	Q01
QUIZALOFOP ETHYL	040-0001	20	0	--	1	ND	ND	Q01
QUIZALOFOP ETHYL	044-0001	21	0	--	1	ND	ND	Q01
QUIZALOFOP ETHYL	048-0001	22	0	--	1	ND	ND	Q01
QUIZALOFOP ETHYL	052-0001	23	0	--	1	ND	ND	Q01
QUIZALOFOP ETHYL	060-0001	24	0	--	1	ND	ND	Q01
QUIZALOFOP ETHYL	062-0001	26	0	--	1	ND	ND	Q01
QUIZALOFOP ETHYL	064-0001	27	0	--	1	ND	ND	Q01
QUIZALOFOP ETHYL	066-0001	28	0	--	1	ND	ND	Q01
QUIZALOFOP ETHYL	068-0001	29	0	--	1	ND	ND	Q01
QUIZALOFOP ETHYL	072-0001	30	0	--	1	ND	ND	Q01
QUIZALOFOP ETHYL	076-0001	31	0	--	1	ND	ND	Q01
QUIZALOFOP ETHYL	080-0001	32	0	--	1	ND	ND	Q01
QUIZALOFOP ETHYL	085-0001	34	0	--	1	ND	ND	Q01
QUIZALOFOP ETHYL	089-0001	35	0	--	1	ND	ND	Q01
QUIZALOFOP ETHYL	093-0001	36	0	--	1	ND	ND	Q01
QUIZALOFOP ETHYL	097-0001	37	0	--	1	ND	ND	Q01

Table 6 (Cont.). Individual Residue Results from Corn Samples

Analyte	Sample ID[a]	Inj. No	Analyte Peak Area	ISTD Peak Area	Dilution Factor	Uncorrected (µg/g)	Reported (µg/g)[b]	Set ID
QUIZALOFOP ETHYL	101-0001	38	0	--	1	ND	ND	Q01
QUIZALOFOP ETHYL	105-0001	39	0	--	1	ND	ND	Q01
QUIZALOFOP ETHYL	109-0001	40	0	--	1	ND	ND	Q01
QUIZALOFOP ETHYL	117-0001	42	298	--	1	0.0011	ND	Q01
QUIZALOFOP ETHYL	119-0001	43	0	--	1	ND	ND	Q01
QUIZALOFOP ETHYL	121-0001	44	0	--	1	ND	ND	Q01
QUIZALOFOP ETHYL	123-0001	45	0	--	1	ND	ND	Q01
QUIZALOFOP ETHYL	125-0001	46	0	--	1	ND	ND	Q01
QUIZALOFOP ETHYL	129-0001	47	0	--	1	ND	ND	Q01
QUIZALOFOP ETHYL	133-0001	48	0	--	1	ND	ND	Q01
QUIZALOFOP ETHYL	137-0001	50	1012	--	1	0.0030	[0.0030]	Q01
QUIZALOFOP ETHYL	139-0001	51	0	--	1	ND	ND	Q01
QUIZALOFOP ETHYL	139-0002	52	0	--	1	ND	ND	Q01
QUIZALOFOP ETHYL	139-0003	53	0	--	1	ND	ND	Q01
QUIZALOFOP ACID	020-0001	3	0	--	1	ND	ND	Q02
QUIZALOFOP ACID	140-0001	10	0	--	1	ND	ND	Q02
QUIZALOFOP ACID	140-0002	11	0	--	1	ND	ND	Q02
QUIZALOFOP ACID	140-0003	12	0	--	1	ND	ND	Q02
QUIZALOFOP ACID	148-0001	13	0	--	1	ND	ND	Q02
QUIZALOFOP ACID	148-0002	14	0	--	1	ND	ND	Q02
QUIZALOFOP ACID	148-0003	15	0	--	1	ND	ND	Q02
QUIZALOFOP ACID	149-0001	16	0	--	1	ND	ND	Q02
QUIZALOFOP ACID	149-0002	17	0	--	1	ND	ND	Q02
QUIZALOFOP ACID	149-0003	18	0	--	1	ND	ND	Q02
QUIZALOFOP ACID	003-0002	19	0	--	1	ND	ND	Q02
QUIZALOFOP ACID	003-0003	20	0	--	1	ND	ND	Q02
QUIZALOFOP ACID	007-0002	21	0	--	1	ND	ND	Q02
QUIZALOFOP ACID	007-0003	22	0	--	1	ND	ND	Q02
QUIZALOFOP ACID	011-0002	23	0	--	1	ND	ND	Q02
QUIZALOFOP ACID	011-0003	25	0	--	1	ND	ND	Q02
QUIZALOFOP ACID	020-0002	26	0	--	1	ND	ND	Q02
QUIZALOFOP ACID	020-0003	27	0	--	1	ND	ND	Q02
QUIZALOFOP ACID	022-0002	28	0	--	1	ND	ND	Q02
QUIZALOFOP ACID	022-0003	29	0	--	1	ND	ND	Q02
QUIZALOFOP ACID	024-0002	30	0	--	1	ND	ND	Q02
QUIZALOFOP ACID	024-0003	31	0	--	1	ND	ND	Q02
QUIZALOFOP ACID	026-0002	33	409	--	1	0.0007	ND	Q02
QUIZALOFOP ACID	026-0003	34	0	--	1	ND	ND	Q02
QUIZALOFOP ACID	028-0002	35	0	--	1	ND	ND	Q02
QUIZALOFOP ACID	028-0003	36	0	--	1	ND	ND	Q02
QUIZALOFOP ACID	032-0002	37	0	--	1	ND	ND	Q02
QUIZALOFOP ACID	032-0003	38	0	--	1	ND	ND	Q02
QUIZALOFOP ACID	036-0002	39	0	--	1	ND	ND	Q02
QUIZALOFOP ACID	036-0003	41	2136	--	1	0.0041	[0.0041]	Q02
QUIZALOFOP ACID	040-0002	42	0	--	1	ND	ND	Q02
QUIZALOFOP ACID	040-0003	43	0	--	1	ND	ND	Q02
QUIZALOFOP ACID	044-0002	44	435	--	1	0.0008	ND	Q02
QUIZALOFOP ACID	044-0003	45	0	--	1	ND	ND	Q02
QUIZALOFOP ACID	048-0002	46	0	--	1	ND	ND	Q02
QUIZALOFOP ACID	048-0003	47	0	--	1	ND	ND	Q02
QUIZALOFOP ACID	052-0002	49	0	--	1	ND	ND	Q02

Table 6 (Cont.). Individual Residue Results from Corn Samples

Analyte	Sample ID[a]	Inj. No	Analyte Peak Area	ISTD Peak Area	Dilution Factor	Uncorrected (µg/g)	Reported (µg/g)[b]	Set ID
QUIZALOFOP ACID	052-0003	50	0	--	1	ND	ND	Q02
QUIZALOFOP ACID	060-0002	51	0	--	1	ND	ND	Q02
QUIZALOFOP ACID	060-0003	52	0	--	1	ND	ND	Q02
QUIZALOFOP ACID	062-0002	53	0	--	1	ND	ND	Q02
QUIZALOFOP ACID	062-0003	54	0	--	1	ND	ND	Q02
QUIZALOFOP ETHYL	020-0001	3	0	--	1	ND	ND	Q02
QUIZALOFOP ETHYL	140-0001	10	0	--	1	ND	ND	Q02
QUIZALOFOP ETHYL	140-0002	11	0	--	1	ND	ND	Q02
QUIZALOFOP ETHYL	140-0003	12	0	--	1	ND	ND	Q02
QUIZALOFOP ETHYL	148-0001	13	0	--	1	ND	ND	Q02
QUIZALOFOP ETHYL	148-0002	14	0	--	1	ND	ND	Q02
QUIZALOFOP ETHYL	148-0003	15	0	--	1	ND	ND	Q02
QUIZALOFOP ETHYL	149-0001	16	0	--	1	ND	ND	Q02
QUIZALOFOP ETHYL	149-0002	17	0	--	1	ND	ND	Q02
QUIZALOFOP ETHYL	149-0003	18	0	--	1	ND	ND	Q02
QUIZALOFOP ETHYL	003-0002	19	0	--	1	ND	ND	Q02
QUIZALOFOP ETHYL	003-0003	20	0	--	1	ND	ND	Q02
QUIZALOFOP ETHYL	007-0002	21	0	--	1	ND	ND	Q02
QUIZALOFOP ETHYL	007-0003	22	0	--	1	ND	ND	Q02
QUIZALOFOP ETHYL	011-0002	23	0	--	1	ND	ND	Q02
QUIZALOFOP ETHYL	011-0003	25	0	--	1	ND	ND	Q02
QUIZALOFOP ETHYL	020-0002	26	0	--	1	ND	ND	Q02
QUIZALOFOP ETHYL	020-0003	27	0	--	1	ND	ND	Q02
QUIZALOFOP ETHYL	022-0002	28	0	--	1	ND	ND	Q02
QUIZALOFOP ETHYL	022-0003	29	0	--	1	ND	ND	Q02
QUIZALOFOP ETHYL	024-0002	30	0	--	1	ND	ND	Q02
QUIZALOFOP ETHYL	024-0003	31	0	--	1	ND	ND	Q02
QUIZALOFOP ETHYL	026-0002	33	336	--	1	0.0002	ND	Q02
QUIZALOFOP ETHYL	026-0003	34	0	--	1	ND	ND	Q02
QUIZALOFOP ETHYL	028-0002	35	0	--	1	ND	ND	Q02
QUIZALOFOP ETHYL	028-0003	36	0	--	1	ND	ND	Q02
QUIZALOFOP ETHYL	032-0002	37	0	--	1	ND	ND	Q02
QUIZALOFOP ETHYL	032-0003	38	0	--	1	ND	ND	Q02
QUIZALOFOP ETHYL	036-0002	39	0	--	1	ND	ND	Q02
QUIZALOFOP ETHYL	036-0003	41	0	--	1	ND	ND	Q02
QUIZALOFOP ETHYL	040-0002	42	0	--	1	ND	ND	Q02
QUIZALOFOP ETHYL	040-0003	43	0	--	1	ND	ND	Q02
QUIZALOFOP ETHYL	044-0002	44	0	--	1	ND	ND	Q02
QUIZALOFOP ETHYL	044-0003	45	0	--	1	ND	ND	Q02
QUIZALOFOP ETHYL	048-0002	46	0	--	1	ND	ND	Q02
QUIZALOFOP ETHYL	048-0003	47	0	--	1	ND	ND	Q02
QUIZALOFOP ETHYL	052-0002	49	1131	--	1	0.0023	ND	Q02
QUIZALOFOP ETHYL	052-0003	50	0	--	1	ND	ND	Q02
QUIZALOFOP ETHYL	060-0002	51	0	--	1	ND	ND	Q02
QUIZALOFOP ETHYL	060-0003	52	0	--	1	ND	ND	Q02
QUIZALOFOP ETHYL	062-0002	53	0	--	1	ND	ND	Q02
QUIZALOFOP ETHYL	062-0003	54	0	--	1	ND	ND	Q02
QUIZALOFOP ACID	020-0001	3	0	--	1	ND	ND	Q03
QUIZALOFOP ACID	064-0002	9	672	--	1	0.0024	ND	Q03
QUIZALOFOP ACID	064-0003	10	0	--	1	ND	ND	Q03
QUIZALOFOP ACID	066-0002	11	0	--	1	ND	ND	Q03

Table 6 (Cont.). Individual Residue Results from Corn Samples

Analyte	Sample ID[a]	Inj. No	Analyte Peak Area	ISTD Peak Area	Dilution Factor	Uncorrected (µg/g)	Reported (µg/g)[b]	Set ID
QUIZALOFOP ACID	066-0003	13	0	--	1	ND	ND	Q03
QUIZALOFOP ACID	068-0002	14	0	--	1	ND	ND	Q03
QUIZALOFOP ACID	068-0003	15	0	--	1	ND	ND	Q03
QUIZALOFOP ACID	072-0002	16	0	--	1	ND	ND	Q03
QUIZALOFOP ACID	072-0003	17	0	--	1	ND	ND	Q03
QUIZALOFOP ACID	076-0002	18	0	--	1	ND	ND	Q03
QUIZALOFOP ACID	076-0003	19	0	--	1	ND	ND	Q03
QUIZALOFOP ACID	080-0002	20	0	--	1	ND	ND	Q03
QUIZALOFOP ACID	080-0003	21	0	--	1	ND	ND	Q03
QUIZALOFOP ACID	085-0002	22	0	--	1	ND	ND	Q03
QUIZALOFOP ACID	085-0003	24	0	--	1	ND	ND	Q03
QUIZALOFOP ACID	089-0002	25	0	--	1	ND	ND	Q03
QUIZALOFOP ACID	089-0003	26	0	--	1	ND	ND	Q03
QUIZALOFOP ACID	093-0002	27	0	--	1	ND	ND	Q03
QUIZALOFOP ACID	093-0003	28	0	--	1	ND	ND	Q03
QUIZALOFOP ACID	097-0002	29	0	--	1	ND	ND	Q03
QUIZALOFOP ACID	097-0003	30	0	--	1	ND	ND	Q03
QUIZALOFOP ACID	101-0002	31	0	--	1	ND	ND	Q03
QUIZALOFOP ACID	101-0003	32	0	--	1	ND	ND	Q03
QUIZALOFOP ACID	105-0002	33	0	--	1	ND	ND	Q03
QUIZALOFOP ACID	105-0003	35	0	--	1	ND	ND	Q03
QUIZALOFOP ACID	109-0002	36	0	--	1	ND	ND	Q03
QUIZALOFOP ACID	109-0003	37	0	--	1	ND	ND	Q03
QUIZALOFOP ACID	117-0002	38	0	--	1	ND	ND	Q03
QUIZALOFOP ACID	117-0003	39	0	--	1	ND	ND	Q03
QUIZALOFOP ACID	119-0002	40	0	--	1	ND	ND	Q03
QUIZALOFOP ACID	119-0003	41	0	--	1	ND	ND	Q03
QUIZALOFOP ACID	121-0002	42	0	--	1	ND	ND	Q03
QUIZALOFOP ACID	121-0003	43	0	--	1	ND	ND	Q03
QUIZALOFOP ACID	123-0002	44	0	--	1	ND	ND	Q03
QUIZALOFOP ACID	123-0003	46	0	--	1	ND	ND	Q03
QUIZALOFOP ACID	125-0002	47	0	--	1	ND	ND	Q03
QUIZALOFOP ACID	125-0003	48	0	--	1	ND	ND	Q03
QUIZALOFOP ACID	129-0002	49	0	--	1	ND	ND	Q03
QUIZALOFOP ACID	129-0003	50	0	--	1	ND	ND	Q03
QUIZALOFOP ACID	133-0002	51	0	--	1	ND	ND	Q03
QUIZALOFOP ACID	133-0003	52	0	--	1	ND	ND	Q03
QUIZALOFOP ACID	137-0002	53	0	--	1	ND	ND	Q03
QUIZALOFOP ACID	137-0003	54	0	--	1	ND	ND	Q03
QUIZALOFOP ACID	011-0006	55	0	--	1	ND	ND	Q03
QUIZALOFOP ACID	011-0007	57	980	--	1	0.0032	[0.0032]	Q03
QUIZALOFOP ACID	080-0006	58	0	--	1	ND	ND	Q03
QUIZALOFOP ACID	080-0007	59	0	--	1	ND	ND	Q03
QUIZALOFOP ACID	139-0004	60	0	--	1	ND	ND	Q03
QUIZALOFOP ACID	139-0005	61	0	--	1	ND	ND	Q03
QUIZALOFOP ACID	139-0006	62	0	--	1	ND	ND	Q03
QUIZALOFOP ACID	140-0004	63	0	--	1	ND	ND	Q03
QUIZALOFOP ACID	140-0005	64	0	--	1	ND	ND	Q03
QUIZALOFOP ACID	140-0006	65	0	--	1	ND	ND	Q03
QUIZALOFOP ACID	148-0004	66	0	--	1	ND	ND	Q03
QUIZALOFOP ACID	148-0005	68	1738	--	1	0.0050	[0.0050]	Q03

Table 6 (Cont.). Individual Residue Results from Corn Samples

Analyte	Sample ID[a]	Inj. No	Analyte Peak Area	ISTD Peak Area	Dilution Factor	Uncorrected (µg/g)	Reported (µg/g)[b]	Set ID
QUIZALOFOP ACID	148-0006	69	0	--	1	ND	ND	Q03
QUIZALOFOP ACID	149-0004	70	0	--	1	ND	ND	Q03
QUIZALOFOP ACID	149-0005	71	0	--	1	ND	ND	Q03
QUIZALOFOP ACID	149-0006	72	0	--	1	ND	ND	Q03
QUIZALOFOP ETHYL	020-0001	3	0	--	1	ND	ND	Q03
QUIZALOFOP ETHYL	064-0002	9	426	--	1	0.0019	ND	Q03
QUIZALOFOP ETHYL	064-0003	10	0	--	1	ND	ND	Q03
QUIZALOFOP ETHYL	066-0002	11	0	--	1	ND	ND	Q03
QUIZALOFOP ETHYL	066-0003	13	0	--	1	ND	ND	Q03
QUIZALOFOP ETHYL	068-0002	14	0	--	1	ND	ND	Q03
QUIZALOFOP ETHYL	068-0003	15	0	--	1	ND	ND	Q03
QUIZALOFOP ETHYL	072-0002	16	0	--	1	ND	ND	Q03
QUIZALOFOP ETHYL	072-0003	17	0	--	1	ND	ND	Q03
QUIZALOFOP ETHYL	076-0002	18	0	--	1	ND	ND	Q03
QUIZALOFOP ETHYL	076-0003	19	0	--	1	ND	ND	Q03
QUIZALOFOP ETHYL	080-0002	20	0	--	1	ND	ND	Q03
QUIZALOFOP ETHYL	080-0003	21	0	--	1	ND	ND	Q03
QUIZALOFOP ETHYL	085-0002	22	0	--	1	ND	ND	Q03
QUIZALOFOP ETHYL	085-0003	24	0	--	1	ND	ND	Q03
QUIZALOFOP ETHYL	089-0002	25	0	--	1	ND	ND	Q03
QUIZALOFOP ETHYL	089-0003	26	0	--	1	ND	ND	Q03
QUIZALOFOP ETHYL	093-0002	27	0	--	1	ND	ND	Q03
QUIZALOFOP ETHYL	093-0003	28	0	--	1	ND	ND	Q03
QUIZALOFOP ETHYL	097-0002	29	0	--	1	ND	ND	Q03
QUIZALOFOP ETHYL	097-0003	30	0	--	1	ND	ND	Q03
QUIZALOFOP ETHYL	101-0002	31	0	--	1	ND	ND	Q03
QUIZALOFOP ETHYL	101-0003	32	0	--	1	ND	ND	Q03
QUIZALOFOP ETHYL	105-0002	33	0	--	1	ND	ND	Q03
QUIZALOFOP ETHYL	105-0003	35	0	--	1	ND	ND	Q03
QUIZALOFOP ETHYL	109-0002	36	0	--	1	ND	ND	Q03
QUIZALOFOP ETHYL	109-0003	37	0	--	1	ND	ND	Q03
QUIZALOFOP ETHYL	117-0002	38	0	--	1	ND	ND	Q03
QUIZALOFOP ETHYL	117-0003	39	0	--	1	ND	ND	Q03
QUIZALOFOP ETHYL	119-0002	40	0	--	1	ND	ND	Q03
QUIZALOFOP ETHYL	119-0003	41	0	--	1	ND	ND	Q03
QUIZALOFOP ETHYL	121-0002	42	0	--	1	ND	ND	Q03
QUIZALOFOP ETHYL	121-0003	43	0	--	1	ND	ND	Q03
QUIZALOFOP ETHYL	123-0002	44	0	--	1	ND	ND	Q03
QUIZALOFOP ETHYL	123-0003	46	0	--	1	ND	ND	Q03
QUIZALOFOP ETHYL	125-0002	47	0	--	1	ND	ND	Q03
QUIZALOFOP ETHYL	125-0003	48	0	--	1	ND	ND	Q03
QUIZALOFOP ETHYL	129-0002	49	0	--	1	ND	ND	Q03
QUIZALOFOP ETHYL	129-0003	50	0	--	1	ND	ND	Q03
QUIZALOFOP ETHYL	133-0002	51	0	--	1	ND	ND	Q03
QUIZALOFOP ETHYL	133-0003	52	0	--	1	ND	ND	Q03
QUIZALOFOP ETHYL	137-0002	53	0	--	1	ND	ND	Q03
QUIZALOFOP ETHYL	137-0003	54	0	--	1	ND	ND	Q03
QUIZALOFOP ETHYL	011-0006	55	0	--	1	ND	ND	Q03
QUIZALOFOP ETHYL	011-0007	57	284	--	1	0.0015	ND	Q03
QUIZALOFOP ETHYL	080-0006	58	0	--	1	ND	ND	Q03
QUIZALOFOP ETHYL	080-0007	59	0	--	1	ND	ND	Q03

Table 6 (Cont.). Individual Residue Results from Corn Samples

Analyte	Sample ID[a]	Inj. No	Analyte Peak Area	ISTD Peak Area	Dilution Factor	Uncorrected (µg/g)	Reported (µg/g)[b]	Set ID
QUIZALOFOP ETHYL	139-0004	60	0	--	1	ND	ND	Q03
QUIZALOFOP ETHYL	139-0005	61	0	--	1	ND	ND	Q03
QUIZALOFOP ETHYL	139-0006	62	0	--	1	ND	ND	Q03
QUIZALOFOP ETHYL	140-0004	63	0	--	1	ND	ND	Q03
QUIZALOFOP ETHYL	140-0005	64	0	--	1	ND	ND	Q03
QUIZALOFOP ETHYL	140-0006	65	0	--	1	ND	ND	Q03
QUIZALOFOP ETHYL	148-0004	66	0	--	1	ND	ND	Q03
QUIZALOFOP ETHYL	148-0005	68	1062	--	1	0.0037	[0.0037]	Q03
QUIZALOFOP ETHYL	148-0006	69	0	--	1	ND	ND	Q03
QUIZALOFOP ETHYL	149-0004	70	0	--	1	ND	ND	Q03
QUIZALOFOP ETHYL	149-0005	71	0	--	1	ND	ND	Q03
QUIZALOFOP ETHYL	149-0006	72	0	--	1	ND	ND	Q03
QUIZALOFOP ACID	157-0001	3	0	--	1	ND	ND	Q04
QUIZALOFOP ACID	001-0001	10	0	--	1	ND	ND	Q04
QUIZALOFOP ACID	002-0001	11	0	--	1	ND	ND	Q04
QUIZALOFOP ACID	005-0001	12	0	--	1	ND	ND	Q04
QUIZALOFOP ACID	006-0001	13	0	--	1	ND	ND	Q04
QUIZALOFOP ACID	009-0001	14	0	--	1	ND	ND	Q04
QUIZALOFOP ACID	010-0001	15	0	--	1	ND	ND	Q04
QUIZALOFOP ACID	014-0001	16	0	--	1	ND	ND	Q04
QUIZALOFOP ACID	015-0001	18	0	--	1	ND	ND	Q04
QUIZALOFOP ACID	016-0001	19	0	--	1	ND	ND	Q04
QUIZALOFOP ACID	017-0001	20	0	--	1	ND	ND	Q04
QUIZALOFOP ACID	018-0001	21	0	--	1	ND	ND	Q04
QUIZALOFOP ACID	019-0001	22	0	--	1	ND	ND	Q04
QUIZALOFOP ACID	030-0001	23	0	--	1	ND	ND	Q04
QUIZALOFOP ACID	031-0001	24	0	--	1	ND	ND	Q04
QUIZALOFOP ACID	034-0001	26	0	--	1	ND	ND	Q04
QUIZALOFOP ACID	035-0001	27	0	--	1	ND	ND	Q04
QUIZALOFOP ACID	038-0001	28	0	--	1	ND	ND	Q04
QUIZALOFOP ACID	039-0001	29	0	--	1	ND	ND	Q04
QUIZALOFOP ACID	042-0001	30	0	--	1	ND	ND	Q04
QUIZALOFOP ACID	043-0001	31	0	--	1	ND	ND	Q04
QUIZALOFOP ACID	046-0001	32	0	--	1	ND	ND	Q04
QUIZALOFOP ACID	047-0001	34	0	--	1	ND	ND	Q04
QUIZALOFOP ACID	050-0001	35	0	--	1	ND	ND	Q04
QUIZALOFOP ACID	051-0001	36	0	--	1	ND	ND	Q04
QUIZALOFOP ACID	054-0001	37	0	--	1	ND	ND	Q04
QUIZALOFOP ACID	055-0001	38	0	--	1	ND	ND	Q04
QUIZALOFOP ACID	056-0001	39	0	--	1	ND	ND	Q04
QUIZALOFOP ACID	057-0001	40	0	--	1	ND	ND	Q04
QUIZALOFOP ACID	058-0001	42	685	--	1	0.0011	ND	Q04
QUIZALOFOP ACID	059-0001	43	0	--	1	ND	ND	Q04
QUIZALOFOP ACID	070-0001	44	0	--	1	ND	ND	Q04
QUIZALOFOP ACID	071-0001	45	0	--	1	ND	ND	Q04
QUIZALOFOP ACID	074-0001	46	344	--	1	ND	ND	Q04
QUIZALOFOP ACID	075-0001	47	0	--	1	ND	ND	Q04
QUIZALOFOP ACID	079-0001	48	0	--	1	ND	ND	Q04
QUIZALOFOP ACID	083-0001	50	550	--	1	0.0006	ND	Q04
QUIZALOFOP ACID	084-0001	51	0	--	1	ND	ND	Q04
QUIZALOFOP ACID	087-0001	52	0	--	1	ND	ND	Q04

Table 6 (Cont.). Individual Residue Results from Corn Samples

Analyte	Sample ID[a]	Inj. No	Analyte Peak Area	ISTD Peak Area	Dilution Factor	Uncorrected (µg/g)	Reported (µg/g)[b]	Set ID
QUIZALOFOP ACID	088-0001	53	0	--	1	ND	ND	Q04
QUIZALOFOP ACID	091-0001	54	0	--	1	ND	ND	Q04
QUIZALOFOP ACID	092-0001	55	0	--	1	ND	ND	Q04
QUIZALOFOP ETHYL	157-0001	3	0	--	1	ND	ND	Q04
QUIZALOFOP ETHYL	001-0001	10	0	--	1	ND	ND	Q04
QUIZALOFOP ETHYL	002-0001	11	0	--	1	ND	ND	Q04
QUIZALOFOP ETHYL	005-0001	12	0	--	1	ND	ND	Q04
QUIZALOFOP ETHYL	006-0001	13	0	--	1	ND	ND	Q04
QUIZALOFOP ETHYL	009-0001	14	0	--	1	ND	ND	Q04
QUIZALOFOP ETHYL	010-0001	15	0	--	1	ND	ND	Q04
QUIZALOFOP ETHYL	014-0001	16	0	--	1	ND	ND	Q04
QUIZALOFOP ETHYL	015-0001	18	0	--	1	ND	ND	Q04
QUIZALOFOP ETHYL	016-0001	19	0	--	1	ND	ND	Q04
QUIZALOFOP ETHYL	017-0001	20	0	--	1	ND	ND	Q04
QUIZALOFOP ETHYL	018-0001	21	0	--	1	ND	ND	Q04
QUIZALOFOP ETHYL	019-0001	22	0	--	1	ND	ND	Q04
QUIZALOFOP ETHYL	030-0001	23	0	--	1	ND	ND	Q04
QUIZALOFOP ETHYL	031-0001	24	0	--	1	ND	ND	Q04
QUIZALOFOP ETHYL	034-0001	26	0	--	1	ND	ND	Q04
QUIZALOFOP ETHYL	035-0001	27	0	--	1	ND	ND	Q04
QUIZALOFOP ETHYL	038-0001	28	0	--	1	ND	ND	Q04
QUIZALOFOP ETHYL	039-0001	29	0	--	1	ND	ND	Q04
QUIZALOFOP ETHYL	042-0001	30	0	--	1	ND	ND	Q04
QUIZALOFOP ETHYL	043-0001	31	0	--	1	ND	ND	Q04
QUIZALOFOP ETHYL	046-0001	32	0	--	1	ND	ND	Q04
QUIZALOFOP ETHYL	047-0001	34	0	--	1	ND	ND	Q04
QUIZALOFOP ETHYL	050-0001	35	0	--	1	ND	ND	Q04
QUIZALOFOP ETHYL	051-0001	36	0	--	1	ND	ND	Q04
QUIZALOFOP ETHYL	054-0001	37	0	--	1	ND	ND	Q04
QUIZALOFOP ETHYL	055-0001	38	0	--	1	ND	ND	Q04
QUIZALOFOP ETHYL	056-0001	39	0	--	1	ND	ND	Q04
QUIZALOFOP ETHYL	057-0001	40	0	--	1	ND	ND	Q04
QUIZALOFOP ETHYL	058-0001	42	627	--	1	0.0011	ND	Q04
QUIZALOFOP ETHYL	059-0001	43	0	--	1	ND	ND	Q04
QUIZALOFOP ETHYL	070-0001	44	0	--	1	ND	ND	Q04
QUIZALOFOP ETHYL	071-0001	45	0	--	1	ND	ND	Q04
QUIZALOFOP ETHYL	074-0001	46	0	--	1	ND	ND	Q04
QUIZALOFOP ETHYL	075-0001	47	0	--	1	ND	ND	Q04
QUIZALOFOP ETHYL	079-0001	48	0	--	1	ND	ND	Q04
QUIZALOFOP ETHYL	083-0001	50	995	--	1	0.0017	ND	Q04
QUIZALOFOP ETHYL	084-0001	51	0	--	1	ND	ND	Q04
QUIZALOFOP ETHYL	087-0001	52	0	--	1	ND	ND	Q04
QUIZALOFOP ETHYL	088-0001	53	0	--	1	ND	ND	Q04
QUIZALOFOP ETHYL	091-0001	54	0	--	1	ND	ND	Q04
QUIZALOFOP ETHYL	092-0001	55	0	--	1	ND	ND	Q04
QUIZALOFOP ACID	157-0001	3	0	--	1	ND	ND	Q05
QUIZALOFOP ACID	095-0001	10	0	--	1	ND	ND	Q05
QUIZALOFOP ACID	096-0001	11	0	--	1	ND	ND	Q05
QUIZALOFOP ACID	099-0001	12	0	--	1	ND	ND	Q05
QUIZALOFOP ACID	100-0001	13	0	--	1	ND	ND	Q05
QUIZALOFOP ACID	103-0001	14	0	--	1	ND	ND	Q05

Table 6 (Cont.). Individual Residue Results from Corn Samples

Analyte	Sample ID[a]	Inj. No	Analyte Peak Area	ISTD Peak Area	Dilution Factor	Uncorrected (µg/g)	Reported (µg/g)[b]	Set ID
QUIZALOFOP ACID	104-0001	15	0	--	1	ND	ND	Q05
QUIZALOFOP ACID	107-0001	16	0	--	1	ND	ND	Q05
QUIZALOFOP ACID	108-0001	18	0	--	1	ND	ND	Q05
QUIZALOFOP ACID	111-0001	19	0	--	1	ND	ND	Q05
QUIZALOFOP ACID	112-0001	20	0	--	1	ND	ND	Q05
QUIZALOFOP ACID	113-0001	21	0	--	1	ND	ND	Q05
QUIZALOFOP ACID	114-0001	22	0	--	1	ND	ND	Q05
QUIZALOFOP ACID	115-0001	23	0	--	1	ND	ND	Q05
QUIZALOFOP ACID	116-0001	24	0	--	1	ND	ND	Q05
QUIZALOFOP ACID	127-0001	26	0	--	1	ND	ND	Q05
QUIZALOFOP ACID	128-0001	27	0	--	1	ND	ND	Q05
QUIZALOFOP ACID	131-0001	28	0	--	1	ND	ND	Q05
QUIZALOFOP ACID	132-0001	29	0	--	1	ND	ND	Q05
QUIZALOFOP ACID	135-0001	30	0	--	1	ND	ND	Q05
QUIZALOFOP ACID	136-0001	31	0	--	1	ND	ND	Q05
QUIZALOFOP ACID	001-0002	32	0	--	1	ND	ND	Q05
QUIZALOFOP ACID	001-0003	34	0	--	1	ND	ND	Q05
QUIZALOFOP ACID	002-0002	35	0	--	1	ND	ND	Q05
QUIZALOFOP ACID	002-0003	36	0	--	1	ND	ND	Q05
QUIZALOFOP ACID	005-0002	37	0	--	1	ND	ND	Q05
QUIZALOFOP ACID	005-0003	38	0	--	1	ND	ND	Q05
QUIZALOFOP ACID	006-0002	39	0	--	1	ND	ND	Q05
QUIZALOFOP ACID	006-0003	40	0	--	1	ND	ND	Q05
QUIZALOFOP ACID	009-0002	42	0	--	1	ND	ND	Q05
QUIZALOFOP ACID	009-0003	43	0	--	1	ND	ND	Q05
QUIZALOFOP ACID	010-0002	44	0	--	1	ND	ND	Q05
QUIZALOFOP ACID	010-0003	45	0	--	1	ND	ND	Q05
QUIZALOFOP ACID	014-0002	46	0	--	1	ND	ND	Q05
QUIZALOFOP ACID	014-0003	47	0	--	1	ND	ND	Q05
QUIZALOFOP ACID	015-0002	48	0	--	1	ND	ND	Q05
QUIZALOFOP ACID	015-0003	50	517	--	1	0.0023	ND	Q05
QUIZALOFOP ACID	016-0002	51	0	--	1	ND	ND	Q05
QUIZALOFOP ACID	016-0003	52	0	--	1	ND	ND	Q05
QUIZALOFOP ACID	017-0002	53	0	--	1	ND	ND	Q05
QUIZALOFOP ACID	017-0003	54	0	--	1	ND	ND	Q05
QUIZALOFOP ETHYL	157-0001	3	0	--	1	ND	ND	Q05
QUIZALOFOP ETHYL	095-0001	10	0	--	1	ND	ND	Q05
QUIZALOFOP ETHYL	096-0001	11	0	--	1	ND	ND	Q05
QUIZALOFOP ETHYL	099-0001	12	0	--	1	ND	ND	Q05
QUIZALOFOP ETHYL	100-0001	13	0	--	1	ND	ND	Q05
QUIZALOFOP ETHYL	103-0001	14	0	--	1	ND	ND	Q05
QUIZALOFOP ETHYL	104-0001	15	0	--	1	ND	ND	Q05
QUIZALOFOP ETHYL	107-0001	16	0	--	1	ND	ND	Q05
QUIZALOFOP ETHYL	108-0001	18	0	--	1	ND	ND	Q05
QUIZALOFOP ETHYL	111-0001	19	0	--	1	ND	ND	Q05
QUIZALOFOP ETHYL	112-0001	20	1183	--	1	0.0031	[0.0031]	Q05
QUIZALOFOP ETHYL	113-0001	21	0	--	1	ND	ND	Q05
QUIZALOFOP ETHYL	114-0001	22	0	--	1	ND	ND	Q05
QUIZALOFOP ETHYL	115-0001	23	0	--	1	ND	ND	Q05
QUIZALOFOP ETHYL	116-0001	24	0	--	1	ND	ND	Q05
QUIZALOFOP ETHYL	127-0001	26	0	--	1	ND	ND	Q05

Table 6 (Cont.). Individual Residue Results from Corn Samples

Analyte	Sample ID[a]	Inj. No	Analyte Peak Area	ISTD Peak Area	Dilution Factor	Uncorrected (µg/g)	Reported (µg/g)[b]	Set ID
QUIZALOFOP ETHYL	128-0001	27	0	--	1	ND	ND	Q05
QUIZALOFOP ETHYL	131-0001	28	0	--	1	ND	ND	Q05
QUIZALOFOP ETHYL	132-0001	29	0	--	1	ND	ND	Q05
QUIZALOFOP ETHYL	135-0001	30	0	--	1	ND	ND	Q05
QUIZALOFOP ETHYL	136-0001	31	0	--	1	ND	ND	Q05
QUIZALOFOP ETHYL	001-0002	32	0	--	1	ND	ND	Q05
QUIZALOFOP ETHYL	001-0003	34	0	--	1	ND	ND	Q05
QUIZALOFOP ETHYL	002-0002	35	0	--	1	ND	ND	Q05
QUIZALOFOP ETHYL	002-0003	36	0	--	1	ND	ND	Q05
QUIZALOFOP ETHYL	005-0002	37	0	--	1	ND	ND	Q05
QUIZALOFOP ETHYL	005-0003	38	0	--	1	ND	ND	Q05
QUIZALOFOP ETHYL	006-0002	39	0	--	1	ND	ND	Q05
QUIZALOFOP ETHYL	006-0003	40	0	--	1	ND	ND	Q05
QUIZALOFOP ETHYL	009-0002	42	531	--	1	0.0018	ND	Q05
QUIZALOFOP ETHYL	009-0003	43	0	--	1	ND	ND	Q05
QUIZALOFOP ETHYL	010-0002	44	0	--	1	ND	ND	Q05
QUIZALOFOP ETHYL	010-0003	45	0	--	1	ND	ND	Q05
QUIZALOFOP ETHYL	014-0002	46	0	--	1	ND	ND	Q05
QUIZALOFOP ETHYL	014-0003	47	0	--	1	ND	ND	Q05
QUIZALOFOP ETHYL	015-0002	48	0	--	1	ND	ND	Q05
QUIZALOFOP ETHYL	015-0003	50	1824	--	1	0.0044	[0.0044]	Q05
QUIZALOFOP ETHYL	016-0002	51	0	--	1	ND	ND	Q05
QUIZALOFOP ETHYL	016-0003	52	0	--	1	ND	ND	Q05
QUIZALOFOP ETHYL	017-0002	53	0	--	1	ND	ND	Q05
QUIZALOFOP ETHYL	017-0003	54	0	--	1	ND	ND	Q05
QUIZALOFOP ACID	157-0001	3	0	--	1	ND	ND	Q06
QUIZALOFOP ACID	018-0002	10	0	--	1	ND	ND	Q06
QUIZALOFOP ACID	018-0003	11	0	--	1	ND	ND	Q06
QUIZALOFOP ACID	019-0002	12	0	--	1	ND	ND	Q06
QUIZALOFOP ACID	019-0003	13	0	--	1	ND	ND	Q06
QUIZALOFOP ACID	030-0002	14	0	--	1	ND	ND	Q06
QUIZALOFOP ACID	030-0003	15	0	--	1	ND	ND	Q06
QUIZALOFOP ACID	031-0002	16	0	--	1	ND	ND	Q06
QUIZALOFOP ACID	031-0003	18	0	--	1	ND	ND	Q06
QUIZALOFOP ACID	034-0002	19	0	--	1	ND	ND	Q06
QUIZALOFOP ACID	034-0003	20	0	--	1	ND	ND	Q06
QUIZALOFOP ACID	035-0002	21	0	--	1	ND	ND	Q06
QUIZALOFOP ACID	035-0003	22	0	--	1	ND	ND	Q06
QUIZALOFOP ACID	038-0002	23	0	--	1	ND	ND	Q06
QUIZALOFOP ACID	038-0003	24	0	--	1	ND	ND	Q06
QUIZALOFOP ACID	039-0002	26	0	--	1	ND	ND	Q06
QUIZALOFOP ACID	039-0003	27	0	--	1	ND	ND	Q06
QUIZALOFOP ACID	042-0002	28	0	--	1	ND	ND	Q06
QUIZALOFOP ACID	042-0003	29	0	--	1	ND	ND	Q06
QUIZALOFOP ACID	043-0002	30	0	--	1	ND	ND	Q06
QUIZALOFOP ACID	043-0003	31	0	--	1	ND	ND	Q06
QUIZALOFOP ACID	046-0002	32	0	--	1	ND	ND	Q06
QUIZALOFOP ACID	046-0003	34	0	--	1	ND	ND	Q06
QUIZALOFOP ACID	047-0002	35	0	--	1	ND	ND	Q06
QUIZALOFOP ACID	047-0003	36	0	--	1	ND	ND	Q06
QUIZALOFOP ACID	050-0002	37	0	--	1	ND	ND	Q06

Table 6 (Cont.). Individual Residue Results from Corn Samples

Analyte	Sample ID[a]	Inj. No	Analyte Peak Area	ISTD Peak Area	Dilution Factor	Uncorrected (µg/g)	Reported (µg/g)[b]	Set ID
QUIZALOFOP ACID	050-0003	38	0	--	1	ND	ND	Q06
QUIZALOFOP ACID	051-0002	39	0	--	1	ND	ND	Q06
QUIZALOFOP ACID	051-0003	40	0	--	1	ND	ND	Q06
QUIZALOFOP ACID	054-0002	42	0	--	1	ND	ND	Q06
QUIZALOFOP ACID	054-0003	43	0	--	1	ND	ND	Q06
QUIZALOFOP ACID	055-0002	44	0	--	1	ND	ND	Q06
QUIZALOFOP ACID	055-0003	45	0	--	1	ND	ND	Q06
QUIZALOFOP ACID	056-0002	46	0	--	1	ND	ND	Q06
QUIZALOFOP ACID	056-0003	47	0	--	1	ND	ND	Q06
QUIZALOFOP ACID	057-0002	48	0	--	1	ND	ND	Q06
QUIZALOFOP ACID	057-0003	50	800	--	1	0.0028	ND	Q06
QUIZALOFOP ACID	058-0002	51	0	--	1	ND	ND	Q06
QUIZALOFOP ACID	058-0003	52	0	--	1	ND	ND	Q06
QUIZALOFOP ACID	059-0002	53	0	--	1	ND	ND	Q06
QUIZALOFOP ACID	059-0003	54	0	--	1	ND	ND	Q06
QUIZALOFOP ETHYL	157-0001	3	0	--	1	ND	ND	Q06
QUIZALOFOP ETHYL	018-0002	10	0	--	1	ND	ND	Q06
QUIZALOFOP ETHYL	018-0003	11	0	--	1	ND	ND	Q06
QUIZALOFOP ETHYL	019-0002	12	0	--	1	ND	ND	Q06
QUIZALOFOP ETHYL	019-0003	13	0	--	1	ND	ND	Q06
QUIZALOFOP ETHYL	030-0002	14	0	--	1	ND	ND	Q06
QUIZALOFOP ETHYL	030-0003	15	0	--	1	ND	ND	Q06
QUIZALOFOP ETHYL	031-0002	16	0	--	1	ND	ND	Q06
QUIZALOFOP ETHYL	031-0003	18	0	--	1	ND	ND	Q06
QUIZALOFOP ETHYL	034-0002	19	0	--	1	ND	ND	Q06
QUIZALOFOP ETHYL	034-0003	20	0	--	1	ND	ND	Q06
QUIZALOFOP ETHYL	035-0002	21	0	--	1	ND	ND	Q06
QUIZALOFOP ETHYL	035-0003	22	0	--	1	ND	ND	Q06
QUIZALOFOP ETHYL	038-0002	23	0	--	1	ND	ND	Q06
QUIZALOFOP ETHYL	038-0003	24	0	--	1	ND	ND	Q06
QUIZALOFOP ETHYL	039-0002	26	0	--	1	ND	ND	Q06
QUIZALOFOP ETHYL	039-0003	27	0	--	1	ND	ND	Q06
QUIZALOFOP ETHYL	042-0002	28	0	--	1	ND	ND	Q06
QUIZALOFOP ETHYL	042-0003	29	0	--	1	ND	ND	Q06
QUIZALOFOP ETHYL	043-0002	30	0	--	1	ND	ND	Q06
QUIZALOFOP ETHYL	043-0003	31	0	--	1	ND	ND	Q06
QUIZALOFOP ETHYL	046-0002	32	0	--	1	ND	ND	Q06
QUIZALOFOP ETHYL	046-0003	34	497	--	1	0.0002	ND	Q06
QUIZALOFOP ETHYL	047-0002	35	0	--	1	ND	ND	Q06
QUIZALOFOP ETHYL	047-0003	36	0	--	1	ND	ND	Q06
QUIZALOFOP ETHYL	050-0002	37	0	--	1	ND	ND	Q06
QUIZALOFOP ETHYL	050-0003	38	0	--	1	ND	ND	Q06
QUIZALOFOP ETHYL	051-0002	39	0	--	1	ND	ND	Q06
QUIZALOFOP ETHYL	051-0003	40	0	--	1	ND	ND	Q06
QUIZALOFOP ETHYL	054-0002	42	640	--	1	0.0005	ND	Q06
QUIZALOFOP ETHYL	054-0003	43	0	--	1	ND	ND	Q06
QUIZALOFOP ETHYL	055-0002	44	0	--	1	ND	ND	Q06
QUIZALOFOP ETHYL	055-0003	45	0	--	1	ND	ND	Q06
QUIZALOFOP ETHYL	056-0002	46	0	--	1	ND	ND	Q06
QUIZALOFOP ETHYL	056-0003	47	0	--	1	ND	ND	Q06
QUIZALOFOP ETHYL	057-0002	48	0	--	1	ND	ND	Q06

Table 6 (Cont.). Individual Residue Results from Corn Samples

Analyte	Sample ID[a]	Inj. No	Analyte Peak Area	ISTD Peak Area	Dilution Factor	Uncorrected (µg/g)	Reported (µg/g)[b]	Set ID
QUIZALOFOP ETHYL	057-0003	50	1652	--	1	0.0024	ND	Q06
QUIZALOFOP ETHYL	058-0002	51	347	--	1	ND	ND	Q06
QUIZALOFOP ETHYL	058-0003	52	0	--	1	ND	ND	Q06
QUIZALOFOP ETHYL	059-0002	53	352	--	1	ND	ND	Q06
QUIZALOFOP ETHYL	059-0003	54	0	--	1	ND	ND	Q06
QUIZALOFOP ACID	157-0001	3	0	--	1	ND	ND	Q07
QUIZALOFOP ACID	070-0002	10	0	--	1	ND	ND	Q07
QUIZALOFOP ACID	070-0003	11	0	--	1	ND	ND	Q07
QUIZALOFOP ACID	071-0002	12	0	--	1	ND	ND	Q07
QUIZALOFOP ACID	071-0003	13	0	--	1	ND	ND	Q07
QUIZALOFOP ACID	074-0002	14	0	--	1	ND	ND	Q07
QUIZALOFOP ACID	074-0003	15	0	--	1	ND	ND	Q07
QUIZALOFOP ACID	075-0002	16	0	--	1	ND	ND	Q07
QUIZALOFOP ACID	075-0003	18	0	--	1	ND	ND	Q07
QUIZALOFOP ACID	079-0002	19	0	--	1	ND	ND	Q07
QUIZALOFOP ACID	079-0003	20	0	--	1	ND	ND	Q07
QUIZALOFOP ACID	083-0002	21	0	--	1	ND	ND	Q07
QUIZALOFOP ACID	083-0003	22	0	--	1	ND	ND	Q07
QUIZALOFOP ACID	084-0002	23	0	--	1	ND	ND	Q07
QUIZALOFOP ACID	084-0003	24	0	--	1	ND	ND	Q07
QUIZALOFOP ACID	087-0002	26	0	--	1	ND	ND	Q07
QUIZALOFOP ACID	087-0003	27	0	--	1	ND	ND	Q07
QUIZALOFOP ACID	088-0002	28	0	--	1	ND	ND	Q07
QUIZALOFOP ACID	088-0003	29	0	--	1	ND	ND	Q07
QUIZALOFOP ACID	091-0002	30	0	--	1	ND	ND	Q07
QUIZALOFOP ACID	091-0003	31	0	--	1	ND	ND	Q07
QUIZALOFOP ACID	092-0002	32	0	--	1	ND	ND	Q07
QUIZALOFOP ACID	092-0003	34	0	--	1	ND	ND	Q07
QUIZALOFOP ACID	095-0002	35	0	--	1	ND	ND	Q07
QUIZALOFOP ACID	095-0003	36	0	--	1	ND	ND	Q07
QUIZALOFOP ACID	096-0002	37	0	--	1	ND	ND	Q07
QUIZALOFOP ACID	096-0003	38	0	--	1	ND	ND	Q07
QUIZALOFOP ACID	099-0002	39	0	--	1	ND	ND	Q07
QUIZALOFOP ACID	099-0003	40	0	--	1	ND	ND	Q07
QUIZALOFOP ACID	100-0002	42	0	--	1	ND	ND	Q07
QUIZALOFOP ACID	100-0003	43	0	--	1	ND	ND	Q07
QUIZALOFOP ACID	103-0002	44	0	--	1	ND	ND	Q07
QUIZALOFOP ACID	103-0003	45	0	--	1	ND	ND	Q07
QUIZALOFOP ACID	104-0002	46	0	--	1	ND	ND	Q07
QUIZALOFOP ACID	104-0003	47	0	--	1	ND	ND	Q07
QUIZALOFOP ACID	107-0002	48	0	--	1	ND	ND	Q07
QUIZALOFOP ACID	107-0003	50	626	--	1	0.0003	ND	Q07
QUIZALOFOP ACID	108-0002	51	0	--	1	ND	ND	Q07
QUIZALOFOP ACID	108-0003	52	0	--	1	ND	ND	Q07
QUIZALOFOP ACID	111-0002	53	0	--	1	ND	ND	Q07
QUIZALOFOP ACID	111-0003	54	0	--	1	ND	ND	Q07
QUIZALOFOP ETHYL	157-0001	3	0	--	1	ND	ND	Q07
QUIZALOFOP ETHYL	070-0002	10	0	--	1	ND	ND	Q07
QUIZALOFOP ETHYL	070-0003	11	0	--	1	ND	ND	Q07
QUIZALOFOP ETHYL	071-0002	12	0	--	1	ND	ND	Q07
QUIZALOFOP ETHYL	071-0003	13	0	--	1	ND	ND	Q07

Table 6 (Cont.). Individual Residue Results from Corn Samples

Analyte	Sample ID[a]	Inj. No	Analyte Peak Area	ISTD Peak Area	Dilution Factor	Uncorrected (µg/g)	Reported (µg/g)[b]	Set ID
QUIZALOFOP ETHYL	074-0002	14	0	--	1	ND	ND	Q07
QUIZALOFOP ETHYL	074-0003	15	0	--	1	ND	ND	Q07
QUIZALOFOP ETHYL	075-0002	16	279	--	1	ND	ND	Q07
QUIZALOFOP ETHYL	075-0003	18	354	--	1	ND	ND	Q07
QUIZALOFOP ETHYL	079-0002	19	0	--	1	ND	ND	Q07
QUIZALOFOP ETHYL	079-0003	20	0	--	1	ND	ND	Q07
QUIZALOFOP ETHYL	083-0002	21	0	--	1	ND	ND	Q07
QUIZALOFOP ETHYL	083-0003	22	0	--	1	ND	ND	Q07
QUIZALOFOP ETHYL	084-0002	23	0	--	1	ND	ND	Q07
QUIZALOFOP ETHYL	084-0003	24	0	--	1	ND	ND	Q07
QUIZALOFOP ETHYL	087-0002	26	0	--	1	ND	ND	Q07
QUIZALOFOP ETHYL	087-0003	27	0	--	1	ND	ND	Q07
QUIZALOFOP ETHYL	088-0002	28	0	--	1	ND	ND	Q07
QUIZALOFOP ETHYL	088-0003	29	0	--	1	ND	ND	Q07
QUIZALOFOP ETHYL	091-0002	30	0	--	1	ND	ND	Q07
QUIZALOFOP ETHYL	091-0003	31	0	--	1	ND	ND	Q07
QUIZALOFOP ETHYL	092-0002	32	0	--	1	ND	ND	Q07
QUIZALOFOP ETHYL	092-0003	34	0	--	1	ND	ND	Q07
QUIZALOFOP ETHYL	095-0002	35	0	--	1	ND	ND	Q07
QUIZALOFOP ETHYL	095-0003	36	0	--	1	ND	ND	Q07
QUIZALOFOP ETHYL	096-0002	37	0	--	1	ND	ND	Q07
QUIZALOFOP ETHYL	096-0003	38	0	--	1	ND	ND	Q07
QUIZALOFOP ETHYL	099-0002	39	0	--	1	ND	ND	Q07
QUIZALOFOP ETHYL	099-0003	40	0	--	1	ND	ND	Q07
QUIZALOFOP ETHYL	100-0002	42	666	--	1	ND	ND	Q07
QUIZALOFOP ETHYL	100-0003	43	0	--	1	ND	ND	Q07
QUIZALOFOP ETHYL	103-0002	44	0	--	1	ND	ND	Q07
QUIZALOFOP ETHYL	103-0003	45	0	--	1	ND	ND	Q07
QUIZALOFOP ETHYL	104-0002	46	0	--	1	ND	ND	Q07
QUIZALOFOP ETHYL	104-0003	47	0	--	1	ND	ND	Q07
QUIZALOFOP ETHYL	107-0002	48	0	--	1	ND	ND	Q07
QUIZALOFOP ETHYL	107-0003	50	1246	--	1	0.0005	ND	Q07
QUIZALOFOP ETHYL	108-0002	51	0	--	1	ND	ND	Q07
QUIZALOFOP ETHYL	108-0003	52	0	--	1	ND	ND	Q07
QUIZALOFOP ETHYL	111-0002	53	0	--	1	ND	ND	Q07
QUIZALOFOP ETHYL	111-0003	54	0	--	1	ND	ND	Q07
QUIZALOFOP ACID	157-0001	3	0	--	1	ND	ND	Q08
QUIZALOFOP ACID	112-0002	10	807	--	1	0.0030	[0.0030]	Q08
QUIZALOFOP ACID	112-0003	11	442	--	1	0.0017	ND	Q08
QUIZALOFOP ACID	113-0002	12	626	--	1	0.0023	ND	Q08
QUIZALOFOP ACID	113-0003	14	1022	--	1	0.0037	[0.0037]	Q08
QUIZALOFOP ACID	114-0002	15	510	--	1	0.0020	ND	Q08
QUIZALOFOP ACID	114-0003	16	491	--	1	0.0019	ND	Q08
QUIZALOFOP ACID	115-0002	17	0	--	1	ND	ND	Q08
QUIZALOFOP ACID	115-0003	18	393	--	1	0.0016	ND	Q08
QUIZALOFOP ACID	116-0002	20	0	--	1	ND	ND	Q08
QUIZALOFOP ACID	116-0003	21	0	--	1	ND	ND	Q08
QUIZALOFOP ACID	127-0002	22	0	--	1	ND	ND	Q08
QUIZALOFOP ACID	127-0003	23	0	--	1	ND	ND	Q08
QUIZALOFOP ACID	128-0002	24	49760	--	1	0.1668	0.1668	Q08
QUIZALOFOP ACID	128-0003	26	33578	--	1	0.1126	0.1126	Q08

Table 6 (Cont.). Individual Residue Results from Corn Samples

Analyte	Sample ID[a]	Inj. No	Analyte Peak Area	ISTD Peak Area	Dilution Factor	Uncorrected (µg/g)	Reported (µg/g)[b]	Set ID
QUIZALOFOP ACID	131-0002	27	0	--	1	ND	ND	Q08
QUIZALOFOP ACID	131-0003	28	0	--	1	ND	ND	Q08
QUIZALOFOP ACID	132-0002	29	0	--	1	ND	ND	Q08
QUIZALOFOP ACID	132-0003	30	0	--	1	ND	ND	Q08
QUIZALOFOP ACID	135-0002	32	0	--	1	ND	ND	Q08
QUIZALOFOP ACID	135-0003	33	0	--	1	ND	ND	Q08
QUIZALOFOP ACID	136-0002	34	0	--	1	ND	ND	Q08
QUIZALOFOP ACID	136-0003	35	0	--	1	ND	ND	Q08
QUIZALOFOP ETHYL	157-0001	3	0	--	1	ND	ND	Q08
QUIZALOFOP ETHYL	112-0002	10	754	--	1	ND	ND	Q08
QUIZALOFOP ETHYL	112-0003	11	468	--	1	ND	ND	Q08
QUIZALOFOP ETHYL	113-0002	12	395	--	1	ND	ND	Q08
QUIZALOFOP ETHYL	113-0003	14	0	--	1	ND	ND	Q08
QUIZALOFOP ETHYL	114-0002	15	0	--	1	ND	ND	Q08
QUIZALOFOP ETHYL	114-0003	16	0	--	1	ND	ND	Q08
QUIZALOFOP ETHYL	115-0002	17	0	--	1	ND	ND	Q08
QUIZALOFOP ETHYL	115-0003	18	0	--	1	ND	ND	Q08
QUIZALOFOP ETHYL	116-0002	20	0	--	1	ND	ND	Q08
QUIZALOFOP ETHYL	116-0003	21	0	--	1	ND	ND	Q08
QUIZALOFOP ETHYL	127-0002	22	0	--	1	ND	ND	Q08
QUIZALOFOP ETHYL	127-0003	23	0	--	1	ND	ND	Q08
QUIZALOFOP ETHYL	128-0002	24	101403	--	1	0.1968	0.1968	Q08
QUIZALOFOP ETHYL	128-0003	26	51379	--	1	0.0989	0.0989	Q08
QUIZALOFOP ETHYL	131-0002	27	0	--	1	ND	ND	Q08
QUIZALOFOP ETHYL	131-0003	28	0	--	1	ND	ND	Q08
QUIZALOFOP ETHYL	132-0002	29	0	--	1	ND	ND	Q08
QUIZALOFOP ETHYL	132-0003	30	0	--	1	ND	ND	Q08
QUIZALOFOP ETHYL	135-0002	32	608	--	1	ND	ND	Q08
QUIZALOFOP ETHYL	135-0003	33	0	--	1	ND	ND	Q08
QUIZALOFOP ETHYL	136-0002	34	0	--	1	ND	ND	Q08
QUIZALOFOP ETHYL	136-0003	35	0	--	1	ND	ND	Q08
QUIZALOFOP ACID	158-0001	3	0	--	1	ND	ND	Q09
QUIZALOFOP ACID	004-0001	10	0	--	1	ND	ND	Q09
QUIZALOFOP ACID	008-0001	11	0	--	1	ND	ND	Q09
QUIZALOFOP ACID	013-0001	12	0	--	1	ND	ND	Q09
QUIZALOFOP ACID	021-0001	13	0	--	1	ND	ND	Q09
QUIZALOFOP ACID	023-0001	14	0	--	1	ND	ND	Q09
QUIZALOFOP ACID	025-0001	15	0	--	1	ND	ND	Q09
QUIZALOFOP ACID	027-0001	16	0	--	1	ND	ND	Q09
QUIZALOFOP ACID	029-0001	18	0	--	1	ND	ND	Q09
QUIZALOFOP ACID	033-0001	19	0	--	1	ND	ND	Q09
QUIZALOFOP ACID	037-0001	20	0	--	1	ND	ND	Q09
QUIZALOFOP ACID	041-0001	21	0	--	1	ND	ND	Q09
QUIZALOFOP ACID	045-0001	22	0	--	1	ND	ND	Q09
QUIZALOFOP ACID	049-0001	23	0	--	1	ND	ND	Q09
QUIZALOFOP ACID	053-0001	24	0	--	1	ND	ND	Q09
QUIZALOFOP ACID	061-0001	26	0	--	1	ND	ND	Q09
QUIZALOFOP ACID	063-0001	27	0	--	1	ND	ND	Q09
QUIZALOFOP ACID	065-0001	28	0	--	1	ND	ND	Q09
QUIZALOFOP ACID	067-0001	29	0	--	1	ND	ND	Q09
QUIZALOFOP ACID	069-0001	30	0	--	1	ND	ND	Q09

Table 6 (Cont.). Individual Residue Results from Corn Samples

Analyte	Sample ID[a]	Inj. No	Analyte Peak Area	ISTD Peak Area	Dilution Factor	Uncorrected (µg/g)	Reported (µg/g)[b]	Set ID
QUIZALOFOP ACID	073-0001	31	0	--	1	ND	ND	Q09
QUIZALOFOP ACID	077-0001	32	0	--	1	ND	ND	Q09
QUIZALOFOP ACID	082-0001	34	0	--	1	ND	ND	Q09
QUIZALOFOP ACID	086-0001	35	0	--	1	ND	ND	Q09
QUIZALOFOP ACID	090-0001	36	0	--	1	ND	ND	Q09
QUIZALOFOP ACID	094-0001	37	0	--	1	ND	ND	Q09
QUIZALOFOP ACID	098-0001	38	0	--	1	ND	ND	Q09
QUIZALOFOP ACID	102-0001	39	0	--	1	ND	ND	Q09
QUIZALOFOP ACID	106-0001	40	0	--	1	ND	ND	Q09
QUIZALOFOP ACID	110-0001	42	0	--	1	ND	ND	Q09
QUIZALOFOP ACID	118-0001	43	0	--	1	ND	ND	Q09
QUIZALOFOP ACID	120-0001	44	0	--	1	ND	ND	Q09
QUIZALOFOP ACID	122-0001	45	0	--	1	ND	ND	Q09
QUIZALOFOP ACID	124-0001	46	0	--	1	ND	ND	Q09
QUIZALOFOP ACID	126-0001	47	0	--	1	ND	ND	Q09
QUIZALOFOP ACID	130-0001	48	0	--	1	ND	ND	Q09
QUIZALOFOP ACID	134-0001	50	506	--	1	0.0003	ND	Q09
QUIZALOFOP ACID	138-0001	51	0	--	1	ND	ND	Q09
QUIZALOFOP ACID	004-0002	52	0	--	1	ND	ND	Q09
QUIZALOFOP ACID	004-0003	53	0	--	1	ND	ND	Q09
QUIZALOFOP ACID	008-0002	54	0	--	1	ND	ND	Q09
QUIZALOFOP ACID	008-0003	55	0	--	1	ND	ND	Q09
QUIZALOFOP ETHYL	158-0001	3	0	--	1	ND	ND	Q09
QUIZALOFOP ETHYL	004-0001	10	0	--	1	ND	ND	Q09
QUIZALOFOP ETHYL	008-0001	11	0	--	1	ND	ND	Q09
QUIZALOFOP ETHYL	013-0001	12	0	--	1	ND	ND	Q09
QUIZALOFOP ETHYL	021-0001	13	0	--	1	ND	ND	Q09
QUIZALOFOP ETHYL	023-0001	14	0	--	1	ND	ND	Q09
QUIZALOFOP ETHYL	025-0001	15	0	--	1	ND	ND	Q09
QUIZALOFOP ETHYL	027-0001	16	0	--	1	ND	ND	Q09
QUIZALOFOP ETHYL	029-0001	18	0	--	1	ND	ND	Q09
QUIZALOFOP ETHYL	033-0001	19	0	--	1	ND	ND	Q09
QUIZALOFOP ETHYL	037-0001	20	0	--	1	ND	ND	Q09
QUIZALOFOP ETHYL	041-0001	21	0	--	1	ND	ND	Q09
QUIZALOFOP ETHYL	045-0001	22	0	--	1	ND	ND	Q09
QUIZALOFOP ETHYL	049-0001	23	0	--	1	ND	ND	Q09
QUIZALOFOP ETHYL	053-0001	24	0	--	1	ND	ND	Q09
QUIZALOFOP ETHYL	061-0001	26	0	--	1	ND	ND	Q09
QUIZALOFOP ETHYL	063-0001	27	0	--	1	ND	ND	Q09
QUIZALOFOP ETHYL	065-0001	28	0	--	1	ND	ND	Q09
QUIZALOFOP ETHYL	067-0001	29	0	--	1	ND	ND	Q09
QUIZALOFOP ETHYL	069-0001	30	0	--	1	ND	ND	Q09
QUIZALOFOP ETHYL	073-0001	31	0	--	1	ND	ND	Q09
QUIZALOFOP ETHYL	077-0001	32	0	--	1	ND	ND	Q09
QUIZALOFOP ETHYL	082-0001	34	0	--	1	ND	ND	Q09
QUIZALOFOP ETHYL	086-0001	35	0	--	1	ND	ND	Q09
QUIZALOFOP ETHYL	090-0001	36	0	--	1	ND	ND	Q09
QUIZALOFOP ETHYL	094-0001	37	0	--	1	ND	ND	Q09
QUIZALOFOP ETHYL	098-0001	38	0	--	1	ND	ND	Q09
QUIZALOFOP ETHYL	102-0001	39	0	--	1	ND	ND	Q09
QUIZALOFOP ETHYL	106-0001	40	0	--	1	ND	ND	Q09

Table 6 (Cont.). Individual Residue Results from Corn Samples

Analyte	Sample ID[a]	Inj. No	Analyte Peak Area	ISTD Peak Area	Dilution Factor	Uncorrected (µg/g)	Reported (µg/g)[b]	Set ID
QUIZALOFOP ETHYL	110-0001	42	624	--	1	ND	ND	Q09
QUIZALOFOP ETHYL	118-0001	43	0	--	1	ND	ND	Q09
QUIZALOFOP ETHYL	120-0001	44	0	--	1	ND	ND	Q09
QUIZALOFOP ETHYL	122-0001	45	0	--	1	ND	ND	Q09
QUIZALOFOP ETHYL	124-0001	46	0	--	1	ND	ND	Q09
QUIZALOFOP ETHYL	126-0001	47	0	--	1	ND	ND	Q09
QUIZALOFOP ETHYL	130-0001	48	0	--	1	ND	ND	Q09
QUIZALOFOP ETHYL	134-0001	50	1548	--	1	0.0009	ND	Q09
QUIZALOFOP ETHYL	138-0001	51	0	--	1	ND	ND	Q09
QUIZALOFOP ETHYL	004-0002	52	0	--	1	ND	ND	Q09
QUIZALOFOP ETHYL	004-0003	53	0	--	1	ND	ND	Q09
QUIZALOFOP ETHYL	008-0002	54	0	--	1	ND	ND	Q09
QUIZALOFOP ETHYL	008-0003	55	0	--	1	ND	ND	Q09
QUIZALOFOP ACID	158-0001	3	0	--	1	ND	ND	Q10
QUIZALOFOP ACID	013-0002	10	547	--	1	0.0011	ND	Q10
QUIZALOFOP ACID	013-0003	11	446	--	1	0.0008	ND	Q10
QUIZALOFOP ACID	021-0002	12	1452	--	1	0.0043	[0.0043]	Q10
QUIZALOFOP ACID	021-0003	13	1325	--	1	0.0039	[0.0039]	Q10
QUIZALOFOP ACID	023-0002	14	741	--	1	0.0018	ND	Q10
QUIZALOFOP ACID	023-0003	15	927	--	1	0.0025	ND	Q10
QUIZALOFOP ACID	025-0002	16	1152	--	1	0.0033	[0.0033]	Q10
QUIZALOFOP ACID	025-0003	18	582	--	1	0.0013	ND	Q10
QUIZALOFOP ACID	027-0002	19	825	--	1	0.0021	ND	Q10
QUIZALOFOP ACID	027-0003	20	673	--	1	0.0016	ND	Q10
QUIZALOFOP ACID	029-0002	21	539	--	1	0.0011	ND	Q10
QUIZALOFOP ACID	029-0003	22	0	--	1	ND	ND	Q10
QUIZALOFOP ACID	033-0002	23	0	--	1	ND	ND	Q10
QUIZALOFOP ACID	033-0003	24	0	--	1	ND	ND	Q10
QUIZALOFOP ACID	037-0002	26	0	--	1	ND	ND	Q10
QUIZALOFOP ACID	037-0003	27	0	--	1	ND	ND	Q10
QUIZALOFOP ACID	041-0002	28	0	--	1	ND	ND	Q10
QUIZALOFOP ACID	041-0003	29	0	--	1	ND	ND	Q10
QUIZALOFOP ACID	045-0002	30	903	--	1	0.0024	ND	Q10
QUIZALOFOP ACID	045-0003	31	0	--	1	ND	ND	Q10
QUIZALOFOP ACID	049-0002	32	0	--	1	ND	ND	Q10
QUIZALOFOP ACID	049-0003	34	500	--	1	0.0010	ND	Q10
QUIZALOFOP ACID	053-0002	35	412	--	1	0.0007	ND	Q10
QUIZALOFOP ACID	053-0003	36	0	--	1	ND	ND	Q10
QUIZALOFOP ACID	061-0002	37	1246	--	1	0.0036	[0.0036]	Q10
QUIZALOFOP ACID	061-0003	38	1241	--	1	0.0036	[0.0036]	Q10
QUIZALOFOP ACID	063-0002	39	1477	--	1	0.0044	[0.0044]	Q10
QUIZALOFOP ACID	063-0003	40	1421	--	1	0.0042	[0.0042]	Q10
QUIZALOFOP ACID	065-0002	42	3079	--	1	0.0100	0.0100	Q10
QUIZALOFOP ACID	065-0003	43	1955	--	1	0.0061	[0.0061]	Q10
QUIZALOFOP ACID	067-0002	44	1809	--	1	0.0055	[0.0055]	Q10
QUIZALOFOP ACID	067-0003	45	2823	--	1	0.0091	[0.0091]	Q10
QUIZALOFOP ACID	069-0002	46	1781	--	1	0.0054	[0.0054]	Q10
QUIZALOFOP ACID	069-0003	47	4479	--	1	0.0148	0.0148	Q10
QUIZALOFOP ACID	073-0002	48	0	--	1	ND	ND	Q10
QUIZALOFOP ACID	073-0003	50	1470	--	1	0.0044	[0.0044]	Q10
QUIZALOFOP ACID	077-0002	51	1085	--	1	0.0030	[0.0030]	Q10

Table 6 (Cont.). Individual Residue Results from Corn Samples

Analyte	Sample ID[a]	Inj. No	Analyte Peak Area	ISTD Peak Area	Dilution Factor	Uncorrected (µg/g)	Reported (µg/g)[b]	Set ID
QUIZALOFOP ACID	077-0003	52	949	--	1	0.0025	ND	Q10
QUIZALOFOP ACID	082-0002	53	0	--	1	ND	ND	Q10
QUIZALOFOP ACID	082-0003	54	0	--	1	ND	ND	Q10
QUIZALOFOP ETHYL	158-0001	3	0	--	1	ND	ND	Q10
QUIZALOFOP ETHYL	013-0002	10	0	--	1	ND	ND	Q10
QUIZALOFOP ETHYL	013-0003	11	0	--	1	ND	ND	Q10
QUIZALOFOP ETHYL	021-0002	12	0	--	1	ND	ND	Q10
QUIZALOFOP ETHYL	021-0003	13	0	--	1	ND	ND	Q10
QUIZALOFOP ETHYL	023-0002	14	0	--	1	ND	ND	Q10
QUIZALOFOP ETHYL	023-0003	15	0	--	1	ND	ND	Q10
QUIZALOFOP ETHYL	025-0002	16	0	--	1	ND	ND	Q10
QUIZALOFOP ETHYL	025-0003	18	0	--	1	ND	ND	Q10
QUIZALOFOP ETHYL	027-0002	19	0	--	1	ND	ND	Q10
QUIZALOFOP ETHYL	027-0003	20	0	--	1	ND	ND	Q10
QUIZALOFOP ETHYL	029-0002	21	0	--	1	ND	ND	Q10
QUIZALOFOP ETHYL	029-0003	22	0	--	1	ND	ND	Q10
QUIZALOFOP ETHYL	033-0002	23	0	--	1	ND	ND	Q10
QUIZALOFOP ETHYL	033-0003	24	0	--	1	ND	ND	Q10
QUIZALOFOP ETHYL	037-0002	26	0	--	1	ND	ND	Q10
QUIZALOFOP ETHYL	037-0003	27	0	--	1	ND	ND	Q10
QUIZALOFOP ETHYL	041-0002	28	0	--	1	ND	ND	Q10
QUIZALOFOP ETHYL	041-0003	29	0	--	1	ND	ND	Q10
QUIZALOFOP ETHYL	045-0002	30	0	--	1	ND	ND	Q10
QUIZALOFOP ETHYL	045-0003	31	0	--	1	ND	ND	Q10
QUIZALOFOP ETHYL	049-0002	32	0	--	1	ND	ND	Q10
QUIZALOFOP ETHYL	049-0003	34	651	--	1	ND	ND	Q10
QUIZALOFOP ETHYL	053-0002	35	265	--	1	ND	ND	Q10
QUIZALOFOP ETHYL	053-0003	36	0	--	1	ND	ND	Q10
QUIZALOFOP ETHYL	061-0002	37	0	--	1	ND	ND	Q10
QUIZALOFOP ETHYL	061-0003	38	619	--	1	ND	ND	Q10
QUIZALOFOP ETHYL	063-0002	39	474	--	1	ND	ND	Q10
QUIZALOFOP ETHYL	063-0003	40	420	--	1	ND	ND	Q10
QUIZALOFOP ETHYL	065-0002	42	1134	--	1	0.0007	ND	Q10
QUIZALOFOP ETHYL	065-0003	43	795	--	1	0	ND	Q10
QUIZALOFOP ETHYL	067-0002	44	0	--	1	ND	ND	Q10
QUIZALOFOP ETHYL	067-0003	45	731	--	1	ND	ND	Q10
QUIZALOFOP ETHYL	069-0002	46	0	--	1	ND	ND	Q10
QUIZALOFOP ETHYL	069-0003	47	0	--	1	ND	ND	Q10
QUIZALOFOP ETHYL	073-0002	48	0	--	1	ND	ND	Q10
QUIZALOFOP ETHYL	073-0003	50	1215	--	1	0.0009	ND	Q10
QUIZALOFOP ETHYL	077-0002	51	460	--	1	ND	ND	Q10
QUIZALOFOP ETHYL	077-0003	52	0	--	1	ND	ND	Q10
QUIZALOFOP ETHYL	082-0002	53	0	--	1	ND	ND	Q10
QUIZALOFOP ETHYL	082-0003	54	0	--	1	ND	ND	Q10
QUIZALOFOP ACID	158-0001	3	0	--	1	ND	ND	Q11
QUIZALOFOP ACID	086-0002	10	0	--	1	ND	ND	Q11
QUIZALOFOP ACID	086-0003	11	0	--	1	ND	ND	Q11
QUIZALOFOP ACID	090-0002	12	0	--	1	ND	ND	Q11
QUIZALOFOP ACID	090-0003	13	0	--	1	ND	ND	Q11
QUIZALOFOP ACID	094-0002	14	0	--	1	ND	ND	Q11
QUIZALOFOP ACID	094-0003	16	0	--	1	ND	ND	Q11

Table 6 (Cont.). Individual Residue Results from Corn Samples

Analyte	Sample ID[a]	Inj. No	Analyte Peak Area	ISTD Peak Area	Dilution Factor	Uncorrected (µg/g)	Reported (µg/g)[b]	Set ID
QUIZALOFOP ACID	098-0002	17	0	--	1	ND	ND	Q11
QUIZALOFOP ACID	098-0003	18	0	--	1	ND	ND	Q11
QUIZALOFOP ACID	102-0002	19	479	--	1	0.0009	ND	Q11
QUIZALOFOP ACID	102-0003	20	625	--	1	0.0014	ND	Q11
QUIZALOFOP ACID	106-0002	21	899	--	1	0.0024	ND	Q11
QUIZALOFOP ACID	106-0003	23	869	--	1	0.0023	ND	Q11
QUIZALOFOP ACID	110-0002	24	0	--	1	ND	ND	Q11
QUIZALOFOP ACID	110-0003	25	0	--	1	ND	ND	Q11
QUIZALOFOP ACID	118-0002	26	553	--	1	0.0011	ND	Q11
QUIZALOFOP ACID	118-0003	27	785	--	1	0.0020	ND	Q11
QUIZALOFOP ACID	120-0002	28	410	--	1	0.0006	ND	Q11
QUIZALOFOP ACID	120-0003	30	447	--	1	0.0007	ND	Q11
QUIZALOFOP ACID	122-0002	31	345	--	1	0.0004	ND	Q11
QUIZALOFOP ACID	122-0003	32	298	--	1	0.0002	ND	Q11
QUIZALOFOP ACID	124-0002	33	0	--	1	ND	ND	Q11
QUIZALOFOP ACID	124-0003	34	432	--	1	0.0007	ND	Q11
QUIZALOFOP ACID	126-0002	35	0	--	1	ND	ND	Q11
QUIZALOFOP ACID	126-0003	37	0	--	1	ND	ND	Q11
QUIZALOFOP ACID	130-0002	38	3004	--	1	0.0101	0.0101	Q11
QUIZALOFOP ACID	130-0003	39	3245	--	1	0.0110	0.0110	Q11
QUIZALOFOP ACID	134-0002	40	0	--	1	ND	ND	Q11
QUIZALOFOP ACID	134-0003	41	0	--	1	ND	ND	Q11
QUIZALOFOP ACID	138-0002	42	0	--	1	ND	ND	Q11
QUIZALOFOP ACID	138-0003	44	852	--	1	0.0022	ND	Q11
QUIZALOFOP ETHYL	158-0001	3	0	--	1	ND	ND	Q11
QUIZALOFOP ETHYL	086-0002	10	0	--	1	ND	ND	Q11
QUIZALOFOP ETHYL	086-0003	11	0	--	1	ND	ND	Q11
QUIZALOFOP ETHYL	090-0002	12	0	--	1	ND	ND	Q11
QUIZALOFOP ETHYL	090-0003	13	0	--	1	ND	ND	Q11
QUIZALOFOP ETHYL	094-0002	14	0	--	1	ND	ND	Q11
QUIZALOFOP ETHYL	094-0003	16	0	--	1	ND	ND	Q11
QUIZALOFOP ETHYL	098-0002	17	0	--	1	ND	ND	Q11
QUIZALOFOP ETHYL	098-0003	18	0	--	1	ND	ND	Q11
QUIZALOFOP ETHYL	102-0002	19	403	--	1	ND	ND	Q11
QUIZALOFOP ETHYL	102-0003	20	518	--	1	0.0001	ND	Q11
QUIZALOFOP ETHYL	106-0002	21	446	--	1	ND	ND	Q11
QUIZALOFOP ETHYL	106-0003	23	0	--	1	ND	ND	Q11
QUIZALOFOP ETHYL	110-0002	24	0	--	1	ND	ND	Q11
QUIZALOFOP ETHYL	110-0003	25	0	--	1	ND	ND	Q11
QUIZALOFOP ETHYL	118-0002	26	0	--	1	ND	ND	Q11
QUIZALOFOP ETHYL	118-0003	27	0	--	1	ND	ND	Q11
QUIZALOFOP ETHYL	120-0002	28	0	--	1	ND	ND	Q11
QUIZALOFOP ETHYL	120-0003	30	0	--	1	ND	ND	Q11
QUIZALOFOP ETHYL	122-0002	31	0	--	1	ND	ND	Q11
QUIZALOFOP ETHYL	122-0003	32	0	--	1	ND	ND	Q11
QUIZALOFOP ETHYL	124-0002	33	0	--	1	ND	ND	Q11
QUIZALOFOP ETHYL	124-0003	34	0	--	1	ND	ND	Q11
QUIZALOFOP ETHYL	126-0002	35	0	--	1	ND	ND	Q11
QUIZALOFOP ETHYL	126-0003	37	710	--	1	0.0004	ND	Q11
QUIZALOFOP ETHYL	130-0002	38	2787	--	1	0.0041	[0.0041]	Q11
QUIZALOFOP ETHYL	130-0003	39	2930	--	1	0.0044	[0.0044]	Q11

Table 6 (Cont.). Individual Residue Results from Corn Samples

Analyte	Sample ID[a]	Inj. No	Analyte Peak Area	ISTD Peak Area	Dilution Factor	Uncorrected (µg/g)	Reported (µg/g)[b]	Set ID
QUIZALOFOP ETHYL	134-0002	40	0	--	1	ND	ND	Q11
QUIZALOFOP ETHYL	134-0003	41	0	--	1	ND	ND	Q11
QUIZALOFOP ETHYL	138-0002	42	0	--	1	ND	ND	Q11
QUIZALOFOP ETHYL	138-0003	44	2582	--	1	0.0038	[0.0038]	Q11
QUIZALOFOP ACID	150-0001	3	0	--	1	ND	ND	Q12
QUIZALOFOP ACID	155-0001	4	0	--	1	ND	ND	Q12
QUIZALOFOP ACID	145-0001	5	0	--	1	ND	ND	Q12
QUIZALOFOP ACID	144-0001	6	0	--	1	ND	ND	Q12
QUIZALOFOP ACID	141-0001	39	606	--	1	0.0020	ND	Q12
QUIZALOFOP ACID	146-0001	40	0	--	1	ND	ND	Q12
QUIZALOFOP ACID	146-0002	42	481	--	1	0.0018	ND	Q12
QUIZALOFOP ACID	155-0002	43	0	--	1	ND	ND	Q12
QUIZALOFOP ACID	145-0002	44	0	--	1	ND	ND	Q12
QUIZALOFOP ACID	154-0001	45	0	--	1	ND	ND	Q12
QUIZALOFOP ACID	154-0002	46	0	--	1	ND	ND	Q12
QUIZALOFOP ACID	144-0002	47	0	--	1	ND	ND	Q12
QUIZALOFOP ACID	153-0001	48	0	--	1	ND	ND	Q12
QUIZALOFOP ACID	153-0002	50	719	--	1	0.0023	ND	Q12
QUIZALOFOP ACID	141-0002	51	0	--	1	ND	ND	Q12
QUIZALOFOP ACID	150-0002	52	0	--	1	ND	ND	Q12
QUIZALOFOP ETHYL	150-0001	3	0	--	1	ND	ND	Q12
QUIZALOFOP ETHYL	155-0001	4	0	--	1	ND	ND	Q12
QUIZALOFOP ETHYL	145-0001	5	0	--	1	ND	ND	Q12
QUIZALOFOP ETHYL	144-0001	6	0	--	1	ND	ND	Q12
QUIZALOFOP ETHYL	141-0001	39	1005	--	1	0.0016	ND	Q12
QUIZALOFOP ETHYL	146-0001	40	539	--	1	0.0007	ND	Q12
QUIZALOFOP ETHYL	146-0002	42	918	--	1	0.0015	ND	Q12
QUIZALOFOP ETHYL	155-0002	43	0	--	1	ND	ND	Q12
QUIZALOFOP ETHYL	145-0002	44	0	--	1	ND	ND	Q12
QUIZALOFOP ETHYL	154-0001	45	0	--	1	ND	ND	Q12
QUIZALOFOP ETHYL	154-0002	46	0	--	1	ND	ND	Q12
QUIZALOFOP ETHYL	144-0002	47	0	--	1	ND	ND	Q12
QUIZALOFOP ETHYL	153-0001	48	0	--	1	ND	ND	Q12
QUIZALOFOP ETHYL	153-0002	50	1744	--	1	0.0031	[0.0031]	Q12
QUIZALOFOP ETHYL	141-0002	51	359	--	1	0.0003	ND	Q12
QUIZALOFOP ETHYL	150-0002	52	0	--	1	ND	ND	Q12
QUIZALOFOP ACID	150-0001	3	0	--	1	ND	ND	Q13
QUIZALOFOP ACID	155-0001	4	0	--	1	ND	ND	Q13
QUIZALOFOP ACID	145-0001	5	0	--	1	ND	ND	Q13
QUIZALOFOP ACID	144-0001	6	0	--	1	ND	ND	Q13
QUIZALOFOP ACID	146-0003	39	682	--	1	0.0020	ND	Q13
QUIZALOFOP ACID	146-0004	40	200	--	1	0.0010	ND	Q13
QUIZALOFOP ACID	155-0003	42	487	--	1	0.0016	ND	Q13
QUIZALOFOP ACID	155-0004	43	165	--	1	0.0010	ND	Q13
QUIZALOFOP ACID	145-0003	44	89	--	1	0.0008	ND	Q13
QUIZALOFOP ACID	145-0004	45	0	--	1	ND	ND	Q13
QUIZALOFOP ACID	154-0003	46	119	--	1	0.0009	ND	Q13
QUIZALOFOP ACID	154-0004	47	0	--	1	ND	ND	Q13
QUIZALOFOP ACID	144-0003	48	0	--	1	ND	ND	Q13
QUIZALOFOP ACID	144-0004	50	870	--	1	0.0023	ND	Q13
QUIZALOFOP ACID	153-0003	51	249	--	1	0.0011	ND	Q13

Table 6 (Cont.). Individual Residue Results from Corn Samples

Analyte	Sample ID[a]	Inj. No	Analyte Peak Area	ISTD Peak Area	Dilution Factor	Uncorrected (µg/g)	Reported (µg/g)[b]	Set ID
QUIZALOFOP ACID	153-0004	52	0	--	1	ND	ND	Q13
QUIZALOFOP ETHYL	150-0001	3	0	--	1	ND	ND	Q13
QUIZALOFOP ETHYL	155-0001	4	0	--	1	ND	ND	Q13
QUIZALOFOP ETHYL	145-0001	5	0	--	1	ND	ND	Q13
QUIZALOFOP ETHYL	144-0001	6	0	--	1	ND	ND	Q13
QUIZALOFOP ETHYL	146-0003	39	1097	--	1	0.0009	ND	Q13
QUIZALOFOP ETHYL	146-0004	40	526	--	1	ND	ND	Q13
QUIZALOFOP ETHYL	155-0003	42	951	--	1	0.0006	ND	Q13
QUIZALOFOP ETHYL	155-0004	43	324	--	1	ND	ND	Q13
QUIZALOFOP ETHYL	145-0003	44	219	--	1	ND	ND	Q13
QUIZALOFOP ETHYL	145-0004	45	0	--	1	ND	ND	Q13
QUIZALOFOP ETHYL	154-0003	46	158	--	1	ND	ND	Q13
QUIZALOFOP ETHYL	154-0004	47	0	--	1	ND	ND	Q13
QUIZALOFOP ETHYL	144-0003	48	0	--	1	ND	ND	Q13
QUIZALOFOP ETHYL	144-0004	50	1871	--	1	0.0025	ND	Q13
QUIZALOFOP ETHYL	153-0003	51	445	--	1	ND	ND	Q13
QUIZALOFOP ETHYL	153-0004	52	128	--	1	ND	ND	Q13
QUIZALOFOP ACID	143-0001	3	0	--	1	ND	ND	Q14
QUIZALOFOP ACID	147-0001	4	0	--	1	ND	ND	Q14
QUIZALOFOP ACID	142-0001	5	0	--	1	ND	ND	Q14
QUIZALOFOP ACID	143-0002	32	534	--	1	ND	ND	Q14
QUIZALOFOP ACID	147-0002	33	0	--	1	ND	ND	Q14
QUIZALOFOP ACID	152-0001	34	147	--	1	ND	ND	Q14
QUIZALOFOP ACID	152-0002	35	0	--	1	ND	ND	Q14
QUIZALOFOP ACID	156-0001	36	0	--	1	ND	ND	Q14
QUIZALOFOP ACID	156-0002	38	606	--	1	ND	ND	Q14
QUIZALOFOP ACID	142-0002	39	284	--	1	ND	ND	Q14
QUIZALOFOP ACID	151-0001	40	167	--	1	ND	ND	Q14
QUIZALOFOP ACID	151-0002	41	0	--	1	ND	ND	Q14
QUIZALOFOP ETHYL	143-0001	3	0	--	1	ND	ND	Q14
QUIZALOFOP ETHYL	147-0001	4	0	--	1	ND	ND	Q14
QUIZALOFOP ETHYL	142-0001	5	0	--	1	ND	ND	Q14
QUIZALOFOP ETHYL	143-0002	32	1525	--	1	0.0034	[0.0034]	Q14
QUIZALOFOP ETHYL	147-0002	33	677	--	1	0.0017	ND	Q14
QUIZALOFOP ETHYL	152-0001	34	1085	--	1	0.0025	ND	Q14
QUIZALOFOP ETHYL	152-0002	35	264	--	1	0.0009	ND	Q14
QUIZALOFOP ETHYL	156-0001	36	241	--	1	0.0008	ND	Q14
QUIZALOFOP ETHYL	156-0002	38	2589	--	1	0.0056	[0.0056]	Q14
QUIZALOFOP ETHYL	142-0002	39	351	--	1	0.0010	ND	Q14
QUIZALOFOP ETHYL	151-0001	40	209	--	1	0.0007	ND	Q14
QUIZALOFOP ETHYL	151-0002	41	166	--	1	0.0007	ND	Q14
QUIZALOFOP ACID	143-0001	3	0	--	1	ND	ND	Q15
QUIZALOFOP ACID	147-0001	4	0	--	1	ND	ND	Q15
QUIZALOFOP ACID	142-0001	5	0	--	1	ND	ND	Q15
QUIZALOFOP ACID	143-0003	31	391	--	1	ND	ND	Q15
QUIZALOFOP ACID	143-0004	32	81	--	1	ND	ND	Q15
QUIZALOFOP ACID	147-0003	33	93	--	1	ND	ND	Q15
QUIZALOFOP ACID	147-0004	34	0	--	1	ND	ND	Q15
QUIZALOFOP ACID	152-0003	35	0	--	1	ND	ND	Q15
QUIZALOFOP ACID	152-0004	37	256	--	1	ND	ND	Q15
QUIZALOFOP ACID	156-0003	38	79	--	1	ND	ND	Q15

Table 6 (Cont.). Individual Residue Results from Corn Samples

Analyte	Sample ID[a]	Inj. No	Analyte Peak Area	ISTD Peak Area	Dilution Factor	Uncorrected (µg/g)	Reported (µg/g)[b]	Set ID
QUIZALOFOP ACID	156-0004	39	0	--	1	ND	ND	Q15
QUIZALOFOP ACID	142-0003	40	0	--	1	ND	ND	Q15
QUIZALOFOP ACID	142-0004	41	0	--	1	ND	ND	Q15
QUIZALOFOP ACID	151-0003	42	0	--	1	ND	ND	Q15
QUIZALOFOP ACID	151-0004	44	668	--	1	0.0006	ND	Q15
QUIZALOFOP ETHYL	143-0001	3	0	--	1	ND	ND	Q15
QUIZALOFOP ETHYL	147-0001	4	0	--	1	ND	ND	Q15
QUIZALOFOP ETHYL	142-0001	5	0	--	1	ND	ND	Q15
QUIZALOFOP ETHYL	143-0003	31	902	--	1	0.0009	ND	Q15
QUIZALOFOP ETHYL	143-0004	32	272	--	1	ND	ND	Q15
QUIZALOFOP ETHYL	147-0003	33	421	--	1	ND	ND	Q15
QUIZALOFOP ETHYL	147-0004	34	179	--	1	ND	ND	Q15
QUIZALOFOP ETHYL	152-0003	35	0	--	1	ND	ND	Q15
QUIZALOFOP ETHYL	152-0004	37	1280	--	1	0.0017	ND	Q15
QUIZALOFOP ETHYL	156-0003	38	303	--	1	ND	ND	Q15
QUIZALOFOP ETHYL	156-0004	39	0	--	1	ND	ND	Q15
QUIZALOFOP ETHYL	142-0003	40	0	--	1	ND	ND	Q15
QUIZALOFOP ETHYL	142-0004	41	0	--	1	ND	ND	Q15
QUIZALOFOP ETHYL	151-0003	42	0	--	1	ND	ND	Q15
QUIZALOFOP ETHYL	151-0004	44	2960	--	1	0.0051	[0.0051]	Q15

[a] Full Sample ID number is preceded by ARA-09-15-10-.
[b] All calculations were done using Microsoft Excel 2003 with full precision.
[c] ND = not detected. The residue was below the 0.003 µg/g limit of detection.
[d] [ ] indicates that the concentration is less than the limit of quantitation of 0.01 µg/g. Values are reported with a lower degree of confidence than values above the limit of quantitation

Table 7. Summarized Field Sample Information and Residue Results

Sample ID[a]	Site	Trial	Trt	Rep	Event	Commodity	2,4-D Reported, µg/g[b]	2,4-DCP Reported, µg/g[b]	Quizalofop Acid Reported, µg/g[b]	Quizalofop Ethyl Reported, µg/g[b]
004-0001A1	NJ	1	T1	1	N/A	Stover	- - - -[c]	- - - -	ND[d]	ND
004-0001A2	NJ	1	T1	1	N/A	Stover	[0.0041][e]	ND	- - - -	- - - -
004-0002A1	NJ	1	T2	1	90	Stover	- - - -	- - - -	ND	ND
004-0002A2	NJ	1	T2	1	90	Stover	8.1905	2.2405	- - - -	- - - -
004-0003A1	NJ	1	T2	2	90	Stover	- - - -	- - - -	ND	ND
004-0003A2	NJ	1	T2	2	90	Stover	5.8767	1.5567	- - - -	- - - -
008-0001A1	PA	2	T1	1	N/A	Stover	- - - -	- - - -	ND	ND
008-0001A2	PA	2	T1	1	N/A	Stover	ND	ND	- - - -	- - - -
008-0002A1	PA	2	T2	1	90	Stover	- - - -	- - - -	ND	ND
008-0002A2	PA	2	T2	1	90	Stover	2.6949	1.6915	- - - -	- - - -
008-0003A1	PA	2	T2	2	90	Stover	- - - -	- - - -	ND	ND
008-0003A2	PA	2	T2	2	90	Stover	1.6267	1.2076	- - - -	- - - -
013-0001A1	GA	3	T1	1	N/A	Stover	- - - -	- - - -	ND	ND
013-0001A2	GA	3	T1	1	N/A	Stover	ND	ND	- - - -	- - - -
013-0002A1	GA	3	T2	1	90	Stover	- - - -	- - - -	ND	ND
013-0002A2	GA	3	T2	1	90	Stover	0.3235	0.2068	- - - -	- - - -
013-0003A1	GA	3	T2	2	90	Stover	- - - -	- - - -	ND	ND
013-0003A2	GA	3	T2	2	90	Stover	0.1617	0.0718	- - - -	- - - -
021-0001A1	GA	4	T1	1	N/A	Stover	- - - -	- - - -	ND	ND
021-0001A10	GA	4	T1	1	N/A	Stover	ND	ND	- - - -	- - - -
021-0001A3	GA	4	T1	1	N/A	Stover	ND	ND	- - - -	- - - -
021-0001A9	GA	4	T1	1	N/A	Stover	ND	ND	- - - -	- - - -
021-0002A1	GA	4	T2	1	76	Stover	- - - -	- - - -	[0.0043]	ND
021-0002A2	GA	4	T2	1	76	Stover	2.8621	1.1387	- - - -	- - - -
021-0003A1	GA	4	T2	2	76	Stover	- - - -	- - - -	[0.0039]	ND
021-0003A2	GA	4	T2	2	76	Stover	2.6386	0.7986	- - - -	- - - -
023-0001A1	GA	4	T1	1	N/A	Stover	- - - -	- - - -	ND	ND
023-0001A2	GA	4	T1	1	N/A	Stover	ND	[0.0036]	- - - -	- - - -
023-0002A1	GA	4	T2	1	83	Stover	- - - -	- - - -	ND	ND
023-0002A2	GA	4	T2	1	83	Stover	0.7854	0.3654	- - - -	- - - -
023-0003A1	GA	4	T2	2	83	Stover	- - - -	- - - -	ND	ND
023-0003A2	GA	4	T2	2	83	Stover	0.8617	0.3683	- - - -	- - - -
025-0001A1	GA	4	T1	1	N/A	Stover	- - - -	- - - -	ND	ND
025-0001A2	GA	4	T1	1	N/A	Stover	ND	ND	- - - -	- - - -
025-0002A1	GA	4	T2	1	90	Stover	- - - -	- - - -	[0.0033]	ND
025-0002A2	GA	4	T2	1	90	Stover	1.3720	0.5220	- - - -	- - - -
025-0003A1	GA	4	T2	2	90	Stover	- - - -	- - - -	ND	ND
025-0003A2	GA	4	T2	2	90	Stover	1.0820	0.4761	- - - -	- - - -
027-0001A1	GA	4	T1	1	N/A	Stover	- - - -	- - - -	ND	ND
027-0001A2	GA	4	T1	1	N/A	Stover	[0.0032]	[0.0031]	- - - -	- - - -
027-0002A1	GA	4	T2	1	97	Stover	- - - -	- - - -	ND	ND
027-0002A2	GA	4	T2	1	97	Stover	1.3267	0.5546	- - - -	- - - -

Table 7 (Cont.). Summarized Field Sample Information and Residue Results

Sample ID[a]	Site	Trial	Trt	Rep	Event	Commodity	2,4-D Reported, µg/g[b]	2,4-DCP Reported, µg/g[b]	Quizalofop Acid Reported, µg/g[b]	Quizalofop Ethyl Reported, µg/g[b]
027-0003A1	GA	4	T2	2	97	Stover	- - - -	- - - -	ND	ND
027-0003A2	GA	4	T2	2	97	Stover	1.2930	0.5369	- - - -	- - - -
029-0001A1	GA	4	T1	1	N/A	Stover	- - - -	- - - -	ND	ND
029-0001A2	GA	4	T1	1	N/A	Stover	ND	ND	- - - -	- - - -
029-0002A1	GA	4	T2	1	104	Stover	- - - -	- - - -	ND	ND
029-0002A2	GA	4	T2	1	104	Stover	0.8775	0.4315	- - - -	- - - -
029-0003A1	GA	4	T2	2	104	Stover	- - - -	- - - -	ND	ND
029-0003A2	GA	4	T2	2	104	Stover	1.0946	0.5992	- - - -	- - - -
033-0001A1	IL	5	T1	1	N/A	Stover	- - - -	- - - -	ND	ND
033-0001A3	IL	5	T1	1	N/A	Stover	ND	[0.0033]	- - - -	- - - -
033-0002A1	IL	5	T2	1	90	Stover	- - - -	- - - -	ND	ND
033-0002A3	IL	5	T2	1	90	Stover	0.1544	0.1475	- - - -	- - - -
033-0003A1	IL	5	T2	2	90	Stover	- - - -	- - - -	ND	ND
033-0003A3	IL	5	T2	2	90	Stover	0.1050	0.0944	- - - -	- - - -
037-0001A1	IL	6	T1	1	N/A	Stover	- - - -	- - - -	ND	ND
037-0001A2	IL	6	T1	1	N/A	Stover	ND	ND	- - - -	- - - -
037-0002A1	IL	6	T2	1	90	Stover	- - - -	- - - -	ND	ND
037-0002A2	IL	6	T2	1	90	Stover	0.2772	0.1682	- - - -	- - - -
037-0003A1	IL	6	T2	2	90	Stover	- - - -	- - - -	ND	ND
037-0003A2	IL	6	T2	2	90	Stover	0.4858	0.3100	- - - -	- - - -
041-0001A1	IN	7	T1	1	N/A	Stover	- - - -	- - - -	ND	ND
041-0001A2	IN	7	T1	1	N/A	Stover	[0.0043]	0.0115	- - - -	- - - -
041-0002A1	IN	7	T2	1	90	Stover	- - - -	- - - -	ND	ND
041-0002A2	IN	7	T2	1	90	Stover	1.4430	1.2784	- - - -	- - - -
041-0003A1	IN	7	T2	2	90	Stover	- - - -	- - - -	ND	ND
041-0003A2	IN	7	T2	2	90	Stover	0.6231	0.5541	- - - -	- - - -
045-0001A1	MO	8	T1	1	N/A	Stover	- - - -	- - - -	ND	ND
045-0001A2	MO	8	T1	1	N/A	Stover	ND	ND	- - - -	- - - -
045-0002A1	MO	8	T2	1	90	Stover	- - - -	- - - -	ND	ND
045-0002A2	MO	8	T2	1	90	Stover	0.0164	[0.0063]	- - - -	- - - -
045-0002A3	MO	8	T2	1	90	Stover	0.0189	[0.0076]	- - - -	- - - -
045-0002A4	MO	8	T2	1	90	Stover	0.0177	[0.0076]	- - - -	- - - -
045-0003A1	MO	8	T2	2	90	Stover	- - - -	- - - -	ND	ND
045-0003A2	MO	8	T2	2	90	Stover	0.0255	0.0192	- - - -	- - - -
049-0001A1	MO	9	T1	1	N/A	Stover	- - - -	- - - -	ND	ND
049-0001A2	MO	9	T1	1	N/A	Stover	[0.0076]	ND	- - - -	- - - -
049-0002A1	MO	9	T2	1	90	Stover	- - - -	- - - -	ND	ND
049-0002A2	MO	9	T2	1	90	Stover	1.1852	0.7407	- - - -	- - - -
049-0003A1	MO	9	T2	2	90	Stover	- - - -	- - - -	ND	ND
049-0003A2	MO	9	T2	2	90	Stover	2.4641	1.1359	- - - -	- - - -

Table 7 (Cont.). Summarized Field Sample Information and Residue Results

Sample ID[a]	Site	Trial	Trt	Rep	Event	Commodity	2,4-D Reported, µg/g[b]	2,4-DCP Reported, µg/g[b]	Quizalofop Acid Reported, µg/g[b]	Quizalofop Ethyl Reported, µg/g[b]
053-0001A1	IL	10	T1	1	N/A	Stover	- - - -	- - - -	ND	ND
053-0001A2	IL	10	T1	1	N/A	Stover	[0.0058]	0.0106	- - - -	- - - -
053-0002A1	IL	10	T2	1	90	Stover	- - - -	- - - -	ND	ND
053-0002A2	IL	10	T2	1	90	Stover	2.5402	1.3063	- - - -	- - - -
053-0003A1	IL	10	T2	2	90	Stover	- - - -	- - - -	ND	ND
053-0003A2	IL	10	T2	2	90	Stover	0.9139	0.4911	- - - -	- - - -
061-0001A1	MI	11	T1	1	N/A	Stover	- - - -	- - - -	ND	ND
061-0001A2	MI	11	T1	1	N/A	Stover	ND	[0.0048]	- - - -	- - - -
061-0002A1	MI	11	T2	1	76	Stover	- - - -	- - - -	[0.0036]	ND
061-0002A2	MI	11	T2	1	76	Stover	3.5459	3.1799	- - - -	- - - -
061-0003A1	MI	11	T2	2	76	Stover	- - - -	- - - -	[0.0036]	ND
061-0003A2	MI	11	T2	2	76	Stover	5.9788	5.4760	- - - -	- - - -
063-0001A1	MI	11	T1	1	N/A	Stover	- - - -	- - - -	ND	ND
063-0001A2	MI	11	T1	1	N/A	Stover	ND	[0.0077]	- - - -	- - - -
063-0002A1	MI	11	T2	1	83	Stover	- - - -	- - - -	[0.0044]	ND
063-0002A2	MI	11	T2	1	83	Stover	4.0802	3.2309	- - - -	- - - -
063-0003A1	MI	11	T2	2	83	Stover	- - - -	- - - -	[0.0042]	ND
063-0003A2	MI	11	T2	2	83	Stover	6.6159	6.1212	- - - -	- - - -
065-0001A1	MI	11	T1	1	N/A	Stover	- - - -	- - - -	ND	ND
065-0001A2	MI	11	T1	1	N/A	Stover	ND	[0.0045]	- - - -	- - - -
065-0002A1	MI	11	T2	1	90	Stover	- - - -	- - - -	0.0100	ND
065-0002A2	MI	11	T2	1	90	Stover	7.2327	4.9117	- - - -	- - - -
065-0003A1	MI	11	T2	2	90	Stover	- - - -	- - - -	[0.0061]	ND
065-0003A2	MI	11	T2	2	90	Stover	8.9195	5.8416	- - - -	- - - -
067-0001A1	MI	11	T1	1	N/A	Stover	- - - -	- - - -	ND	ND
067-0001A3	MI	11	T1	1	N/A	Stover	ND	[0.0059]	- - - -	- - - -
067-0002A1	MI	11	T2	1	97	Stover	- - - -	- - - -	[0.0055]	ND
067-0002A2	MI	11	T2	1	97	Stover	4.7351	3.8709	- - - -	- - - -
067-0003A1	MI	11	T2	2	97	Stover	- - - -	- - - -	[0.0091]	ND
067-0003A2	MI	11	T2	2	97	Stover	7.0505	4.4368	- - - -	- - - -
069-0001A1	MI	11	T1	1	N/A	Stover	- - - -	- - - -	ND	ND
069-0001A2	MI	11	T1	1	N/A	Stover	ND	0.0146	- - - -	- - - -
069-0002A1	MI	11	T2	1	104	Stover	- - - -	- - - -	[0.0054]	ND
069-0002A2	MI	11	T2	1	104	Stover	5.7977	3.7686	- - - -	- - - -
069-0003A1	MI	11	T2	2	104	Stover	- - - -	- - - -	0.0148	ND
069-0003A2	MI	11	T2	2	104	Stover	3.2277	2.6079	- - - -	- - - -
073-0001A1	OH	12	T1	1	N/A	Stover	- - - -	- - - -	ND	ND
073-0001A2	OH	12	T1	1	N/A	Stover	ND	[0.0055]	- - - -	- - - -
073-0002A1	OH	12	T2	1	90	Stover	- - - -	- - - -	ND	ND
073-0002A2	OH	12	T2	1	90	Stover	1.6763	1.1874	- - - -	- - - -
073-0003A1	OH	12	T2	2	90	Stover	- - - -	- - - -	[0.0044]	ND
073-0003A2	OH	12	T2	2	90	Stover	2.4219	1.7679	- - - -	- - - -

Table 7 (Cont.). Summarized Field Sample Information and Residue Results

Sample ID[a]	Site	Trial	Trt	Rep	Event	Commodity	2,4-D Reported, µg/g[b]	2,4-DCP Reported, µg/g[b]	Quizalofop Acid Reported, µg/g[b]	Quizalofop Ethyl Reported, µg/g[b]
077-0001A1	IN	13	T1	1	N/A	Stover	- - - -	- - - -	ND	ND
077-0001A2	IN	13	T1	1	N/A	Stover	ND	[0.0072]	- - - -	- - - -
077-0002A1	IN	13	T2	1	90	Stover	- - - -	- - - -	[0.0030]	ND
077-0002A2	IN	13	T2	1	90	Stover	3.1011	3.1748	- - - -	- - - -
077-0003A1	IN	13	T2	2	90	Stover	- - - -	- - - -	ND	ND
077-0003A2	IN	13	T2	2	90	Stover	3.1342	3.0319	- - - -	- - - -
082-0001A1	NE	14	T1	1	N/A	Stover	- - - -	- - - -	ND	ND
082-0001A2	NE	14	T1	1	N/A	Stover	ND	[0.0054]	- - - -	- - - -
082-0002A1	NE	14	T2	1	90	Stover	- - - -	- - - -	ND	ND
082-0002A2	NE	14	T2	1	90	Stover	0.4050	0.3978	- - - -	- - - -
082-0003A1	NE	14	T2	2	90	Stover	- - - -	- - - -	ND	ND
082-0003A2	NE	14	T2	2	90	Stover	0.2504	0.2473	- - - -	- - - -
086-0001A1	NE	15	T1	1	N/A	Stover	- - - -	- - - -	ND	ND
086-0001A2	NE	15	T1	1	N/A	Stover	ND	[0.0048]	- - - -	- - - -
086-0002A1	NE	15	T2	1	90	Stover	- - - -	- - - -	ND	ND
086-0002A2	NE	15	T2	1	90	Stover	[0.0035]	[0.0070]	- - - -	- - - -
086-0003A1	NE	15	T2	2	90	Stover	- - - -	- - - -	ND	ND
086-0003A2	NE	15	T2	2	90	Stover	0.0111	0.0315	- - - -	- - - -
090-0001A1	KS	16	T1	1	N/A	Stover	- - - -	- - - -	ND	ND
090-0001A2	KS	16	T1	1	N/A	Stover	ND	[0.0057]	- - - -	- - - -
090-0002A1	KS	16	T2	1	90	Stover	- - - -	- - - -	ND	ND
090-0002A2	KS	16	T2	1	90	Stover	0.0272	0.0237	- - - -	- - - -
090-0003A1	KS	16	T2	2	90	Stover	- - - -	- - - -	ND	ND
090-0003A2	KS	16	T2	2	90	Stover	0.0295	0.0350	- - - -	- - - -
094-0001A1	IA	17	T1	1	N/A	Stover	- - - -	- - - -	ND	ND
094-0001A2	IA	17	T1	1	N/A	Stover	ND	[0.0033]	- - - -	- - - -
094-0002A1	IA	17	T2	1	90	Stover	- - - -	- - - -	ND	ND
094-0002A2	IA	17	T2	1	90	Stover	0.1586	0.1914	- - - -	- - - -
094-0003A1	IA	17	T2	2	90	Stover	- - - -	- - - -	ND	ND
094-0003A2	IA	17	T2	2	90	Stover	0.0919	0.1267	- - - -	- - - -
098-0001A1	MO	18	T1	1	N/A	Stover	- - - -	- - - -	ND	ND
098-0001A2	MO	18	T1	1	N/A	Stover	ND	[0.0049]	- - - -	- - - -
098-0002A1	MO	18	T2	1	90	Stover	- - - -	- - - -	ND	ND
098-0002A2	MO	18	T2	1	90	Stover	ND	ND	- - - -	- - - -
098-0003A1	MO	18	T2	2	90	Stover	- - - -	- - - -	ND	ND
098-0003A2	MO	18	T2	2	90	Stover	ND	[0.0038]	- - - -	- - - -
102-0001A1	IA	19	T1	1	N/A	Stover	- - - -	- - - -	ND	ND
102-0001A2	IA	19	T1	1	N/A	Stover	ND	ND	- - - -	- - - -
102-0002A1	IA	19	T2	1	90	Stover	- - - -	- - - -	ND	ND
102-0002A2	IA	19	T2	1	90	Stover	1.3106	2.4465	- - - -	- - - -
102-0003A1	IA	19	T2	2	90	Stover	- - - -	- - - -	ND	ND
102-0003A2	IA	19	T2	2	90	Stover	0.7546	1.5763	- - - -	- - - -

Table 7 (Cont.). Summarized Field Sample Information and Residue Results

Sample ID[a]	Site	Trial	Trt	Rep	Event	Commodity	2,4-D Reported, µg/g[b]	2,4-DCP Reported, µg/g[b]	Quizalofop Acid Reported, µg/g[b]	Quizalofop Ethyl Reported, µg/g[b]
106-0001A1	MN	20	T1	1	N/A	Stover	- - - -	- - - -	ND	ND
106-0001A2	MN	20	T1	1	N/A	Stover	ND	[0.0034]	- - - -	- - - -
106-0002A1	MN	20	T2	1	90	Stover	- - - -	- - - -	ND	ND
106-0002A2	MN	20	T2	1	90	Stover	1.1957	1.7164	- - - -	- - - -
106-0003A1	MN	20	T2	2	90	Stover	- - - -	- - - -	ND	ND
106-0003A2	MN	20	T2	2	90	Stover	0.8746	1.1589	- - - -	- - - -
110-0001A1	MO	21	T1	1	N/A	Stover	- - - -	- - - -	ND	ND
110-0001A2	MO	21	T1	1	N/A	Stover	ND	[0.0070]	- - - -	- - - -
110-0002A1	MO	21	T2	1	90	Stover	- - - -	- - - -	ND	ND
110-0002A2	MO	21	T2	1	90	Stover	0.7935	0.2710	- - - -	- - - -
110-0003A1	MO	21	T2	2	90	Stover	- - - -	- - - -	ND	ND
110-0003A2	MO	21	T2	2	90	Stover	0.7685	0.3048	- - - -	- - - -
118-0001A1	OK	22	T1	1	N/A	Stover	- - - -	- - - -	ND	ND
118-0001A2	OK	22	T1	1	N/A	Stover	ND	[0.0030]	- - - -	- - - -
118-0002A1	OK	22	T2	1	76	Stover	- - - -	- - - -	ND	ND
118-0002A2	OK	22	T2	1	76	Stover	0.2865	0.0584	- - - -	- - - -
118-0003A1	OK	22	T2	2	76	Stover	- - - -	- - - -	ND	ND
118-0003A2	OK	22	T2	2	76	Stover	0.2770	0.0550	- - - -	- - - -
120-0001A1	OK	22	T1	1	N/A	Stover	- - - -	- - - -	ND	ND
120-0001A2	OK	22	T1	1	N/A	Stover	ND	[0.0094]	- - - -	- - - -
120-0002A1	OK	22	T2	1	83	Stover	- - - -	- - - -	ND	ND
120-0002A2	OK	22	T2	1	83	Stover	0.2580	0.0427	- - - -	- - - -
120-0003A1	OK	22	T2	2	83	Stover	- - - -	- - - -	ND	ND
120-0003A2	OK	22	T2	2	83	Stover	0.1644	0.0334	- - - -	- - - -
122-0001A1	OK	22	T1	1	N/A	Stover	- - - -	- - - -	ND	ND
122-0001A2	OK	22	T1	1	N/A	Stover	ND	ND	- - - -	- - - -
122-0002A1	OK	22	T2	1	90	Stover	- - - -	- - - -	ND	ND
122-0002A2	OK	22	T2	1	90	Stover	0.1769	0.0352	- - - -	- - - -
122-0003A1	OK	22	T2	2	90	Stover	- - - -	- - - -	ND	ND
122-0003A2	OK	22	T2	2	90	Stover	0.1763	0.0329	- - - -	- - - -
124-0001A1	OK	22	T1	1	N/A	Stover	- - - -	- - - -	ND	ND
124-0001A2	OK	22	T1	1	N/A	Stover	ND	ND	- - - -	- - - -
124-0002A1	OK	22	T2	1	97	Stover	- - - -	- - - -	ND	ND
124-0002A2	OK	22	T2	1	97	Stover	0.2091	0.0396	- - - -	- - - -
124-0003A1	OK	22	T2	2	97	Stover	- - - -	- - - -	ND	ND
124-0003A2	OK	22	T2	2	97	Stover	0.1879	0.0314	- - - -	- - - -
126-0001A1	OK	22	T1	1	N/A	Stover	- - - -	- - - -	ND	ND
126-0001A3	OK	22	T1	1	N/A	Stover	ND	[0.0030]	- - - -	- - - -
126-0002A1	OK	22	T2	1	104	Stover	- - - -	- - - -	ND	ND
126-0002A2	OK	22	T2	1	104	Stover	0.0962	0.0220	- - - -	- - - -
126-0003A1	OK	22	T2	2	104	Stover	- - - -	- - - -	ND	ND
126-0003A2	OK	22	T2	2	104	Stover	0.0985	0.0184	- - - -	- - - -

Table 7 (Cont.). Summarized Field Sample Information and Residue Results

Sample ID[a]	Site	Trial	Trt	Rep	Event	Commodity	2,4-D Reported, µg/g[b]	2,4-DCP Reported, µg/g[b]	Quizalofop Acid Reported, µg/g[b]	Quizalofop Ethyl Reported, µg/g[b]
130-0001A1	TX	23	T1	1	N/A	Stover	- - - -	- - - -	ND	ND
130-0001A3	TX	23	T1	1	N/A	Stover	ND	[0.0060]	- - - -	- - - -
130-0002A1	TX	23	T2	1	90	Stover	- - - -	- - - -	0.0101	[0.0041]
130-0002A2	TX	23	T2	1	90	Stover	0.6024	0.2287	- - - -	- - - -
130-0003A1	TX	23	T2	2	90	Stover	- - - -	- - - -	0.0110	[0.0044]
130-0003A2	TX	23	T2	2	90	Stover	0.8740	0.5168	- - - -	- - - -
134-0001A1	ON	24	T1	1	N/A	Stover	- - - -	- - - -	ND	ND
134-0001A3	ON	24	T1	1	N/A	Stover	ND	[0.0064]	- - - -	- - - -
134-0002A1	ON	24	T2	1	90	Stover	- - - -	- - - -	ND	ND
134-0002A2	ON	24	T2	1	90	Stover	0.2307	0.3140	- - - -	- - - -
134-0003A1	ON	24	T2	2	90	Stover	- - - -	- - - -	ND	ND
134-0003A2	ON	24	T2	2	90	Stover	0.3555	0.4303	- - - -	- - - -
138-0001A1	ON	25	T1	1	N/A	Stover	- - - -	- - - -	ND	ND
138-0001A3	ON	25	T1	1	N/A	Stover	ND	ND	- - - -	- - - -
138-0002A1	ON	25	T2	1	90	Stover	- - - -	- - - -	ND	ND
138-0002A2	ON	25	T2	1	90	Stover	0.1271	0.6180	- - - -	- - - -
138-0003A1	ON	25	T2	2	90	Stover	- - - -	- - - -	ND	[0.0038]
138-0003A2	ON	25	T2	2	90	Stover	0.1704	0.6744	- - - -	- - - -
158-0001A1	GA	4	T1	1	N/A	Stover	- - - -	- - - -	ND	ND
158-0001A13	GA	4	T1	1	N/A	Stover	- - - -	- - - -	ND	ND
158-0001A7	GA	4	T1	1	N/A	Stover	- - - -	- - - -	ND	ND
142-0001A1	GA	3	T1	1	N/A	Starch	- - - -	- - - -	ND	ND
142-0001A30	GA	3	T1	1	N/A	Starch	ND	ND	- - - -	- - - -
142-0001A9	GA	3	T1	1	N/A	Starch	- - - -	- - - -	ND	ND
142-0002A1	GA	3	T1	2	N/A	Starch	- - - -	- - - -	ND	ND
142-0002A2	GA	3	T1	2	N/A	Starch	ND	ND	- - - -	- - - -
142-0003A1	GA	3	T4	1	90	Starch	- - - -	- - - -	ND	ND
142-0003A3	GA	3	T4	1	90	Starch	ND	ND	- - - -	- - - -
142-0004A1	GA	14	T4	2	90	Starch	- - - -	- - - -	ND	ND
142-0004A4	GA	3	T4	2	90	Starch	ND	ND	- - - -	- - - -
151-0001A1	NE	14	T1	1	N/A	Starch	- - - -	- - - -	ND	ND
151-0001A2	NE	14	T1	1	N/A	Starch	ND	ND	- - - -	- - - -
151-0002A1	NE	14	T1	2	N/A	Starch	- - - -	- - - -	ND	ND
151-0002A2	NE	14	T1	2	N/A	Starch	ND	ND	- - - -	- - - -
151-0003A1	NE	14	T4	1	90	Starch	- - - -	- - - -	ND	ND
151-0003A4	NE	14	T4	1	90	Starch	ND	ND	- - - -	- - - -
151-0004A1	NE	3	T4	2	90	Starch	- - - -	- - - -	ND	[0.0051]
151-0004A4	NE	14	T4	2	90	Starch	ND	ND	- - - -	- - - -
143-0001A1	GA	3	T1	1	N/A	Oil, refined	- - - -	- - - -	ND	ND
143-0001A30	GA	3	T1	1	N/A	Oil, refined	ND	ND	- - - -	- - - -
143-0001A9	GA	3	T1	1	N/A	Oil, refined	- - - -	- - - -	ND	ND
143-0002A1	GA	3	T1	2	N/A	Oil, refined	- - - -	- - - -	ND	[0.0034]

Table 7 (Cont.). Summarized Field Sample Information and Residue Results

Sample ID[a]	Site	Trial	Trt	Rep	Event	Commodity	2,4-D Reported, µg/g[b]	2,4-DCP Reported, µg/g[b]	Quizalofop Acid Reported, µg/g[b]	Quizalofop Ethyl Reported, µg/g[b]
143-0002A2	GA	3	T1	2	N/A	Oil, refined	ND	ND	- - - -	- - - -
143-0003A1	GA	3	T4	1	90	Oil, refined	- - - -	- - - -	ND	ND
143-0003A3	GA	3	T4	1	90	Oil, refined	ND	ND	- - - -	- - - -
143-0004A1	GA	3	T4	2	90	Oil, refined	- - - -	- - - -	ND	ND
143-0004A3	GA	3	T4	2	90	Oil, refined	ND	ND	- - - -	- - - -
147-0001A1	GA	3	T1	1	N/A	Oil, refined	- - - -	- - - -	ND	ND
147-0001A30	GA	3	T1	1	N/A	Oil, refined	ND	ND	- - - -	- - - -
147-0001A9	GA	3	T1	1	N/A	Oil, refined	- - - -	- - - -	ND	ND
147-0002A1	GA	3	T1	2	N/A	Oil, refined	- - - -	- - - -	ND	ND
147-0002A2	GA	3	T1	2	N/A	Oil, refined	ND	ND	- - - -	- - - -
147-0003A1	GA	3	T4	1	90	Oil, refined	- - - -	- - - -	ND	ND
147-0003A3	GA	3	T4	1	90	Oil, refined	ND	ND	- - - -	- - - -
147-0004A1	GA	3	T4	2	90	Oil, refined	- - - -	- - - -	ND	ND
147-0004A3	GA	3	T4	2	90	Oil, refined	ND	ND	- - - -	- - - -
152-0001A1	NE	14	T1	1	N/A	Oil, refined	- - - -	- - - -	ND	ND
152-0001A2	NE	14	T1	1	N/A	Oil, refined	ND	ND	- - - -	- - - -
152-0002A1	NE	14	T1	2	N/A	Oil, refined	- - - -	- - - -	ND	ND
152-0002A2	NE	14	T1	2	N/A	Oil, refined	ND	ND	- - - -	- - - -
152-0003A1	NE	14	T4	1	90	Oil, refined	- - - -	- - - -	ND	ND
152-0003A3	NE	14	T4	1	90	Oil, refined	ND	ND	- - - -	- - - -
152-0004A1	NE	14	T4	2	90	Oil, refined	- - - -	- - - -	ND	ND
152-0004A3	NE	14	T4	2	90	Oil, refined	ND	ND	- - - -	- - - -
156-0001A1	NE	14	T1	1	N/A	Oil, refined	- - - -	- - - -	ND	ND
156-0001A2	NE	14	T1	1	N/A	Oil, refined	ND	ND	- - - -	- - - -
156-0002A1	NE	14	T1	2	N/A	Oil, refined	- - - -	- - - -	ND	[0.0056]
156-0002A2	NE	14	T1	2	N/A	Oil, refined	ND	ND	- - - -	- - - -
156-0003A1	NE	14	T4	1	90	Oil, refined	- - - -	- - - -	ND	ND
156-0003A3	NE	14	T4	1	90	Oil, refined	ND	ND	- - - -	- - - -
156-0004A1	NE	14	T4	2	90	Oil, refined	- - - -	- - - -	ND	ND
156-0004A3	NE	14	T4	2	90	Oil, refined	ND	ND	- - - -	- - - -
144-0001A1	GA	3	T1	1	N/A	Meal	- - - -	- - - -	ND	ND
144-0001A22	GA	3	T1	1	N/A	Meal	ND	ND	- - - -	- - - -
144-0001A9	GA	3	T1	1	N/A	Meal	- - - -	- - - -	ND	ND
144-0002A1	GA	3	T1	2	N/A	Meal	- - - -	- - - -	ND	ND
144-0002A2	GA	3	T1	2	N/A	Meal	ND	ND	- - - -	- - - -
144-0003A1	GA	3	T4	1	90	Meal	- - - -	- - - -	ND	ND
144-0003A2	GA	3	T4	1	90	Meal	ND	ND	- - - -	- - - -
144-0004A1	GA	3	T4	2	90	Meal	- - - -	- - - -	ND	ND
144-0004A2	GA	3	T4	2	90	Meal	ND	ND	- - - -	- - - -
153-0001A1	NE	14	T1	1	N/A	Meal	- - - -	- - - -	ND	ND
153-0001A2	NE	14	T1	1	N/A	Meal	ND	ND	- - - -	- - - -
153-0002A1	NE	14	T1	2	N/A	Meal	- - - -	- - - -	ND	[0.0031]

Table 7 (Cont.). Summarized Field Sample Information and Residue Results

Sample ID[a]	Site	Trial	Trt	Rep	Event	Commodity	2,4-D Reported, µg/g[b]	2,4-DCP Reported, µg/g[b]	Quizalofop Acid Reported, µg/g[b]	Quizalofop Ethyl Reported, µg/g[b]
153-0002A2	NE	14	T1	2	N/A	Meal	ND	ND	- - - -	- - - -
153-0003A1	NE	14	T4	1	90	Meal	- - - -	- - - -	ND	ND
153-0003A2	NE	14	T4	1	90	Meal	ND	ND	- - - -	- - - -
153-0004A1	NE	14	T4	2	90	Meal	- - - -	- - - -	ND	ND
153-0004A2	NE	14	T4	2	90	Meal	ND	ND	- - - -	- - - -
145-0001A1	GA	3	T1	1	N/A	Grits	- - - -	- - - -	ND	ND
145-0001A22	GA	3	T1	1	N/A	Grits	ND	ND	- - - -	- - - -
145-0001A9	GA	3	T1	1	N/A	Grits	- - - -	- - - -	ND	ND
145-0002A1	GA	3	T1	2	N/A	Grits	- - - -	- - - -	ND	ND
145-0002A2	GA	3	T1	2	N/A	Grits	ND	ND	- - - -	- - - -
145-0003A1	GA	3	T4	1	90	Grits	- - - -	- - - -	ND	ND
145-0003A2	GA	3	T4	1	90	Grits	ND	ND	- - - -	- - - -
145-0004A1	GA	3	T4	2	90	Grits	- - - -	- - - -	ND	ND
145-0004A2	GA	3	T4	2	90	Grits	ND	ND	- - - -	- - - -
154-0001A1	NE	14	T1	1	N/A	Grits	- - - -	- - - -	ND	ND
154-0001A32	NE	14	T1	1	N/A	Grits	ND	ND	- - - -	- - - -
154-0002A1	NE	14	T1	2	N/A	Grits	- - - -	- - - -	ND	ND
154-0002A2	NE	14	T1	2	N/A	Grits	ND	ND	- - - -	- - - -
154-0003A1	NE	14	T4	1	90	Grits	- - - -	- - - -	ND	ND
154-0003A2	NE	14	T4	1	90	Grits	ND	ND	- - - -	- - - -
154-0004A1	NE	14	T4	2	90	Grits	- - - -	- - - -	ND	ND
154-0004A2	NE	14	T4	2	90	Grits	ND	ND	- - - -	- - - -
003-0001A1	NJ	1	T1	1	N/A	Grain	- - - -	- - - -	ND	ND
003-0001A2	NJ	1	T1	1	N/A	Grain	ND	ND	- - - -	- - - -
003-0002A1	NJ	1	T2	1	90	Grain	- - - -	- - - -	ND	ND
003-0002A2	NJ	1	T2	1	90	Grain	ND	ND	- - - -	- - - -
003-0003A1	NJ	1	T2	2	90	Grain	- - - -	- - - -	ND	ND
003-0003A2	NJ	1	T2	2	90	Grain	ND	ND	- - - -	- - - -
007-0001A1	PA	2	T1	1	N/A	Grain	- - - -	- - - -	ND	ND
007-0001A2	PA	2	T1	1	N/A	Grain	ND	ND	- - - -	- - - -
007-0002A1	PA	2	T2	1	90	Grain	- - - -	- - - -	ND	ND
007-0002A2	PA	2	T2	1	90	Grain	ND	ND	- - - -	- - - -
007-0003A1	PA	2	T2	2	90	Grain	- - - -	- - - -	ND	ND
007-0003A2	PA	2	T2	2	90	Grain	ND	ND	- - - -	- - - -
011-0001A1	GA	3	T1	1	N/A	Grain	- - - -	- - - -	ND	ND
011-0001A14	GA	3	T1	1	N/A	Grain	ND	ND	- - - -	- - - -
011-0001A2	GA	3	T1	1	N/A	Grain	ND	ND	- - - -	- - - -
011-0001A8	GA	3	T1	1	N/A	Grain	ND	ND	- - - -	- - - -
011-0002A1	GA	3	T2	1	90	Grain	- - - -	- - - -	ND	ND
011-0002A2	GA	3	T2	1	90	Grain	ND	ND	- - - -	- - - -
011-0003A1	GA	3	T2	2	90	Grain	- - - -	- - - -	ND	ND
011-0003A2	GA	3	T2	2	90	Grain	ND	ND	- - - -	- - - -

Table 7 (Cont.). Summarized Field Sample Information and Residue Results

Sample ID[a]	Site	Trial	Trt	Rep	Event	Commodity	2,4-D Reported, µg/g[b]	2,4-DCP Reported, µg/g[b]	Quizalofop Acid Reported, µg/g[b]	Quizalofop Ethyl Reported, µg/g[b]
011-0006A1	GA	3	T4	1	90	Grain	- - - -	- - - -	ND	ND
011-0006A2	GA	3	T4	1	90	Grain	ND	ND	- - - -	- - - -
011-0007A1	GA	3	T4	2	90	Grain	- - - -	- - - -	[0.0032]	ND
011-0007A2	GA	3	T4	2	90	Grain	ND	ND	- - - -	- - - -
020-0001A14	GA	4	T1	1	N/A	Grain	- - - -	- - - -	ND	ND
020-0001A2	GA	4	T1	1	N/A	Grain	- - - -	- - - -	ND	ND
020-0001A44	GA	4	T1	1	N/A	Grain	ND	ND	- - - -	- - - -
020-0001A8	GA	4	T1	1	N/A	Grain	- - - -	- - - -	ND	ND
020-0002A1	GA	4	T2	1	76	Grain	- - - -	- - - -	ND	ND
020-0002A2	GA	4	T2	1	76	Grain	ND	ND	- - - -	- - - -
020-0003A1	GA	4	T2	2	76	Grain	- - - -	- - - -	ND	ND
020-0003A2	GA	4	T2	2	76	Grain	ND	ND	- - - -	- - - -
022-0001A1	GA	4	T1	1	N/A	Grain	- - - -	- - - -	ND	ND
022-0001A2	GA	4	T1	1	N/A	Grain	ND	ND	- - - -	- - - -
022-0002A1	GA	4	T2	1	83	Grain	- - - -	- - - -	ND	ND
022-0002A2	GA	4	T2	1	83	Grain	ND	ND	- - - -	- - - -
022-0003A1	GA	4	T2	2	83	Grain	- - - -	- - - -	ND	ND
022-0003A2	GA	4	T2	2	83	Grain	ND	ND	- - - -	- - - -
024-0001A1	GA	4	T1	1	N/A	Grain	- - - -	- - - -	ND	ND
024-0001A2	GA	4	T1	1	N/A	Grain	ND	ND	- - - -	- - - -
024-0002A1	GA	4	T2	1	90	Grain	- - - -	- - - -	ND	ND
024-0002A2	GA	4	T2	1	90	Grain	ND	ND	- - - -	- - - -
024-0003A1	GA	4	T2	2	90	Grain	- - - -	- - - -	ND	ND
024-0003A2	GA	4	T2	2	90	Grain	ND	ND	- - - -	- - - -
026-0001A1	GA	4	T1	1	N/A	Grain	- - - -	- - - -	ND	ND
026-0001A2	GA	4	T1	1	N/A	Grain	ND	ND	- - - -	- - - -
026-0002A1	GA	4	T2	1	97	Grain	- - - -	- - - -	ND	ND
026-0002A2	GA	4	T2	1	97	Grain	ND	ND	- - - -	- - - -
026-0003A1	GA	4	T2	2	97	Grain	- - - -	- - - -	ND	ND
026-0003A2	GA	4	T2	2	97	Grain	ND	ND	- - - -	- - - -
028-0001A1	GA	4	T1	1	N/A	Grain	- - - -	- - - -	ND	ND
028-0001A2	GA	4	T1	1	N/A	Grain	ND	ND	- - - -	- - - -
028-0002A1	GA	4	T2	1	104	Grain	- - - -	- - - -	ND	ND
028-0002A2	GA	4	T2	1	104	Grain	ND	ND	- - - -	- - - -
028-0003A1	GA	4	T2	2	104	Grain	- - - -	- - - -	ND	ND
028-0003A2	GA	4	T2	2	104	Grain	ND	ND	- - - -	- - - -
032-0001A1	IL	5	T1	1	N/A	Grain	- - - -	- - - -	ND	ND
032-0001A2	IL	5	T1	1	N/A	Grain	ND	ND	- - - -	- - - -
032-0002A1	IL	5	T2	1	90	Grain	- - - -	- - - -	ND	ND
032-0002A2	IL	5	T2	1	90	Grain	ND	ND	- - - -	- - - -
032-0002A3	IL	5	T2	1	90	Grain	ND	ND	- - - -	- - - -
032-0002A4	IL	5	T2	1	90	Grain	ND	ND	- - - -	- - - -
032-0003A1	IL	5	T2	2	90	Grain	- - - -	- - - -	ND	ND
032-0003A2	IL	5	T2	2	90	Grain	ND	ND	- - - -	- - - -

Table 7 (Cont.). Summarized Field Sample Information and Residue Results

Sample ID[a]	Site	Trial	Trt	Rep	Event	Commodity	2,4-D Reported, µg/g[b]	2,4-DCP Reported, µg/g[b]	Quizalofop Acid Reported, µg/g[b]	Quizalofop Ethyl Reported, µg/g[b]
036-0001A1	IL	6	T1	1	N/A	Grain	- - - -	- - - -	ND	ND
036-0001A2	IL	6	T1	1	N/A	Grain	ND	ND	- - - -	- - - -
036-0002A1	IL	6	T2	1	90	Grain	- - - -	- - - -	ND	ND
036-0002A2	IL	6	T2	1	90	Grain	ND	ND	- - - -	- - - -
036-0003A1	IL	6	T2	2	90	Grain	- - - -	- - - -	[0.0041]	ND
036-0003A2	IL	6	T2	2	90	Grain	ND	ND	- - - -	- - - -
040-0001A1	IN	7	T1	1	N/A	Grain	- - - -	- - - -	ND	ND
040-0001A2	IN	7	T1	1	N/A	Grain	ND	ND	- - - -	- - - -
040-0002A1	IN	7	T2	1	90	Grain	- - - -	- - - -	ND	ND
040-0002A2	IN	7	T2	1	90	Grain	ND	ND	- - - -	- - - -
040-0003A1	IN	7	T2	2	90	Grain	- - - -	- - - -	ND	ND
040-0003A2	IN	7	T2	2	90	Grain	ND	ND	- - - -	- - - -
044-0001A1	MO	8	T1	1	N/A	Grain	- - - -	- - - -	ND	ND
044-0001A2	MO	8	T1	1	N/A	Grain	ND	ND	- - - -	- - - -
044-0002A1	MO	8	T2	1	90	Grain	- - - -	- - - -	ND	ND
044-0002A4	MO	8	T2	1	90	Grain	0.0671	ND	- - - -	- - - -
044-0003A1	MO	8	T2	2	90	Grain	- - - -	- - - -	ND	ND
044-0003A4	MO	8	T2	2	90	Grain	ND	ND	- - - -	- - - -
048-0001A1	MO	9	T1	1	N/A	Grain	- - - -	- - - -	ND	ND
048-0001A2	MO	9	T1	1	N/A	Grain	ND	ND	- - - -	- - - -
048-0002A1	MO	9	T2	1	90	Grain	- - - -	- - - -	ND	ND
048-0002A2	MO	9	T2	1	90	Grain	ND	ND	- - - -	- - - -
048-0003A1	MO	9	T2	2	90	Grain	- - - -	- - - -	ND	ND
048-0003A2	MO	9	T2	2	90	Grain	ND	ND	- - - -	- - - -
052-0001A1	IL	10	T1	1	N/A	Grain	- - - -	- - - -	ND	ND
052-0001A2	IL	10	T1	1	N/A	Grain	ND	[0.0067]	- - - -	- - - -
052-0002A1	IL	10	T2	1	90	Grain	- - - -	- - - -	ND	ND
052-0002A2	IL	10	T2	1	90	Grain	ND	ND	- - - -	- - - -
052-0003A1	IL	10	T2	2	90	Grain	- - - -	- - - -	ND	ND
052-0003A2	IL	10	T2	2	90	Grain	ND	ND	- - - -	- - - -
060-0001A1	MI	11	T1	1	N/A	Grain	- - - -	- - - -	ND	ND
060-0001A2	MI	11	T1	1	N/A	Grain	ND	ND	- - - -	- - - -
060-0002A1	MI	11	T2	1	76	Grain	- - - -	- - - -	ND	ND
060-0002A2	MI	11	T2	1	76	Grain	ND	[0.0042]	- - - -	- - - -
060-0003A1	MI	11	T2	2	76	Grain	- - - -	- - - -	ND	ND
060-0003A2	MI	11	T2	2	76	Grain	ND	[0.0036]	- - - -	- - - -
062-0001A1	MI	11	T1	1	N/A	Grain	- - - -	- - - -	ND	ND
062-0001A2	MI	11	T1	1	N/A	Grain	ND	ND	- - - -	- - - -
062-0002A1	MI	11	T2	1	83	Grain	- - - -	- - - -	ND	ND
062-0002A2	MI	11	T2	1	83	Grain	ND	[0.0031]	- - - -	- - - -
062-0003A1	MI	11	T2	2	83	Grain	- - - -	- - - -	ND	ND
062-0003A2	MI	11	T2	2	83	Grain	ND	[0.0033]	- - - -	- - - -

Table 7 (Cont.). Summarized Field Sample Information and Residue Results

Sample ID[a]	Site	Trial	Trt	Rep	Event	Commodity	2,4-D Reported, µg/g[b]	2,4-DCP Reported, µg/g[b]	Quizalofop Acid Reported, µg/g[b]	Quizalofop Ethyl Reported, µg/g[b]
064-0001A1	MI	11	T1	1	N/A	Grain	- - - -	- - - -	ND	ND
064-0001A2	MI	11	T1	1	N/A	Grain	ND	ND	- - - -	- - - -
064-0002A1	MI	11	T2	1	90	Grain	- - - -	- - - -	ND	ND
064-0002A2	MI	11	T2	1	90	Grain	ND	ND	- - - -	- - - -
064-0003A1	MI	11	T2	2	90	Grain	- - - -	- - - -	ND	ND
064-0003A2	MI	11	T2	2	90	Grain	ND	[0.0033]	- - - -	- - - -
066-0001A1	MI	11	T1	1	N/A	Grain	- - - -	- - - -	ND	ND
066-0001A2	MI	11	T1	1	N/A	Grain	ND	ND	- - - -	- - - -
066-0002A1	MI	11	T2	1	97	Grain	- - - -	- - - -	ND	ND
066-0002A2	MI	11	T2	1	97	Grain	ND	[0.0044]	- - - -	- - - -
066-0003A1	MI	11	T2	2	97	Grain	- - - -	- - - -	ND	ND
066-0003A2	MI	11	T2	2	97	Grain	ND	[0.0036]	- - - -	- - - -
068-0001A1	MI	11	T1	1	N/A	Grain	- - - -	- - - -	ND	ND
068-0001A2	MI	11	T1	1	N/A	Grain	ND	ND	- - - -	- - - -
068-0002A1	MI	11	T2	1	104	Grain	- - - -	- - - -	ND	ND
068-0002A2	MI	11	T2	1	104	Grain	ND	ND	- - - -	- - - -
068-0003A1	MI	11	T2	2	104	Grain	- - - -	- - - -	ND	ND
068-0003A2	MI	11	T2	2	104	Grain	ND	[0.0043]	- - - -	- - - -
072-0001A1	OH	12	T1	1	N/A	Grain	- - - -	- - - -	ND	ND
072-0001A2	OH	12	T1	1	N/A	Grain	ND	ND	- - - -	- - - -
072-0002A1	OH	12	T2	1	90	Grain	- - - -	- - - -	ND	ND
072-0002A2	OH	12	T2	1	90	Grain	[0.0037]	ND	- - - -	- - - -
072-0003A1	OH	12	T2	2	90	Grain	- - - -	- - - -	ND	ND
072-0003A2	OH	12	T2	2	90	Grain	ND	ND	- - - -	- - - -
076-0001A1	IN	13	T1	1	N/A	Grain	- - - -	- - - -	ND	ND
076-0001A2	IN	13	T1	1	N/A	Grain	ND	ND	- - - -	- - - -
076-0002A1	IN	13	T2	1	90	Grain	- - - -	- - - -	ND	ND
076-0002A2	IN	13	T2	1	90	Grain	ND	ND	- - - -	- - - -
076-0003A1	IN	13	T2	2	90	Grain	- - - -	- - - -	ND	ND
076-0003A2	IN	13	T2	2	90	Grain	ND	ND	- - - -	- - - -
080-0001A1	NE	14	T1	1	N/A	Grain	- - - -	- - - -	ND	ND
080-0001A2	NE	14	T1	1	N/A	Grain	ND	ND	- - - -	- - - -
080-0002A1	NE	14	T2	1	90	Grain	- - - -	- - - -	ND	ND
080-0002A2	NE	14	T2	1	90	Grain	ND	ND	- - - -	- - - -
080-0003A1	NE	14	T2	2	90	Grain	- - - -	- - - -	ND	ND
080-0003A2	NE	14	T2	2	90	Grain	ND	ND	- - - -	- - - -
080-0006A1	NE	14	T4	1	90	Grain	- - - -	- - - -	ND	ND
080-0006A2	NE	14	T4	1	90	Grain	ND	ND	- - - -	- - - -
080-0007A1	NE	14	T4	2	90	Grain	- - - -	- - - -	ND	ND
080-0007A2	NE	14	T4	2	90	Grain	ND	ND	- - - -	- - - -
085-0001A1	NE	15	T1	1	N/A	Grain	- - - -	- - - -	ND	ND
085-0001A2	NE	15	T1	1	N/A	Grain	ND	ND	- - - -	- - - -

Table 7 (Cont.). Summarized Field Sample Information and Residue Results

Sample ID[a]	Site	Trial	Trt	Rep	Event	Commodity	2,4-D Reported, µg/g[b]	2,4-DCP Reported, µg/g[b]	Quizalofop Acid Reported, µg/g[b]	Quizalofop Ethyl Reported, µg/g[b]
085-0002A1	NE	15	T2	1	90	Grain	- - - -	- - - -	ND	ND
085-0002A2	NE	15	T2	1	90	Grain	ND	ND	- - - -	- - - -
085-0003A1	NE	15	T2	2	90	Grain	- - - -	- - - -	ND	ND
085-0003A2	NE	15	T2	2	90	Grain	ND	ND	- - - -	- - - -
089-0001A1	KS	16	T1	1	N/A	Grain	- - - -	- - - -	ND	ND
089-0001A2	KS	16	T1	1	N/A	Grain	ND	[0.0033]	- - - -	- - - -
089-0002A1	KS	16	T2	1	90	Grain	- - - -	- - - -	ND	ND
089-0002A2	KS	16	T2	1	90	Grain	ND	ND	- - - -	- - - -
089-0003A1	KS	16	T2	2	90	Grain	- - - -	- - - -	ND	ND
089-0003A2	KS	16	T2	2	90	Grain	ND	ND	- - - -	- - - -
093-0001A1	IA	17	T1	1	N/A	Grain	- - - -	- - - -	ND	ND
093-0001A2	IA	17	T1	1	N/A	Grain	ND	ND	- - - -	- - - -
093-0002A1	IA	17	T2	1	90	Grain	- - - -	- - - -	ND	ND
093-0002A2	IA	17	T2	1	90	Grain	ND	ND	- - - -	- - - -
093-0003A1	IA	17	T2	2	90	Grain	- - - -	- - - -	ND	ND
093-0003A2	IA	17	T2	2	90	Grain	ND	ND	- - - -	- - - -
097-0001A1	MO	18	T1	1	N/A	Grain	- - - -	- - - -	ND	ND
097-0001A2	MO	18	T1	1	N/A	Grain	ND	ND	- - - -	- - - -
097-0002A1	MO	18	T2	1	90	Grain	- - - -	- - - -	ND	ND
097-0002A2	MO	18	T2	1	90	Grain	ND	ND	- - - -	- - - -
097-0003A1	MO	18	T2	2	90	Grain	- - - -	- - - -	ND	ND
097-0003A2	MO	18	T2	2	90	Grain	ND	ND	- - - -	- - - -
101-0001A1	IA	19	T1	1	N/A	Grain	- - - -	- - - -	ND	ND
101-0001A2	IA	19	T1	1	N/A	Grain	ND	ND	- - - -	- - - -
101-0002A1	IA	19	T2	1	90	Grain	- - - -	- - - -	ND	ND
101-0002A2	IA	19	T2	1	90	Grain	ND	ND	- - - -	- - - -
101-0003A1	IA	19	T2	2	90	Grain	- - - -	- - - -	ND	ND
101-0003A2	IA	19	T2	2	90	Grain	ND	ND	- - - -	- - - -
105-0001A1	MN	20	T1	1	N/A	Grain	- - - -	- - - -	ND	ND
105-0001A3	MN	20	T1	1	N/A	Grain	ND	ND	- - - -	- - - -
105-0002A1	MN	20	T2	1	90	Grain	- - - -	- - - -	ND	ND
105-0002A3	MN	20	T2	1	90	Grain	ND	ND	- - - -	- - - -
105-0003A1	MN	20	T2	2	90	Grain	- - - -	- - - -	ND	ND
105-0003A3	MN	20	T2	2	90	Grain	ND	ND	- - - -	- - - -
109-0001A1	MO	21	T1	1	N/A	Grain	- - - -	- - - -	ND	ND
109-0001A2	MO	21	T1	1	N/A	Grain	ND	ND	- - - -	- - - -
109-0002A1	MO	21	T2	1	90	Grain	- - - -	- - - -	ND	ND
109-0002A2	MO	21	T2	1	90	Grain	ND	ND	- - - -	- - - -
109-0003A1	MO	21	T2	2	90	Grain	- - - -	- - - -	ND	ND
109-0003A2	MO	21	T2	2	90	Grain	ND	ND	- - - -	- - - -
117-0001A1	OK	22	T1	1	N/A	Grain	- - - -	- - - -	ND	ND
117-0001A2	OK	22	T1	1	N/A	Grain	ND	ND	- - - -	- - - -

Table 7 (Cont.). Summarized Field Sample Information and Residue Results

Sample ID[a]	Site	Trial	Trt	Rep	Event	Commodity	2,4-D Reported, µg/g[b]	2,4-DCP Reported, µg/g[b]	Quizalofop Acid Reported, µg/g[b]	Quizalofop Ethyl Reported, µg/g[b]
117-0002A1	OK	22	T2	1	76	Grain	- - - -	- - - -	ND	ND
117-0002A2	OK	22	T2	1	76	Grain	ND	ND	- - - -	- - - -
117-0003A1	OK	22	T2	2	76	Grain	- - - -	- - - -	ND	ND
117-0003A2	OK	22	T2	2	76	Grain	ND	ND	- - - -	- - - -
119-0001A1	OK	22	T1	1	N/A	Grain	- - - -	- - - -	ND	ND
119-0001A2	OK	22	T1	1	N/A	Grain	ND	ND	- - - -	- - - -
119-0002A1	OK	22	T2	1	83	Grain	- - - -	- - - -	ND	ND
119-0002A2	OK	22	T2	1	83	Grain	ND	ND	- - - -	- - - -
119-0003A1	OK	22	T2	2	83	Grain	- - - -	- - - -	ND	ND
119-0003A2	OK	22	T2	2	83	Grain	ND	ND	- - - -	- - - -
121-0001A1	OK	22	T1	1	N/A	Grain	- - - -	- - - -	ND	ND
121-0001A2	OK	22	T1	1	N/A	Grain	ND	ND	- - - -	- - - -
121-0002A1	OK	22	T2	1	90	Grain	- - - -	- - - -	ND	ND
121-0002A2	OK	22	T2	1	90	Grain	[0.0036]	ND	- - - -	- - - -
121-0003A1	OK	22	T2	2	90	Grain	- - - -	- - - -	ND	ND
121-0003A2	OK	22	T2	2	90	Grain	[0.0040]	ND	- - - -	- - - -
123-0001A1	OK	22	T1	1	N/A	Grain	- - - -	- - - -	ND	ND
123-0001A2	OK	22	T1	1	N/A	Grain	ND	ND	- - - -	- - - -
123-0002A1	OK	22	T2	1	97	Grain	- - - -	- - - -	ND	ND
123-0002A2	OK	22	T2	1	97	Grain	ND	ND	- - - -	- - - -
123-0003A1	OK	22	T2	2	97	Grain	- - - -	- - - -	ND	ND
123-0003A2	OK	22	T2	2	97	Grain	ND	ND	- - - -	- - - -
125-0001A1	OK	22	T1	1	N/A	Grain	- - - -	- - - -	ND	ND
125-0001A2	OK	22	T1	1	N/A	Grain	ND	ND	- - - -	- - - -
125-0002A1	OK	22	T2	1	104	Grain	- - - -	- - - -	ND	ND
125-0002A2	OK	22	T2	1	104	Grain	ND	ND	- - - -	- - - -
125-0003A1	OK	22	T2	2	104	Grain	- - - -	- - - -	ND	ND
125-0003A2	OK	22	T2	2	104	Grain	ND	ND	- - - -	- - - -
129-0001A1	TX	23	T1	1	N/A	Grain	- - - -	- - - -	ND	ND
129-0001A2	TX	23	T1	1	N/A	Grain	ND	ND	- - - -	- - - -
129-0002A1	TX	23	T2	1	90	Grain	- - - -	- - - -	ND	ND
129-0002A2	TX	23	T2	1	90	Grain	ND	ND	- - - -	- - - -
129-0003A1	TX	23	T2	2	90	Grain	- - - -	- - - -	ND	ND
129-0003A2	TX	23	T2	2	90	Grain	ND	ND	- - - -	- - - -
133-0001A1	ON	24	T1	1	N/A	Grain	- - - -	- - - -	ND	ND
133-0001A2	ON	24	T1	1	N/A	Grain	ND	ND	- - - -	- - - -
133-0002A1	ON	24	T2	1	90	Grain	- - - -	- - - -	ND	ND
133-0002A2	ON	24	T2	1	90	Grain	ND	ND	- - - -	- - - -
133-0003A1	ON	24	T2	2	90	Grain	- - - -	- - - -	ND	ND
133-0003A2	ON	24	T2	2	90	Grain	ND	ND	- - - -	- - - -
137-0001A1	ON	25	T1	1	N/A	Grain	- - - -	- - - -	[0.0061]	[0.0030]
137-0001A2	ON	25	T1	1	N/A	Grain	ND	ND	- - - -	- - - -

Table 7 (Cont.). Summarized Field Sample Information and Residue Results

Sample ID[a]	Site	Trial	Trt	Rep	Event	Commodity	2,4-D Reported, µg/g[b]	2,4-DCP Reported, µg/g[b]	Quizalofop Acid Reported, µg/g[b]	Quizalofop Ethyl Reported, µg/g[b]
137-0002A1	ON	25	T2	1	90	Grain	- - - -	- - - -	ND	ND
137-0002A2	ON	25	T2	1	90	Grain	ND	ND	- - - -	- - - -
137-0003A1	ON	25	T2	2	90	Grain	- - - -	- - - -	ND	ND
137-0003A2	ON	25	T2	2	90	Grain	ND	ND	- - - -	- - - -
139-0001A1	GA	3	T1	1	N/A	Grain	- - - -	- - - -	ND	ND
139-0001A2	GA	3	T1	1	N/A	Grain	ND	ND	- - - -	- - - -
139-0002A1	GA	3	T1	2	N/A	Grain	- - - -	- - - -	ND	ND
139-0002A2	GA	3	T1	2	N/A	Grain	ND	ND	- - - -	- - - -
139-0003A1	GA	3	T1	3	N/A	Grain	- - - -	- - - -	ND	ND
139-0003A2	GA	3	T1	3	N/A	Grain	ND	ND	- - - -	- - - -
139-0004A1	GA	3	T4	1	90	Grain	- - - -	- - - -	ND	ND
139-0004A3	GA	3	T4	1	90	Grain	ND	[0.0030]	- - - -	- - - -
139-0005A1	GA	3	T4	2	90	Grain	- - - -	- - - -	ND	ND
139-0005A2	GA	3	T4	2	90	Grain	ND	ND	- - - -	- - - -
139-0006A1	GA	3	T4	3	90	Grain	- - - -	- - - -	ND	ND
139-0006A2	GA	3	T4	3	90	Grain	ND	ND	- - - -	- - - -
140-0001A1	GA	3	T1	1	N/A	Grain	- - - -	- - - -	ND	ND
140-0001A2	GA	3	T1	1	N/A	Grain	ND	ND	- - - -	- - - -
140-0002A1	GA	3	T1	2	N/A	Grain	- - - -	- - - -	ND	ND
140-0002A2	GA	3	T1	2	N/A	Grain	ND	ND	- - - -	- - - -
140-0003A1	GA	3	T1	3	N/A	Grain	- - - -	- - - -	ND	ND
140-0003A2	GA	3	T1	3	N/A	Grain	ND	ND	- - - -	- - - -
140-0004A1	GA	3	T4	1	90	Grain	- - - -	- - - -	ND	ND
140-0004A3	GA	3	T4	1	90	Grain	ND	ND	- - - -	- - - -
140-0005A1	GA	3	T4	2	90	Grain	- - - -	- - - -	ND	ND
140-0005A2	GA	3	T4	2	90	Grain	ND	ND	- - - -	- - - -
140-0006A1	GA	3	T4	3	90	Grain	- - - -	- - - -	ND	ND
140-0006A2	GA	3	T4	3	90	Grain	ND	ND	- - - -	- - - -
148-0001A1	NE	14	T1	1	N/A	Grain	- - - -	- - - -	ND	ND
148-0001A2	NE	14	T1	1	N/A	Grain	ND	ND	- - - -	- - - -
148-0002A1	NE	14	T1	2	N/A	Grain	- - - -	- - - -	ND	ND
148-0002A2	NE	14	T1	2	N/A	Grain	ND	ND	- - - -	- - - -
148-0003A1	NE	14	T1	3	N/A	Grain	- - - -	- - - -	ND	ND
148-0003A2	NE	14	T1	3	N/A	Grain	ND	ND	- - - -	- - - -
148-0004A1	NE	14	T4	1	90	Grain	- - - -	- - - -	ND	ND
148-0004A3	NE	14	T4	1	90	Grain	ND	0.0138	- - - -	- - - -
148-0005A1	NE	14	T4	2	90	Grain	- - - -	- - - -	[0.0050]	[0.0037]
148-0005A2	NE	14	T4	2	90	Grain	ND	[0.0070]	- - - -	- - - -
148-0006A1	NE	14	T4	3	90	Grain	- - - -	- - - -	ND	ND
148-0006A2	NE	14	T4	3	90	Grain	ND	[0.0037]	- - - -	- - - -
149-0001A1	NE	14	T1	1	N/A	Grain	- - - -	- - - -	ND	ND
149-0001A2	NE	14	T1	1	N/A	Grain	ND	ND	- - - -	- - - -

Table 7 (Cont.). Summarized Field Sample Information and Residue Results

Sample ID[a]	Site	Trial	Trt	Rep	Event	Commodity	2,4-D Reported, µg/g[b]	2,4-DCP Reported, µg/g[b]	Quizalofop Acid Reported, µg/g[b]	Quizalofop Ethyl Reported, µg/g[b]
149-0002A1	NE	14	T1	2	N/A	Grain	- - - -	- - - -	ND	ND
149-0002A2	NE	14	T1	2	N/A	Grain	ND	ND	- - - -	- - - -
149-0003A1	NE	14	T1	3	N/A	Grain	- - - -	- - - -	ND	ND
149-0003A2	NE	14	T1	3	N/A	Grain	ND	ND	- - - -	- - - -
149-0004A1	NE	14	T4	1	90	Grain	- - - -	- - - -	ND	ND
149-0004A3	NE	14	T4	1	90	Grain	ND	ND	- - - -	- - - -
149-0005A1	NE	14	T4	2	90	Grain	- - - -	- - - -	ND	ND
149-0005A2	NE	14	T4	2	90	Grain	ND	ND	- - - -	- - - -
149-0006A1	NE	14	T4	3	90	Grain	- - - -	- - - -	ND	ND
149-0006A2	NE	14	T4	3	90	Grain	ND	ND	- - - -	- - - -
001-0001A1	NJ	1	T1	1	N/A	Forage	- - - -	- - - -	ND	ND
001-0001A2	NJ	1	T1	1	N/A	Forage	ND	ND	- - - -	- - - -
001-0002A1	NJ	1	T2	1	40	Forage	- - - -	- - - -	ND	ND
001-0002A2	NJ	1	T2	1	40	Forage	2.2363	1.7781	- - - -	- - - -
001-0003A1	NJ	1	T2	2	40	Forage	- - - -	- - - -	ND	ND
001-0003A2	NJ	1	T2	2	40	Forage	2.9883	2.1899	- - - -	- - - -
002-0001A1	NJ	1	T1	1	N/A	Forage	- - - -	- - - -	ND	ND
002-0001A2	NJ	1	T1	1	N/A	Forage	ND	ND	- - - -	- - - -
002-0002A1	NJ	1	T2	1	60	Forage	- - - -	- - - -	ND	ND
002-0002A2	NJ	1	T2	1	60	Forage	3.3531	1.9824	- - - -	- - - -
002-0003A1	NJ	1	T2	2	60	Forage	- - - -	- - - -	ND	ND
002-0003A2	NJ	1	T2	2	60	Forage	2.0687	1.3658	- - - -	- - - -
005-0001A1	PA	2	T1	1	N/A	Forage	- - - -	- - - -	ND	ND
005-0001A2	PA	2	T1	1	N/A	Forage	ND	ND	- - - -	- - - -
005-0002A1	PA	2	T2	1	40	Forage	- - - -	- - - -	ND	ND
005-0002A2	PA	2	T2	1	40	Forage	2.2267	3.9970	- - - -	- - - -
005-0003A1	PA	2	T2	2	40	Forage	- - - -	- - - -	ND	ND
005-0003A2	PA	2	T2	2	40	Forage	1.5660	2.1151	- - - -	- - - -
006-0001A1	PA	2	T1	1	N/A	Forage	- - - -	- - - -	ND	ND
006-0001A2	PA	2	T1	1	N/A	Forage	ND	[0.0072]	- - - -	- - - -
006-0002A1	PA	2	T2	1	60	Forage	- - - -	- - - -	ND	ND
006-0002A2	PA	2	T2	1	60	Forage	1.7752	3.1016	- - - -	- - - -
006-0003A1	PA	2	T2	2	60	Forage	- - - -	- - - -	ND	ND
006-0003A2	PA	2	T2	2	60	Forage	1.8045	2.6889	- - - -	- - - -
009-0001A1	GA	3	T1	1	N/A	Forage	- - - -	- - - -	ND	ND
009-0001A2	GA	3	T1	1	N/A	Forage	ND	ND	- - - -	- - - -
009-0002A1	GA	3	T2	1	40	Forage	- - - -	- - - -	ND	ND
009-0002A2	GA	3	T2	1	40	Forage	0.6159	0.3946	- - - -	- - - -
009-0003A1	GA	3	T2	2	40	Forage	- - - -	- - - -	ND	ND
009-0003A2	GA	3	T2	2	40	Forage	0.6883	0.5511	- - - -	- - - -
010-0001A1	GA	3	T1	1	N/A	Forage	- - - -	- - - -	ND	ND
010-0001A2	GA	3	T1	1	N/A	Forage	ND	[0.0043]	- - - -	- - - -

Table 7 (Cont.). Summarized Field Sample Information and Residue Results

Sample ID[a]	Site	Trial	Trt	Rep	Event	Commodity	2,4-D Reported, µg/g[b]	2,4-DCP Reported, µg/g[b]	Quizalofop Acid Reported, µg/g[b]	Quizalofop Ethyl Reported, µg/g[b]
010-0002A1	GA	3	T2	1	60	Forage	- - - -	- - - -	ND	ND
010-0002A2	GA	3	T2	1	60	Forage	0.3362	0.1685	- - - -	- - - -
010-0003A1	GA	3	T2	2	60	Forage	- - - -	- - - -	ND	ND
010-0003A2	GA	3	T2	2	60	Forage	0.4805	0.1740	- - - -	- - - -
014-0001A1	GA	4	T1	1	N/A	Forage	- - - -	- - - -	ND	ND
014-0001A2	GA	4	T1	1	N/A	Forage	ND	ND	- - - -	- - - -
014-0002A1	GA	4	T2	1	40	Forage	- - - -	- - - -	ND	ND
014-0002A2	GA	4	T2	1	40	Forage	0.8884	0.6915	- - - -	- - - -
014-0003A1	GA	4	T2	2	40	Forage	- - - -	- - - -	ND	ND
014-0003A2	GA	4	T2	2	40	Forage	1.1682	0.9267	- - - -	- - - -
015-0001A1	GA	4	T1	1	N/A	Forage	- - - -	- - - -	ND	ND
015-0001A2	GA	4	T1	1	N/A	Forage	ND	ND	- - - -	- - - -
015-0002A1	GA	4	T2	1	46	Forage	- - - -	- - - -	ND	ND
015-0002A2	GA	4	T2	1	46	Forage	0.5212	0.4705	- - - -	- - - -
015-0003A1	GA	4	T2	2	46	Forage	- - - -	- - - -	ND	[0.0044]
015-0003A2	GA	4	T2	2	46	Forage	0.6547	0.5233	- - - -	- - - -
016-0001A1	GA	4	T1	1	N/A	Forage	- - - -	- - - -	ND	ND
016-0001A2	GA	4	T1	1	N/A	Forage	ND	ND	- - - -	- - - -
016-0002A1	GA	4	T2	1	53	Forage	- - - -	- - - -	ND	ND
016-0002A2	GA	4	T2	1	53	Forage	0.4470	0.3939	- - - -	- - - -
016-0003A1	GA	4	T2	2	53	Forage	- - - -	- - - -	ND	ND
016-0003A2	GA	4	T2	2	53	Forage	0.1243	0.0997	- - - -	- - - -
017-0001A1	GA	4	T1	1	N/A	Forage	- - - -	- - - -	ND	ND
017-0001A2	GA	4	T1	1	N/A	Forage	ND	ND	- - - -	- - - -
017-0002A1	GA	4	T2	1	60	Forage	- - - -	- - - -	ND	ND
017-0002A2	GA	4	T2	1	60	Forage	[0.0059]	[0.0053]	- - - -	- - - -
017-0003A1	GA	4	T2	2	60	Forage	- - - -	- - - -	ND	ND
017-0003A2	GA	4	T2	2	60	Forage	ND	[0.0042]	- - - -	- - - -
018-0001A1	GA	4	T1	1	N/A	Forage	- - - -	- - - -	ND	ND
018-0001A2	GA	4	T1	1	N/A	Forage	ND	ND	- - - -	- - - -
018-0002A1	GA	4	T2	1	67	Forage	- - - -	- - - -	ND	ND
018-0002A2	GA	4	T2	1	67	Forage	ND	ND	- - - -	- - - -
018-0003A1	GA	4	T2	2	67	Forage	- - - -	- - - -	ND	ND
018-0003A2	GA	4	T2	2	67	Forage	ND	[0.0040]	- - - -	- - - -
019-0001A1	GA	4	T1	1	N/A	Forage	- - - -	- - - -	ND	ND
019-0001A2	GA	4	T1	1	N/A	Forage	ND	ND	- - - -	- - - -
019-0002A1	GA	4	T2	1	74	Forage	- - - -	- - - -	ND	ND
019-0002A2	GA	4	T2	1	74	Forage	ND	[0.0046]	- - - -	- - - -
019-0003A1	GA	4	T2	2	74	Forage	- - - -	- - - -	ND	ND
019-0003A2	GA	4	T2	2	74	Forage	ND	[0.0058]	- - - -	- - - -
030-0001A1	IL	5	T1	1	N/A	Forage	- - - -	- - - -	ND	ND
030-0001A3	IL	5	T1	1	N/A	Forage	ND	ND	- - - -	- - - -

Table 7 (Cont.). Summarized Field Sample Information and Residue Results

Sample ID[a]	Site	Trial	Trt	Rep	Event	Commodity	2,4-D Reported, µg/g[b]	2,4-DCP Reported, µg/g[b]	Quizalofop Acid Reported, µg/g[b]	Quizalofop Ethyl Reported, µg/g[b]
030-0002A1	IL	5	T2	1	40	Forage	- - - -	- - - -	ND	ND
030-0002A3	IL	5	T2	1	40	Forage	0.4492	0.4226	- - - -	- - - -
030-0003A1	IL	5	T2	2	40	Forage	- - - -	- - - -	ND	ND
030-0003A3	IL	5	T2	2	40	Forage	0.3028	0.4778	- - - -	- - - -
031-0001A1	IL	5	T1	1	N/A	Forage	- - - -	- - - -	ND	ND
031-0001A2	IL	5	T1	1	N/A	Forage	ND	ND	- - - -	- - - -
031-0002A1	IL	5	T2	1	60	Forage	- - - -	- - - -	ND	ND
031-0002A2	IL	5	T2	1	60	Forage	0.5712	0.6140	- - - -	- - - -
031-0003A1	IL	5	T2	2	60	Forage	- - - -	- - - -	ND	ND
031-0003A2	IL	5	T2	2	60	Forage	0.9030	1.1988	- - - -	- - - -
034-0001A1	IL	6	T1	1	N/A	Forage	- - - -	- - - -	ND	ND
034-0001A2	IL	6	T1	1	N/A	Forage	ND	ND	- - - -	- - - -
034-0002A1	IL	6	T2	1	40	Forage	- - - -	- - - -	ND	ND
034-0002A2	IL	6	T2	1	40	Forage	0.4361	0.8889	- - - -	- - - -
034-0003A1	IL	6	T2	2	40	Forage	- - - -	- - - -	ND	ND
034-0003A2	IL	6	T2	2	40	Forage	0.4689	0.8479	- - - -	- - - -
035-0001A1	IL	6	T1	1	N/A	Forage	- - - -	- - - -	ND	ND
035-0001A2	IL	6	T1	1	N/A	Forage	ND	ND	- - - -	- - - -
035-0002A1	IL	6	T2	1	60	Forage	- - - -	- - - -	ND	ND
035-0002A2	IL	6	T2	1	60	Forage	0.2227	0.4369	- - - -	- - - -
035-0003A1	IL	6	T2	2	60	Forage	- - - -	- - - -	ND	ND
035-0003A2	IL	6	T2	2	60	Forage	0.2188	0.5279	- - - -	- - - -
038-0001A1	IN	7	T1	1	N/A	Forage	- - - -	- - - -	ND	ND
038-0001A2	IN	7	T1	1	N/A	Forage	ND	ND	- - - -	- - - -
038-0002A1	IN	7	T2	1	40	Forage	- - - -	- - - -	ND	ND
038-0002A2	IN	7	T2	1	40	Forage	1.0544	1.9137	- - - -	- - - -
038-0003A1	IN	7	T2	2	40	Forage	- - - -	- - - -	ND	ND
038-0003A2	IN	7	T2	2	40	Forage	1.2968	2.1148	- - - -	- - - -
039-0001A1	IN	7	T1	1	N/A	Forage	- - - -	- - - -	ND	ND
039-0001A2	IN	7	T1	1	N/A	Forage	ND	ND	- - - -	- - - -
039-0002A1	IN	7	T2	1	60	Forage	- - - -	- - - -	ND	ND
039-0002A2	IN	7	T2	1	60	Forage	1.5772	2.4305	- - - -	- - - -
039-0003A1	IN	7	T2	2	60	Forage	- - - -	- - - -	ND	ND
039-0003A2	IN	7	T2	2	60	Forage	0.5949	0.9874	- - - -	- - - -
042-0001A1	MO	8	T1	1	N/A	Forage	- - - -	- - - -	ND	ND
042-0001A2	MO	8	T1	1	N/A	Forage	ND	[0.0049]	- - - -	- - - -
042-0002A1	MO	8	T2	1	40	Forage	- - - -	- - - -	ND	ND
042-0002A2	MO	8	T2	1	40	Forage	[0.0099]	0.0115	- - - -	- - - -
042-0003A1	MO	8	T2	2	40	Forage	- - - -	- - - -	ND	ND
042-0003A2	MO	8	T2	2	40	Forage	ND	[0.0030]	- - - -	- - - -
043-0001A1	MO	8	T1	1	N/A	Forage	- - - -	- - - -	ND	ND
043-0001A2	MO	8	T1	1	N/A	Forage	ND	[0.0056]	- - - -	- - - -

Table 7 (Cont.). Summarized Field Sample Information and Residue Results

Sample ID[a]	Site	Trial	Trt	Rep	Event	Commodity	2,4-D Reported, µg/g[b]	2,4-DCP Reported, µg/g[b]	Quizalofop Acid Reported, µg/g[b]	Quizalofop Ethyl Reported, µg/g[b]
043-0002A1	MO	8	T2	1	60	Forage	- - - -	- - - -	ND	ND
043-0002A2	MO	8	T2	1	60	Forage	0.0644	0.0835	- - - -	- - - -
043-0003A1	MO	8	T2	2	60	Forage	- - - -	- - - -	ND	ND
043-0003A2	MO	8	T2	2	60	Forage	0.0295	0.0518	- - - -	- - - -
046-0001A1	MO	9	T1	1	N/A	Forage	- - - -	- - - -	ND	ND
046-0001A15	MO	9	T1	1	N/A	Forage	ND	ND	- - - -	- - - -
046-0001A2	MO	9	T1	1	N/A	Forage	ND	ND	- - - -	- - - -
046-0001A21	MO	9	T1	1	N/A	Forage	ND	ND	- - - -	- - - -
046-0001A3	MO	9	T1	1	N/A	Forage	ND	ND	- - - -	- - - -
046-0001A9	MO	9	T1	1	N/A	Forage	ND	ND	- - - -	- - - -
046-0002A1	MO	9	T2	1	40	Forage	- - - -	- - - -	ND	ND
046-0002A2	MO	9	T2	1	40	Forage	1.5970	1.6850	- - - -	- - - -
046-0003A1	MO	9	T2	2	40	Forage	- - - -	- - - -	ND	ND
046-0003A2	MO	9	T2	2	40	Forage	0.9675	1.4721	- - - -	- - - -
047-0001A1	MO	9	T1	1	N/A	Forage	- - - -	- - - -	ND	ND
047-0001A2	MO	9	T1	1	N/A	Forage	ND	[0.0035]	- - - -	- - - -
047-0002A1	MO	9	T2	1	60	Forage	- - - -	- - - -	ND	ND
047-0002A2	MO	9	T2	1	60	Forage	1.2850	1.5588	- - - -	- - - -
047-0003A1	MO	9	T2	2	60	Forage	- - - -	- - - -	ND	ND
047-0003A2	MO	9	T2	2	60	Forage	0.6283	1.0050	- - - -	- - - -
050-0001A1	IL	10	T1	1	N/A	Forage	- - - -	- - - -	ND	ND
050-0001A2	IL	10	T1	1	N/A	Forage	ND	[0.0094]	- - - -	- - - -
050-0002A1	IL	10	T2	1	40	Forage	- - - -	- - - -	ND	ND
050-0002A2	IL	10	T2	1	40	Forage	ND	[0.0046]	- - - -	- - - -
050-0003A1	IL	10	T2	2	40	Forage	- - - -	- - - -	ND	ND
050-0003A2	IL	10	T2	2	40	Forage	0.0843	0.1134	- - - -	- - - -
051-0001A1	IL	10	T1	1	N/A	Forage	- - - -	- - - -	ND	ND
051-0001A2	IL	10	T1	1	N/A	Forage	ND	[0.0052]	- - - -	- - - -
051-0002A1	IL	10	T2	1	60	Forage	- - - -	- - - -	ND	ND
051-0002A2	IL	10	T2	1	60	Forage	0.0183	0.0289	- - - -	- - - -
051-0003A1	IL	10	T2	2	60	Forage	- - - -	- - - -	ND	ND
051-0003A2	IL	10	T2	2	60	Forage	0.0235	0.0250	- - - -	- - - -
054-0001A1	MI	11	T1	1	N/A	Forage	- - - -	- - - -	ND	ND
054-0001A2	MI	11	T1	1	N/A	Forage	ND	[0.0084]	- - - -	- - - -
054-0002A1	MI	11	T2	1	40	Forage	- - - -	- - - -	ND	ND
054-0002A2	MI	11	T2	1	40	Forage	1.4753	3.0053	- - - -	- - - -
054-0003A1	MI	11	T2	2	40	Forage	- - - -	- - - -	ND	ND
054-0003A2	MI	11	T2	2	40	Forage	1.5986	2.6725	- - - -	- - - -
055-0001A1	MI	11	T1	1	N/A	Forage	- - - -	- - - -	ND	ND
055-0001A2	MI	11	T1	1	N/A	Forage	ND	[0.0035]	- - - -	- - - -
055-0002A1	MI	11	T2	1	46	Forage	- - - -	- - - -	ND	ND
055-0002A2	MI	11	T2	1	46	Forage	0.7119	1.4708	- - - -	- - - -

Table 7 (Cont.). Summarized Field Sample Information and Residue Results

Sample ID[a]	Site	Trial	Trt	Rep	Event	Commodity	2,4-D Reported, µg/g[b]	2,4-DCP Reported, µg/g[b]	Quizalofop Acid Reported, µg/g[b]	Quizalofop Ethyl Reported, µg/g[b]
055-0003A1	MI	11	T2	2	46	Forage	- - - -	- - - -	ND	ND
055-0003A2	MI	11	T2	2	46	Forage	0.9129	2.1439	- - - -	- - - -
056-0001A1	MI	11	T1	1	N/A	Forage	- - - -	- - - -	ND	ND
056-0001A3	MI	11	T1	1	N/A	Forage	ND	ND	- - - -	- - - -
056-0002A1	MI	11	T2	1	53	Forage	- - - -	- - - -	ND	ND
056-0002A3	MI	11	T2	1	53	Forage	1.4192	2.1370	- - - -	- - - -
056-0003A1	MI	11	T2	2	53	Forage	- - - -	- - - -	ND	ND
056-0003A3	MI	11	T2	2	53	Forage	2.5099	4.0840	- - - -	- - - -
057-0001A1	MI	11	T1	1	N/A	Forage	- - - -	- - - -	ND	ND
057-0001A2	MI	11	T1	1	N/A	Forage	ND	[0.0030]	- - - -	- - - -
057-0002A1	MI	11	T2	1	60	Forage	- - - -	- - - -	ND	ND
057-0002A2	MI	11	T2	1	60	Forage	3.1673	4.1470	- - - -	- - - -
057-0003A1	MI	11	T2	2	60	Forage	- - - -	- - - -	ND	ND
057-0003A2	MI	11	T2	2	60	Forage	2.0255	3.2044	- - - -	- - - -
058-0001A1	MI	11	T1	1	N/A	Forage	- - - -	- - - -	ND	ND
058-0001A2	MI	11	T1	1	N/A	Forage	ND	[0.0030]	- - - -	- - - -
058-0002A1	MI	11	T2	1	67	Forage	- - - -	- - - -	ND	ND
058-0002A2	MI	11	T2	1	67	Forage	1.9372	3.2731	- - - -	- - - -
058-0003A1	MI	11	T2	2	67	Forage	- - - -	- - - -	ND	ND
058-0003A2	MI	11	T2	2	67	Forage	2.1153	4.2914	- - - -	- - - -
059-0001A1	MI	11	T1	1	N/A	Forage	- - - -	- - - -	ND	ND
059-0001A2	MI	11	T1	1	N/A	Forage	ND	ND	- - - -	- - - -
059-0002A1	MI	11	T2	1	74	Forage	- - - -	- - - -	ND	ND
059-0002A2	MI	11	T2	1	74	Forage	0.8910	1.5874	- - - -	- - - -
059-0003A1	MI	11	T2	2	74	Forage	- - - -	- - - -	ND	ND
059-0003A2	MI	11	T2	2	74	Forage	0.8267	1.7421	- - - -	- - - -
070-0001A1	OH	12	T1	1	N/A	Forage	- - - -	- - - -	ND	ND
070-0001A2	OH	12	T1	1	N/A	Forage	ND	ND	- - - -	- - - -
070-0002A1	OH	12	T2	1	40	Forage	- - - -	- - - -	ND	ND
070-0002A2	OH	12	T2	1	40	Forage	0.8143	1.1786	- - - -	- - - -
070-0003A1	OH	12	T2	2	40	Forage	- - - -	- - - -	ND	ND
070-0003A2	OH	12	T2	2	40	Forage	0.8935	1.5655	- - - -	- - - -
071-0001A1	OH	12	T1	1	N/A	Forage	- - - -	- - - -	ND	ND
071-0001A2	OH	12	T1	1	N/A	Forage	ND	ND	- - - -	- - - -
071-0002A1	OH	12	T2	1	60	Forage	- - - -	- - - -	ND	ND
071-0002A2	OH	12	T2	1	60	Forage	0.3462	0.5641	- - - -	- - - -
071-0003A1	OH	12	T2	2	60	Forage	- - - -	- - - -	ND	ND
071-0003A2	OH	12	T2	2	60	Forage	0.2223	0.7512	- - - -	- - - -
074-0001A1	IN	13	T1	1	N/A	Forage	- - - -	- - - -	ND	ND
074-0001A2	IN	13	T1	1	N/A	Forage	ND	[0.0030]	- - - -	- - - -
074-0002A1	IN	13	T2	1	40	Forage	- - - -	- - - -	ND	ND
074-0002A2	IN	13	T2	1	40	Forage	1.8308	3.2328	- - - -	- - - -

Table 7 (Cont.). Summarized Field Sample Information and Residue Results

Sample ID[a]	Site	Trial	Trt	Rep	Event	Commodity	2,4-D Reported, µg/g[b]	2,4-DCP Reported, µg/g[b]	Quizalofop Acid Reported, µg/g[b]	Quizalofop Ethyl Reported, µg/g[b]
074-0003A1	IN	13	T2	2	40	Forage	- - - -	- - - -	ND	ND
074-0003A2	IN	13	T2	2	40	Forage	0.6086	2.0079	- - - -	- - - -
075-0001A1	IN	13	T1	1	N/A	Forage	- - - -	- - - -	ND	ND
075-0001A2	IN	13	T1	1	N/A	Forage	ND	ND	- - - -	- - - -
075-0002A1	IN	13	T2	1	60	Forage	- - - -	- - - -	ND	ND
075-0002A2	IN	13	T2	1	60	Forage	1.3388	2.7916	- - - -	- - - -
075-0003A1	IN	13	T2	2	60	Forage	- - - -	- - - -	ND	ND
075-0003A2	IN	13	T2	2	60	Forage	2.4445	5.0073	- - - -	- - - -
079-0001A1	NE	14	T1	1	N/A	Forage	- - - -	- - - -	ND	ND
079-0001A2	NE	14	T1	1	N/A	Forage	ND	ND	- - - -	- - - -
079-0002A1	NE	14	T2	1	60	Forage	- - - -	- - - -	ND	ND
079-0002A2	NE	14	T2	1	60	Forage	0.0405	0.1635	- - - -	- - - -
079-0003A1	NE	14	T2	2	60	Forage	- - - -	- - - -	ND	ND
079-0003A2	NE	14	T2	2	60	Forage	0.0818	0.2868	- - - -	- - - -
083-0001A1	NE	15	T1	1	N/A	Forage	- - - -	- - - -	ND	ND
083-0001A2	NE	15	T1	1	N/A	Forage	ND	ND	- - - -	- - - -
083-0002A1	NE	15	T2	1	40	Forage	- - - -	- - - -	ND	ND
083-0002A2	NE	15	T2	1	40	Forage	0.0220	0.0123	- - - -	- - - -
083-0003A1	NE	15	T2	2	40	Forage	- - - -	- - - -	ND	ND
083-0003A2	NE	15	T2	2	40	Forage	[0.0070]	0.0469	- - - -	- - - -
084-0001A1	NE	15	T1	1	N/A	Forage	- - - -	- - - -	ND	ND
084-0001A2	NE	15	T1	1	N/A	Forage	ND	ND	- - - -	- - - -
084-0002A1	NE	15	T2	1	60	Forage	- - - -	- - - -	ND	ND
084-0002A2	NE	15	T2	1	60	Forage	[0.0045]	0.0380	- - - -	- - - -
084-0003A1	NE	15	T2	2	60	Forage	- - - -	- - - -	ND	ND
084-0003A2	NE	15	T2	2	60	Forage	ND	0.0158	- - - -	- - - -
087-0001A1	KS	16	T1	1	N/A	Forage	- - - -	- - - -	ND	ND
087-0001A2	KS	16	T1	1	N/A	Forage	ND	[0.0072]	- - - -	- - - -
087-0002A1	KS	16	T2	1	40	Forage	- - - -	- - - -	ND	ND
087-0002A2	KS	16	T2	1	40	Forage	0.0347	0.0777	- - - -	- - - -
087-0003A1	KS	16	T2	2	40	Forage	- - - -	- - - -	ND	ND
087-0003A2	KS	16	T2	2	40	Forage	[0.0083]	0.0271	- - - -	- - - -
088-0001A1	KS	16	T1	1	N/A	Forage	- - - -	- - - -	ND	ND
088-0001A2	KS	16	T1	1	N/A	Forage	ND	[0.0035]	- - - -	- - - -
088-0002A1	KS	16	T2	1	60	Forage	- - - -	- - - -	ND	ND
088-0002A2	KS	16	T2	1	60	Forage	ND	0.0159	- - - -	- - - -
088-0003A1	KS	16	T2	2	60	Forage	- - - -	- - - -	ND	ND
088-0003A2	KS	16	T2	2	60	Forage	0.0127	0.0233	- - - -	- - - -
091-0001A1	IA	17	T1	1	N/A	Forage	- - - -	- - - -	ND	ND
091-0001A3	IA	17	T1	1	N/A	Forage	ND	ND	- - - -	- - - -
091-0002A1	IA	17	T2	1	40	Forage	- - - -	- - - -	ND	ND
091-0002A3	IA	17	T2	1	40	Forage	0.3515	0.8970	- - - -	- - - -

Table 7 (Cont.). Summarized Field Sample Information and Residue Results

Sample ID[a]	Site	Trial	Trt	Rep	Event	Commodity	2,4-D Reported, µg/g[b]	2,4-DCP Reported, µg/g[b]	Quizalofop Acid Reported, µg/g[b]	Quizalofop Ethyl Reported, µg/g[b]
091-0003A1	IA	17	T2	2	40	Forage	- - - -	- - - -	ND	ND
091-0003A3	IA	17	T2	2	40	Forage	0.4933	1.1569	- - - -	- - - -
092-0001A1	IA	17	T1	1	N/A	Forage	- - - -	- - - -	ND	ND
092-0001A2	IA	17	T1	1	N/A	Forage	ND	ND	- - - -	- - - -
092-0002A1	IA	17	T2	1	60	Forage	- - - -	- - - -	ND	ND
092-0002A2	IA	17	T2	1	60	Forage	0.1603	0.4587	- - - -	- - - -
092-0003A1	IA	17	T2	2	60	Forage	- - - -	- - - -	ND	ND
092-0003A2	IA	17	T2	2	60	Forage	0.2110	0.6650	- - - -	- - - -
095-0001A1	MO	18	T1	1	N/A	Forage	- - - -	- - - -	ND	ND
095-0001A3	MO	18	T1	1	N/A	Forage	ND	[0.0099]	- - - -	- - - -
095-0002A1	MO	18	T2	1	40	Forage	- - - -	- - - -	ND	ND
095-0002A3	MO	18	T2	1	40	Forage	[0.0049]	0.0135	- - - -	- - - -
095-0003A1	MO	18	T2	2	40	Forage	- - - -	- - - -	ND	ND
095-0003A3	MO	18	T2	2	40	Forage	ND	[0.0030]	- - - -	- - - -
096-0001A1	MO	18	T1	1	N/A	Forage	- - - -	- - - -	ND	ND
096-0001A2	MO	18	T1	1	N/A	Forage	ND	ND	- - - -	- - - -
096-0002A1	MO	18	T2	1	60	Forage	- - - -	- - - -	ND	ND
096-0002A2	MO	18	T2	1	60	Forage	0.0124	0.0398	- - - -	- - - -
096-0003A1	MO	18	T2	2	60	Forage	- - - -	- - - -	ND	ND
096-0003A2	MO	18	T2	2	60	Forage	[0.0060]	0.0162	- - - -	- - - -
099-0001A1	IA	19	T1	1	N/A	Forage	- - - -	- - - -	ND	ND
099-0001A2	IA	19	T1	1	N/A	Forage	ND	[0.0070]	- - - -	- - - -
099-0002A1	IA	19	T2	1	40	Forage	- - - -	- - - -	ND	ND
099-0002A2	IA	19	T2	1	40	Forage	0.2731	1.5473	- - - -	- - - -
099-0003A1	IA	19	T2	2	40	Forage	- - - -	- - - -	ND	ND
099-0003A2	IA	19	T2	2	40	Forage	0.5231	2.1519	- - - -	- - - -
100-0001A1	IA	19	T1	1	N/A	Forage	- - - -	- - - -	ND	ND
100-0001A2	IA	19	T1	1	N/A	Forage	ND	ND	- - - -	- - - -
100-0002A1	IA	19	T2	1	60	Forage	- - - -	- - - -	ND	ND
100-0002A2	IA	19	T2	1	60	Forage	0.3083	1.3010	- - - -	- - - -
100-0003A1	IA	19	T2	2	60	Forage	- - - -	- - - -	ND	ND
100-0003A2	IA	19	T2	2	60	Forage	0.1853	0.8464	- - - -	- - - -
103-0001A1	MN	20	T1	1	N/A	Forage	- - - -	- - - -	ND	ND
103-0001A2	MN	20	T1	1	N/A	Forage	ND	ND	- - - -	- - - -
103-0002A1	MN	20	T2	1	40	Forage	- - - -	- - - -	ND	ND
103-0002A2	MN	20	T2	1	40	Forage	0.4800	1.4962	- - - -	- - - -
103-0003A1	MN	20	T2	2	40	Forage	- - - -	- - - -	ND	ND
103-0003A2	MN	20	T2	2	40	Forage	0.7315	2.2111	- - - -	- - - -
104-0001A1	MN	20	T1	1	N/A	Forage	- - - -	- - - -	ND	ND
104-0001A3	MN	20	T1	1	N/A	Forage	ND	ND	- - - -	- - - -
104-0002A1	MN	20	T2	1	60	Forage	- - - -	- - - -	ND	ND
104-0002A2	MN	20	T2	1	60	Forage	0.2926	1.0194	- - - -	- - - -

Table 7 (Cont.). Summarized Field Sample Information and Residue Results

Sample ID[a]	Site	Trial	Trt	Rep	Event	Commodity	2,4-D Reported, µg/g[b]	2,4-DCP Reported, µg/g[b]	Quizalofop Acid Reported, µg/g[b]	Quizalofop Ethyl Reported, µg/g[b]
104-0003A1	MN	20	T2	2	60	Forage	- - - -	- - - -	ND	ND
104-0003A2	MN	20	T2	2	60	Forage	0.2931	0.8123	- - - -	- - - -
107-0001A1	MO	21	T1	1	N/A	Forage	- - - -	- - - -	ND	ND
107-0001A3	MO	21	T1	1	N/A	Forage	ND	ND	- - - -	- - - -
107-0002A1	MO	21	T2	1	40	Forage	- - - -	- - - -	ND	ND
107-0002A2	MO	21	T2	1	40	Forage	0.3311	0.3232	- - - -	- - - -
107-0003A1	MO	21	T2	2	40	Forage	- - - -	- - - -	ND	ND
107-0003A2	MO	21	T2	2	40	Forage	0.3770	0.3451	- - - -	- - - -
108-0001A1	MO	21	T1	1	N/A	Forage	- - - -	- - - -	ND	ND
108-0001A2	MO	21	T1	1	N/A	Forage	ND	ND	- - - -	- - - -
108-0002A1	MO	21	T2	1	60	Forage	- - - -	- - - -	ND	ND
108-0002A2	MO	21	T2	1	60	Forage	0.6157	0.3694	- - - -	- - - -
108-0003A1	MO	21	T2	2	60	Forage	- - - -	- - - -	ND	ND
108-0003A2	MO	21	T2	2	60	Forage	0.2811	0.1846	- - - -	- - - -
111-0001A1	OK	22	T1	1	N/A	Forage	- - - -	- - - -	ND	ND
111-0001A2	OK	22	T1	1	N/A	Forage	ND	[0.0032]	- - - -	- - - -
111-0002A1	OK	22	T2	1	40	Forage	- - - -	- - - -	ND	ND
111-0002A2	OK	22	T2	1	40	Forage	0.0206	0.0316	- - - -	- - - -
111-0003A1	OK	22	T2	2	40	Forage	- - - -	- - - -	ND	ND
111-0003A2	OK	22	T2	2	40	Forage	0.2533	0.1301	- - - -	- - - -
112-0001A1	OK	22	T1	1	N/A	Forage	- - - -	- - - -	ND	[0.0031]
112-0001A2	OK	22	T1	1	N/A	Forage	ND	ND	- - - -	- - - -
112-0002A1	OK	22	T2	1	46	Forage	- - - -	- - - -	[0.0030]	ND
112-0002A2	OK	22	T2	1	46	Forage	0.2515	0.0940	- - - -	- - - -
112-0003A1	OK	22	T2	2	46	Forage	- - - -	- - - -	ND	ND
112-0003A2	OK	22	T2	2	46	Forage	0.2734	0.0727	- - - -	- - - -
113-0001A1	OK	22	T1	1	N/A	Forage	- - - -	- - - -	ND	ND
113-0001A2	OK	22	T1	1	N/A	Forage	ND	[0.0047]	- - - -	- - - -
113-0002A1	OK	22	T2	1	53	Forage	- - - -	- - - -	ND	ND
113-0002A2	OK	22	T2	1	53	Forage	0.1873	0.0788	- - - -	- - - -
113-0003A1	OK	22	T2	2	53	Forage	- - - -	- - - -	[0.0037]	ND
113-0003A2	OK	22	T2	2	53	Forage	1.0952	0.2067	- - - -	- - - -
114-0001A1	OK	22	T1	1	N/A	Forage	- - - -	- - - -	ND	ND
114-0001A2	OK	22	T1	1	N/A	Forage	ND	ND	- - - -	- - - -
114-0002A1	OK	22	T2	1	60	Forage	- - - -	- - - -	ND	ND
114-0002A2	OK	22	T2	1	60	Forage	0.2945	0.0584	- - - -	- - - -
114-0003A1	OK	22	T2	2	60	Forage	- - - -	- - - -	ND	ND
114-0003A2	OK	22	T2	2	60	Forage	0.2129	0.0561	- - - -	- - - -
115-0001A1	OK	22	T1	1	N/A	Forage	- - - -	- - - -	ND	ND
115-0001A2	OK	22	T1	1	N/A	Forage	ND	[0.0044]	- - - -	- - - -
115-0002A1	OK	22	T2	1	67	Forage	- - - -	- - - -	ND	ND
115-0002A2	OK	22	T2	1	67	Forage	0.1915	0.0323	- - - -	- - - -

Table 7 (Cont.). Summarized Field Sample Information and Residue Results

Sample ID[a]	Site	Trial	Trt	Rep	Event	Commodity	2,4-D Reported, µg/g[b]	2,4-DCP Reported, µg/g[b]	Quizalofop Acid Reported, µg/g[b]	Quizalofop Ethyl Reported, µg/g[b]
115-0003A1	OK	22	T2	2	67	Forage	- - - -	- - - -	ND	ND
115-0003A2	OK	22	T2	2	67	Forage	0.1005	0.0361	- - - -	- - - -
115-0003A3	OK	22	T2	2	67	Forage	0.1119	0.0388	- - - -	- - - -
115-0003A4	OK	22	T2	2	67	Forage	0.1001	0.0352	- - - -	- - - -
115-0003A5	OK	22	T2	2	67	Forage	0.1072	0.0352	- - - -	- - - -
115-0003A6	OK	22	T2	2	67	Forage	0.1161	0.0402	- - - -	- - - -
115-0003A7	OK	22	T2	2	67	Forage	0.1047	0.0371	- - - -	- - - -
116-0001A1	OK	22	T1	1	N/A	Forage	- - - -	- - - -	ND	ND
116-0001A2	OK	22	T1	1	N/A	Forage	ND	[0.0050]	- - - -	- - - -
116-0002A1	OK	22	T2	1	74	Forage	- - - -	- - - -	ND	ND
116-0002A2	OK	22	T2	1	74	Forage	0.0585	0.0360	- - - -	- - - -
116-0003A1	OK	22	T2	2	74	Forage	- - - -	- - - -	ND	ND
116-0003A2	OK	22	T2	2	74	Forage	0.0840	0.0297	- - - -	- - - -
127-0001A1	TX	23	T1	1	N/A	Forage	- - - -	- - - -	ND	ND
127-0001A2	TX	23	T1	1	N/A	Forage	ND	ND	- - - -	- - - -
127-0002A1	TX	23	T2	1	40	Forage	- - - -	- - - -	ND	ND
127-0002A2	TX	23	T2	1	40	Forage	ND	ND	- - - -	- - - -
127-0003A1	TX	23	T2	2	40	Forage	- - - -	- - - -	ND	ND
127-0003A2	TX	23	T2	2	40	Forage	ND	ND	- - - -	- - - -
128-0001A1	TX	23	T1	1	N/A	Forage	- - - -	- - - -	ND	ND
128-0001A2	TX	23	T1	1	N/A	Forage	ND	ND	- - - -	- - - -
128-0002A1	TX	23	T2	1	60	Forage	- - - -	- - - -	0.1668	0.1968
128-0002A2	TX	23	T2	1	60	Forage	4.3733	1.1629	- - - -	- - - -
128-0003A1	TX	23	T2	2	60	Forage	- - - -	- - - -	0.1126	0.0989
128-0003A2	TX	23	T2	2	60	Forage	1.8756	0.7773	- - - -	- - - -
131-0001A1	ON	24	T1	1	N/A	Forage	- - - -	- - - -	ND	ND
131-0001A2	ON	24	T1	1	N/A	Forage	ND	[0.0031]	- - - -	- - - -
131-0002A1	ON	24	T2	1	40	Forage	- - - -	- - - -	ND	ND
131-0002A2	ON	24	T2	1	40	Forage	0.2017	0.6971	- - - -	- - - -
131-0003A1	ON	24	T2	2	40	Forage	- - - -	- - - -	ND	ND
131-0003A2	ON	24	T2	2	40	Forage	0.2297	0.9205	- - - -	- - - -
132-0001A1	ON	24	T1	1	N/A	Forage	- - - -	- - - -	ND	ND
132-0001A3	ON	24	T1	1	N/A	Forage	ND	ND	- - - -	- - - -
132-0002A1	ON	24	T2	1	60	Forage	- - - -	- - - -	ND	ND
132-0002A2	ON	24	T2	1	60	Forage	0.0129	0.0404	- - - -	- - - -
132-0003A1	ON	24	T2	2	60	Forage	- - - -	- - - -	ND	ND
132-0003A2	ON	24	T2	2	60	Forage	0.0273	0.0886	- - - -	- - - -
135-0001A1	ON	25	T1	1	N/A	Forage	- - - -	- - - -	ND	ND
135-0001A3	ON	25	T1	1	N/A	Forage	ND	ND	- - - -	- - - -
135-0002A1	ON	25	T2	1	40	Forage	- - - -	- - - -	ND	ND
135-0002A2	ON	25	T2	1	40	Forage	0.0734	0.6035	- - - -	- - - -
135-0003A1	ON	25	T2	2	40	Forage	- - - -	- - - -	ND	ND

Table 7 (Cont.). Summarized Field Sample Information and Residue Results

Sample ID[a]	Site	Trial	Trt	Rep	Event	Commodity	2,4-D Reported, µg/g[b]	2,4-DCP Reported, µg/g[b]	Quizalofop Acid Reported, µg/g[b]	Quizalofop Ethyl Reported, µg/g[b]
135-0003A2	ON	25	T2	2	40	Forage	0.0661	0.5683	- - - -	- - - -
136-0001A1	ON	25	T1	1	N/A	Forage	- - - -	- - - -	ND	ND
136-0001A3	ON	25	T1	1	N/A	Forage	ND	ND	- - - -	- - - -
136-0002A1	ON	25	T2	1	60	Forage	- - - -	- - - -	ND	ND
136-0002A2	ON	25	T2	1	60	Forage	0.0304	0.2001	- - - -	- - - -
136-0003A1	ON	25	T2	2	60	Forage	- - - -	- - - -	ND	ND
136-0003A2	ON	25	T2	2	60	Forage	0.0299	0.2745	- - - -	- - - -
157-0001A1	GA	4	T1	1	N/A	Forage	- - - -	- - - -	ND	ND
157-0001A13	GA	4	T1	1	N/A	Forage	- - - -	- - - -	ND	ND
157-0001A19	GA	4	T1	1	N/A	Forage	- - - -	- - - -	ND	ND
157-0001A25	GA	4	T1	1	N/A	Forage	- - - -	- - - -	ND	ND
157-0001A7	GA	4	T1	1	N/A	Forage	- - - -	- - - -	ND	ND
146-0001A1	GA	3	T1	1	N/A	Flour	- - - -	- - - -	ND	ND
146-0001A3	GA	3	T1	1	N/A	Flour	ND	ND	- - - -	- - - -
146-0002A1	GA	3	T1	2	N/A	Flour	- - - -	- - - -	ND	ND
146-0002A3	GA	3	T1	2	N/A	Flour	ND	ND	- - - -	- - - -
146-0003A1	GA	3	T4	1	90	Flour	- - - -	- - - -	ND	ND
146-0003A3	GA	3	T4	1	90	Flour	ND	ND	- - - -	- - - -
146-0004A1	GA	3	T4	2	90	Flour	- - - -	- - - -	ND	ND
146-0004A3	GA	3	T4	2	90	Flour	ND	ND	- - - -	- - - -
155-0001A1	NE	14	T1	1	N/A	Flour	- - - -	- - - -	ND	ND
155-0001A22	NE	14	T1	1	N/A	Flour	ND	ND	- - - -	- - - -
155-0001A9	NE	14	T1	1	N/A	Flour	- - - -	- - - -	ND	ND
155-0002A1	NE	14	T1	2	N/A	Flour	- - - -	- - - -	ND	ND
155-0002A2	NE	14	T1	2	N/A	Flour	ND	ND	- - - -	- - - -
155-0003A1	NE	14	T4	1	90	Flour	- - - -	- - - -	ND	ND
155-0003A2	NE	14	T4	1	90	Flour	ND	ND	- - - -	- - - -
155-0004A1	NE	14	T4	2	90	Flour	- - - -	- - - -	ND	ND
155-0004A2	NE	14	T4	2	90	Flour	ND	ND	- - - -	- - - -
141-0001A1	GA	3	T1	1	N/A	Aspirated grain fractions	- - - -	- - - -	ND	ND
141-0001A2	GA	3	T1	1	N/A	Aspirated grain fractions	ND	ND	- - - -	- - - -
141-0002A1	GA	3	T4	1	90	Aspirated grain fractions	- - - -	- - - -	ND	ND
141-0002A2	GA	3	T4	1	90	Aspirated grain fractions	0.0211	[0.0060]	- - - -	- - - -
150-0001A1	NE	14	T1	1	N/A	Aspirated grain fractions	- - - -	- - - -	ND	ND
150-0001A22	NE	14	T1	1	N/A	Aspirated grain fractions	ND	ND	- - - -	- - - -
150-0001A9	NE	14	T1	1	N/A	Aspirated grain fractions	- - - -	- - - -	ND	ND
150-0002A1	NE	14	T4	1	90	Aspirated grain fractions	- - - -	- - - -	ND	ND
150-0002A2	NE	14	T4	1	90	Aspirated grain fractions	ND	ND	- - - -	- - - -

[a] Full Sample ID number is preceded by ARA-09-15-10-.
[b] All calculations were done using Microsoft Excel 2003 with full precision.
[c] Results not reported for this analyte, it was not monitored in this particular analysis .
[d] [ ] indicates that the concentration is less than the limit of quantitation of 0.01 µg/g. Values are reported with a lower degree of confidence than values above the limit of quantitation.
[e] ND = not detected. The residue was below the 0.003 µg/g limit of detection.

REPORT

EPL-BAS STUDY NUMBER: 205G416
SPONSOR STUDY NUMBER: FAPC07-152704

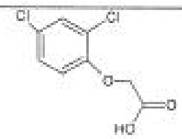
CERTIFICATE OF ANALYSIS FOR TEST/REFERENCE/CONTROL SUBSTANCES

TITLE/OBJECTIVE: Determination of the purity and/or identity of the following test/reference/control substance for use in a study.

TEST/REFERENCE/CONTROL SUBSTANCE:
TEST SUBSTANCE NUMBER: AGR275828
LOT NUMBER: MORRIS/1710
DESCRIPTION: 2,4-DICHLOROPHENOXYACETIC ACID ANALYTICAL STANDARD

REFERENCE SUBSTANCE USED: NONE PURITY: NA

INITIATION DATE: 3-JANUARY-2008

	MOLECULAR FORMULA: $C_8H_6Cl_2O_3$
	MOLECULAR WEIGHT: 221.04

CHEMICAL NAME: 2,4-DICHLOROPHENOXYACETIC ACID

METHODS USED: PURITY: HPLC/UV IDENTIFICATION: ESI-/MS

RESULTS and CONCLUSIONS:

x **IDENTITY:**
Spectroscopic identification is consistent with the proposed structure

x **RECERTIFICATION: UNCHANGED**
Current value of 99.5% is within experimental variation of previously established purity of 99.5%. The purity is unchanged and remains 99.5%.

NA **OTHER:** NA

RECERTIFICATION DATE: 15-JANUARY-2012

CALCULATIONS:
Internal Standard: NA External Standard: NA Area Normalized: x
Other: NA

STUDY DIRECTOR SIGNATURE: L S
Li Sheng

STUDY COMPLETION DATE: 1/21/08

PEER REVIEWER SIGNATURE: Michelle Smith

DATE: 1/21/08

TESTING FACILITY ADDRESS:
EPL Bio-Analytical Services (EPL-BAS)
9095 West Harristown Blvd.
Niantic, IL 62551

SPONSOR ADDRESS:
Dow AgroSciences LLC
9330 Zionsville Road
Indianapolis, Indiana 46268

All raw data and retainer samples associated with this study will be archived in the Sponsor archive. Only descriptive statistics were used unless otherwise noted in the results. This study was conducted in accordance with the Good Laboratory Practice Standard, 40 CFR Part 160.135 (b).

Figure 1. Certificate of Analysis for 2,4-D Reference Standard AGR275828

REPORT

EPL-BAS STUDY NUMBER: 205G354
SPONSOR STUDY NUMBER: FAPC073503

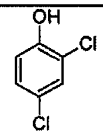
CERTIFICATE OF ANALYSIS FOR TEST/REFERENCE/CONTROL SUBSTANCES

TITLE/OBJECTIVE: Determination of the purity and/or identity of the following test/reference/control substance for use in a study.

TEST/REFERENCE/CONTROL SUBSTANCE:
TEST SUBSTANCE NUMBER: AGR182992
LOT NUMBER: QCR 696-132-1
DESCRIPTION: 2,4-DICHLOROPHENOL
REFERENCE COMPOUND – QUANTITATIVE

REFERENCE SUBSTANCE USED: NONE PURITY: NA

INITIATION DATE: 28-AUGUST-2007

	MOLECULAR FORMULA: $C_6H_4Cl_2O$
	MOLECULAR WEIGHT: 163.0

CHEMICAL NAME: 2,4-DICHLOROPHENOL

METHODS USED: PURITY: GC/FID IDENTIFICATION: GC/MS

RESULTS and CONCLUSIONS:

x **IDENTITY:**
Spectroscopic identification is consistent with the proposed structure

x **RECERTIFICATION: UNCHANGED**
Current value of 100% is within experimental variation of previously established purity of >99%. The purity is unchanged and remains >99%.

NA **OTHER:** NA

RECERTIFICATION DATE: 27-SEPTEMBER-2011

CALCULATIONS:
Internal Standard: NA External Standard: NA Area Normalized: x
Other: NA

STUDY DIRECTOR SIGNATURE: Sara Bendler
Sara Bendler

STUDY COMPLETION DATE: 10/2/07

PEER REVIEWER SIGNATURE: Michelle Smith

DATE: 10/2/07

TESTING FACILITY ADDRESS:
EPL Bio-Analytical Services (EPL-BAS)
395 N. Memorial Parkway
Harristown, Illinois 62537

SPONSOR ADDRESS:
Dow AgroSciences LLC
9330 Zionsville Road
Indianapolis, Indiana 46268

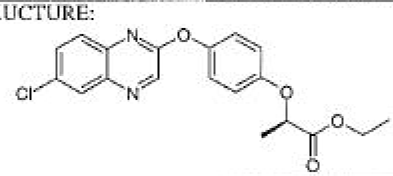
All raw data and retainer samples associated with this study will be archived in the Sponsor archive. Only descriptive statistics were used unless otherwise noted in the results. This study was conducted in accordance with the Good Laboratory Practice Standard, 40 CFR Part 160.135 (b).

Figure 2. Certificate of Analysis for 2,4-DCP Reference Standard AGR182992

REPORT **FAPC NUMBER** 09-225778

CERTIFICATE OF ANALYSIS FOR TEST/REFERENCE/CONTROL SUBSTANCES

TITLE OBJECTIVE: Determination of purity and/or identity of the following test/reference/control substance for use in a study.

	TEST/REFERENCE/CONTROL SUBSTANCE:		
TEST SUBSTANCE NO:	TSN106317		
LOT NO:	36891-47		
DESCRIPTION:	R (+) Quizalofop-ethyl		
	Reference Compound-Quantitative		
STRUCTURE: 		MOLECULAR FORMULA: $C_{19}H_{17}ClN_2O_4$ MOLECULAR WEIGHT: 372.81	
REFERENCE SUBSTANCE USED:	N/A	PURITY:	N/A
CHEMICAL NAME: (R)-2-[4-(6-chloro-quinoxalin-2-yloxy)-phenoxy]-propionic acid ethyl ester			

INITIATION DATE: June 9, 2009

METHODS USED:

PURITY:	IDENTIFICATION:
LC	IR

RESULTS and CONCLUSIONS:

X **RECERTIFICATION: UNCHANGED**
Current value of 92% R (+) Quizalofop-ethyl is within experimental variation of previously determined value of 92%. Purity is unchanged and remains 92% R (+) Quizalofop-ethyl.

X **IDENTITY:**
Spectroscopic analysis was consistent with the proposed structure.

X **OTHER:**
Optical purity = 93%, Chemical purity = 99%

RE-CERTIFICATION DATE: June 18, 2011

CALCULATIONS:

Area Normalized: X Internal Standard: N/A External Standard: N/A
Other (explain): R (+) Quizalofop-ethyl purity = Optical purity x Chemical purity

STUDY DIRECTOR SIGNATURE: Jennifer H. Jones
Jennifer H. Jones

STUDY COMPLETION DATE: August 28, 2009

PEER REVIEWER SIGNATURE: Rose M Nelson

DATE: August 25, 2009

TESTING FACILITY ADDRESS:
Dow AgroSciences LLC
Crop Protection R&D Analytical/Product Chemistry Center of Expertise
9330 Zionsville Road
Indianapolis, Indiana 46268

All raw data and retainer samples associated with this study will be archived in the testing facility archive. Only descriptive statistics were used unless otherwise noted in the results. This study was conducted in accordance with the Good Laboratory Practice Standard, 40 CFR Part 160.135 (b).

Figure 3. Certificate of Analysis for Quizalofop-Ethyl Reference Standard TSN106317

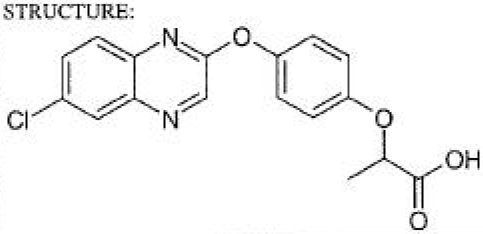
REPORT **FAPC NUMBER** 09-227492

CERTIFICATE OF ANALYSIS FOR TEST/REFERENCE/CONTROL SUBSTANCES

TITLE OBJECTIVE: Determination of purity and/or identity of the following test/reference/control substance for use in a study.

	TEST/REFERENCE/CONTROL SUBSTANCE:
TEST SUBSTANCE NO:	TSN106172
LOT NO:	36602-91
DESCRIPTION:	(R)-2-[4-(6-chloroquinoxalin-2-yloxy)phenoxy]propionic acid
	Reference Compound- Quantitative

STRUCTURE:



MOLECULAR FORMULA: $C_{17}H_{13}ClN_2O_4$

MOLECULAR WEIGHT: 344.76

REFERENCE SUBSTANCE USED:	N/A	PURITY:	N/A

INITIATION DATE: July 13, 2009

METHODS USED:

PURITY:	IDENTIFICATION:
LC	IR

RESULTS and CONCLUSIONS:

X **RECERTIFICATION: UNCHANGED**
Current value of 99% is within experimental variation of previously determined value of 96%. Purity is unchanged and remains 96%.

X **IDENTITY:**
Spectroscopic analysis was consistent with the proposed structure.

N/A **OTHER:**
N/A

RE-CERTIFICATION DATE: August 13, 2011

CALCULATIONS:

Area Normalized: X Internal Standard: N/A External Standard: N/A
Other (explain): N/A

STUDY DIRECTOR SIGNATURE: Jennifer H. Jones
Jennifer H. Jones

STUDY COMPLETION DATE: November 17, 2009

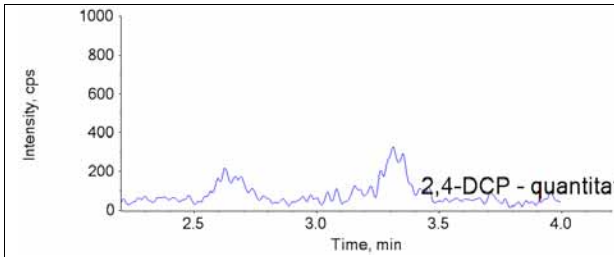
PEER REVIEWER SIGNATURE: Ashleigh Frank

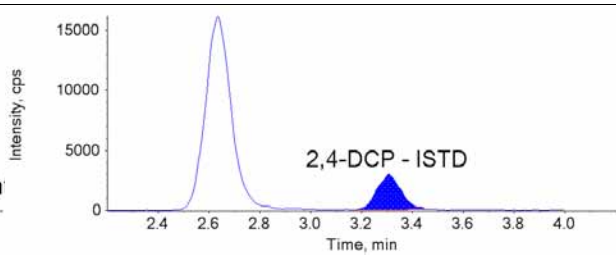
DATE: November 13, 2009

TESTING FACILITY ADDRESS:
Dow AgroSciences LLC
Crop Protection R&D Analytical/Product Chemistry Center of Expertise
9330 Zionsville Road
Indianapolis, Indiana 46268

All raw data and retainer samples associated with this study will be archived in the testing facility archive. Only descriptive statistics were used unless otherwise noted in the results. This study was conducted in accordance with the Good Laboratory Practice Standard, 40 CFR Part 160.135 (b).

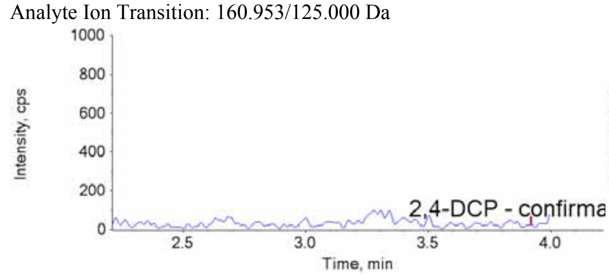
Figure 4. Certificate of Analysis for Quizalofop Acid Reference Standard TSN106172

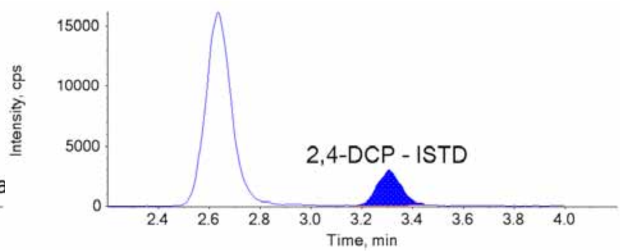




Sample Name: Reagent Blank
Data File: ARA-09-15-10 D16.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 0
Analyte Ion Transition: 160.953/125.000 Da

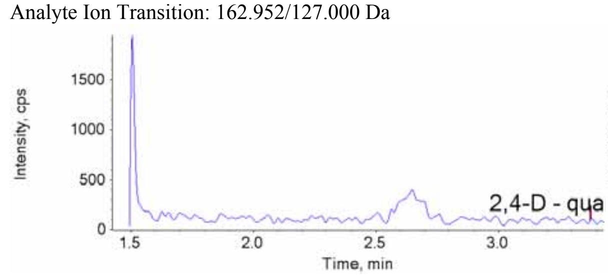
Acquired: 8/10/2010 6:57:42 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 19166
IS Ion Transition: 167.000/131.000 Da

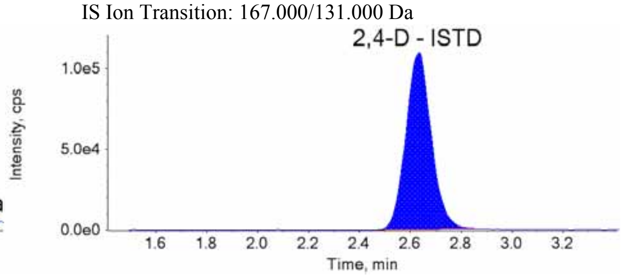




Sample Name: Reagent Blank
Data File: ARA-09-15-10 D16.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 0
Analyte Ion Transition: 162.952/127.000 Da

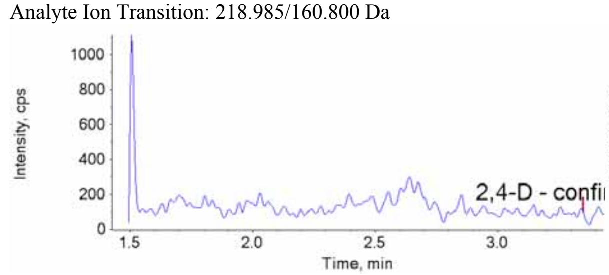
Acquired: 8/10/2010 6:57:42 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 19166
IS Ion Transition: 167.000/131.000 Da

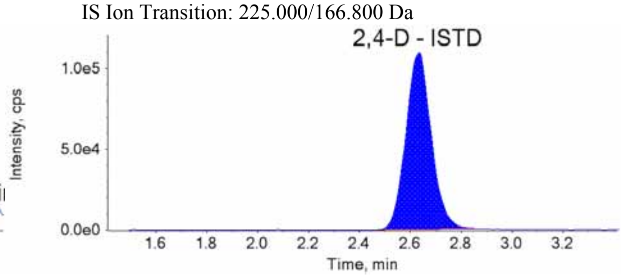




Sample Name: Reagent Blank
Data File: ARA-09-15-10 D16.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 0
Analyte Ion Transition: 218.985/160.800 Da

Acquired: 8/10/2010 6:57:42 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 759783
IS Ion Transition: 225.000/166.800 Da

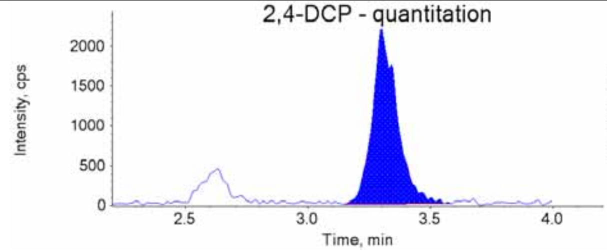


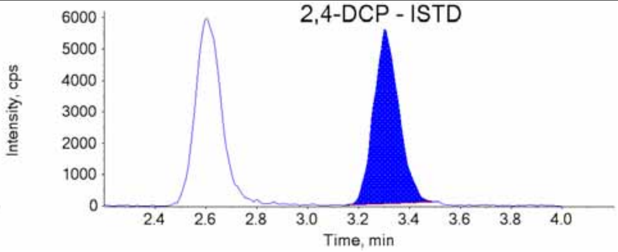


Sample Name: Reagent Blank
Data File: ARA-09-15-10 D16.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 0
Analyte Ion Transition: 220.968/163.000 Da

Acquired: 8/10/2010 6:57:42 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 759783
IS Ion Transition: 225.000/166.800 Da

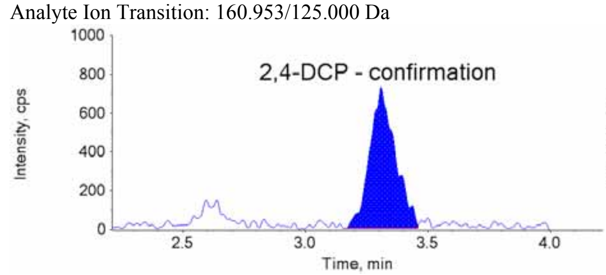
Figure 5. Typical Chromatogram of 2,4-D Analysis, Reagent Blank

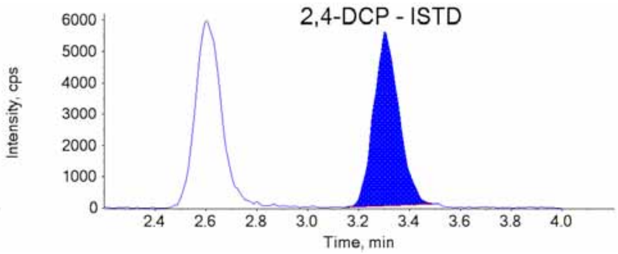




Sample Name: 0.5 std
Data File: ARA-09-15-10 D16.wiff
Analyte Integration Type: Base To Base
Analyte Peak Area: 15608
Analyte Ion Transition: 160.953/125.000 Da

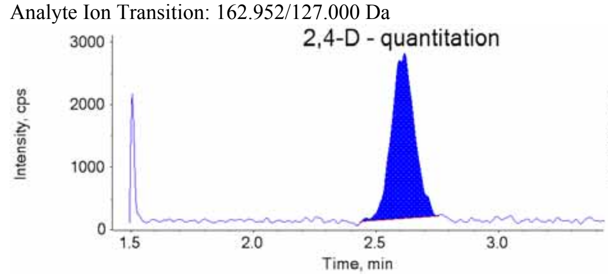
Acquired: 8/10/2010 7:49:21 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 38207
IS Ion Transition: 167.000/131.000 Da

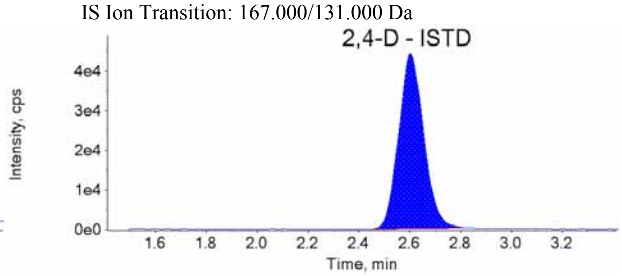




Sample Name: 0.5 std
Data File: ARA-09-15-10 D16.wiff
Analyte Integration Type: Exponential Skim
Analyte Peak Area: 5309
Analyte Ion Transition: 162.952/127.000 Da

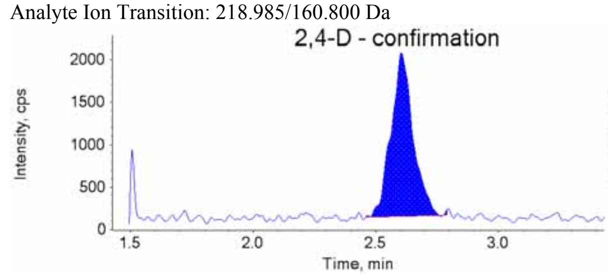
Acquired: 8/10/2010 7:49:21 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 38207
IS Ion Transition: 167.000/131.000 Da

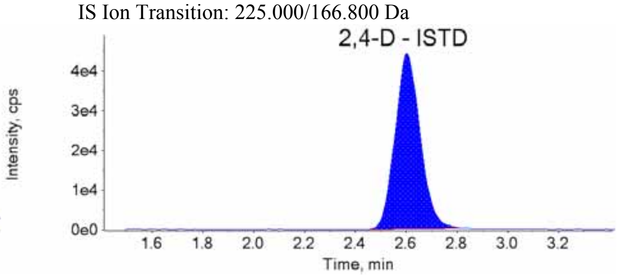




Sample Name: 0.5 std
Data File: ARA-09-15-10 D16.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 17499
Analyte Ion Transition: 218.985/160.800 Da

Acquired: 8/10/2010 7:49:21 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 301735
IS Ion Transition: 225.000/166.800 Da

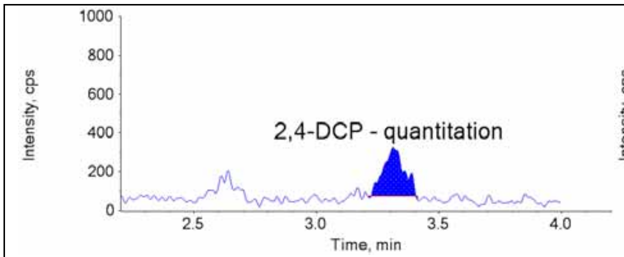


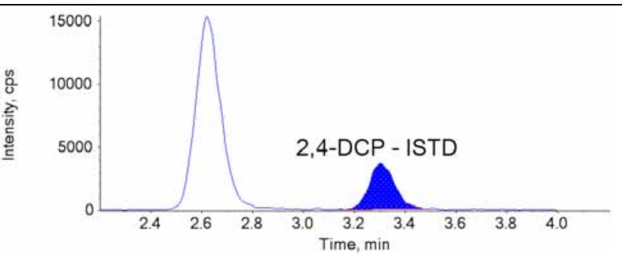


Sample Name: 0.5 std
Data File: ARA-09-15-10 D16.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 12029
Analyte Ion Transition: 220.968/163.000 Da

Acquired: 8/10/2010 7:49:21 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 301735
IS Ion Transition: 225.000/166.800 Da

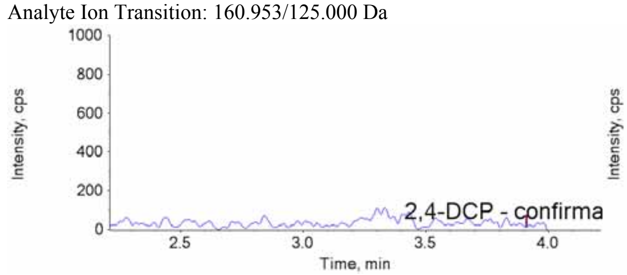
Figure 6. Typical Chromatogram of 2,4-D Analysis, 0.5 ng/mL Calibration Standard

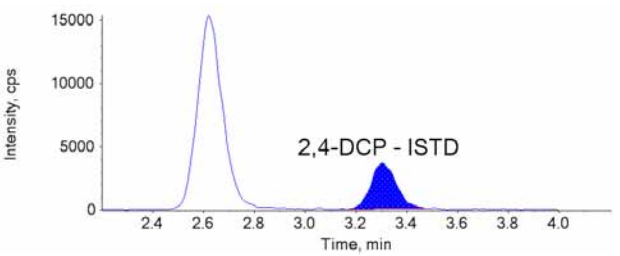




Sample Name: ARA-09-15-10-020-0001A44
Data File: ARA-09-15-10 D16.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 1470
Analyte Ion Transition: 160.953/125.000 Da

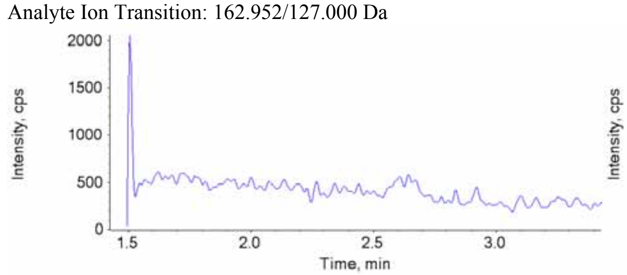
Acquired: 8/10/2010 7:23:31 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 25355
IS Ion Transition: 167.000/131.000 Da

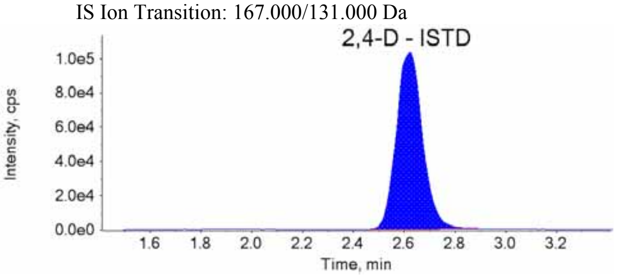




Sample Name: ARA-09-15-10-020-0001A44
Data File: ARA-09-15-10 D16.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 0
Analyte Ion Transition: 162.952/127.000 Da

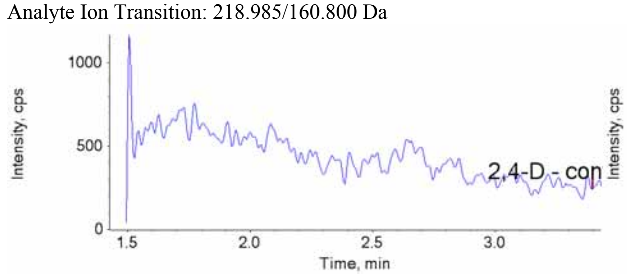
Acquired: 8/10/2010 7:23:31 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 25355
IS Ion Transition: 167.000/131.000 Da

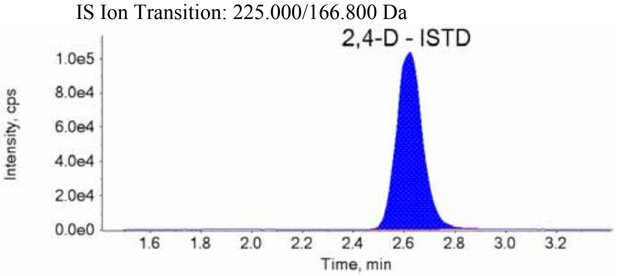




Sample Name: ARA-09-15-10-020-0001A44
Data File: ARA-09-15-10 D16.wiff
Analyte Integration Type: No Peak
Analyte Peak Area: 0
Analyte Ion Transition: 218.985/160.800 Da

Acquired: 8/10/2010 7:23:31 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 741989
IS Ion Transition: 225.000/166.800 Da

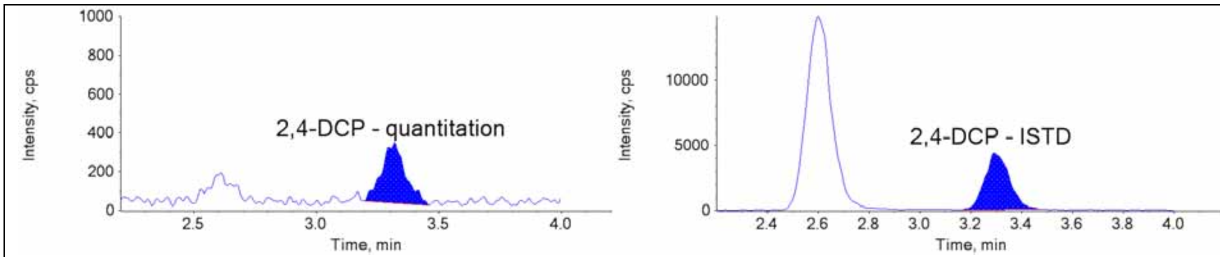




Sample Name: ARA-09-15-10-020-0001A44
Data File: ARA-09-15-10 D16.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 0
Analyte Ion Transition: 220.968/163.000 Da

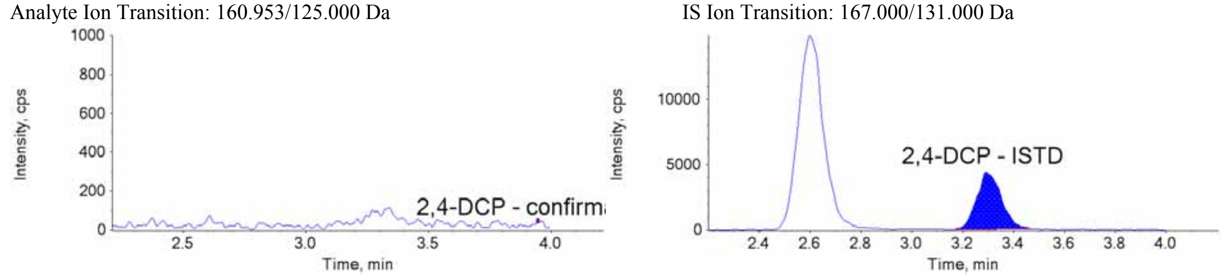
Acquired: 8/10/2010 7:23:31 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 741989
IS Ion Transition: 225.000/166.800 Da

Figure 7. Typical Chromatogram of 2,4-D Analysis, Control Corn Grain (020-0001)



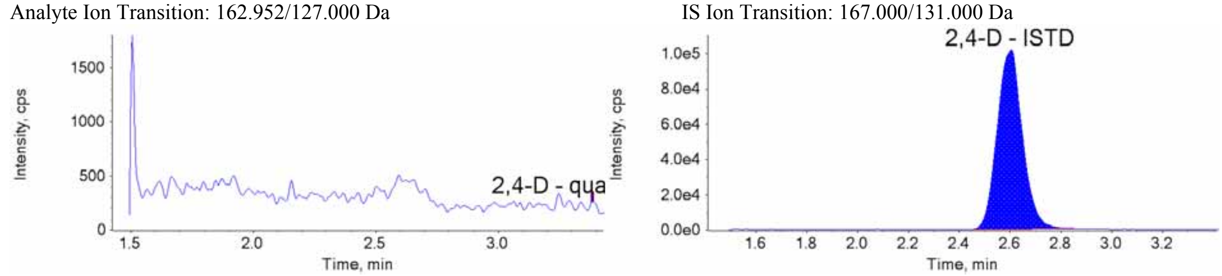
Sample Name: ARA-09-15-10-022-0001A2
Data File: ARA-09-15-10 D16.wiff
Analyte Integration Type: Base To Base
Analyte Peak Area: 2102
Analyte Ion Transition: 160.953/125.000 Da

Acquired: 8/10/2010 7:30:00 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 29408
IS Ion Transition: 167.000/131.000 Da



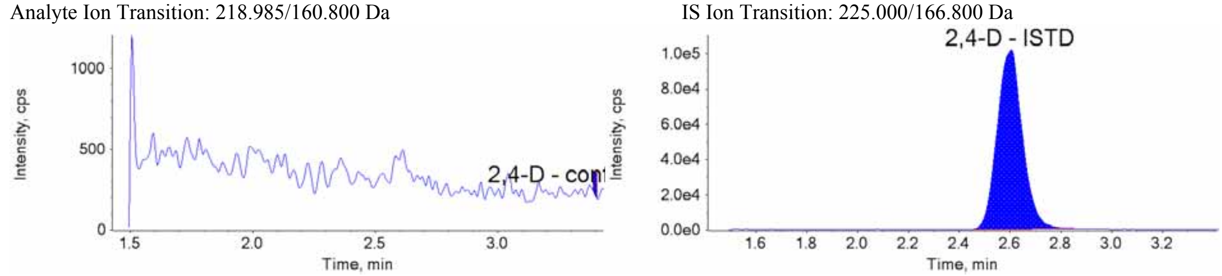
Sample Name: ARA-09-15-10-022-0001A2
Data File: ARA-09-15-10 D16.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 0
Analyte Ion Transition: 162.952/127.000 Da

Acquired: 8/10/2010 7:30:00 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 29408
IS Ion Transition: 167.000/131.000 Da



Sample Name: ARA-09-15-10-022-0001A2
Data File: ARA-09-15-10 D16.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 0
Analyte Ion Transition: 218.985/160.800 Da

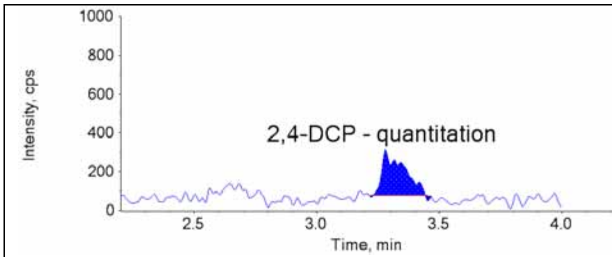
Acquired: 8/10/2010 7:30:00 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 703412
IS Ion Transition: 225.000/166.800 Da

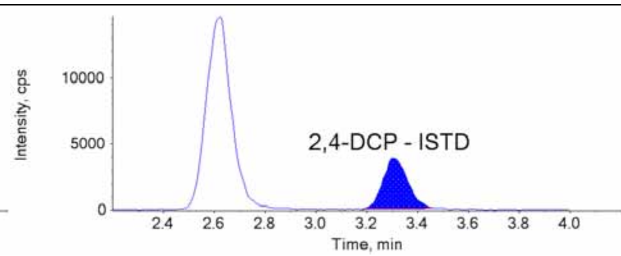


Sample Name: ARA-09-15-10-022-0001A2
Data File: ARA-09-15-10 D16.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 0
Analyte Ion Transition: 220.968/163.000 Da

Acquired: 8/10/2010 7:30:00 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 703412
IS Ion Transition: 225.000/166.800 Da

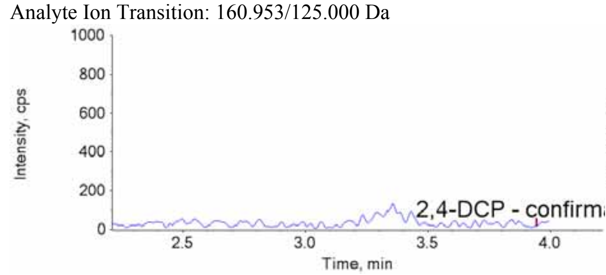
Figure 8. Typical Chromatogram of 2,4-D Analysis, Control Corn Grain (022-0001)

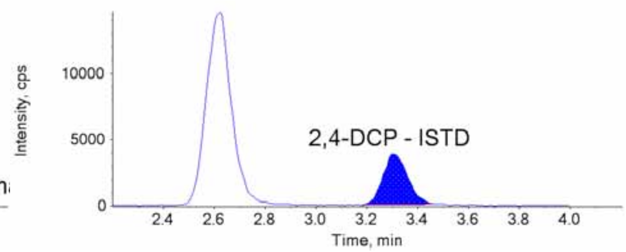




Sample Name: ARA-09-15-10-024-0001A2
Data File: ARA-09-15-10 D16.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 1439
Analyte Ion Transition: 160.953/125.000 Da

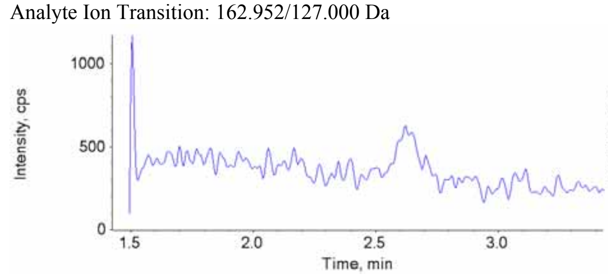
Acquired: 8/10/2010 7:36:26 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 26258
IS Ion Transition: 167.000/131.000 Da

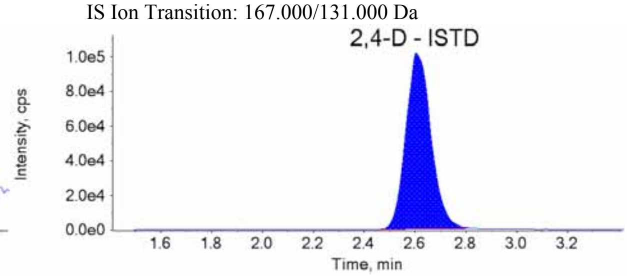




Sample Name: ARA-09-15-10-024-0001A2
Data File: ARA-09-15-10 D16.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 0
Analyte Ion Transition: 162.952/127.000 Da

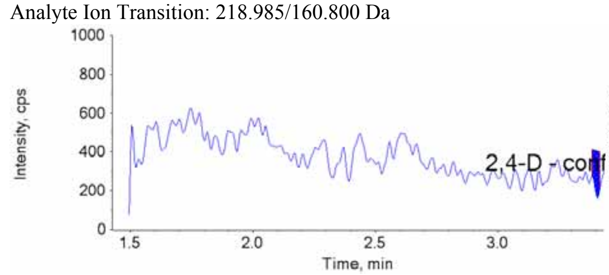
Acquired: 8/10/2010 7:36:26 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 26258
IS Ion Transition: 167.000/131.000 Da

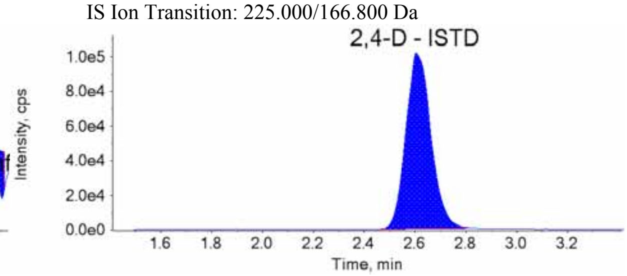




Sample Name: ARA-09-15-10-024-0001A2
Data File: ARA-09-15-10 D16.wiff
Analyte Integration Type: No Peak
Analyte Peak Area: 0
Analyte Ion Transition: 218.985/160.800 Da

Acquired: 8/10/2010 7:36:26 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 698561
IS Ion Transition: 225.000/166.800 Da

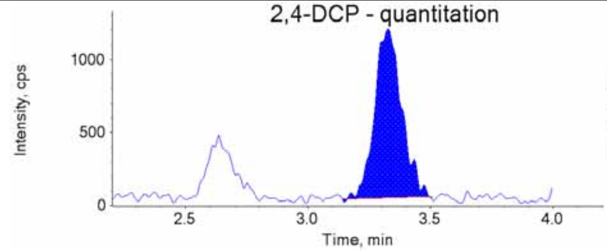


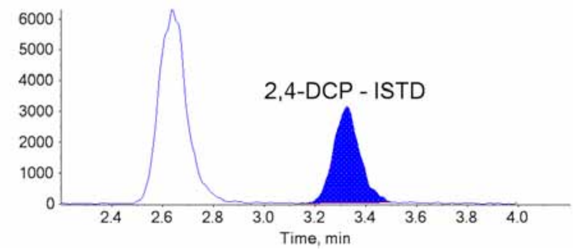


Sample Name: ARA-09-15-10-024-0001A2
Data File: ARA-09-15-10 D16.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 0
Analyte Ion Transition: 220.968/163.000 Da

Acquired: 8/10/2010 7:36:26 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 698561
IS Ion Transition: 225.000/166.800 Da

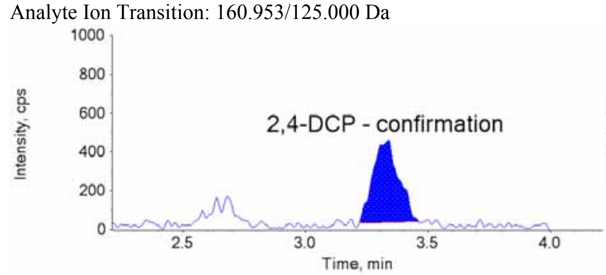
Figure 9. Typical Chromatogram of 2,4-D Analysis, Control Corn Grain (024-0001)

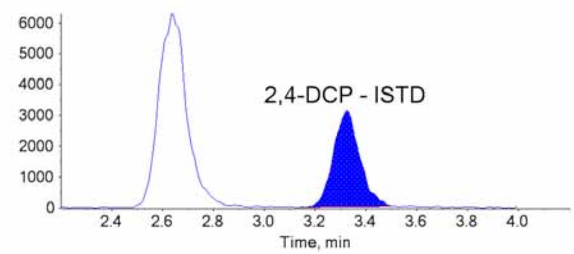




Sample Name: ARA-09-15-10-011-0001A4 + 0.01 µg/g
Data File: ARA-09-15-10 D16.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 8911
Analyte Ion Transition: 160.953/125.000 Da

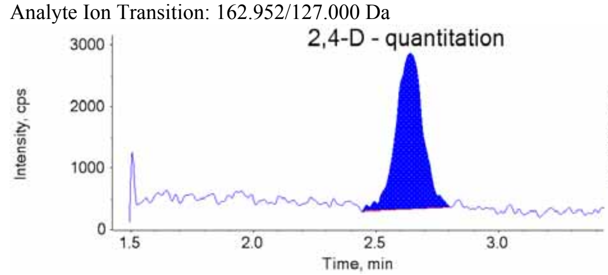
Acquired: 8/11/2010 12:59:48 AM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 22417
IS Ion Transition: 167.000/131.000 Da

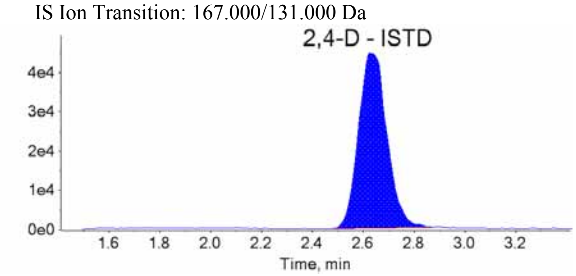




Sample Name: ARA-09-15-10-011-0001A4 + 0.01 µg/g
Data File: ARA-09-15-10 D16.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 3112
Analyte Ion Transition: 162.952/127.000 Da

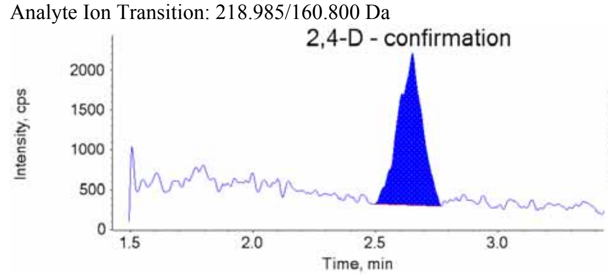
Acquired: 8/11/2010 12:59:48 AM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 22417
IS Ion Transition: 167.000/131.000 Da

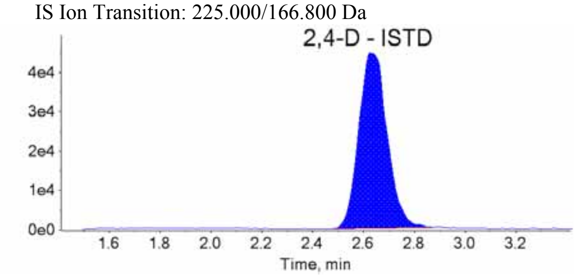




Sample Name: ARA-09-15-10-011-0001A4 + 0.01 µg/g
Data File: ARA-09-15-10 D16.wiff
Analyte Integration Type: Base To Base
Analyte Peak Area: 19594
Analyte Ion Transition: 218.985/160.800 Da

Acquired: 8/11/2010 12:59:48 AM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 344637
IS Ion Transition: 225.000/166.800 Da

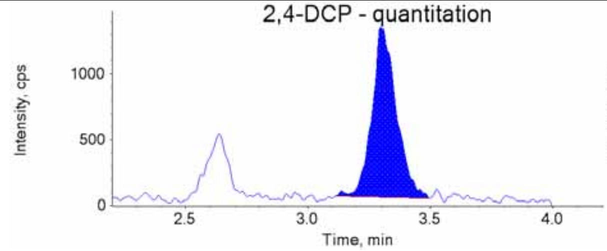


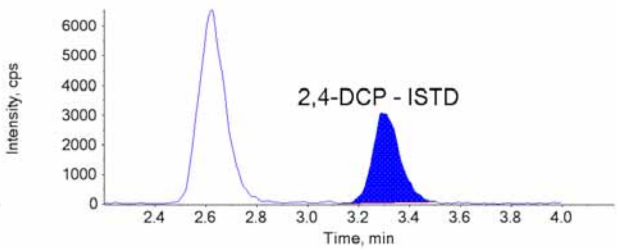


Sample Name: ARA-09-15-10-011-0001A4 + 0.01 µg/g
Data File: ARA-09-15-10 D16.wiff
Analyte Integration Type: Base To Base
Analyte Peak Area: 13001
Analyte Ion Transition: 220.968/163.000 Da

Acquired: 8/11/2010 12:59:48 AM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 344637
IS Ion Transition: 225.000/166.800 Da

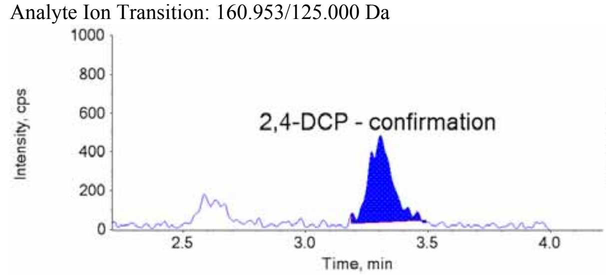
Figure 10. Typical Chromatogram of 2,4-D Analysis, Corn Grain 0.01 µg/g Recovery

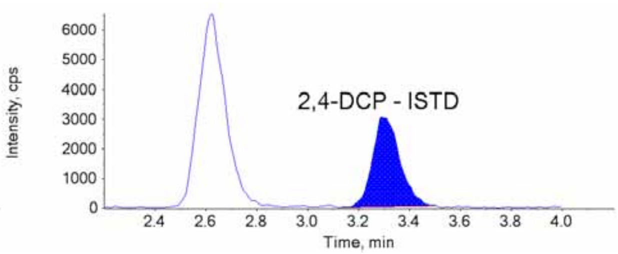




Sample Name: ARA-09-15-10-011-0001A5 + 0.01 µg/g
Data File: ARA-09-15-10 D16.wiff
Analyte Integration Type: Base To Base
Analyte Peak Area: 9027
Analyte Ion Transition: 160.953/125.000 Da

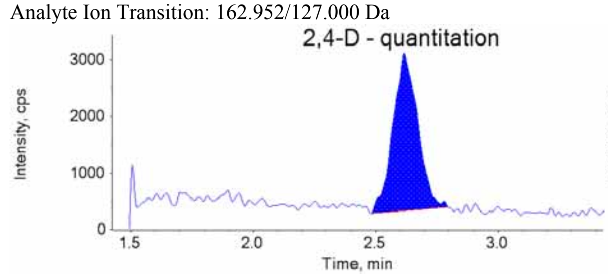
Acquired: 8/11/2010 1:06:19 AM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 22682
IS Ion Transition: 167.000/131.000 Da

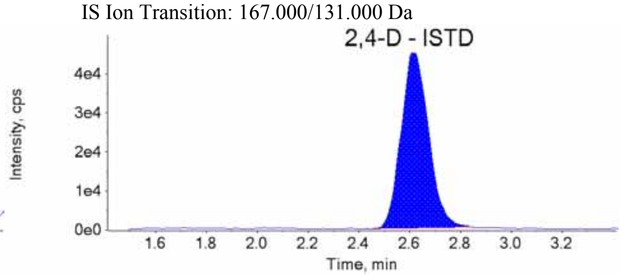




Sample Name: ARA-09-15-10-011-0001A5 + 0.01 µg/g
Data File: ARA-09-15-10 D16.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 2981
Analyte Ion Transition: 162.952/127.000 Da

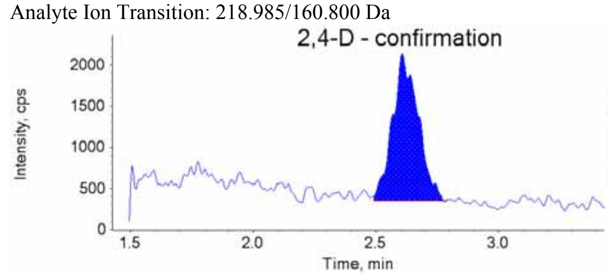
Acquired: 8/11/2010 1:06:19 AM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 22682
IS Ion Transition: 167.000/131.000 Da

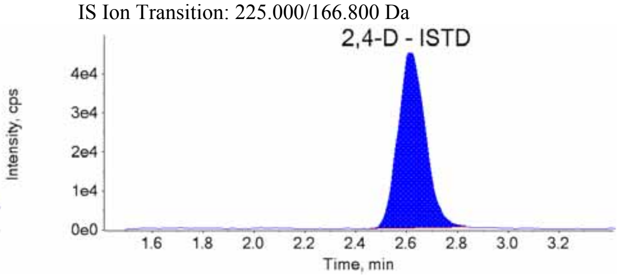




Sample Name: ARA-09-15-10-011-0001A5 + 0.01 µg/g
Data File: ARA-09-15-10 D16.wiff
Analyte Integration Type: Base To Base
Analyte Peak Area: 19493
Analyte Ion Transition: 218.985/160.800 Da

Acquired: 8/11/2010 1:06:19 AM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 337652
IS Ion Transition: 225.000/166.800 Da

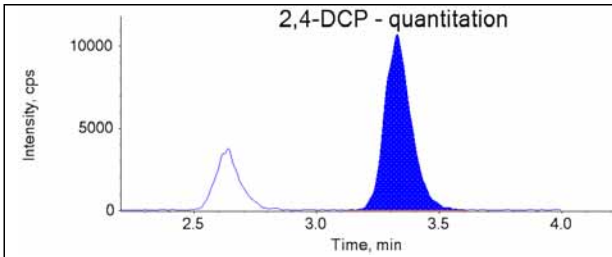


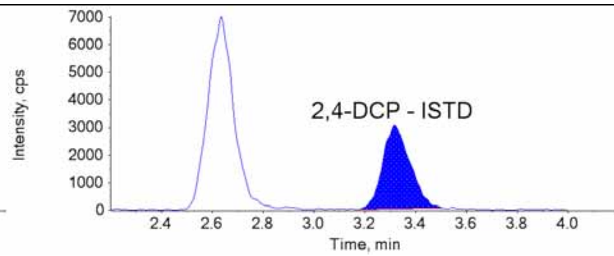


Sample Name: ARA-09-15-10-011-0001A5 + 0.01 µg/g
Data File: ARA-09-15-10 D16.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 13053
Analyte Ion Transition: 220.968/163.000 Da

Acquired: 8/11/2010 1:06:19 AM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 337652
IS Ion Transition: 225.000/166.800 Da

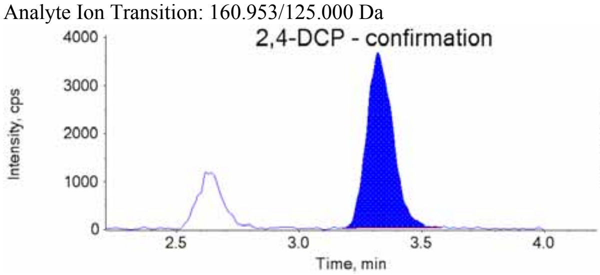
Figure 11. Typical Chromatogram of 2,4-D Analysis, Corn Grain 0.01 µg/g Recovery

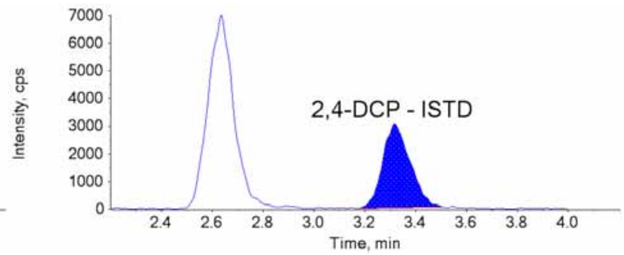




Sample Name: ARA-09-15-10-011-0001A6 + 0.10 µg/g
Data File: ARA-09-15-10 D16.wiff
Analyte Integration Type: Base To Base
Analyte Peak Area: 79842
Analyte Ion Transition: 160.953/125.000 Da

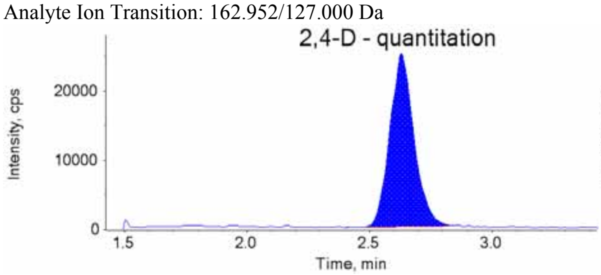
Acquired: 8/11/2010 1:12:47 AM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 22757
IS Ion Transition: 167.000/131.000 Da

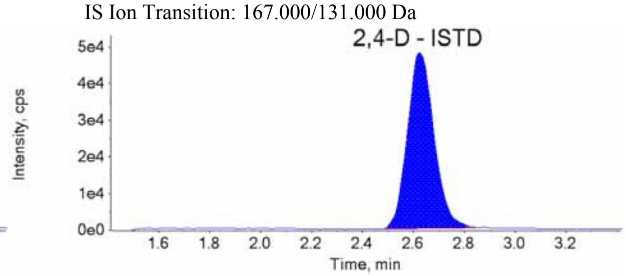




Sample Name: ARA-09-15-10-011-0001A6 + 0.10 µg/g
Data File: ARA-09-15-10 D16.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 27153
Analyte Ion Transition: 162.952/127.000 Da

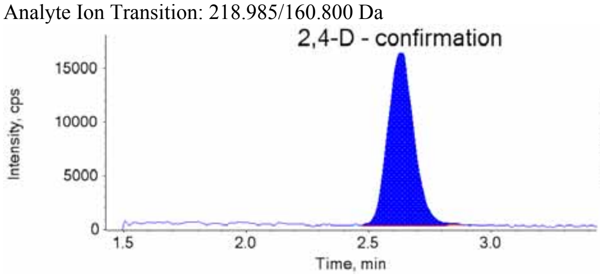
Acquired: 8/11/2010 1:12:47 AM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 22757
IS Ion Transition: 167.000/131.000 Da

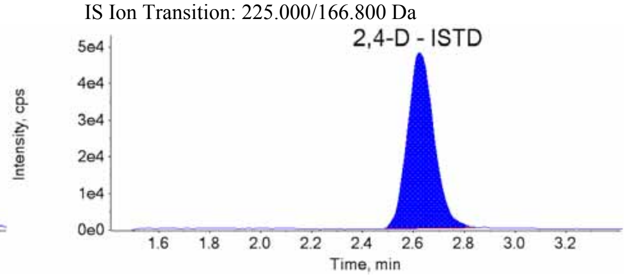




Sample Name: ARA-09-15-10-011-0001A6 + 0.10 µg/g
Data File: ARA-09-15-10 D16.wiff
Analyte Integration Type: Base To Base
Analyte Peak Area: 170938
Analyte Ion Transition: 218.985/160.800 Da

Acquired: 8/11/2010 1:12:47 AM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 348600
IS Ion Transition: 225.000/166.800 Da

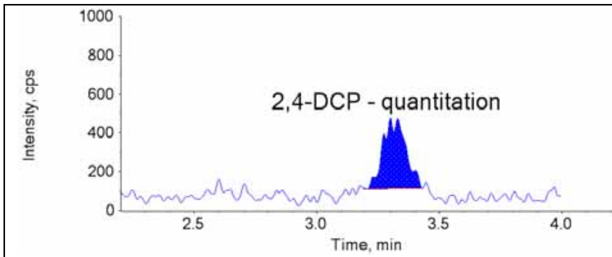


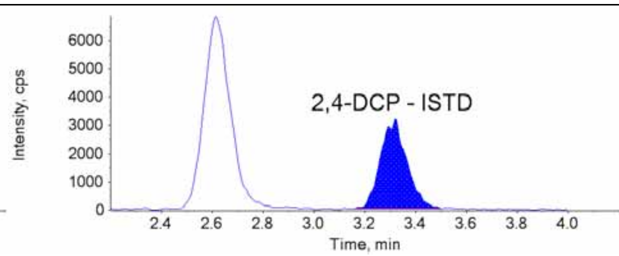


Sample Name: ARA-09-15-10-011-0001A6 + 0.10 µg/g
Data File: ARA-09-15-10 D16.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 119283
Analyte Ion Transition: 220.968/163.000 Da

Acquired: 8/11/2010 1:12:47 AM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 348600
IS Ion Transition: 225.000/166.800 Da

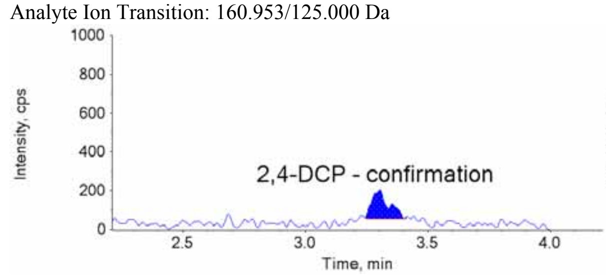
Figure 12. Typical Chromatogram of 2,4-D Analysis, Corn Grain 0.10 µg/g Recovery

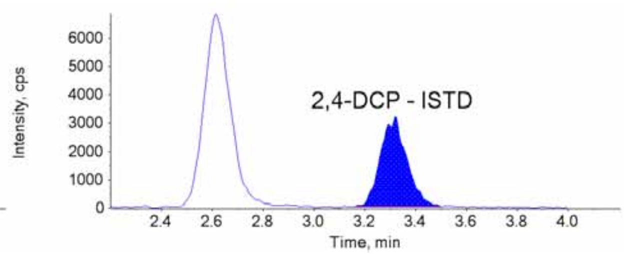




Sample Name: ARA-09-15-10-148-0002A2
Data File: ARA-09-15-10 D16.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 2370
Analyte Ion Transition: 160.953/125.000 Da

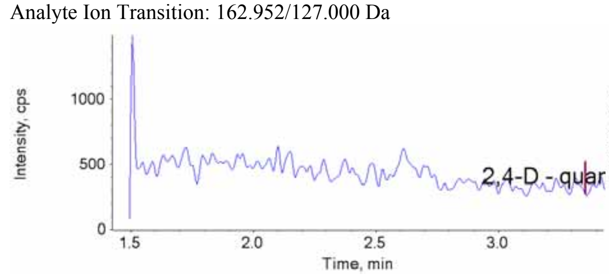
Acquired: 8/11/2010 12:21:03 AM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 22931
IS Ion Transition: 167.000/131.000 Da

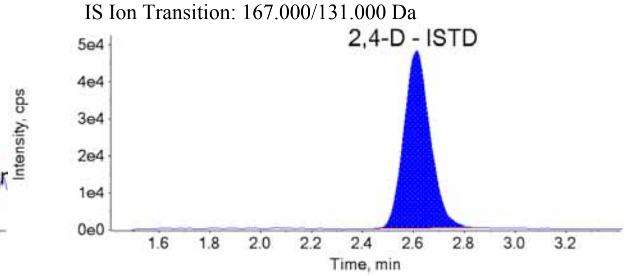




Sample Name: ARA-09-15-10-148-0002A2
Data File: ARA-09-15-10 D16.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 726
Analyte Ion Transition: 162.952/127.000 Da

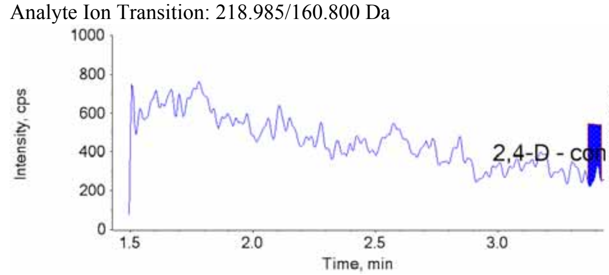
Acquired: 8/11/2010 12:21:03 AM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 22931
IS Ion Transition: 167.000/131.000 Da

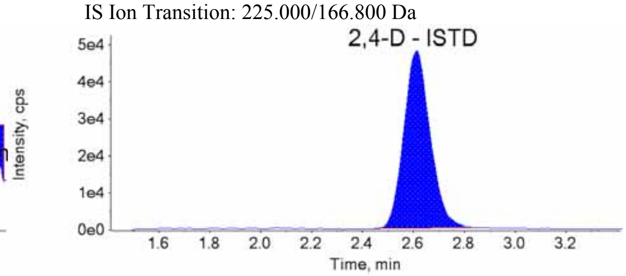




Sample Name: ARA-09-15-10-148-0002A2
Data File: ARA-09-15-10 D16.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 0
Analyte Ion Transition: 218.985/160.800 Da

Acquired: 8/11/2010 12:21:03 AM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 341475
IS Ion Transition: 225.000/166.800 Da

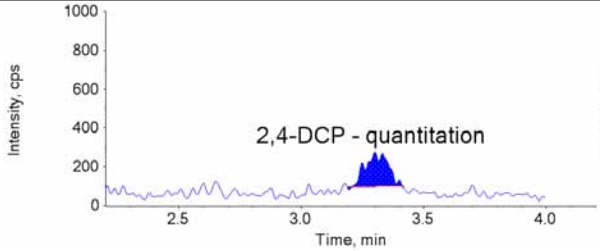


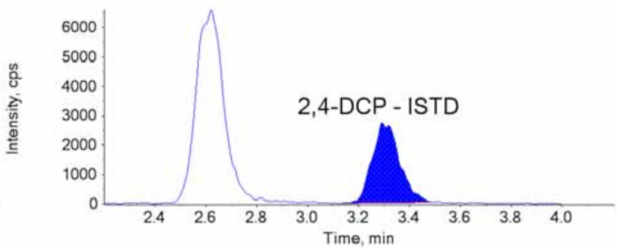


Sample Name: ARA-09-15-10-148-0002A2
Data File: ARA-09-15-10 D16.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 0
Analyte Ion Transition: 220.968/163.000 Da

Acquired: 8/11/2010 12:21:03 AM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 341475
IS Ion Transition: 225.000/166.800 Da

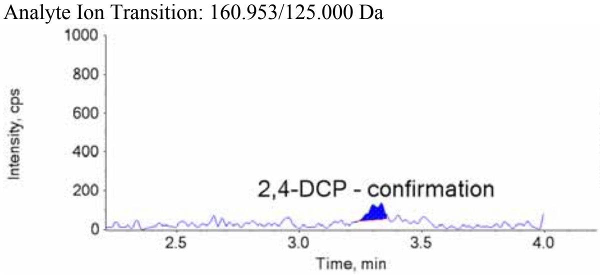
Figure 13. Typical Chromatogram of 2,4-D Analysis, Treated Corn Grain (148-0002)

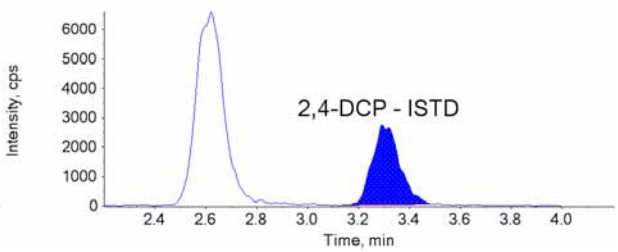




Sample Name: ARA-09-15-10-148-0003A2
Data File: ARA-09-15-10 D16.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 1146
Analyte Ion Transition: 160.953/125.000 Da

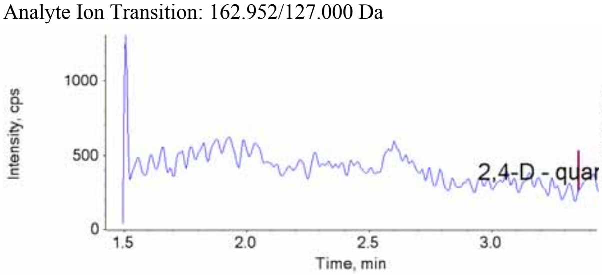
Acquired: 8/11/2010 12:27:30 AM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 20422
IS Ion Transition: 167.000/131.000 Da

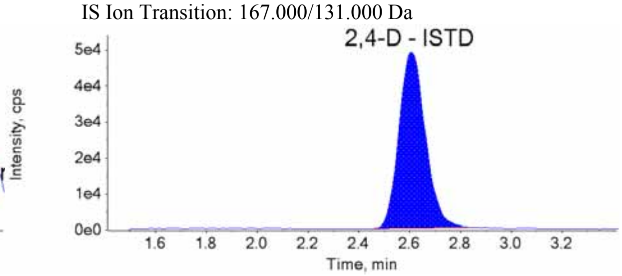




Sample Name: ARA-09-15-10-148-0003A2
Data File: ARA-09-15-10 D16.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 334
Analyte Ion Transition: 162.952/127.000 Da

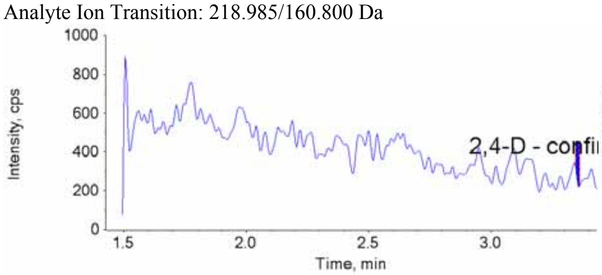
Acquired: 8/11/2010 12:27:30 AM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 20422
IS Ion Transition: 167.000/131.000 Da

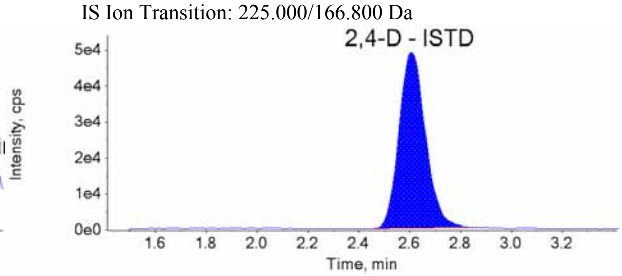




Sample Name: ARA-09-15-10-148-0003A2
Data File: ARA-09-15-10 D16.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 0
Analyte Ion Transition: 218.985/160.800 Da

Acquired: 8/11/2010 12:27:30 AM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 352870
IS Ion Transition: 225.000/166.800 Da

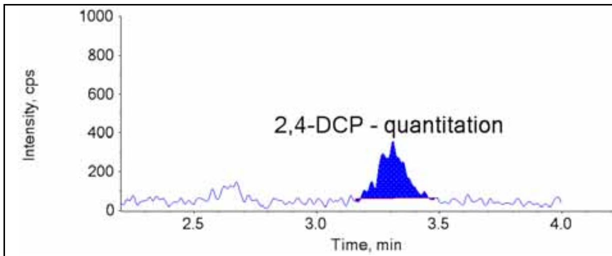


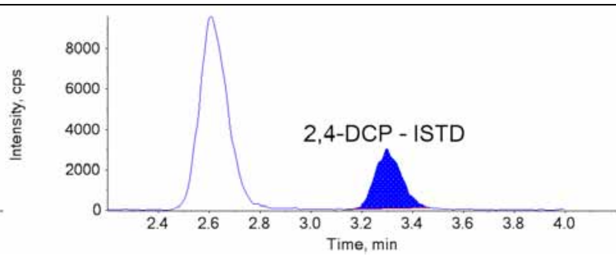


Sample Name: ARA-09-15-10-148-0003A2
Data File: ARA-09-15-10 D16.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 0
Analyte Ion Transition: 220.968/163.000 Da

Acquired: 8/11/2010 12:27:30 AM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 352870
IS Ion Transition: 225.000/166.800 Da

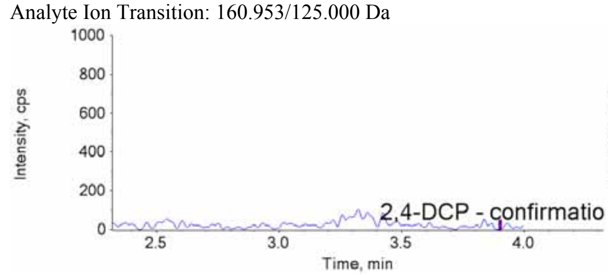
Figure 14. Typical Chromatogram of 2,4-D Analysis, Treated Corn Grain (148-0003)

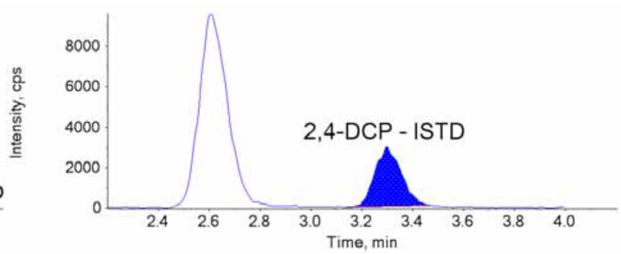




Sample Name: ARA-09-15-10-020-0002A2
Data File: ARA-09-15-10 D17.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 1963
Analyte Ion Transition: 160.953/125.000 Da

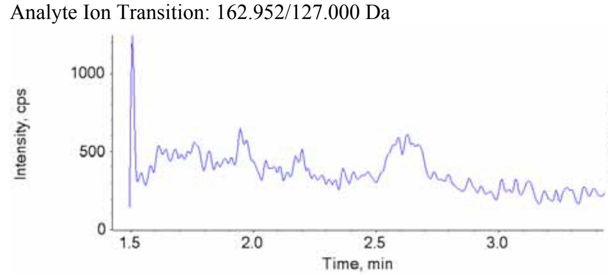
Acquired: 8/11/2010 6:45:24 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 21207
IS Ion Transition: 167.000/131.000 Da

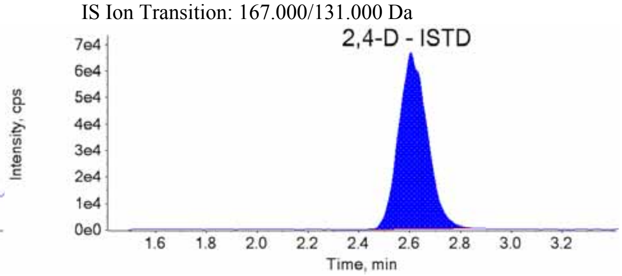




Sample Name: ARA-09-15-10-020-0002A2
Data File: ARA-09-15-10 D17.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 0
Analyte Ion Transition: 162.952/127.000 Da

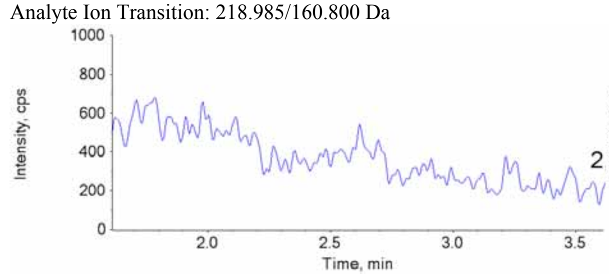
Acquired: 8/11/2010 6:45:24 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 21207
IS Ion Transition: 167.000/131.000 Da

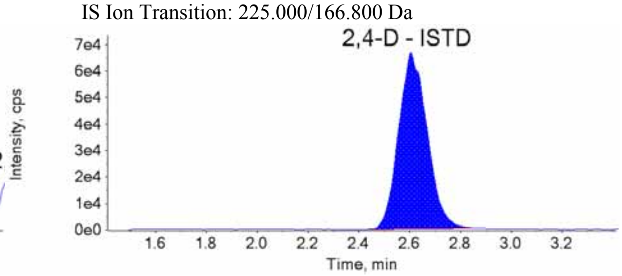




Sample Name: ARA-09-15-10-020-0002A2
Data File: ARA-09-15-10 D17.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 0
Analyte Ion Transition: 218.985/160.800 Da

Acquired: 8/11/2010 6:45:24 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 519815
IS Ion Transition: 225.000/166.800 Da

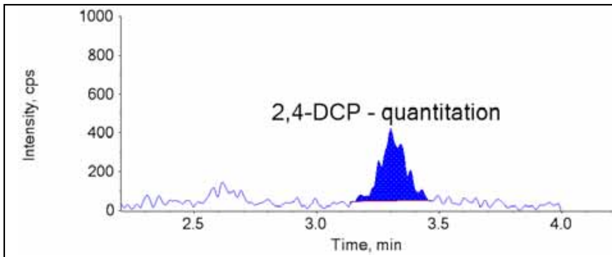


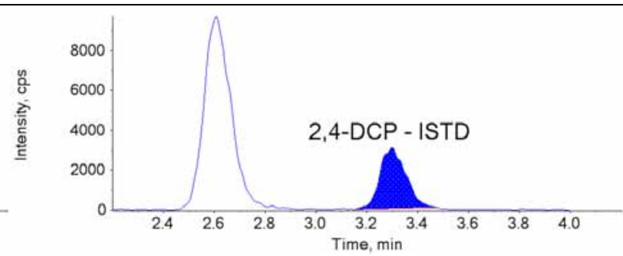


Sample Name: ARA-09-15-10-020-0002A2
Data File: ARA-09-15-10 D17.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 0
Analyte Ion Transition: 220.968/163.000 Da

Acquired: 8/11/2010 6:45:24 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 519815
IS Ion Transition: 225.000/166.800 Da

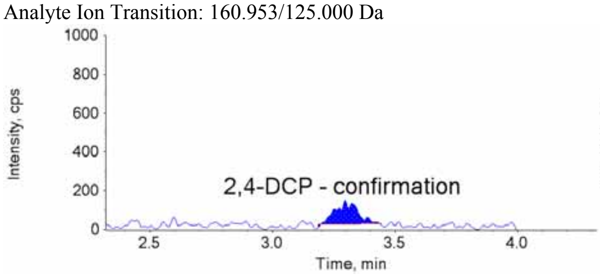
Figure 15. Typical Chromatogram of 2,4-D Analysis, Treated Corn Grain (020-0002)

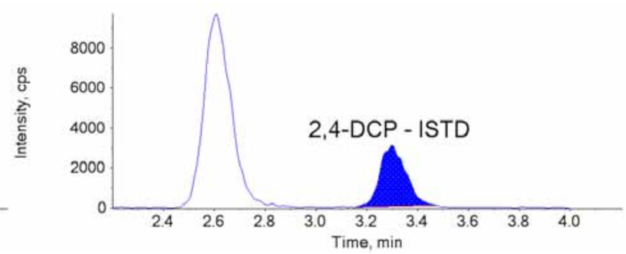




Sample Name: ARA-09-15-10-020-0003A2
Data File: ARA-09-15-10 D17.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 2503
Analyte Ion Transition: 160.953/125.000 Da

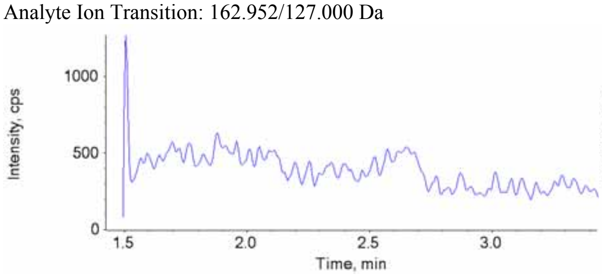
Acquired: 8/11/2010 6:51:52 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 21450
IS Ion Transition: 167.000/131.000 Da

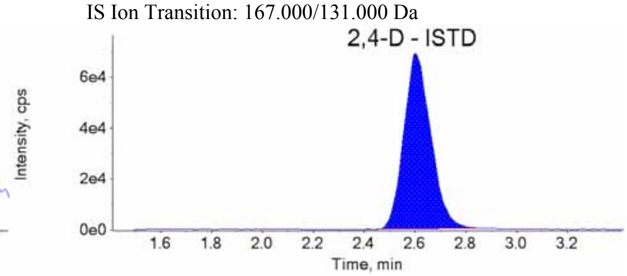




Sample Name: ARA-09-15-10-020-0003A2
Data File: ARA-09-15-10 D17.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 753
Analyte Ion Transition: 162.952/127.000 Da

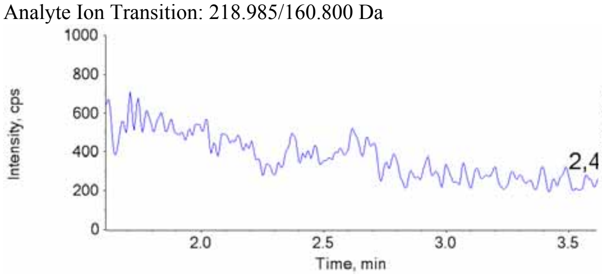
Acquired: 8/11/2010 6:51:52 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 21450
IS Ion Transition: 167.000/131.000 Da

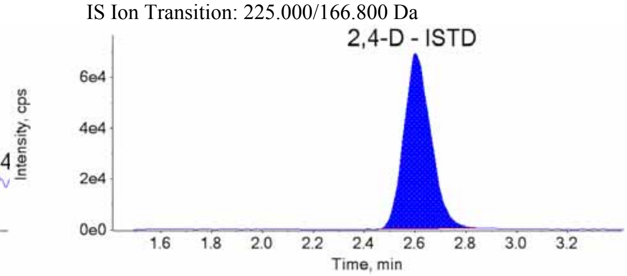




Sample Name: ARA-09-15-10-020-0003A2
Data File: ARA-09-15-10 D17.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 0
Analyte Ion Transition: 218.985/160.800 Da

Acquired: 8/11/2010 6:51:52 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 506196
IS Ion Transition: 225.000/166.800 Da

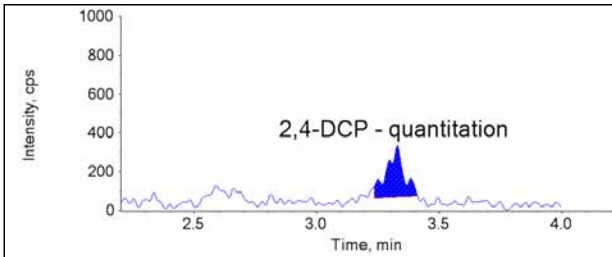


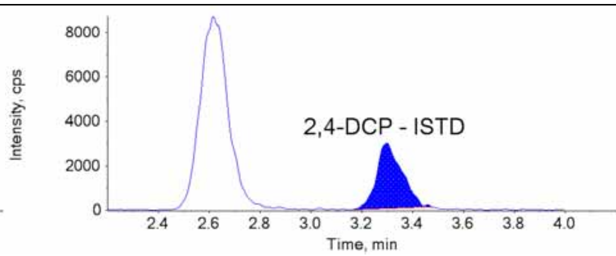


Sample Name: ARA-09-15-10-020-0003A2
Data File: ARA-09-15-10 D17.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 0
Analyte Ion Transition: 220.968/163.000 Da

Acquired: 8/11/2010 6:51:52 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 506196
IS Ion Transition: 225.000/166.800 Da

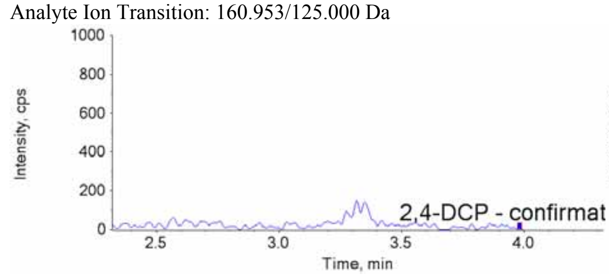
Figure 16. Typical Chromatogram of 2,4-D Analysis, Treated Corn Grain (020-0003)

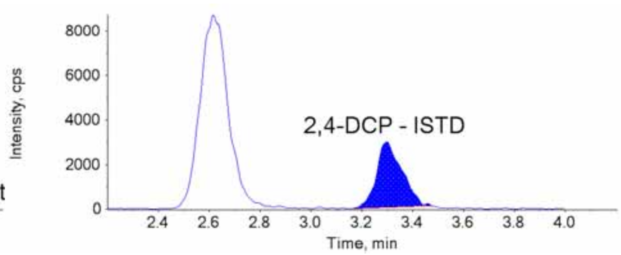




Sample Name: ARA-09-15-10-022-0002A2
Data File: ARA-09-15-10 D17.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 1267
Analyte Ion Transition: 160.953/125.000 Da

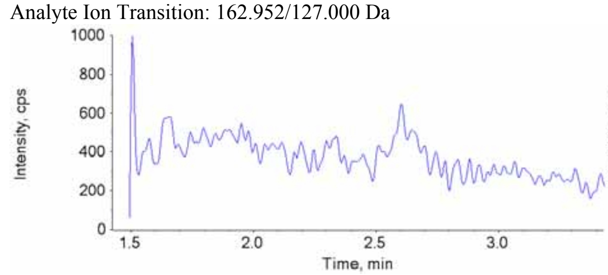
Acquired: 8/11/2010 6:58:19 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 20335
IS Ion Transition: 167.000/131.000 Da

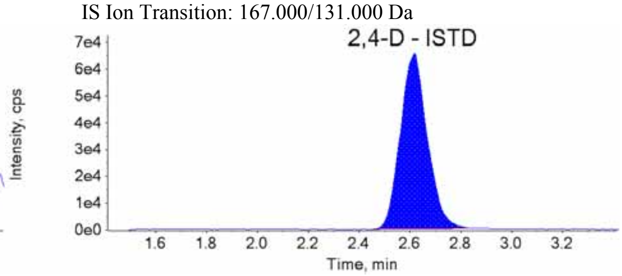




Sample Name: ARA-09-15-10-022-0002A2
Data File: ARA-09-15-10 D17.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 0
Analyte Ion Transition: 162.952/127.000 Da

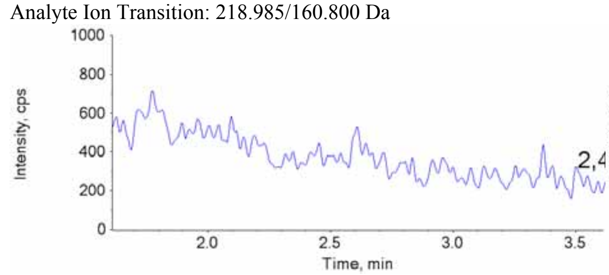
Acquired: 8/11/2010 6:58:19 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 20335
IS Ion Transition: 167.000/131.000 Da

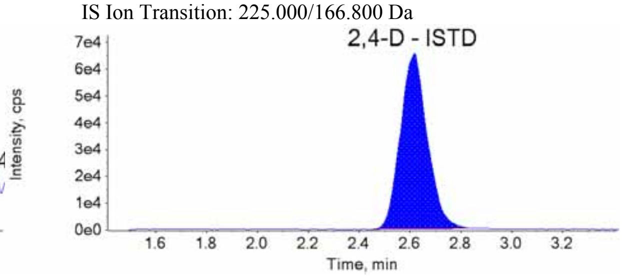




Sample Name: ARA-09-15-10-022-0002A2
Data File: ARA-09-15-10 D17.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 0
Analyte Ion Transition: 218.985/160.800 Da

Acquired: 8/11/2010 6:58:19 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 479250
IS Ion Transition: 225.000/166.800 Da

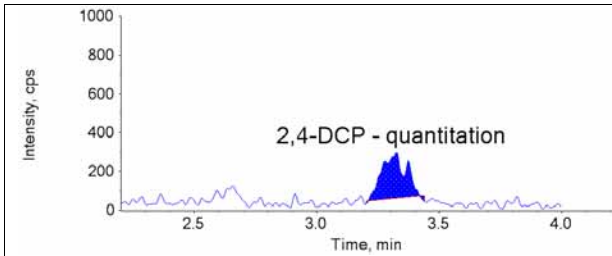


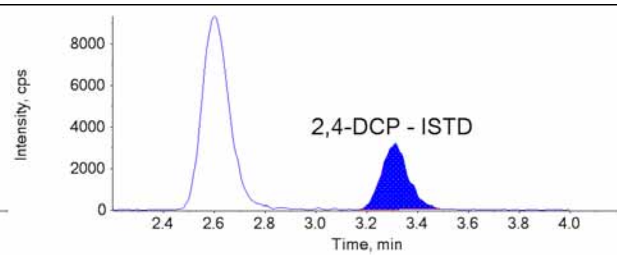


Sample Name: ARA-09-15-10-022-0002A2
Data File: ARA-09-15-10 D17.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 0
Analyte Ion Transition: 220.968/163.000 Da

Acquired: 8/11/2010 6:58:19 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 479250
IS Ion Transition: 225.000/166.800 Da

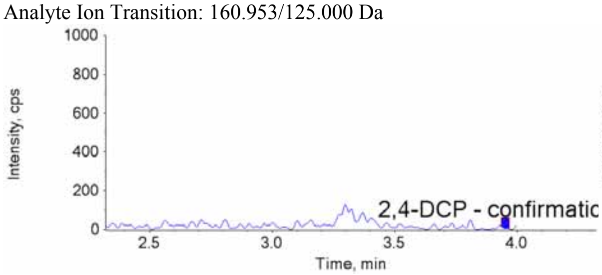
Figure 17. Typical Chromatogram of 2,4-D Analysis, Treated Corn Grain (022-0002)

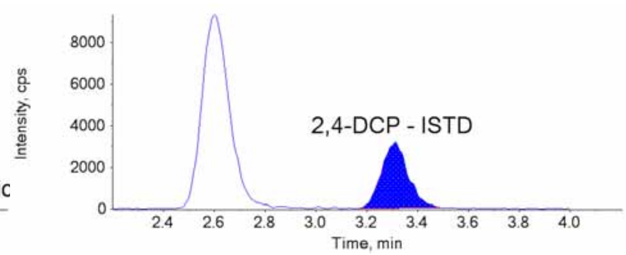




Sample Name: ARA-09-15-10-022-0003A2
Data File: ARA-09-15-10 D17.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 1607
Analyte Ion Transition: 160.953/125.000 Da

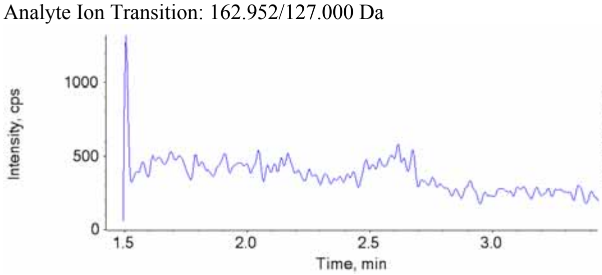
Acquired: 8/11/2010 7:04:47 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 23019
IS Ion Transition: 167.000/131.000 Da

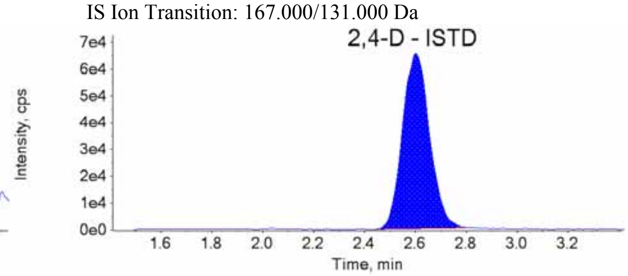




Sample Name: ARA-09-15-10-022-0003A2
Data File: ARA-09-15-10 D17.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 0
Analyte Ion Transition: 162.952/127.000 Da

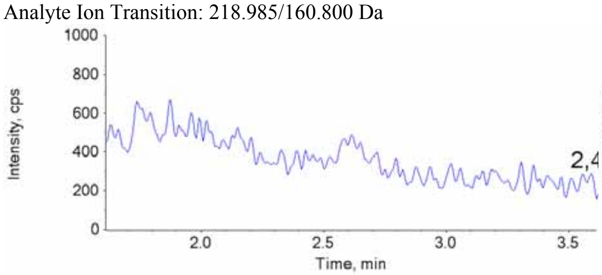
Acquired: 8/11/2010 7:04:47 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 23019
IS Ion Transition: 167.000/131.000 Da

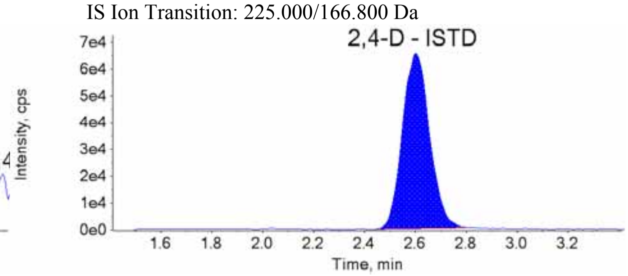




Sample Name: ARA-09-15-10-022-0003A2
Data File: ARA-09-15-10 D17.wiff
Analyte Integration Type: No Peak
Analyte Peak Area: 0
Analyte Ion Transition: 218.985/160.800 Da

Acquired: 8/11/2010 7:04:47 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 483446
IS Ion Transition: 225.000/166.800 Da

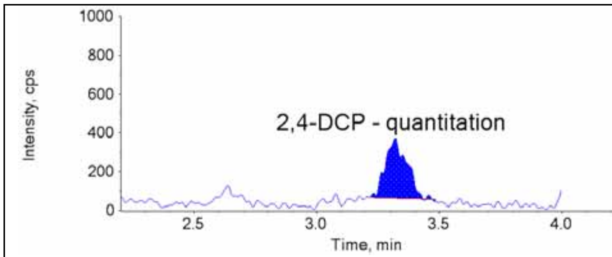


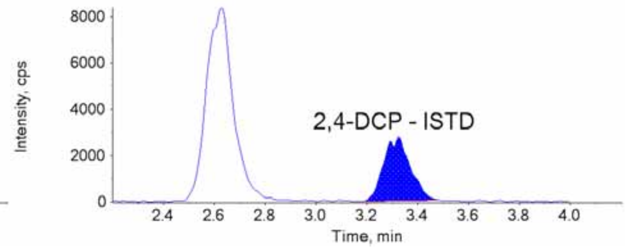


Sample Name: ARA-09-15-10-022-0003A2
Data File: ARA-09-15-10 D17.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 0
Analyte Ion Transition: 220.968/163.000 Da

Acquired: 8/11/2010 7:04:47 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 483446
IS Ion Transition: 225.000/166.800 Da

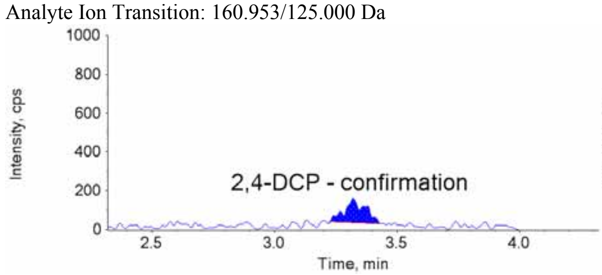
Figure 18. Typical Chromatogram of 2,4-D Analysis, Treated Corn Grain (022-0003)

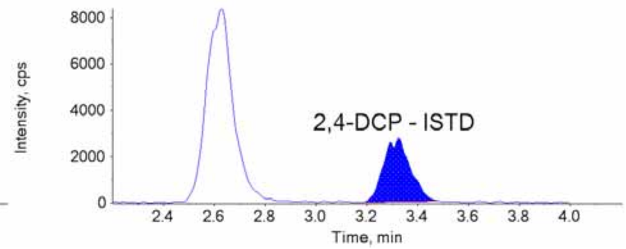




Sample Name: ARA-09-15-10-032-0002A3
Data File: ARA-09-15-10 D17.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 1881
Analyte Ion Transition: 160.953/125.000 Da

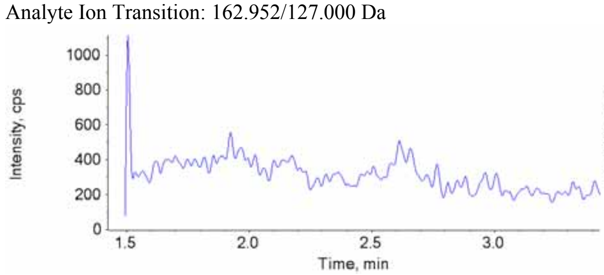
Acquired: 8/11/2010 7:56:29 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 20177
IS Ion Transition: 167.000/131.000 Da

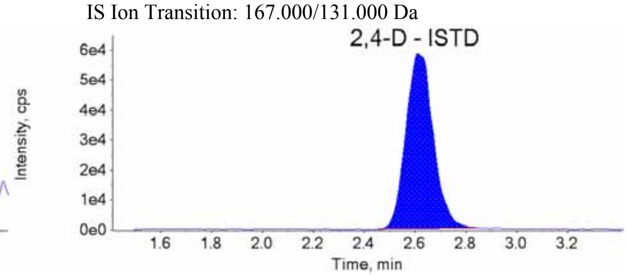




Sample Name: ARA-09-15-10-032-0002A3
Data File: ARA-09-15-10 D17.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 738
Analyte Ion Transition: 162.952/127.000 Da

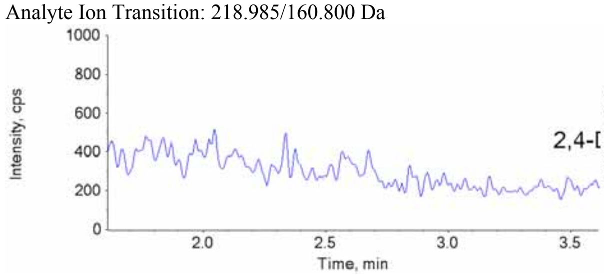
Acquired: 8/11/2010 7:56:29 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 20177
IS Ion Transition: 167.000/131.000 Da

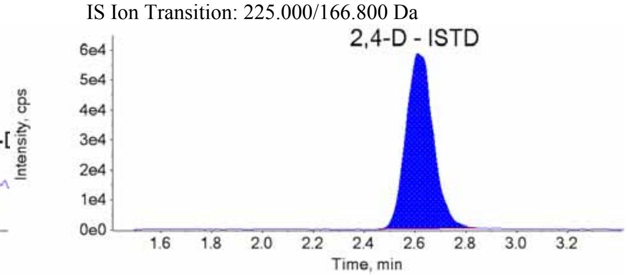




Sample Name: ARA-09-15-10-032-0002A3
Data File: ARA-09-15-10 D17.wiff
Analyte Integration Type: No Peak
Analyte Peak Area: 0
Analyte Ion Transition: 218.985/160.800 Da

Acquired: 8/11/2010 7:56:29 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 438382
IS Ion Transition: 225.000/166.800 Da

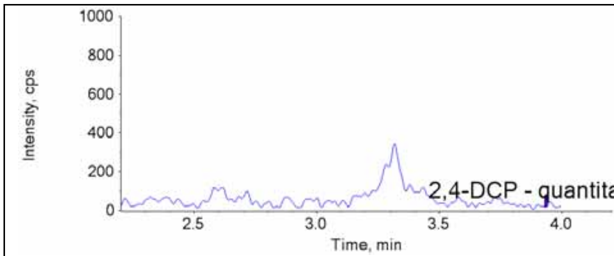


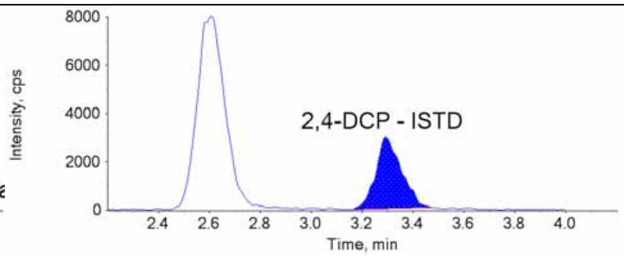


Sample Name: ARA-09-15-10-032-0002A3
Data File: ARA-09-15-10 D17.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 0
Analyte Ion Transition: 220.968/163.000 Da

Acquired: 8/11/2010 7:56:29 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 438382
IS Ion Transition: 225.000/166.800 Da

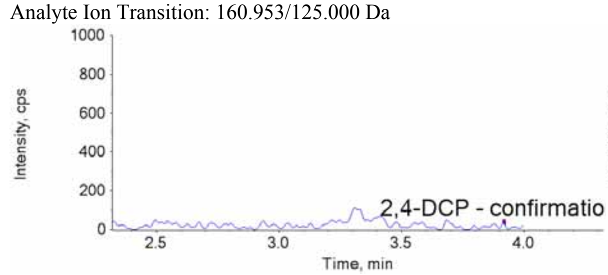
Figure 19. Typical Chromatogram of 2,4-D Analysis, Treated Corn Grain (032-0002)

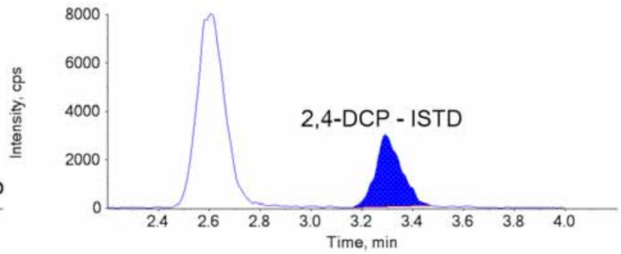




Sample Name: ARA-09-15-10-032-0003A2
Data File: ARA-09-15-10 D17.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 0
Analyte Ion Transition: 160.953/125.000 Da

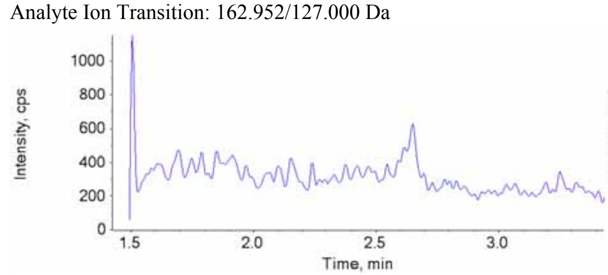
Acquired: 8/11/2010 8:02:58 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 20666
IS Ion Transition: 167.000/131.000 Da

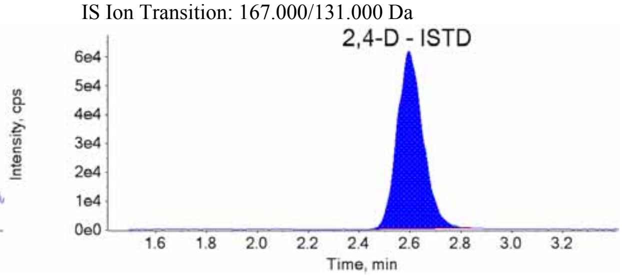




Sample Name: ARA-09-15-10-032-0003A2
Data File: ARA-09-15-10 D17.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 0
Analyte Ion Transition: 162.952/127.000 Da

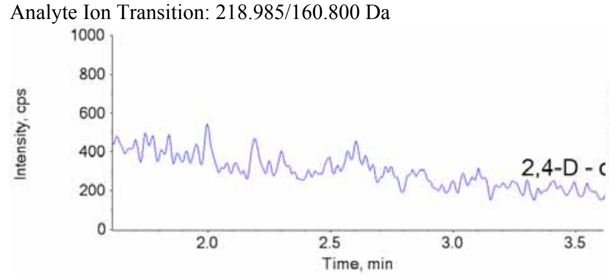
Acquired: 8/11/2010 8:02:58 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 20666
IS Ion Transition: 167.000/131.000 Da

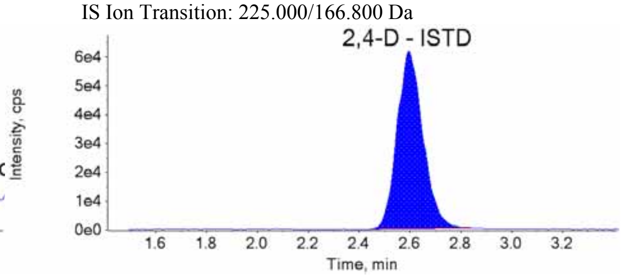




Sample Name: ARA-09-15-10-032-0003A2
Data File: ARA-09-15-10 D17.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 0
Analyte Ion Transition: 218.985/160.800 Da

Acquired: 8/11/2010 8:02:58 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 432903
IS Ion Transition: 225.000/166.800 Da

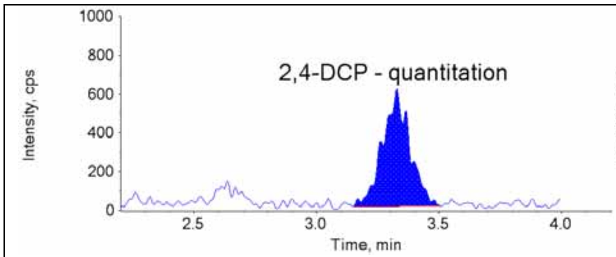


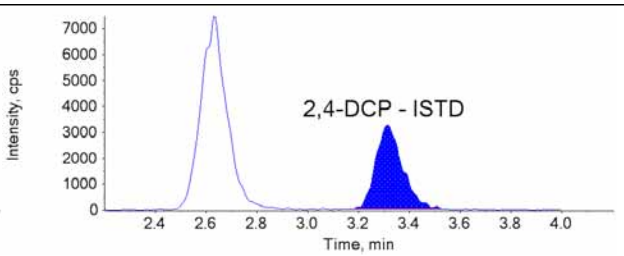


Sample Name: ARA-09-15-10-032-0003A2
Data File: ARA-09-15-10 D17.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 0
Analyte Ion Transition: 220.968/163.000 Da

Acquired: 8/11/2010 8:02:58 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 432903
IS Ion Transition: 225.000/166.800 Da

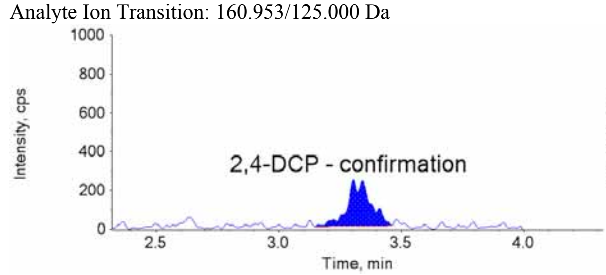
Figure 20. Typical Chromatogram of 2,4-D Analysis, Treated Corn Grain (032-0003)

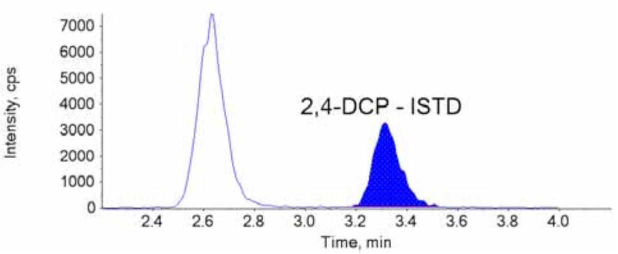




Sample Name: ARA-09-15-10-060-0002A2
Data File: ARA-09-15-10 D17.wiff
Analyte Integration Type: Base To Base
Analyte Peak Area: 4624
Analyte Ion Transition: 160.953/125.000 Da

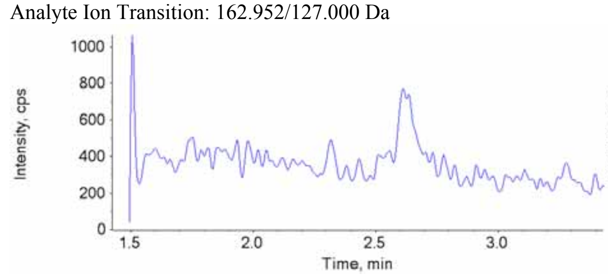
Acquired: 8/11/2010 9:20:37 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 23897
IS Ion Transition: 167.000/131.000 Da

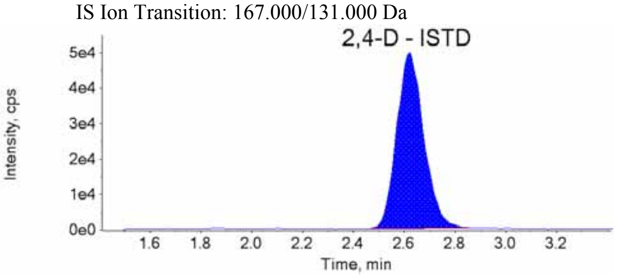




Sample Name: ARA-09-15-10-060-0002A2
Data File: ARA-09-15-10 D17.wiff
Analyte Integration Type: Base To Base
Analyte Peak Area: 1580
Analyte Ion Transition: 162.952/127.000 Da

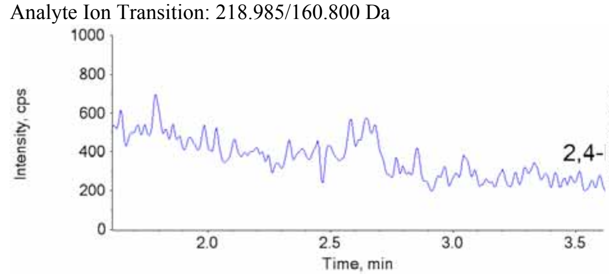
Acquired: 8/11/2010 9:20:37 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 23897
IS Ion Transition: 167.000/131.000 Da

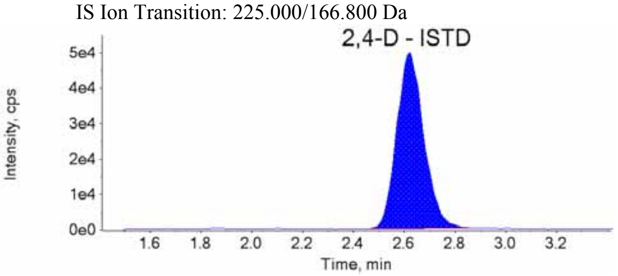




Sample Name: ARA-09-15-10-060-0002A2
Data File: ARA-09-15-10 D17.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 0
Analyte Ion Transition: 218.985/160.800 Da

Acquired: 8/11/2010 9:20:37 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 368352
IS Ion Transition: 225.000/166.800 Da

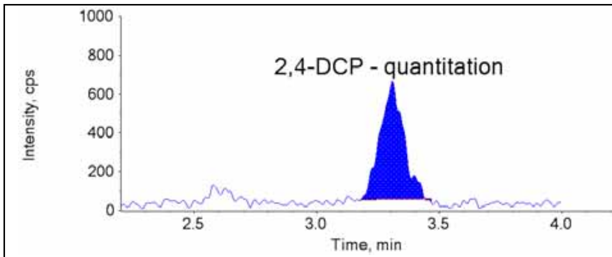


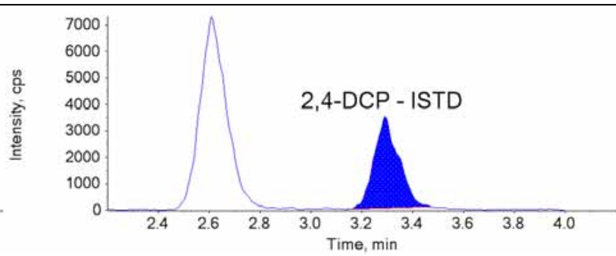


Sample Name: ARA-09-15-10-060-0002A2
Data File: ARA-09-15-10 D17.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 0
Analyte Ion Transition: 220.968/163.000 Da

Acquired: 8/11/2010 9:20:37 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 368352
IS Ion Transition: 225.000/166.800 Da

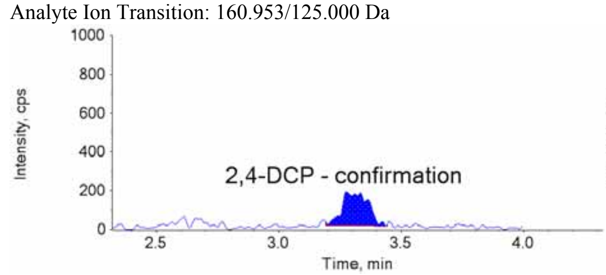
Figure 21. Typical Chromatogram of 2,4-D Analysis, Treated Corn Grain (060-0002)

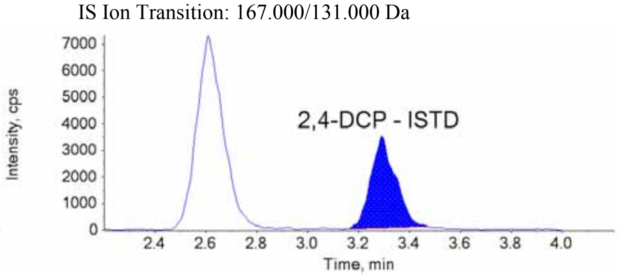




Sample Name: ARA-09-15-10-060-0003A2
Data File: ARA-09-15-10 D17.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 4045
Analyte Ion Transition: 160.953/125.000 Da

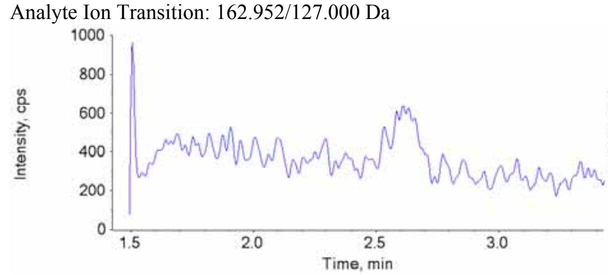
Acquired: 8/11/2010 9:27:07 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 23202
IS Ion Transition: 167.000/131.000 Da

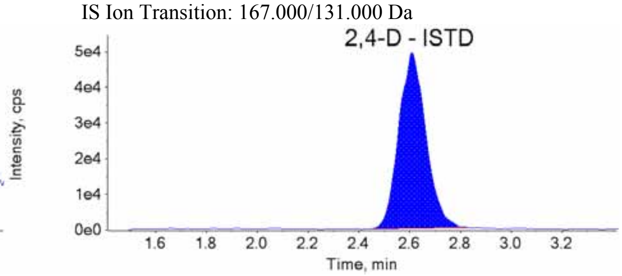




Sample Name: ARA-09-15-10-060-0003A2
Data File: ARA-09-15-10 D17.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 1316
Analyte Ion Transition: 162.952/127.000 Da

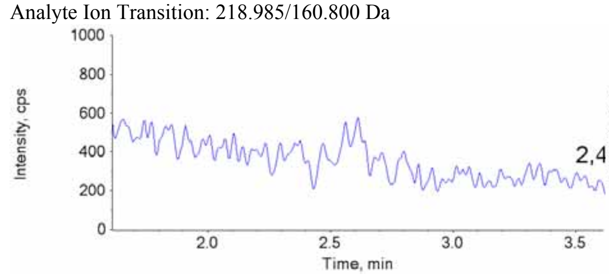
Acquired: 8/11/2010 9:27:07 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 23202
IS Ion Transition: 167.000/131.000 Da

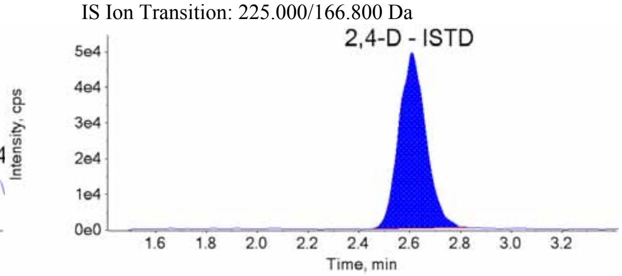




Sample Name: ARA-09-15-10-060-0003A2
Data File: ARA-09-15-10 D17.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 0
Analyte Ion Transition: 218.985/160.800 Da

Acquired: 8/11/2010 9:27:07 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 363983
IS Ion Transition: 225.000/166.800 Da

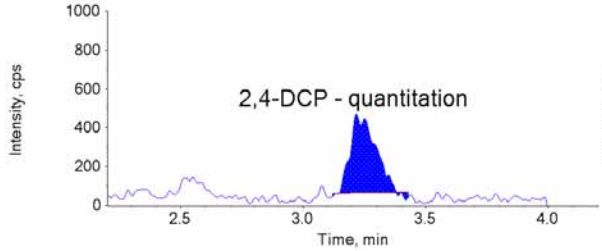


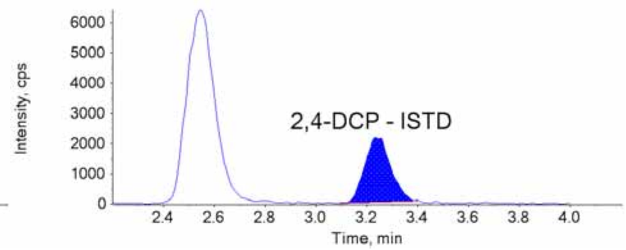


Sample Name: ARA-09-15-10-060-0003A2
Data File: ARA-09-15-10 D17.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 0
Analyte Ion Transition: 220.968/163.000 Da

Acquired: 8/11/2010 9:27:07 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 363983
IS Ion Transition: 225.000/166.800 Da

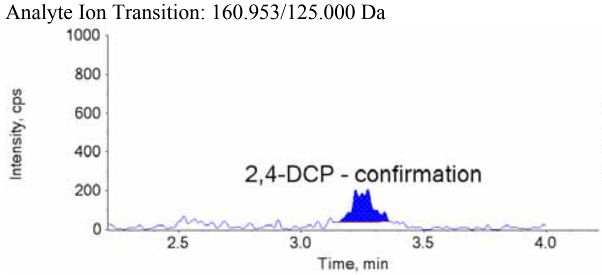
Figure 22. Typical Chromatogram of 2,4-D Analysis, Treated Corn Grain (060-0003)

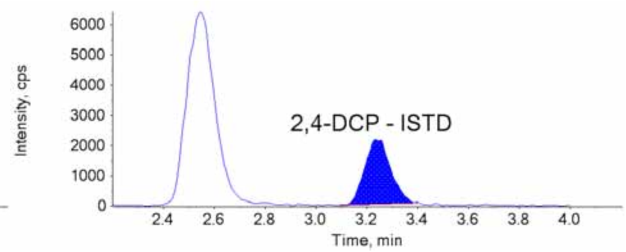




Sample Name: ARA-09-15-10-046-0001A2
Data File: ARA-09-15-10 D21.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 2909
Analyte Ion Transition: 160.953/125.000 Da

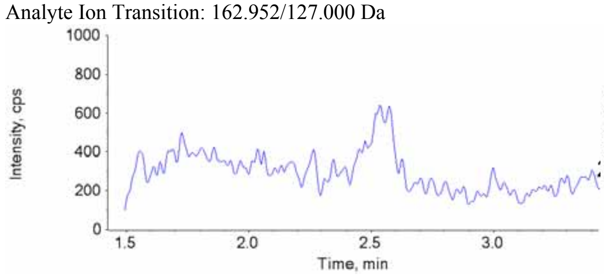
Acquired: 8/19/2010 10:59:05 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 15380
IS Ion Transition: 167.000/131.000 Da

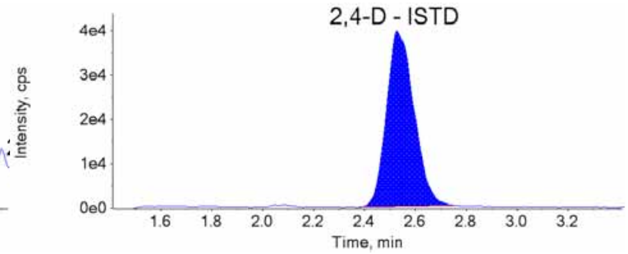




Sample Name: ARA-09-15-10-046-0001A2
Data File: ARA-09-15-10 D21.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 968
Analyte Ion Transition: 162.952/127.000 Da

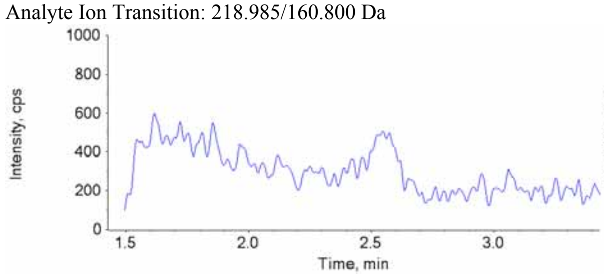
Acquired: 8/19/2010 10:59:05 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 15380
IS Ion Transition: 167.000/131.000 Da

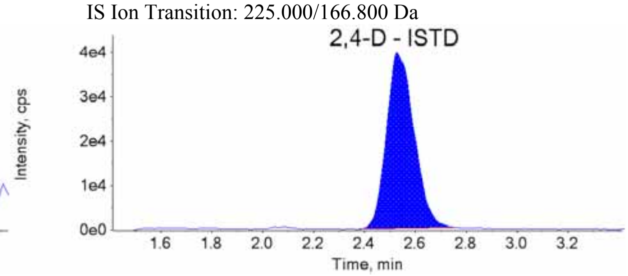




Sample Name: ARA-09-15-10-046-0001A2
Data File: ARA-09-15-10 D21.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 0
Analyte Ion Transition: 218.985/160.800 Da

Acquired: 8/19/2010 10:59:05 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 297466
IS Ion Transition: 225.000/166.800 Da

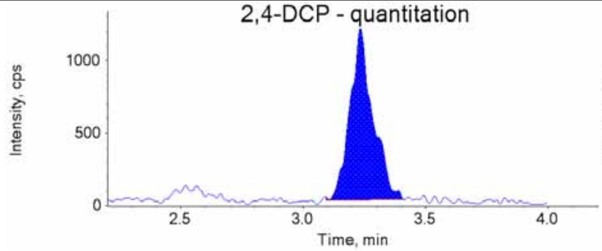


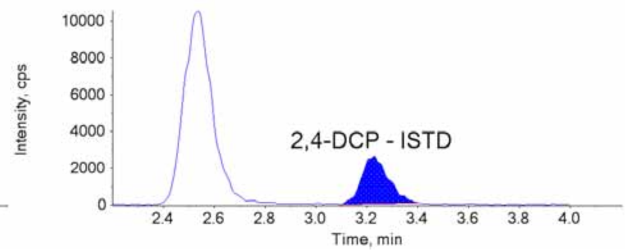


Sample Name: ARA-09-15-10-046-0001A2
Data File: ARA-09-15-10 D21.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 0
Analyte Ion Transition: 220.968/163.000 Da

Acquired: 8/19/2010 10:59:05 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 297466
IS Ion Transition: 225.000/166.800 Da

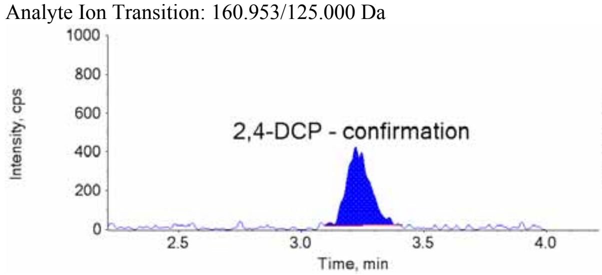
Figure 23. Typical Chromatogram of 2,4-D Analysis, Control Corn Forage (046-0001)

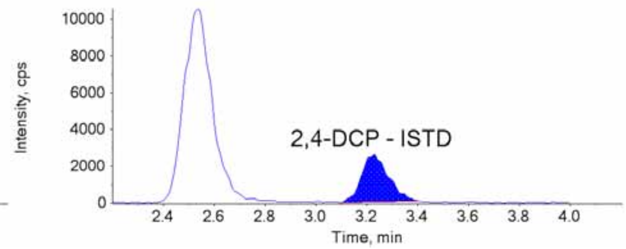




Sample Name: ARA-09-15-10-095-0001A3
Data File: ARA-09-15-10 D22.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 7707
Analyte Ion Transition: 160.953/125.000 Da

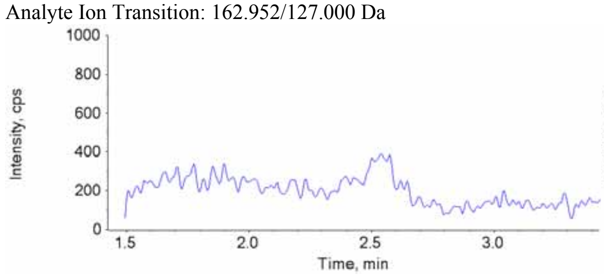
Acquired: 8/20/2010 4:52:44 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 18959
IS Ion Transition: 167.000/131.000 Da

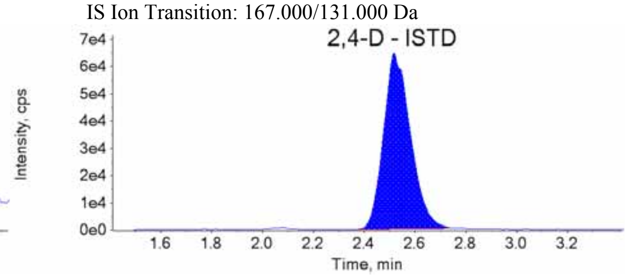




Sample Name: ARA-09-15-10-095-0001A3
Data File: ARA-09-15-10 D22.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 2637
Analyte Ion Transition: 162.952/127.000 Da

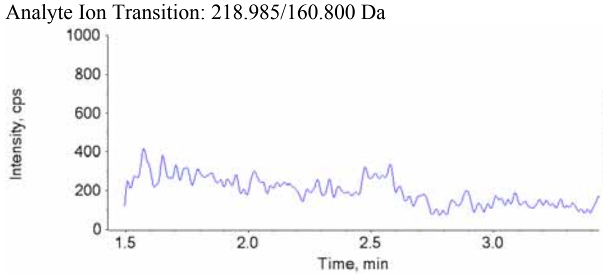
Acquired: 8/20/2010 4:52:44 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 18959
IS Ion Transition: 167.000/131.000 Da

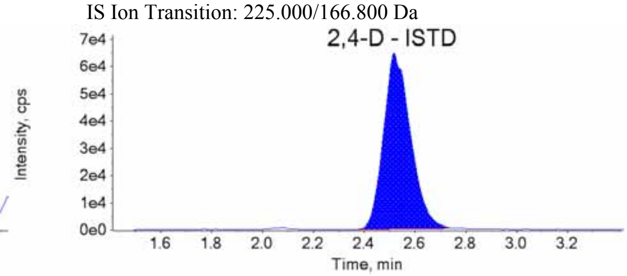




Sample Name: ARA-09-15-10-095-0001A3
Data File: ARA-09-15-10 D22.wiff
Analyte Integration Type: No Peak
Analyte Peak Area: 0
Analyte Ion Transition: 218.985/160.800 Da

Acquired: 8/20/2010 4:52:44 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 476215
IS Ion Transition: 225.000/166.800 Da

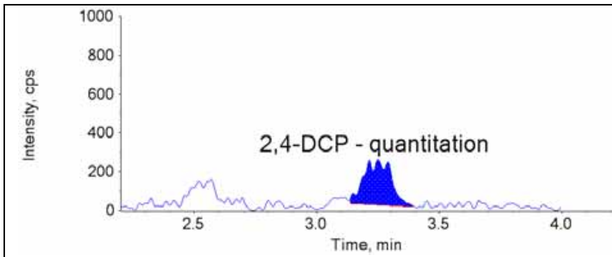


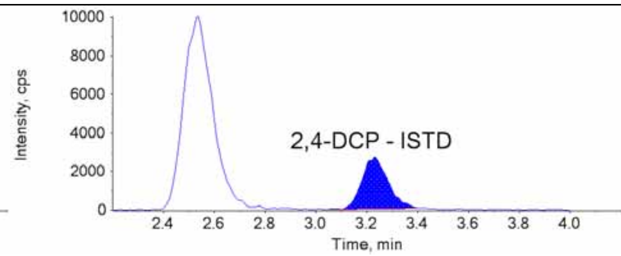


Sample Name: ARA-09-15-10-095-0001A3
Data File: ARA-09-15-10 D22.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 0
Analyte Ion Transition: 220.968/163.000 Da

Acquired: 8/20/2010 4:52:44 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 476215
IS Ion Transition: 225.000/166.800 Da

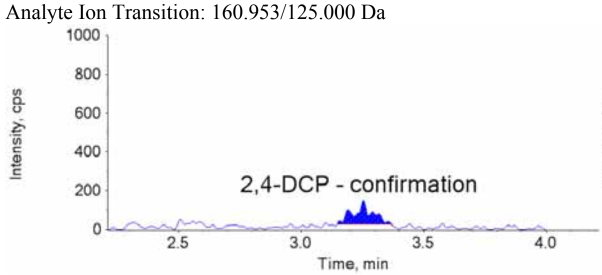
Figure 24. Typical Chromatogram of 2,4-D Analysis, Control Corn Forage (095-0001)

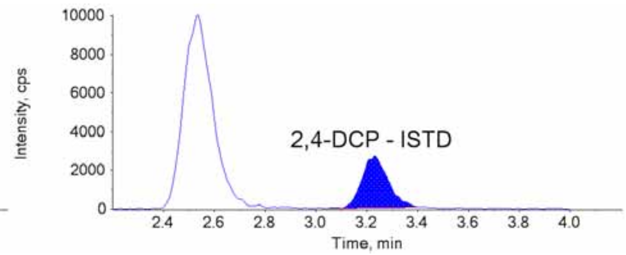




Sample Name: ARA-09-15-10-096-0001A2
Data File: ARA-09-15-10 D22.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 1833
Analyte Ion Transition: 160.953/125.000 Da

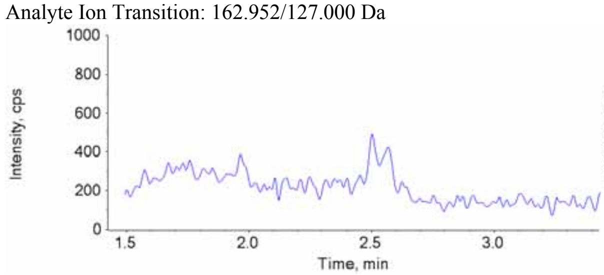
Acquired: 8/20/2010 4:59:12 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 19303
IS Ion Transition: 167.000/131.000 Da

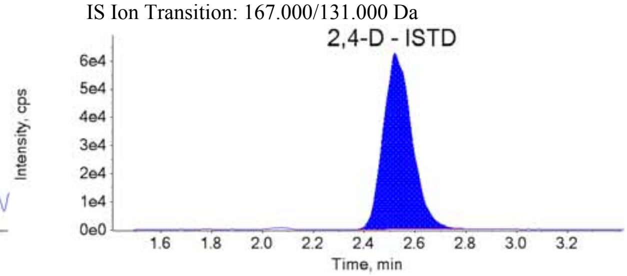




Sample Name: ARA-09-15-10-096-0001A2
Data File: ARA-09-15-10 D22.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 630
Analyte Ion Transition: 162.952/127.000 Da

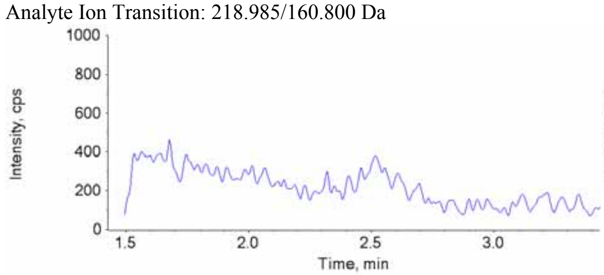
Acquired: 8/20/2010 4:59:12 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 19303
IS Ion Transition: 167.000/131.000 Da

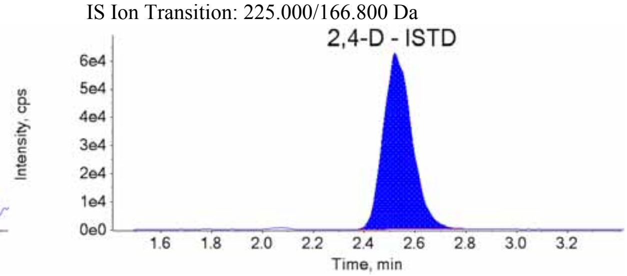




Sample Name: ARA-09-15-10-096-0001A2
Data File: ARA-09-15-10 D22.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 0
Analyte Ion Transition: 218.985/160.800 Da

Acquired: 8/20/2010 4:59:12 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 484253
IS Ion Transition: 225.000/166.800 Da

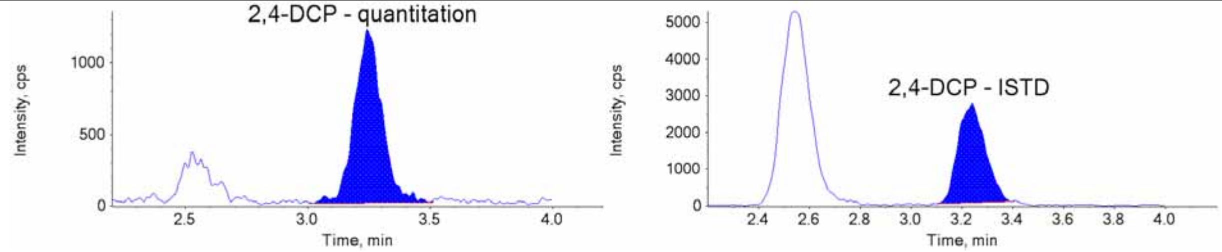




Sample Name: ARA-09-15-10-096-0001A2
Data File: ARA-09-15-10 D22.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 0
Analyte Ion Transition: 220.968/163.000 Da

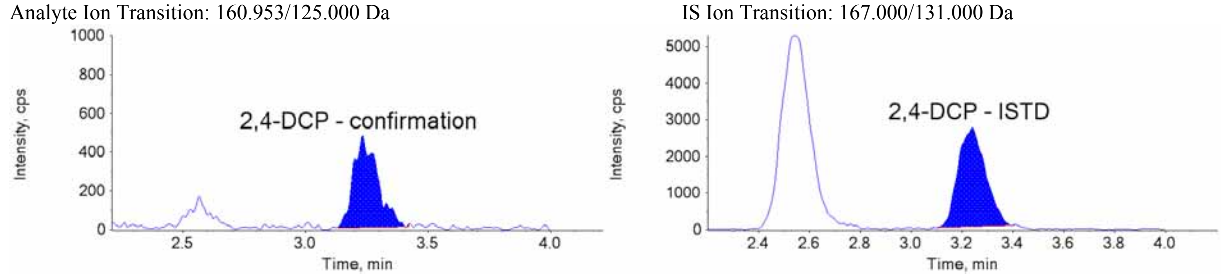
Acquired: 8/20/2010 4:59:12 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 484253
IS Ion Transition: 225.000/166.800 Da

Figure 25. Typical Chromatogram of 2,4-D Analysis, Control Corn Forage (096-0001)



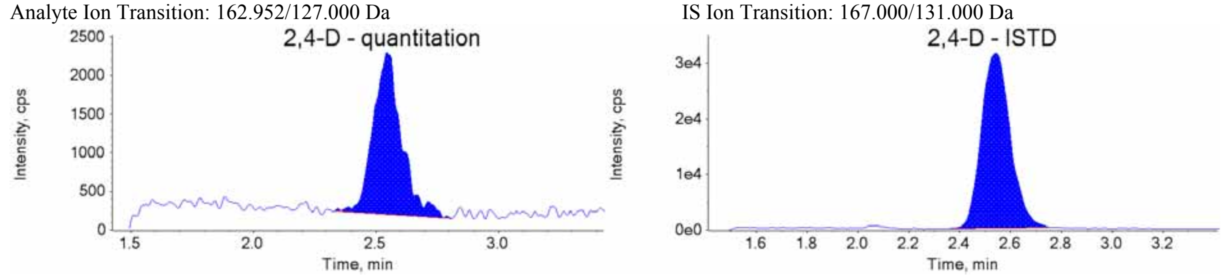
Sample Name: ARA-09-15-10-046-0001A5 + 0.01 µg/g
Data File: ARA-09-15-10 D21.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 9611
Analyte Ion Transition: 160.953/125.000 Da

Acquired: 8/20/2010 1:15:03 AM
Injection Volume (µL): 30
IS Integration Type: Manual
IS Peak Area: 20258
IS Ion Transition: 167.000/131.000 Da



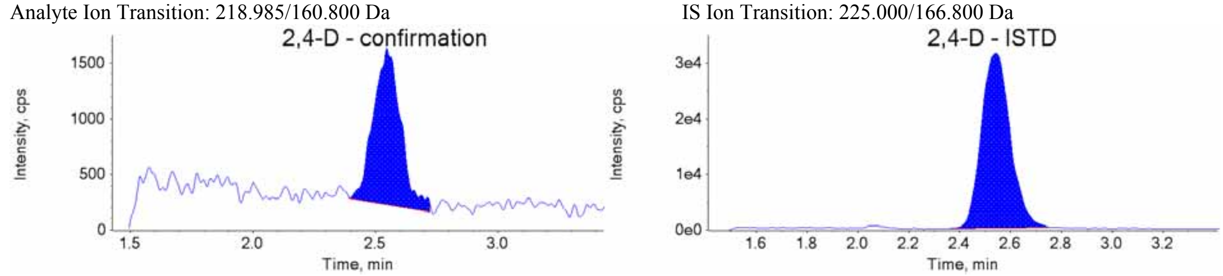
Sample Name: ARA-09-15-10-046-0001A5 + 0.01 µg/g
Data File: ARA-09-15-10 D21.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 3231
Analyte Ion Transition: 162.952/127.000 Da

Acquired: 8/20/2010 1:15:03 AM
Injection Volume (µL): 30
IS Integration Type: Manual
IS Peak Area: 20258
IS Ion Transition: 167.000/131.000 Da



Sample Name: ARA-09-15-10-046-0001A5 + 0.01 µg/g
Data File: ARA-09-15-10 D21.wiff
Analyte Integration Type: Base To Base
Analyte Peak Area: 16956
Analyte Ion Transition: 218.985/160.800 Da

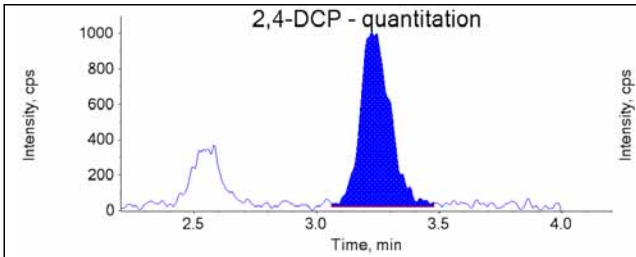
Acquired: 8/20/2010 1:15:03 AM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 249962
IS Ion Transition: 225.000/166.800 Da

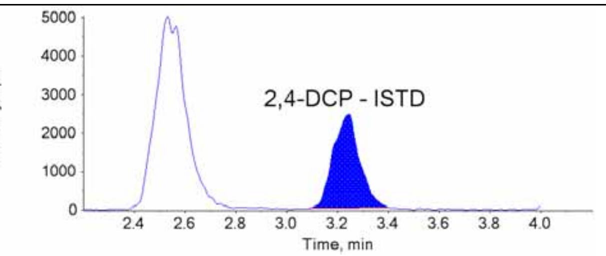


Sample Name: ARA-09-15-10-046-0001A5 + 0.01 µg/g
Data File: ARA-09-15-10 D21.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 10971
Analyte Ion Transition: 220.968/163.000 Da

Acquired: 8/20/2010 1:15:03 AM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 249962
IS Ion Transition: 225.000/166.800 Da

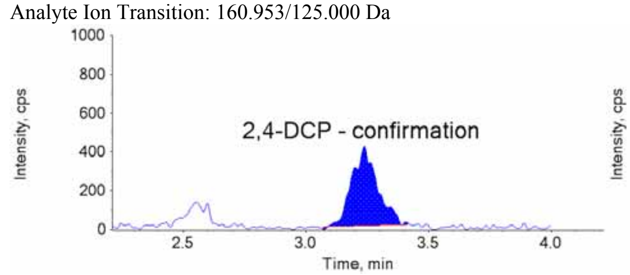
Figure 26. Typical Chromatogram of 2,4-D Analysis, Corn Forage 0.01 µg/g Recovery

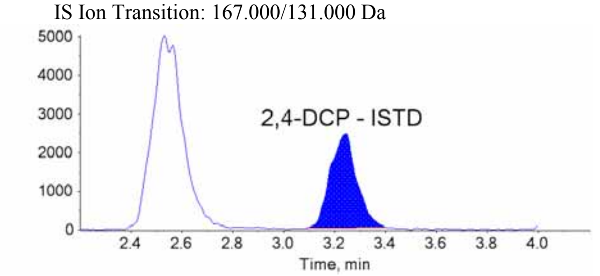




Sample Name: ARA-09-15-10-046-0001A6 + 0.01 µg/g
Data File: ARA-09-15-10 D21.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 8490
Analyte Ion Transition: 160.953/125.000 Da

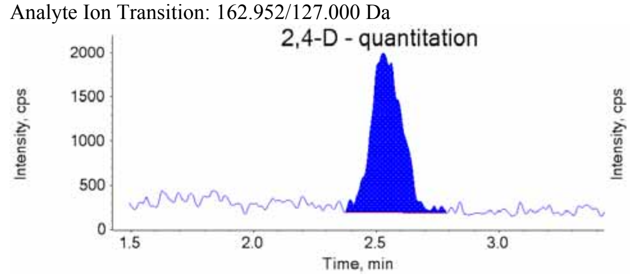
Acquired: 8/20/2010 1:21:32 AM
Injection Volume (µL): 30
IS Integration Type: Manual
IS Peak Area: 17666
IS Ion Transition: 167.000/131.000 Da

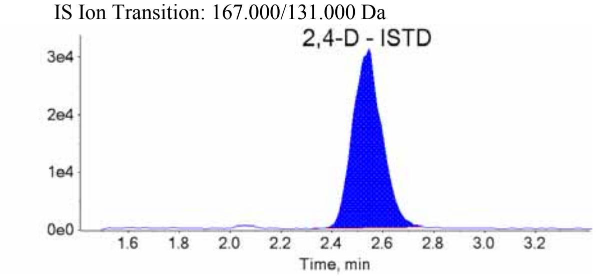




Sample Name: ARA-09-15-10-046-0001A6 + 0.01 µg/g
Data File: ARA-09-15-10 D21.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 2960
Analyte Ion Transition: 162.952/127.000 Da

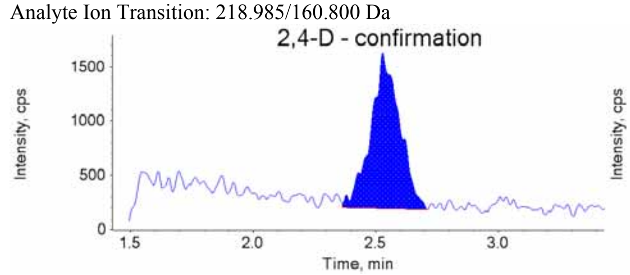
Acquired: 8/20/2010 1:21:32 AM
Injection Volume (µL): 30
IS Integration Type: Manual
IS Peak Area: 17666
IS Ion Transition: 167.000/131.000 Da

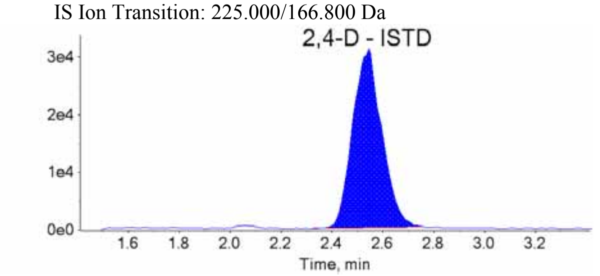




Sample Name: ARA-09-15-10-046-0001A6 + 0.01 µg/g
Data File: ARA-09-15-10 D21.wiff
Analyte Integration Type: Base To Base
Analyte Peak Area: 16108
Analyte Ion Transition: 218.985/160.800 Da

Acquired: 8/20/2010 1:21:32 AM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 246722
IS Ion Transition: 225.000/166.800 Da

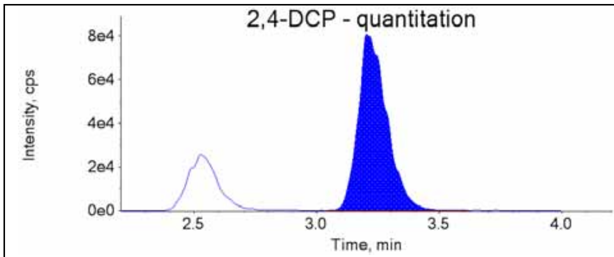


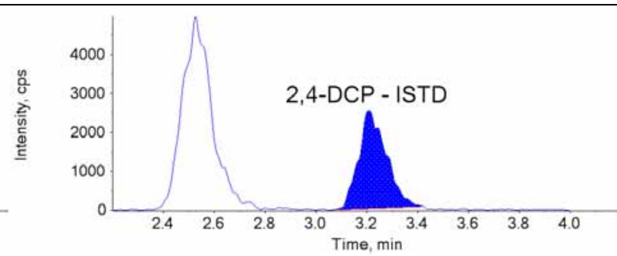


Sample Name: ARA-09-15-10-046-0001A6 + 0.01 µg/g
Data File: ARA-09-15-10 D21.wiff
Analyte Integration Type: Valley
Analyte Peak Area: 11835
Analyte Ion Transition: 220.968/163.000 Da

Acquired: 8/20/2010 1:21:32 AM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 246722
IS Ion Transition: 225.000/166.800 Da

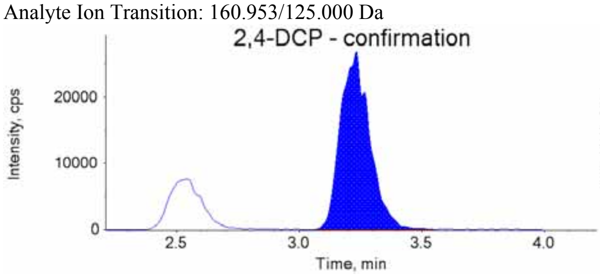
Figure 27. Typical Chromatogram of 2,4-D Analysis, Corn Forage 0.01 µg/g Recovery

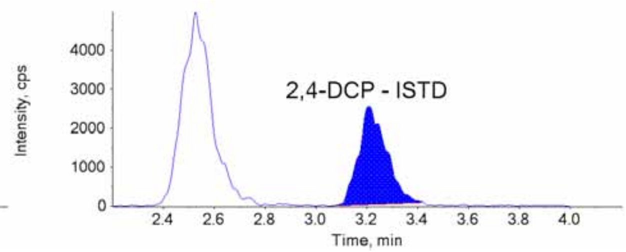




Sample Name: ARA-09-15-10-046-0001A30 + 10.0 µg/g
Data File: ARA-09-15-10 D25.wiff
Analyte Integration Type: Base To Base
Analyte Peak Area: 659880
Analyte Ion Transition: 160.953/125.000 Da

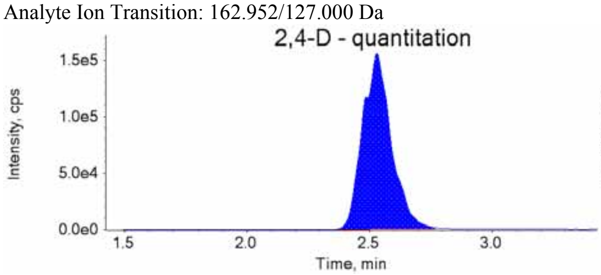
Acquired: 8/25/2010 4:06:57 AM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 19137
IS Ion Transition: 167.000/131.000 Da

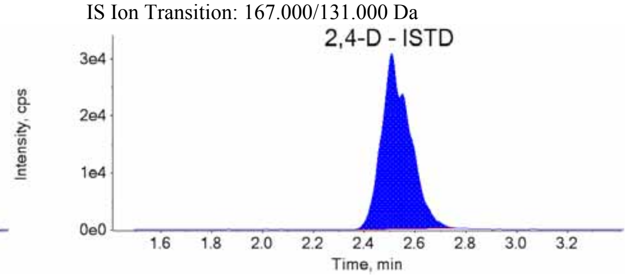




Sample Name: ARA-09-15-10-046-0001A30 + 10.0 µg/g
Data File: ARA-09-15-10 D25.wiff
Analyte Integration Type: Valley
Analyte Peak Area: 222921
Analyte Ion Transition: 162.952/127.000 Da

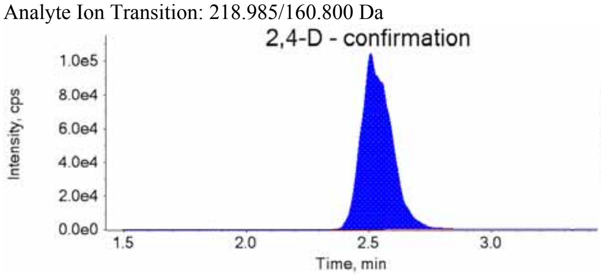
Acquired: 8/25/2010 4:06:57 AM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 19137
IS Ion Transition: 167.000/131.000 Da

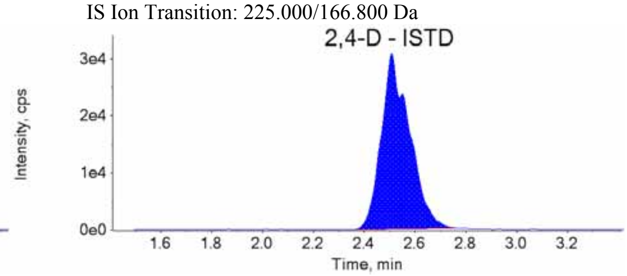




Sample Name: ARA-09-15-10-046-0001A30 + 10.0 µg/g
Data File: ARA-09-15-10 D25.wiff
Analyte Integration Type: Exponential Skim
Analyte Peak Area: 1276262
Analyte Ion Transition: 218.985/160.800 Da

Acquired: 8/25/2010 4:06:57 AM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 237807
IS Ion Transition: 225.000/166.800 Da

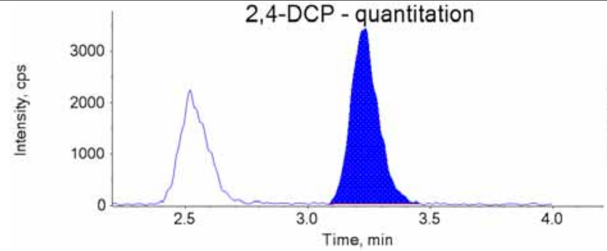


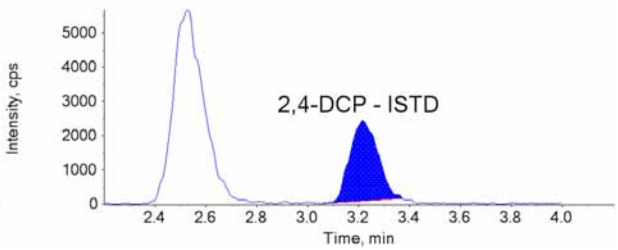


Sample Name: ARA-09-15-10-046-0001A30 + 10.0 µg/g
Data File: ARA-09-15-10 D25.wiff
Analyte Integration Type: Valley
Analyte Peak Area: 884699
Analyte Ion Transition: 220.968/163.000 Da

Acquired: 8/25/2010 4:06:57 AM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 237807
IS Ion Transition: 225.000/166.800 Da

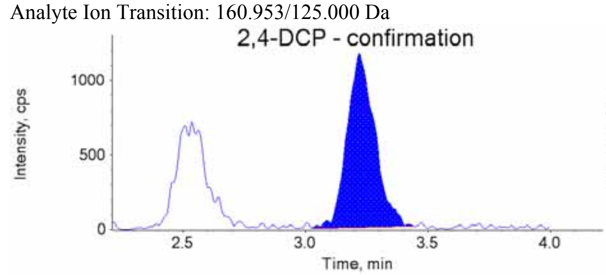
Figure 28. Typical Chromatogram of 2,4-D Analysis, Corn Forage 10.0 µg/g Recovery

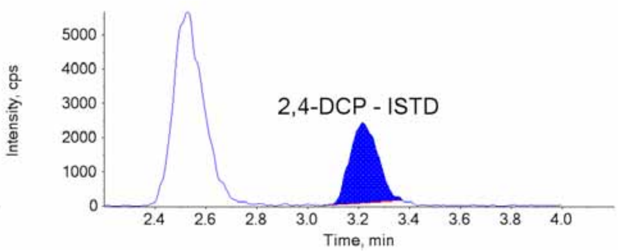




Sample Name: ARA-09-15-10-009-0002A2
Data File: ARA-09-15-10 D24.wiff
Analyte Integration Type: Base To Base
Analyte Peak Area: 26366
Analyte Ion Transition: 160.953/125.000 Da

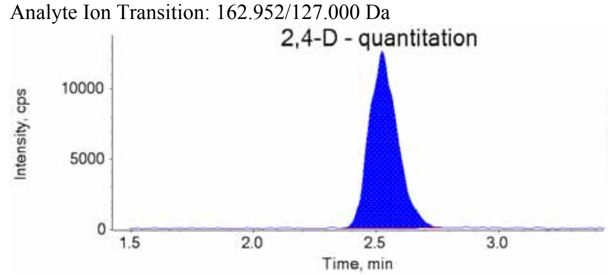
Acquired: 8/24/2010 9:04:17 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 18013
IS Ion Transition: 167.000/131.000 Da

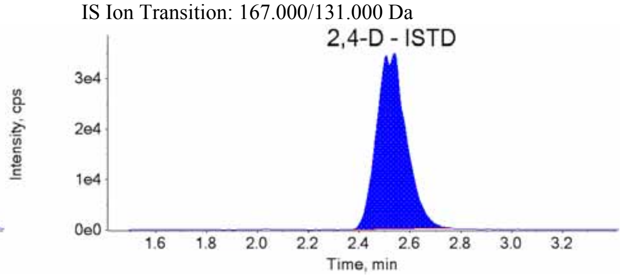




Sample Name: ARA-09-15-10-009-0002A2
Data File: ARA-09-15-10 D24.wiff
Analyte Integration Type: Valley
Analyte Peak Area: 9239
Analyte Ion Transition: 162.952/127.000 Da

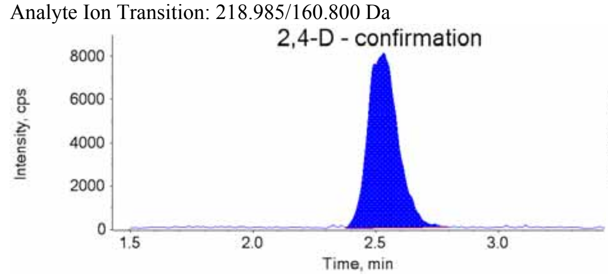
Acquired: 8/24/2010 9:04:17 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 18013
IS Ion Transition: 167.000/131.000 Da

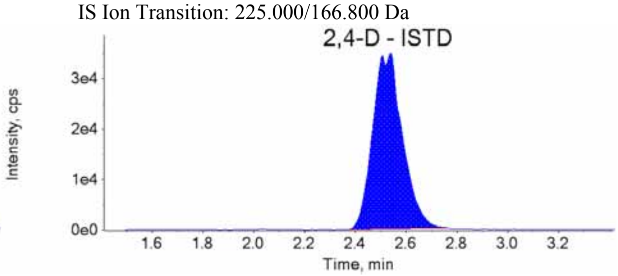




Sample Name: ARA-09-15-10-009-0002A2
Data File: ARA-09-15-10 D24.wiff
Analyte Integration Type: Base To Base
Analyte Peak Area: 99476
Analyte Ion Transition: 218.985/160.800 Da

Acquired: 8/24/2010 9:04:17 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 284533
IS Ion Transition: 225.000/166.800 Da

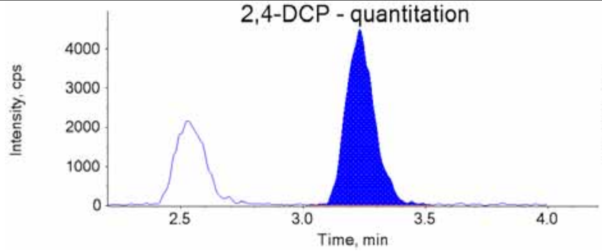


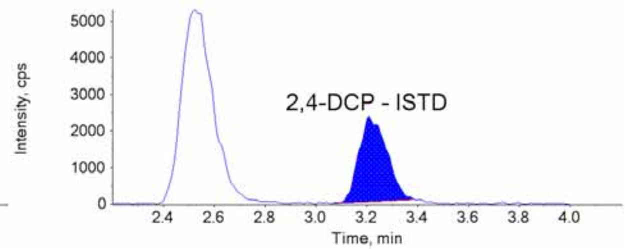


Sample Name: ARA-09-15-10-009-0002A2
Data File: ARA-09-15-10 D24.wiff
Analyte Integration Type: Valley
Analyte Peak Area: 69781
Analyte Ion Transition: 220.968/163.000 Da

Acquired: 8/24/2010 9:04:17 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 284533
IS Ion Transition: 225.000/166.800 Da

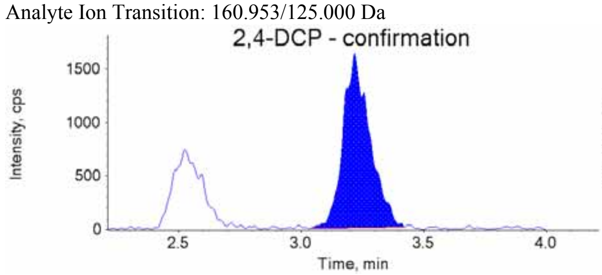
Figure 29. Typical Chromatogram of 2,4-D Analysis, Treated Corn Forage (009-0002)

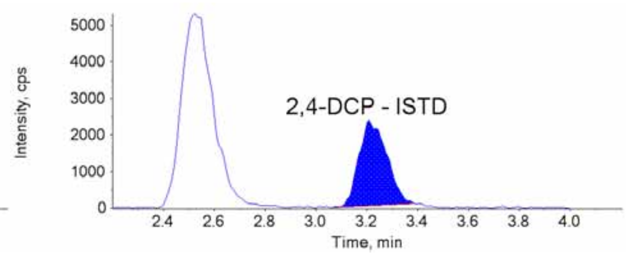




Sample Name: ARA-09-15-10-009-0003A2
Data File: ARA-09-15-10 D24.wiff
Analyte Integration Type: Base To Base
Analyte Peak Area: 36064
Analyte Ion Transition: 160.953/125.000 Da

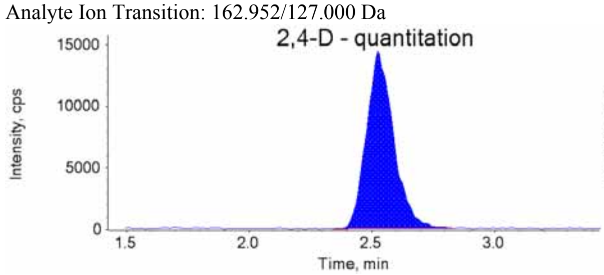
Acquired: 8/24/2010 9:36:34 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 17829
IS Ion Transition: 167.000/131.000 Da

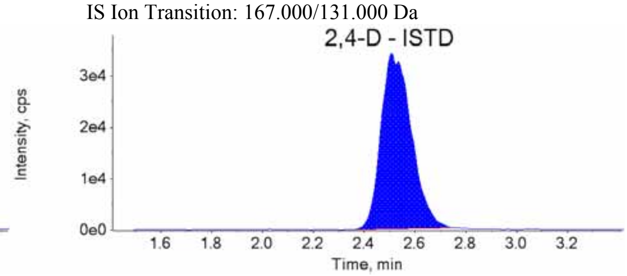




Sample Name: ARA-09-15-10-009-0003A2
Data File: ARA-09-15-10 D24.wiff
Analyte Integration Type: Base To Base
Analyte Peak Area: 12342
Analyte Ion Transition: 162.952/127.000 Da

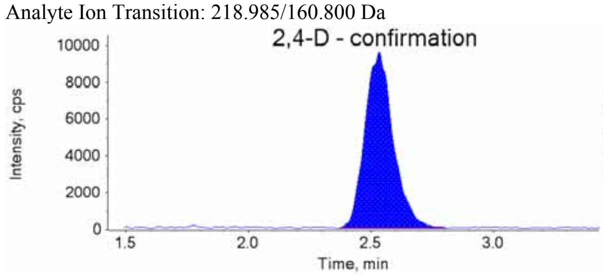
Acquired: 8/24/2010 9:36:34 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 17829
IS Ion Transition: 167.000/131.000 Da

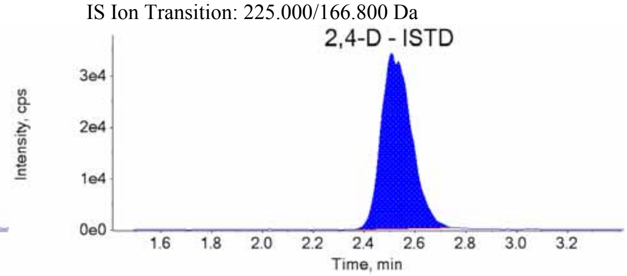




Sample Name: ARA-09-15-10-009-0003A2
Data File: ARA-09-15-10 D24.wiff
Analyte Integration Type: Base To Base
Analyte Peak Area: 110645
Analyte Ion Transition: 218.985/160.800 Da

Acquired: 8/24/2010 9:36:34 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 283920
IS Ion Transition: 225.000/166.800 Da

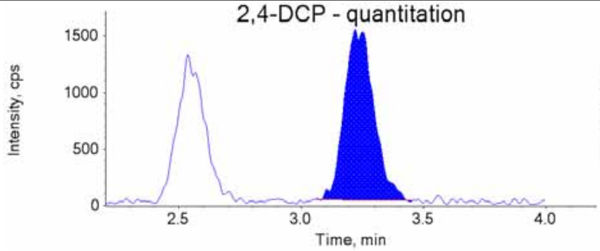


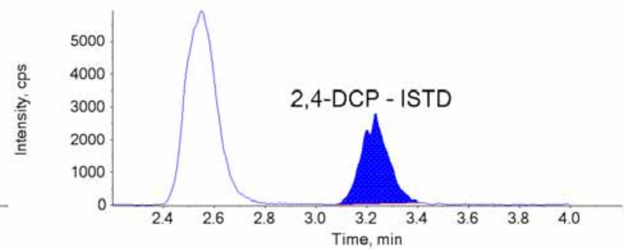


Sample Name: ARA-09-15-10-009-0003A2
Data File: ARA-09-15-10 D24.wiff
Analyte Integration Type: Base To Base
Analyte Peak Area: 77054
Analyte Ion Transition: 220.968/163.000 Da

Acquired: 8/24/2010 9:36:34 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 283920
IS Ion Transition: 225.000/166.800 Da

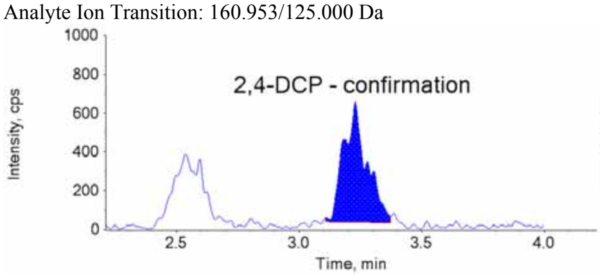
Figure 30. Typical Chromatogram of 2,4-D Analysis, Treated Corn Forage (009-0003)

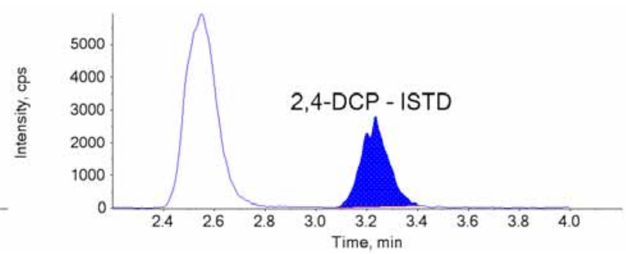




Sample Name: ARA-09-15-10-010-0002A2
Data File: ARA-09-15-10 D23.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 12776
Analyte Ion Transition: 160.953/125.000 Da

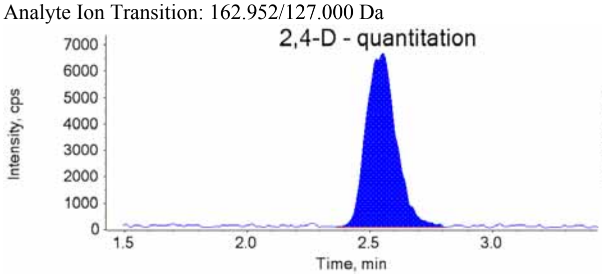
Acquired: 8/23/2010 10:00:53 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 19884
IS Ion Transition: 167.000/131.000 Da

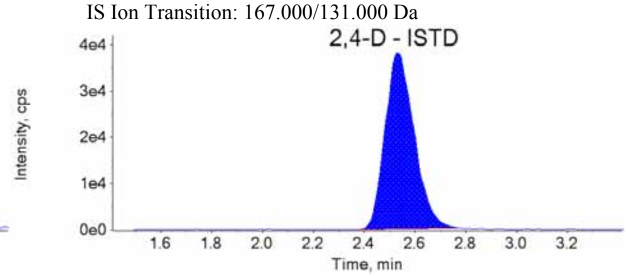




Sample Name: ARA-09-15-10-010-0002A2
Data File: ARA-09-15-10 D23.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 4082
Analyte Ion Transition: 162.952/127.000 Da

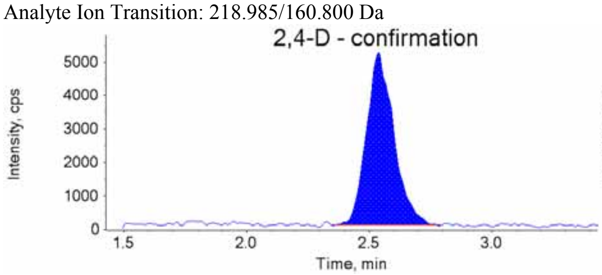
Acquired: 8/23/2010 10:00:53 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 19884
IS Ion Transition: 167.000/131.000 Da

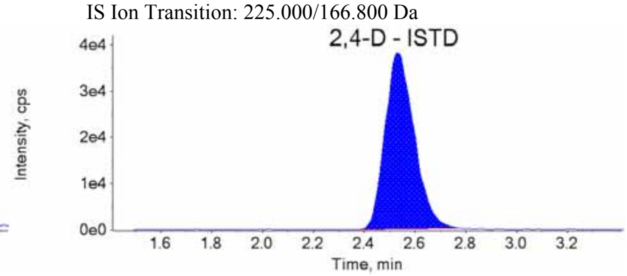




Sample Name: ARA-09-15-10-010-0002A2
Data File: ARA-09-15-10 D23.wiff
Analyte Integration Type: Base To Base
Analyte Peak Area: 56368
Analyte Ion Transition: 218.985/160.800 Da

Acquired: 8/23/2010 10:00:53 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 303327
IS Ion Transition: 225.000/166.800 Da

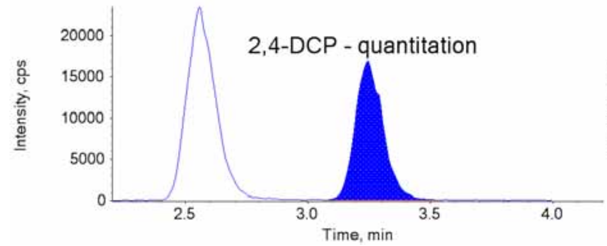


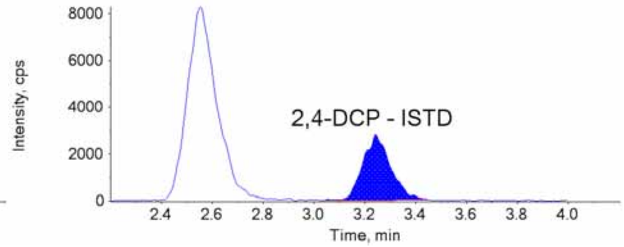


Sample Name: ARA-09-15-10-010-0002A2
Data File: ARA-09-15-10 D23.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 40096
Analyte Ion Transition: 220.968/163.000 Da

Acquired: 8/23/2010 10:00:53 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 303327
IS Ion Transition: 225.000/166.800 Da

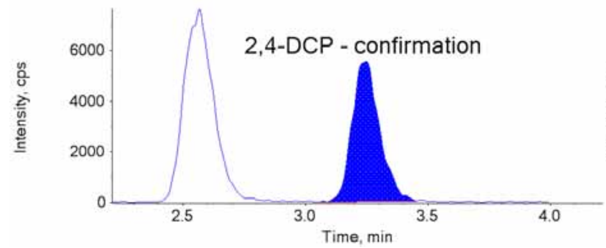
Figure 31. Typical Chromatogram of 2,4-D Analysis, Treated Corn Forage (010-0002)

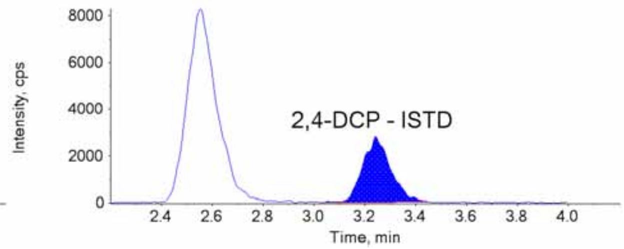




Sample Name: ARA-09-15-10-010-0003A2
Data File: ARA-09-15-10 D24.wiff
Analyte Integration Type: Base To Base
Analyte Peak Area: 132515
Analyte Ion Transition: 160.953/125.000 Da

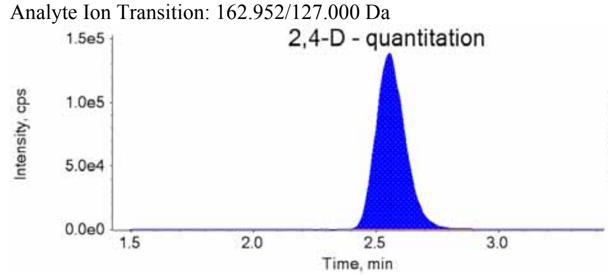
Acquired: 8/24/2010 6:48:20 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 21134
IS Ion Transition: 167.000/131.000 Da

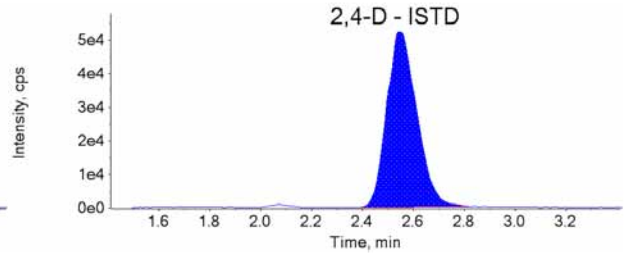




Sample Name: ARA-09-15-10-010-0003A2
Data File: ARA-09-15-10 D24.wiff
Analyte Integration Type: Base To Base
Analyte Peak Area: 43568
Analyte Ion Transition: 162.952/127.000 Da

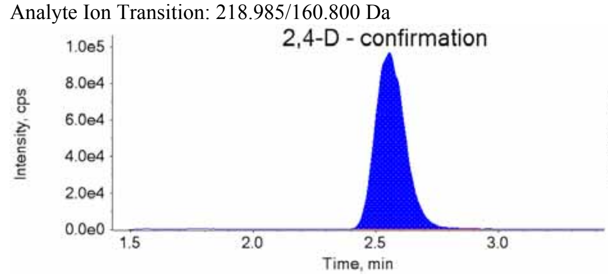
Acquired: 8/24/2010 6:48:20 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 21134
IS Ion Transition: 167.000/131.000 Da

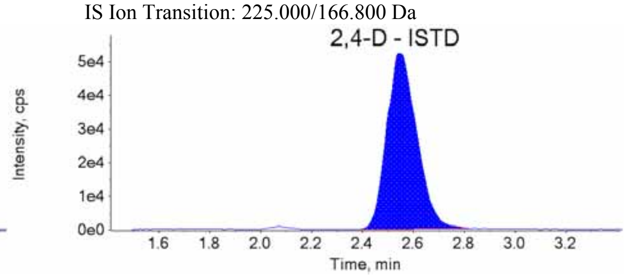




Sample Name: ARA-09-15-10-010-0003A2
Data File: ARA-09-15-10 D24.wiff
Analyte Integration Type: Base To Base
Analyte Peak Area: 1152787
Analyte Ion Transition: 218.985/160.800 Da

Acquired: 8/24/2010 6:48:20 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 431732
IS Ion Transition: 225.000/166.800 Da

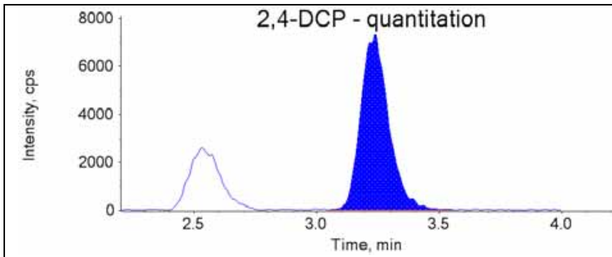


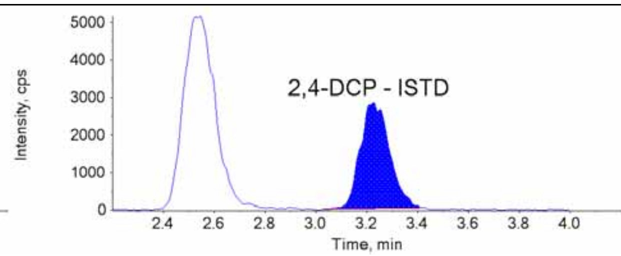


Sample Name: ARA-09-15-10-010-0003A2
Data File: ARA-09-15-10 D24.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 826888
Analyte Ion Transition: 220.968/163.000 Da

Acquired: 8/24/2010 6:48:20 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 431732
IS Ion Transition: 225.000/166.800 Da

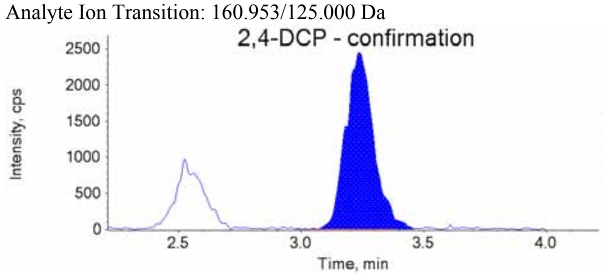
Figure 32. Typical Chromatogram of 2,4-D Analysis, Treated Corn Forage (010-0003)

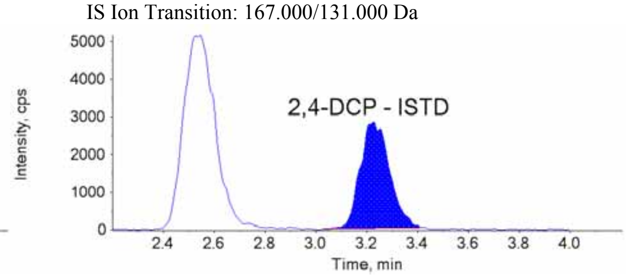




Sample Name: ARA-09-15-10-014-0002A2
Data File: ARA-09-15-10 D24.wiff
Analyte Integration Type: Base To Base
Analyte Peak Area: 57819
Analyte Ion Transition: 160.953/125.000 Da

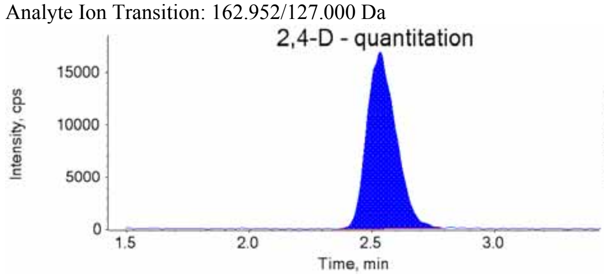
Acquired: 8/24/2010 10:21:48 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 22904
IS Ion Transition: 167.000/131.000 Da

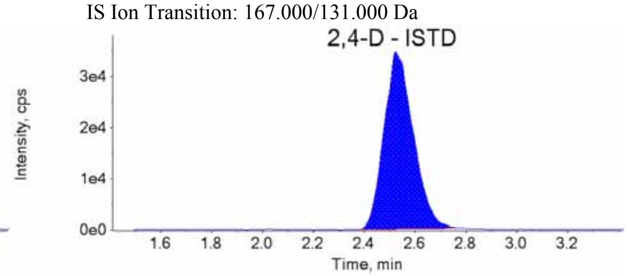




Sample Name: ARA-09-15-10-014-0002A2
Data File: ARA-09-15-10 D24.wiff
Analyte Integration Type: Exponential Skim
Analyte Peak Area: 19657
Analyte Ion Transition: 162.952/127.000 Da

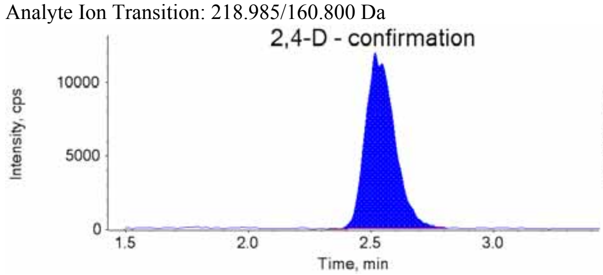
Acquired: 8/24/2010 10:21:48 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 22904
IS Ion Transition: 167.000/131.000 Da

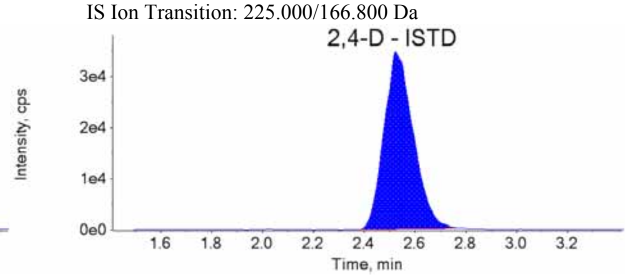




Sample Name: ARA-09-15-10-014-0002A2
Data File: ARA-09-15-10 D24.wiff
Analyte Integration Type: Base To Base
Analyte Peak Area: 138860
Analyte Ion Transition: 218.985/160.800 Da

Acquired: 8/24/2010 10:21:48 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 277417
IS Ion Transition: 225.000/166.800 Da

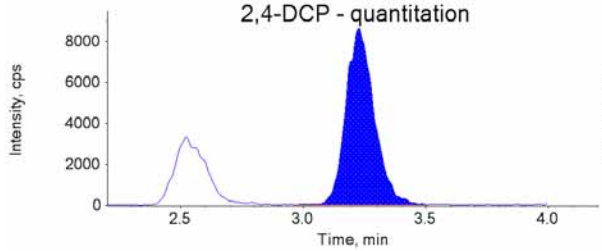


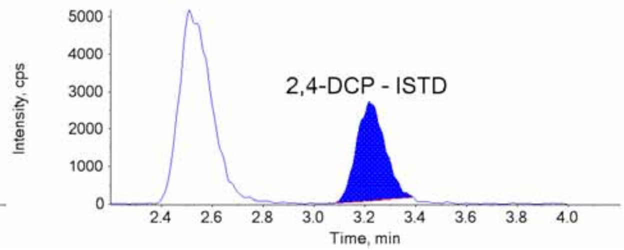


Sample Name: ARA-09-15-10-014-0002A2
Data File: ARA-09-15-10 D24.wiff
Analyte Integration Type: Base To Base
Analyte Peak Area: 98597
Analyte Ion Transition: 220.968/163.000 Da

Acquired: 8/24/2010 10:21:48 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 277417
IS Ion Transition: 225.000/166.800 Da

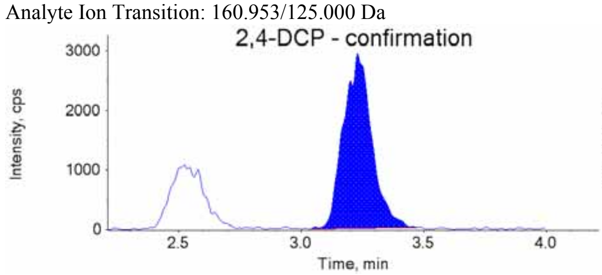
Figure 33. Typical Chromatogram of 2,4-D Analysis, Treated Corn Forage (014-0002)

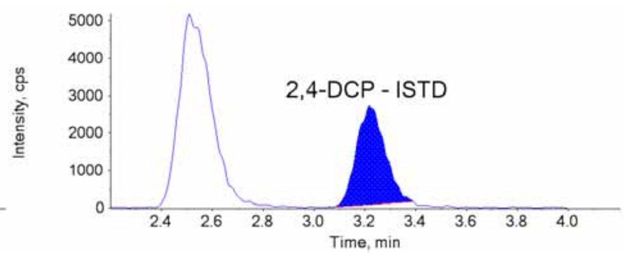




Sample Name: ARA-09-15-10-014-0003A2
Data File: ARA-09-15-10 D24.wiff
Analyte Integration Type: Base To Base
Analyte Peak Area: 69024
Analyte Ion Transition: 160.953/125.000 Da

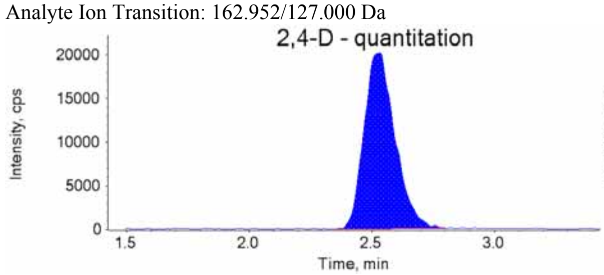
Acquired: 8/24/2010 10:28:16 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 20514
IS Ion Transition: 167.000/131.000 Da

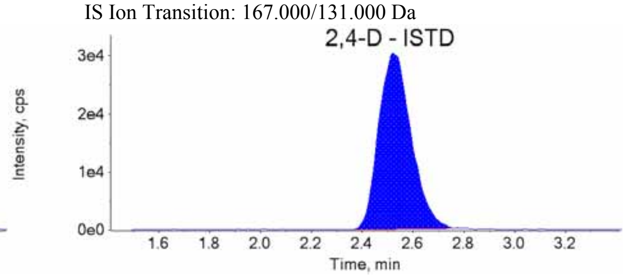




Sample Name: ARA-09-15-10-014-0003A2
Data File: ARA-09-15-10 D24.wiff
Analyte Integration Type: Base To Base
Analyte Peak Area: 23658
Analyte Ion Transition: 162.952/127.000 Da

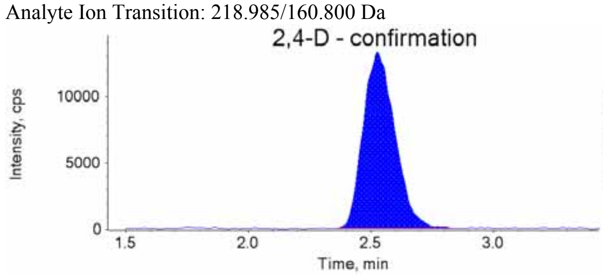
Acquired: 8/24/2010 10:28:16 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 20514
IS Ion Transition: 167.000/131.000 Da

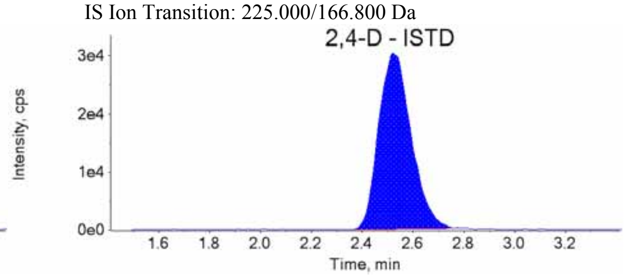




Sample Name: ARA-09-15-10-014-0003A2
Data File: ARA-09-15-10 D24.wiff
Analyte Integration Type: Base To Base
Analyte Peak Area: 176845
Analyte Ion Transition: 218.985/160.800 Da

Acquired: 8/24/2010 10:28:16 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 269780
IS Ion Transition: 225.000/166.800 Da

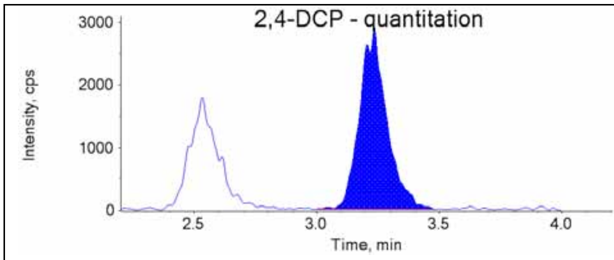


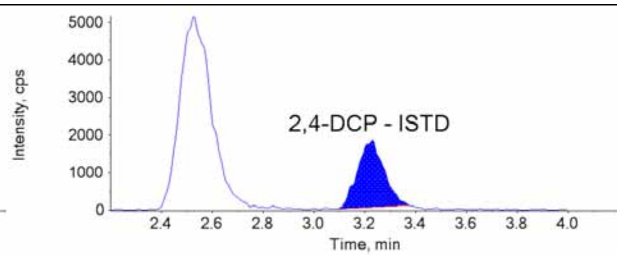


Sample Name: ARA-09-15-10-014-0003A2
Data File: ARA-09-15-10 D24.wiff
Analyte Integration Type: Base To Base
Analyte Peak Area: 119433
Analyte Ion Transition: 220.968/163.000 Da

Acquired: 8/24/2010 10:28:16 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 269780
IS Ion Transition: 225.000/166.800 Da

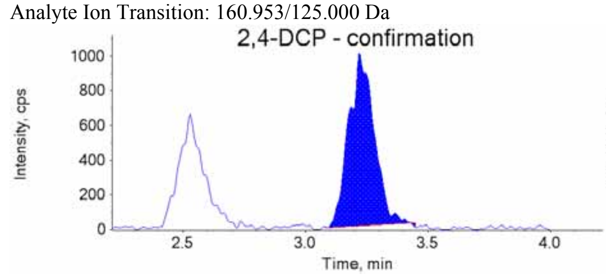
Figure 34. Typical Chromatogram of 2,4-D Analysis, Treated Corn Forage (014-0003)

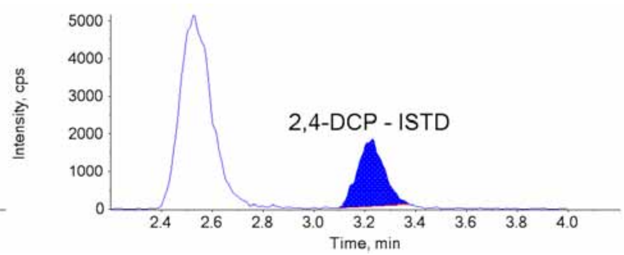




Sample Name: ARA-09-15-10-015-0002A2
Data File: ARA-09-15-10 D24.wiff
Analyte Integration Type: Base To Base
Analyte Peak Area: 22384
Analyte Ion Transition: 160.953/125.000 Da

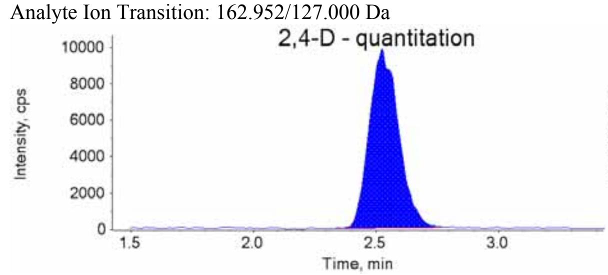
Acquired: 8/24/2010 9:10:44 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 12901
IS Ion Transition: 167.000/131.000 Da

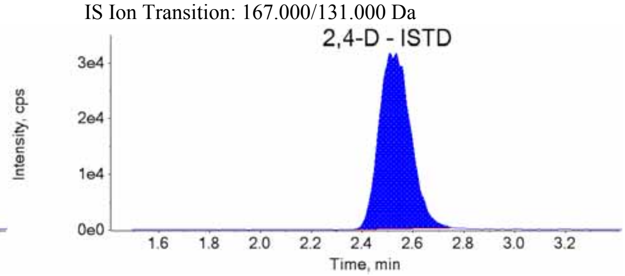




Sample Name: ARA-09-15-10-015-0002A2
Data File: ARA-09-15-10 D24.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 7224
Analyte Ion Transition: 162.952/127.000 Da

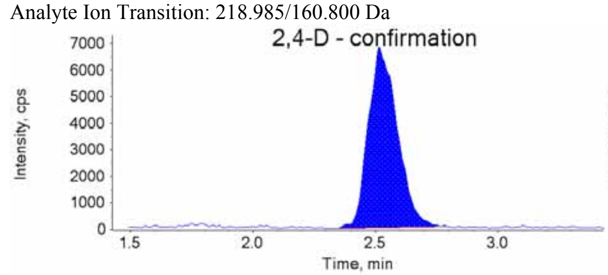
Acquired: 8/24/2010 9:10:44 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 12901
IS Ion Transition: 167.000/131.000 Da

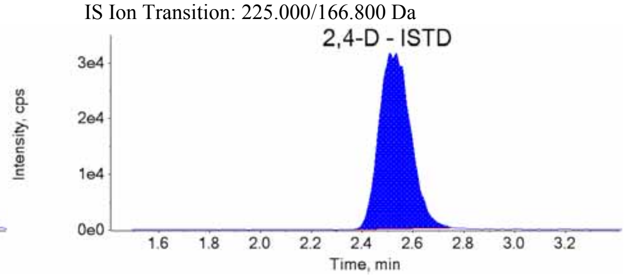




Sample Name: ARA-09-15-10-015-0002A2
Data File: ARA-09-15-10 D24.wiff
Analyte Integration Type: Base To Base
Analyte Peak Area: 81714
Analyte Ion Transition: 218.985/160.800 Da

Acquired: 8/24/2010 9:10:44 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 275024
IS Ion Transition: 225.000/166.800 Da

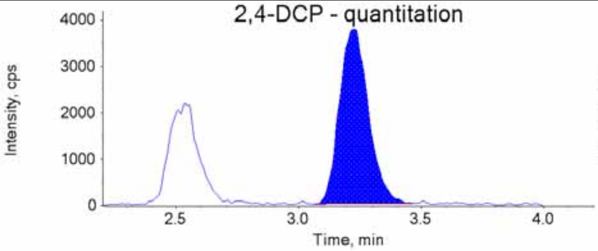


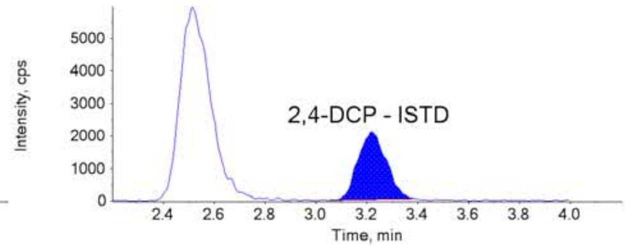


Sample Name: ARA-09-15-10-015-0002A2
Data File: ARA-09-15-10 D24.wiff
Analyte Integration Type: Base To Base
Analyte Peak Area: 56188
Analyte Ion Transition: 220.968/163.000 Da

Acquired: 8/24/2010 9:10:44 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 275024
IS Ion Transition: 225.000/166.800 Da

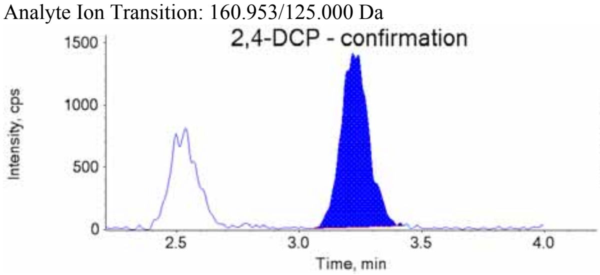
Figure 35. Typical Chromatogram of 2,4-D Analysis, Treated Corn Forage (015-0002)

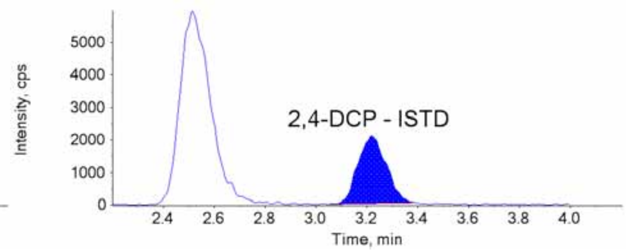




Sample Name: ARA-09-15-10-015-0003A2
Data File: ARA-09-15-10 D24.wiff
Analyte Integration Type: Base To Base
Analyte Peak Area: 31076
Analyte Ion Transition: 160.953/125.000 Da

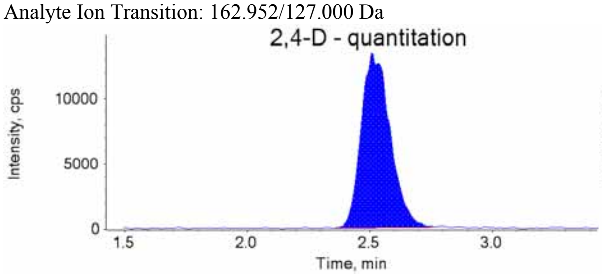
Acquired: 8/24/2010 8:44:53 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 16155
IS Ion Transition: 167.000/131.000 Da

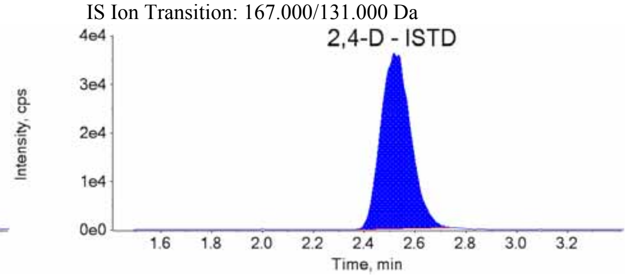




Sample Name: ARA-09-15-10-015-0003A2
Data File: ARA-09-15-10 D24.wiff
Analyte Integration Type: Base To Base
Analyte Peak Area: 11124
Analyte Ion Transition: 162.952/127.000 Da

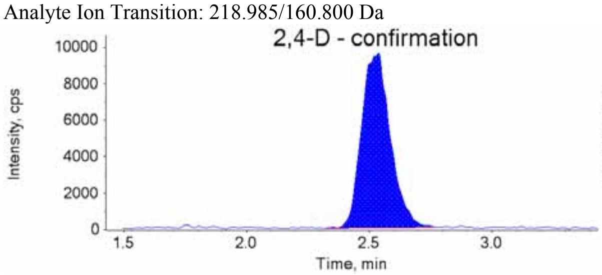
Acquired: 8/24/2010 8:44:53 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 16155
IS Ion Transition: 167.000/131.000 Da

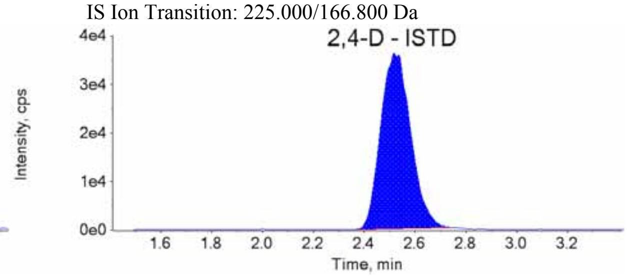




Sample Name: ARA-09-15-10-015-0003A2
Data File: ARA-09-15-10 D24.wiff
Analyte Integration Type: Base To Base
Analyte Peak Area: 110093
Analyte Ion Transition: 218.985/160.800 Da

Acquired: 8/24/2010 8:44:53 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 296680
IS Ion Transition: 225.000/166.800 Da

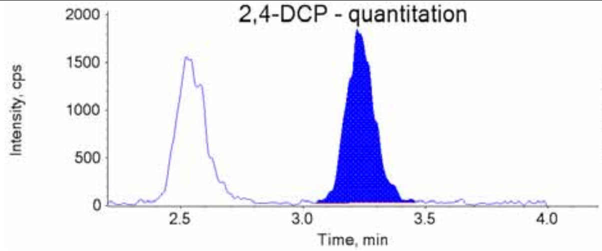


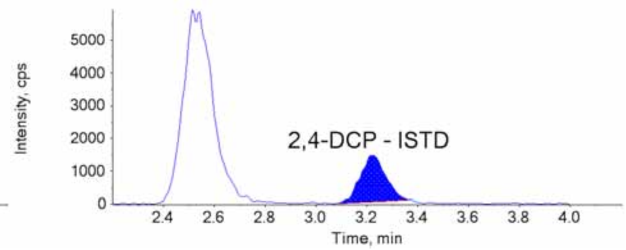


Sample Name: ARA-09-15-10-015-0003A2
Data File: ARA-09-15-10 D24.wiff
Analyte Integration Type: Base To Base
Analyte Peak Area: 78287
Analyte Ion Transition: 220.968/163.000 Da

Acquired: 8/24/2010 8:44:53 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 296680
IS Ion Transition: 225.000/166.800 Da

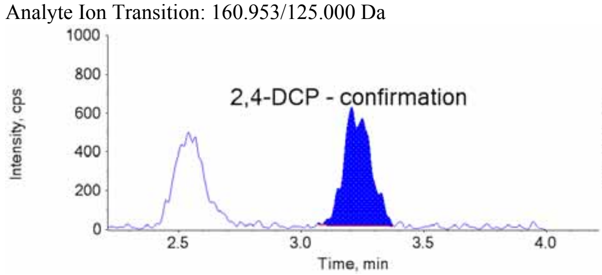
Figure 36. Typical Chromatogram of 2,4-D Analysis, Treated Corn Forage (015-0003)

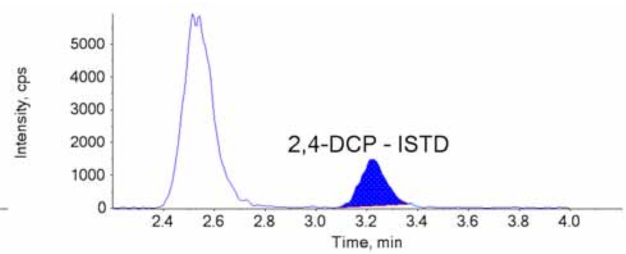




Sample Name: ARA-09-15-10-016-0002A2
Data File: ARA-09-15-10 D24.wiff
Analyte Integration Type: Base To Base
Analyte Peak Area: 14454
Analyte Ion Transition: 160.953/125.000 Da

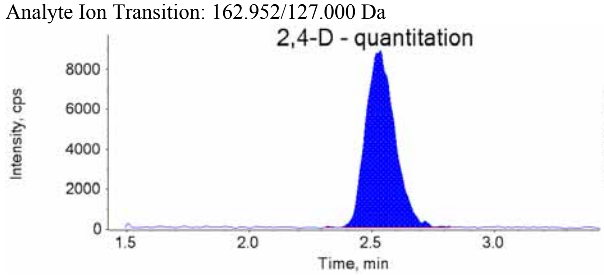
Acquired: 8/24/2010 8:57:49 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 9891
IS Ion Transition: 167.000/131.000 Da

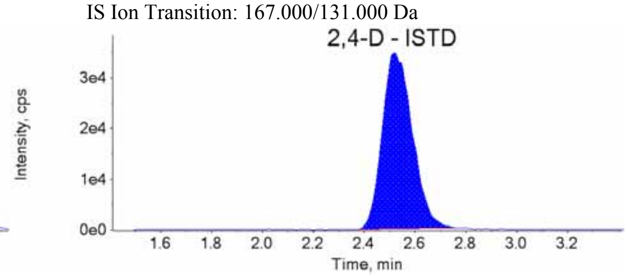




Sample Name: ARA-09-15-10-016-0002A2
Data File: ARA-09-15-10 D24.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 4661
Analyte Ion Transition: 162.952/127.000 Da

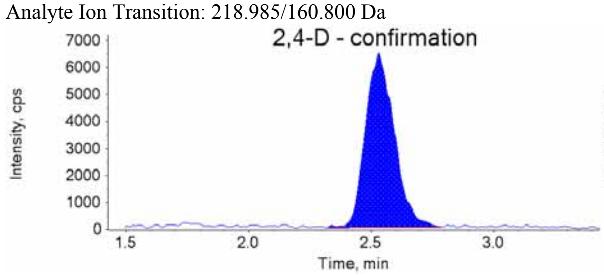
Acquired: 8/24/2010 8:57:49 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 9891
IS Ion Transition: 167.000/131.000 Da

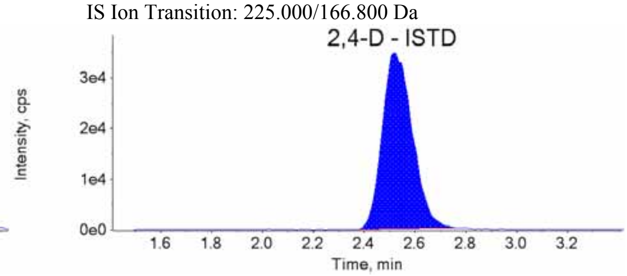




Sample Name: ARA-09-15-10-016-0002A2
Data File: ARA-09-15-10 D24.wiff
Analyte Integration Type: Base To Base
Analyte Peak Area: 73313
Analyte Ion Transition: 218.985/160.800 Da

Acquired: 8/24/2010 8:57:49 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 286310
IS Ion Transition: 225.000/166.800 Da

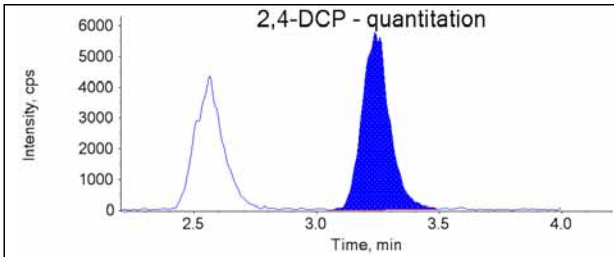


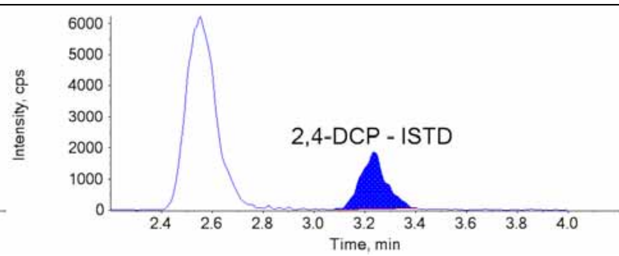


Sample Name: ARA-09-15-10-016-0002A2
Data File: ARA-09-15-10 D24.wiff
Analyte Integration Type: Base To Base
Analyte Peak Area: 53021
Analyte Ion Transition: 220.968/163.000 Da

Acquired: 8/24/2010 8:57:49 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 286310
IS Ion Transition: 225.000/166.800 Da

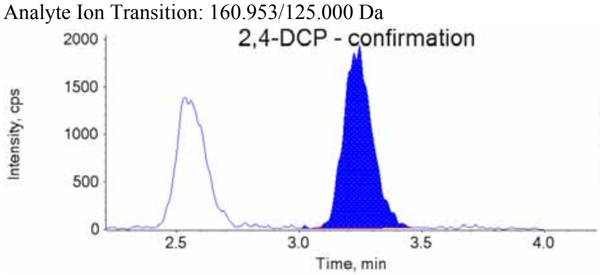
Figure 37. Typical Chromatogram of 2,4-D Analysis, Treated Corn Forage (016-0002)

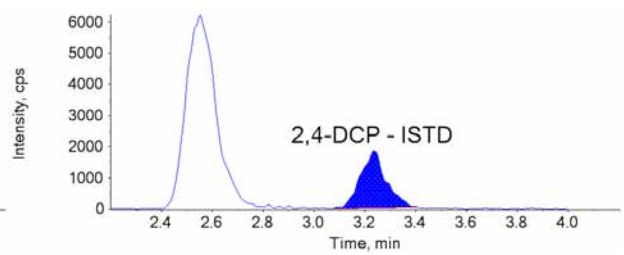




Sample Name: ARA-09-15-10-016-0003A2
Data File: ARA-09-15-10 D23.wiff
Analyte Integration Type: Base To Base
Analyte Peak Area: 44934
Analyte Ion Transition: 160.953/125.000 Da

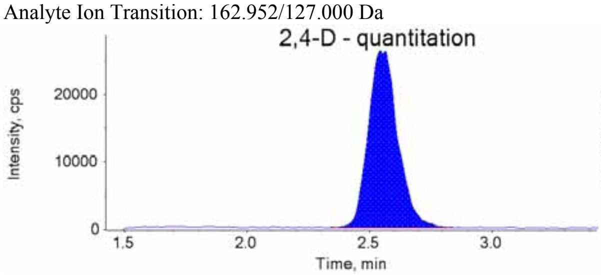
Acquired: 8/23/2010 9:09:04 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 12786
IS Ion Transition: 167.000/131.000 Da

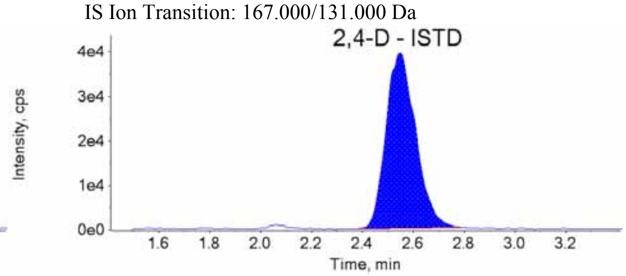




Sample Name: ARA-09-15-10-016-0003A2
Data File: ARA-09-15-10 D23.wiff
Analyte Integration Type: Base To Base
Analyte Peak Area: 15473
Analyte Ion Transition: 162.952/127.000 Da

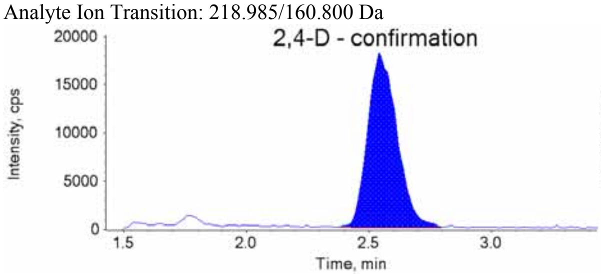
Acquired: 8/23/2010 9:09:04 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 12786
IS Ion Transition: 167.000/131.000 Da

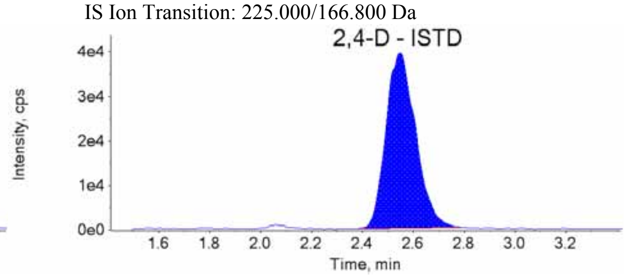




Sample Name: ARA-09-15-10-016-0003A2
Data File: ARA-09-15-10 D23.wiff
Analyte Integration Type: Base To Base
Analyte Peak Area: 220047
Analyte Ion Transition: 218.985/160.800 Da

Acquired: 8/23/2010 9:09:04 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 320679
IS Ion Transition: 225.000/166.800 Da

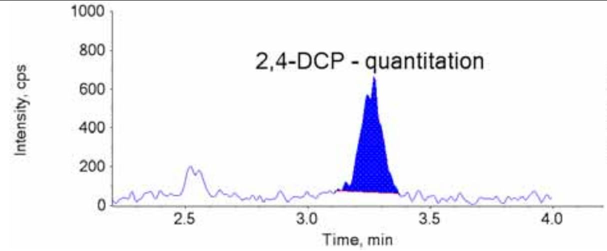


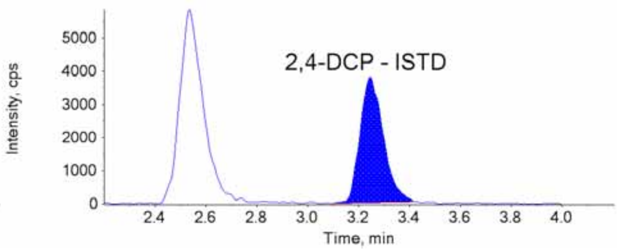


Sample Name: ARA-09-15-10-016-0003A2
Data File: ARA-09-15-10 D23.wiff
Analyte Integration Type: Exponential Skim
Analyte Peak Area: 150332
Analyte Ion Transition: 220.968/163.000 Da

Acquired: 8/23/2010 9:09:04 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 320679
IS Ion Transition: 225.000/166.800 Da

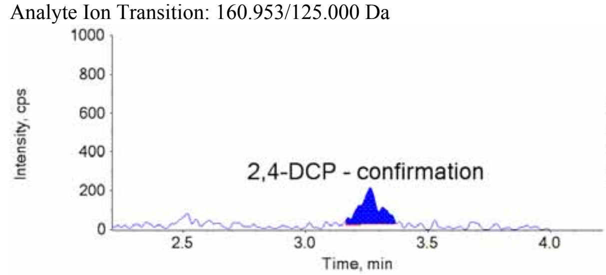
Figure 38. Typical Chromatogram of 2,4-D Analysis, Treated Corn Forage (016-0003)

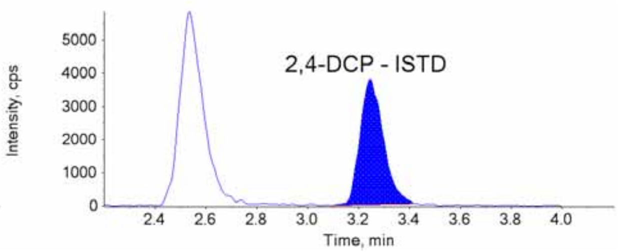




Sample Name: ARA-09-15-10-021-0001A10
Data File: ARA-09-15-10 D26.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 3323
Analyte Ion Transition: 160.953/125.000 Da

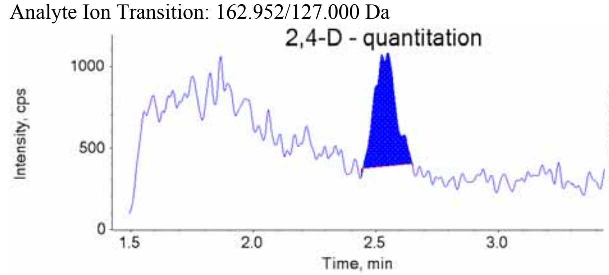
Acquired: 8/27/2010 10:47:50 AM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 24075
IS Ion Transition: 167.000/131.000 Da

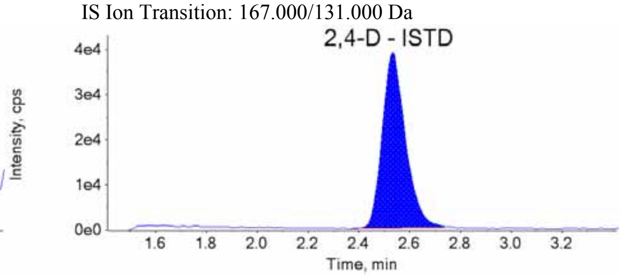




Sample Name: ARA-09-15-10-021-0001A10
Data File: ARA-09-15-10 D26.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 1075
Analyte Ion Transition: 162.952/127.000 Da

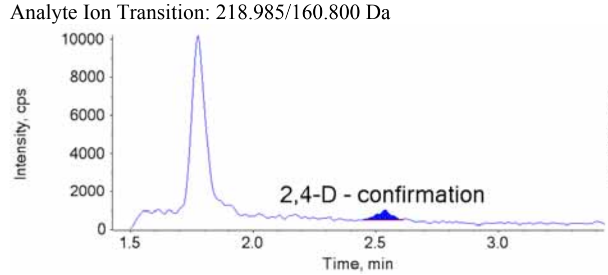
Acquired: 8/27/2010 10:47:50 AM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 24075
IS Ion Transition: 167.000/131.000 Da

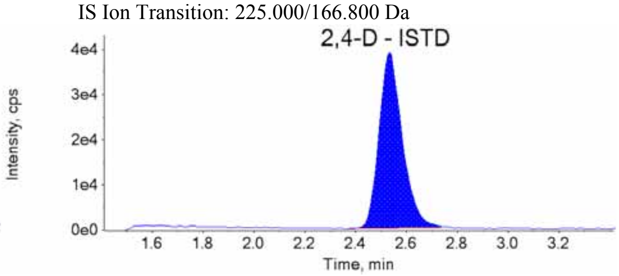




Sample Name: ARA-09-15-10-021-0001A10
Data File: ARA-09-15-10 D26.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 4259
Analyte Ion Transition: 218.985/160.800 Da

Acquired: 8/27/2010 10:47:50 AM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 245162
IS Ion Transition: 225.000/166.800 Da

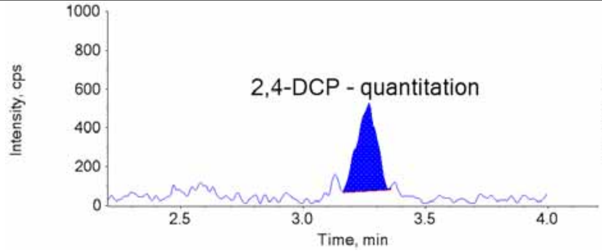


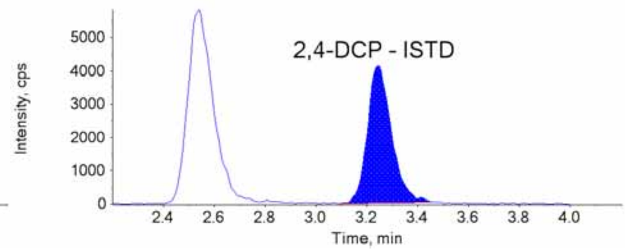


Sample Name: ARA-09-15-10-021-0001A10
Data File: ARA-09-15-10 D26.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 2362
Analyte Ion Transition: 220.968/163.000 Da

Acquired: 8/27/2010 10:47:50 AM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 245162
IS Ion Transition: 225.000/166.800 Da

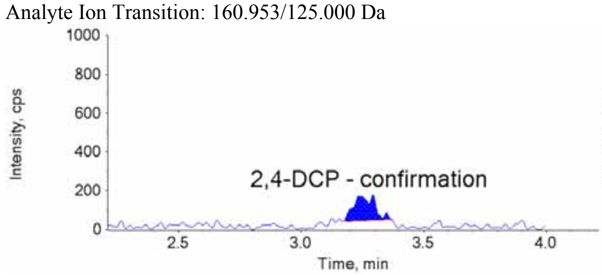
Figure 39. Typical Chromatogram of 2,4-D Analysis, Control Corn Stover (021-0001)

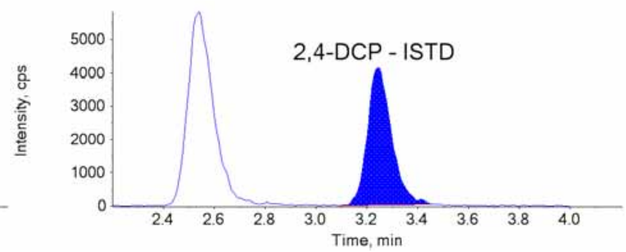




Sample Name: ARA-09-15-10-008-0001A2
Data File: ARA-09-15-10 D26.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 2516
Analyte Ion Transition: 160.953/125.000 Da

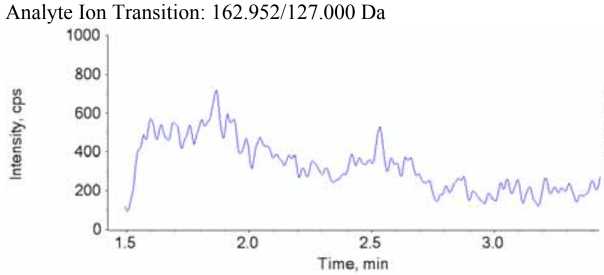
Acquired: 8/27/2010 10:34:53 AM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 27231
IS Ion Transition: 167.000/131.000 Da

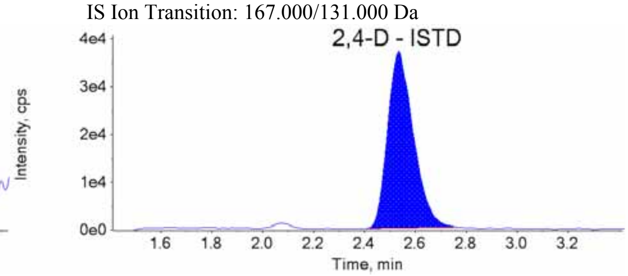




Sample Name: ARA-09-15-10-008-0001A2
Data File: ARA-09-15-10 D26.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 793
Analyte Ion Transition: 162.952/127.000 Da

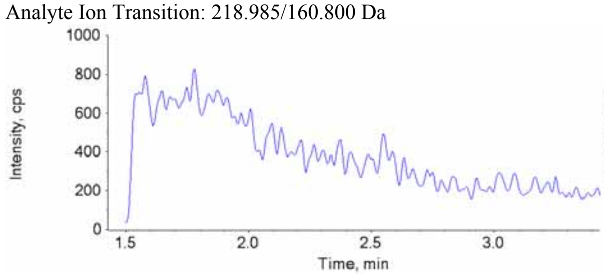
Acquired: 8/27/2010 10:34:53 AM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 27231
IS Ion Transition: 167.000/131.000 Da

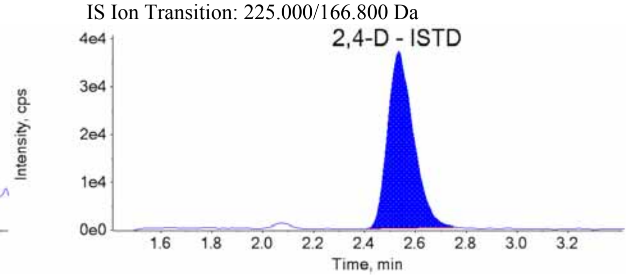




Sample Name: ARA-09-15-10-008-0001A2
Data File: ARA-09-15-10 D26.wiff
Analyte Integration Type: No Peak
Analyte Peak Area: 0
Analyte Ion Transition: 218.985/160.800 Da

Acquired: 8/27/2010 10:34:53 AM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 259482
IS Ion Transition: 225.000/166.800 Da

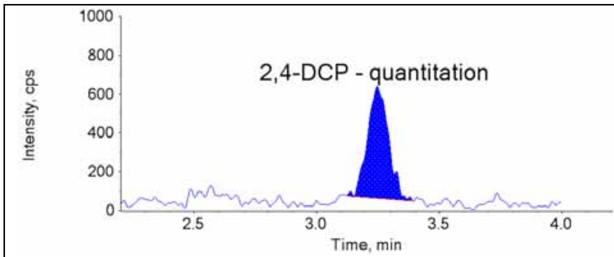


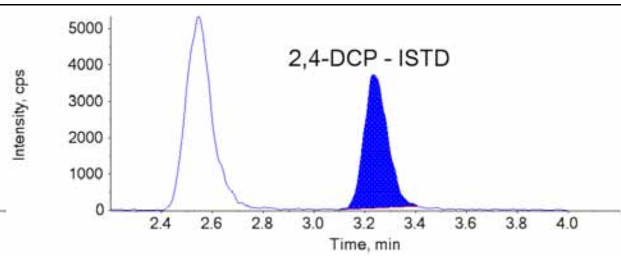


Sample Name: ARA-09-15-10-008-0001A2
Data File: ARA-09-15-10 D26.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 0
Analyte Ion Transition: 220.968/163.000 Da

Acquired: 8/27/2010 10:34:53 AM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 259482
IS Ion Transition: 225.000/166.800 Da

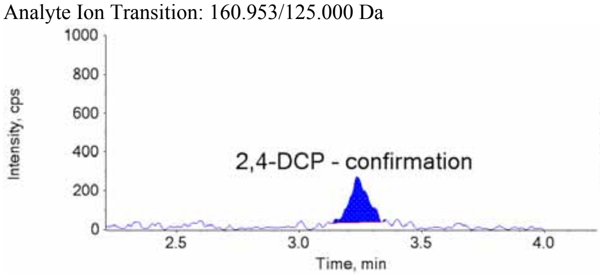
Figure 40. Typical Chromatogram of 2,4-D Analysis, Control Corn Stover (008-0001)

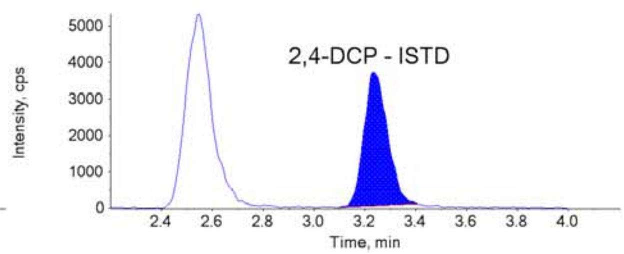




Sample Name: ARA-09-15-10-013-0001A2
Data File: ARA-09-15-10 D26.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 3448
Analyte Ion Transition: 160.953/125.000 Da

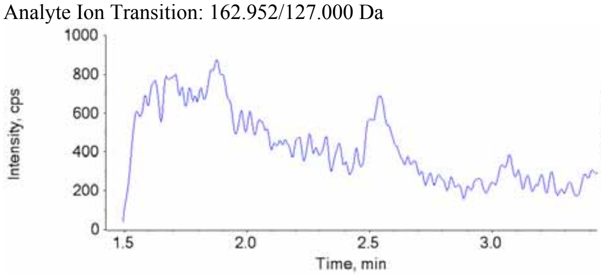
Acquired: 8/27/2010 10:41:22 AM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 23835
IS Ion Transition: 167.000/131.000 Da

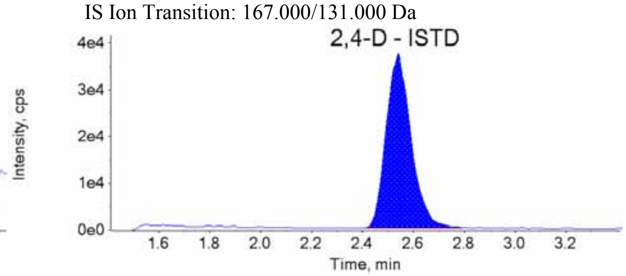




Sample Name: ARA-09-15-10-013-0001A2
Data File: ARA-09-15-10 D26.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 1209
Analyte Ion Transition: 162.952/127.000 Da

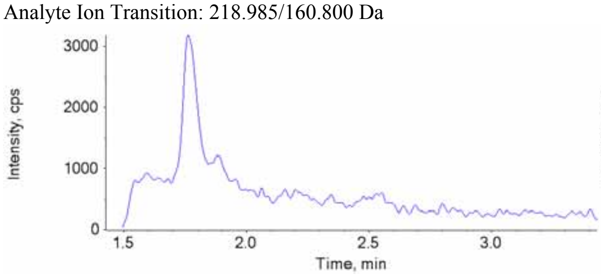
Acquired: 8/27/2010 10:41:22 AM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 23835
IS Ion Transition: 167.000/131.000 Da

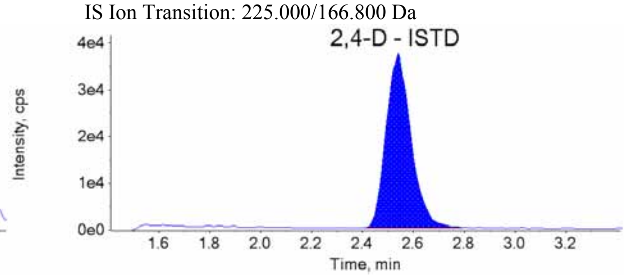




Sample Name: ARA-09-15-10-013-0001A2
Data File: ARA-09-15-10 D26.wiff
Analyte Integration Type: No Peak
Analyte Peak Area: 0
Analyte Ion Transition: 218.985/160.800 Da

Acquired: 8/27/2010 10:41:22 AM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 254676
IS Ion Transition: 225.000/166.800 Da

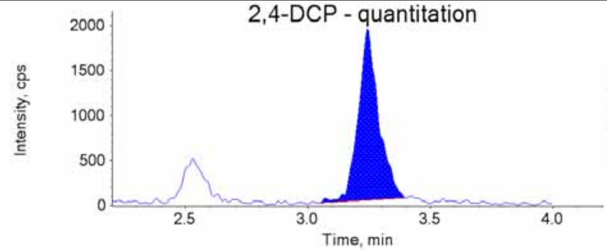


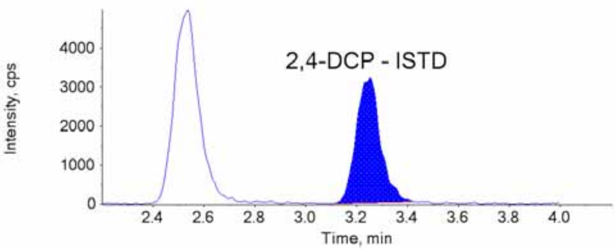


Sample Name: ARA-09-15-10-013-0001A2
Data File: ARA-09-15-10 D26.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 0
Analyte Ion Transition: 220.968/163.000 Da

Acquired: 8/27/2010 10:41:22 AM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 254676
IS Ion Transition: 225.000/166.800 Da

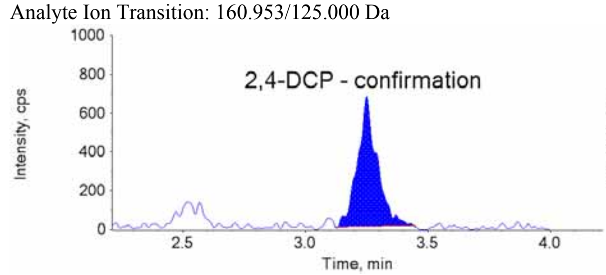
Figure 41. Typical Chromatogram of 2,4-D Analysis, Control Corn Stover (013-0001)

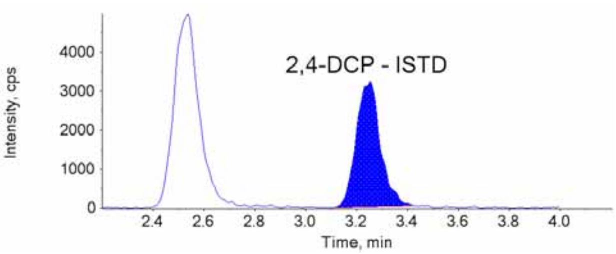




Sample Name: ARA-09-15-10-021-0001A12 + 0.01 µg/g
Data File: ARA-09-15-10 D26.wiff
Analyte Integration Type: Base To Base
Analyte Peak Area: 11021
Analyte Ion Transition: 160.953/125.000 Da

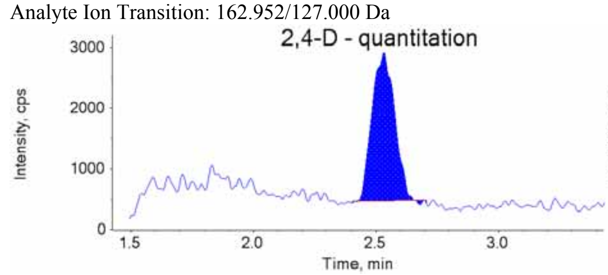
Acquired: 8/27/2010 3:19:56 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 21266
IS Ion Transition: 167.000/131.000 Da

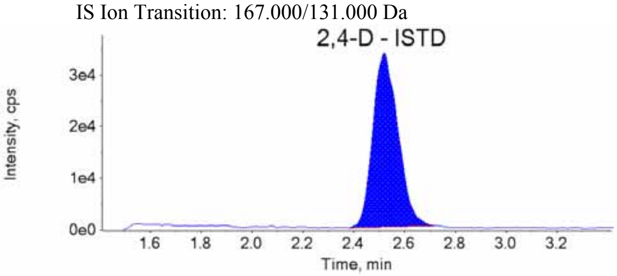




Sample Name: ARA-09-15-10-021-0001A12 + 0.01 µg/g
Data File: ARA-09-15-10 D26.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 3677
Analyte Ion Transition: 162.952/127.000 Da

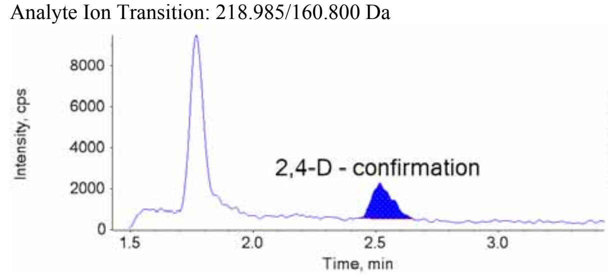
Acquired: 8/27/2010 3:19:56 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 21266
IS Ion Transition: 167.000/131.000 Da

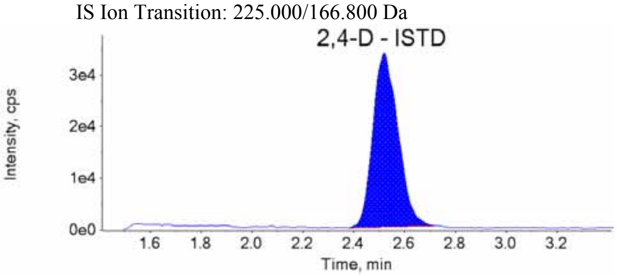




Sample Name: ARA-09-15-10-021-0001A12 + 0.01 µg/g
Data File: ARA-09-15-10 D26.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 15587
Analyte Ion Transition: 218.985/160.800 Da

Acquired: 8/27/2010 3:19:56 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 231003
IS Ion Transition: 225.000/166.800 Da

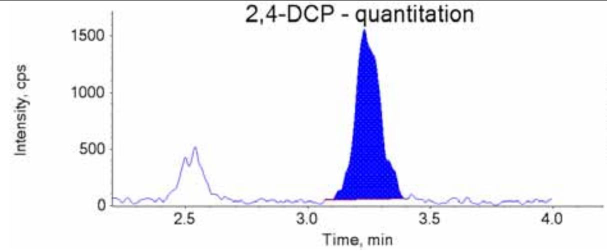


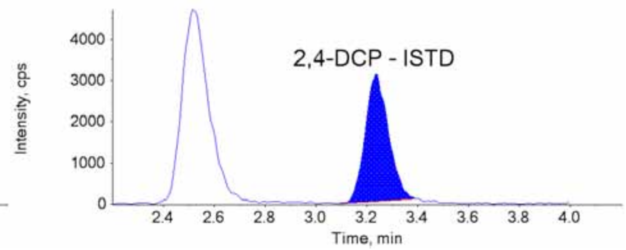


Sample Name: ARA-09-15-10-021-0001A12 + 0.01 µg/g
Data File: ARA-09-15-10 D26.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 10017
Analyte Ion Transition: 220.968/163.000 Da

Acquired: 8/27/2010 3:19:56 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 231003
IS Ion Transition: 225.000/166.800 Da

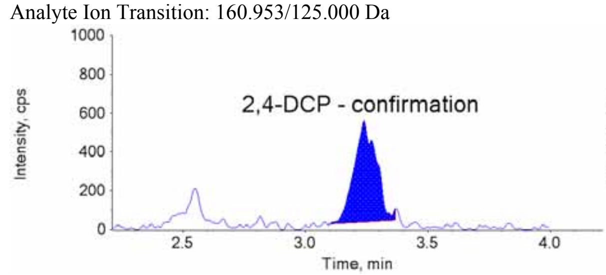
Figure 42. Typical Chromatogram of 2,4-D Analysis, Corn Stover 0.01 µg/g Recovery

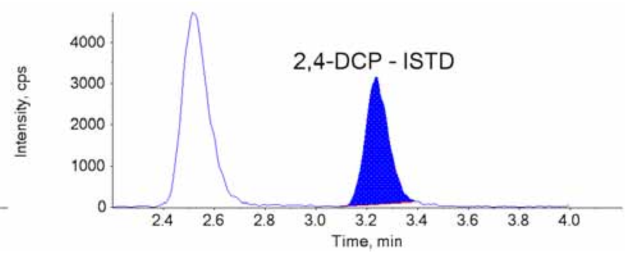




Sample Name: ARA-09-15-10-021-0001A13 + 0.01 µg/g
Data File: ARA-09-15-10 D26.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 10714
Analyte Ion Transition: 160.953/125.000 Da

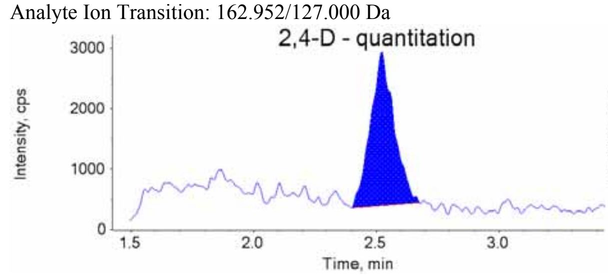
Acquired: 8/27/2010 3:26:25 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 19657
IS Ion Transition: 167.000/131.000 Da

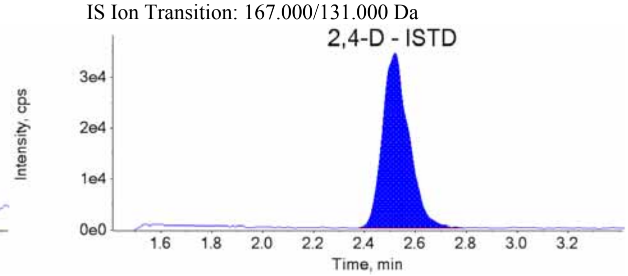




Sample Name: ARA-09-15-10-021-0001A13 + 0.01 µg/g
Data File: ARA-09-15-10 D26.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 3314
Analyte Ion Transition: 162.952/127.000 Da

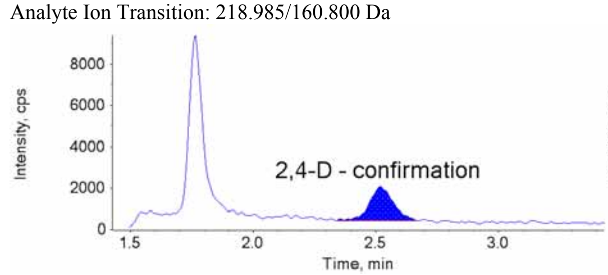
Acquired: 8/27/2010 3:26:25 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 19657
IS Ion Transition: 167.000/131.000 Da

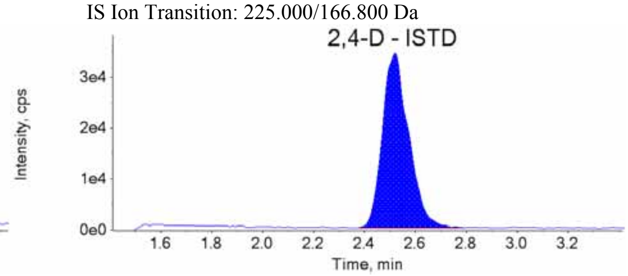




Sample Name: ARA-09-15-10-021-0001A13 + 0.01 µg/g
Data File: ARA-09-15-10 D26.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 16962
Analyte Ion Transition: 218.985/160.800 Da

Acquired: 8/27/2010 3:26:25 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 247143
IS Ion Transition: 225.000/166.800 Da

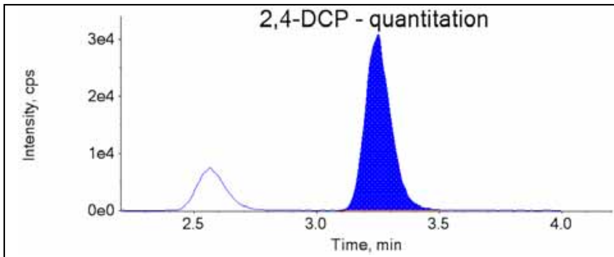


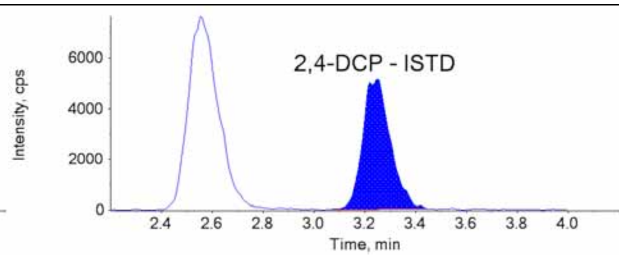


Sample Name: ARA-09-15-10-021-0001A13 + 0.01 µg/g
Data File: ARA-09-15-10 D26.wiff
Analyte Integration Type: Base To Base
Analyte Peak Area: 11565
Analyte Ion Transition: 220.968/163.000 Da

Acquired: 8/27/2010 3:26:25 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 247143
IS Ion Transition: 225.000/166.800 Da

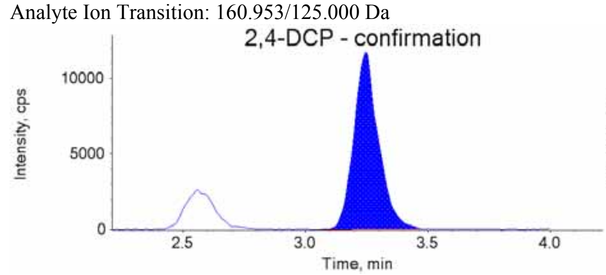
Figure 43. Typical Chromatogram of 2,4-D Analysis, Corn Stover 0.01 µg/g Recovery

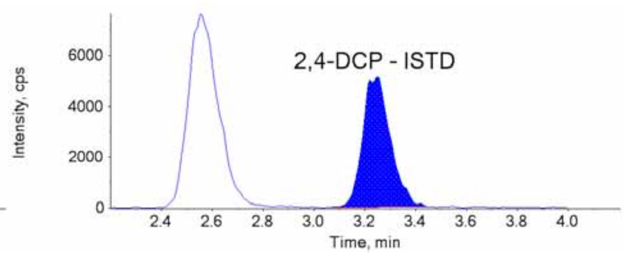




Sample Name: ARA-09-15-10-021-0001A19 + 20.0 µg/g
Data File: ARA-09-15-10 D28c.wiff
Analyte Integration Type: Base To Base
Analyte Peak Area: 223042
Analyte Ion Transition: 160.953/125.000 Da

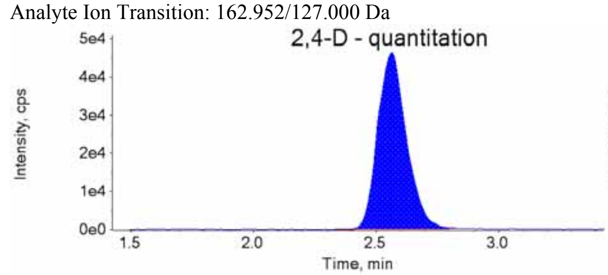
Acquired: 9/1/2010 6:26:31 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 39241
IS Ion Transition: 167.000/131.000 Da

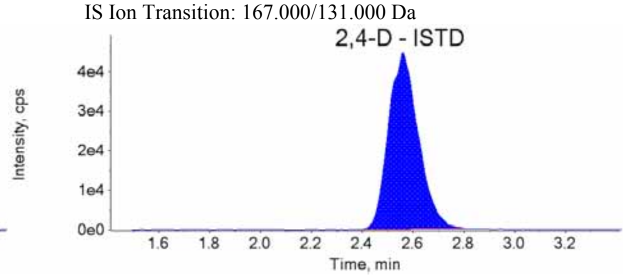




Sample Name: ARA-09-15-10-021-0001A19 + 20.0 µg/g
Data File: ARA-09-15-10 D28c.wiff
Analyte Integration Type: Base To Base
Analyte Peak Area: 81434
Analyte Ion Transition: 162.952/127.000 Da

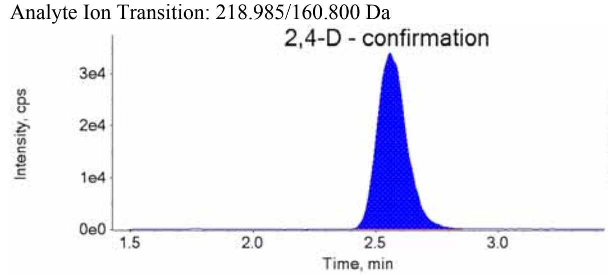
Acquired: 9/1/2010 6:26:31 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 39241
IS Ion Transition: 167.000/131.000 Da

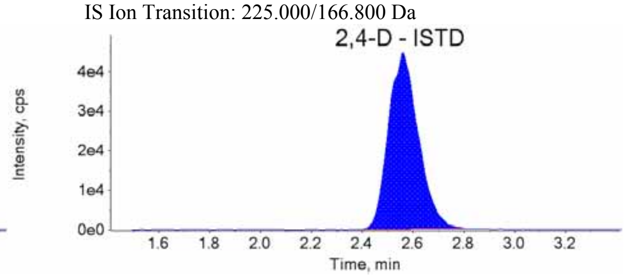




Sample Name: ARA-09-15-10-021-0001A19 + 20.0 µg/g
Data File: ARA-09-15-10 D28c.wiff
Analyte Integration Type: Base To Base
Analyte Peak Area: 376499
Analyte Ion Transition: 218.985/160.800 Da

Acquired: 9/1/2010 6:26:31 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 364200
IS Ion Transition: 225.000/166.800 Da

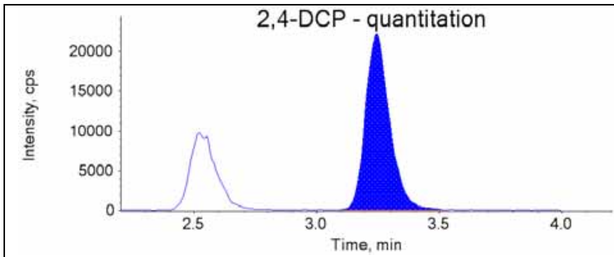


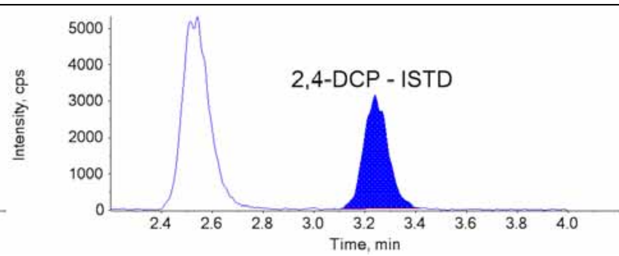


Sample Name: ARA-09-15-10-021-0001A19 + 20.0 µg/g
Data File: ARA-09-15-10 D28c.wiff
Analyte Integration Type: Base To Base
Analyte Peak Area: 282454
Analyte Ion Transition: 220.968/163.000 Da

Acquired: 9/1/2010 6:26:31 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 364200
IS Ion Transition: 225.000/166.800 Da

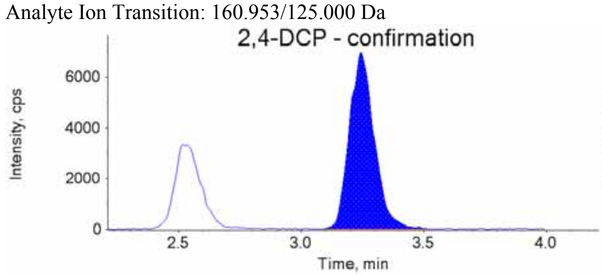
Figure 44. Typical Chromatogram of 2,4-D Analysis, Corn Stover 20.0 µg/g Recovery

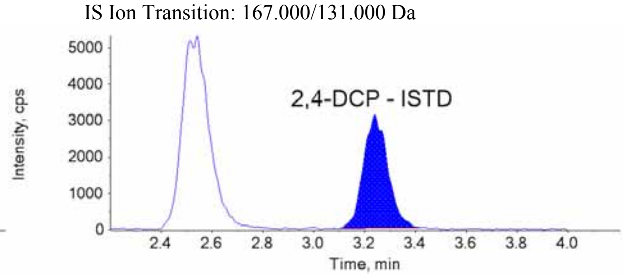




Sample Name: ARA-09-15-10-013-0002A2
Data File: ARA-09-15-10 D27.wiff
Analyte Integration Type: Base To Base
Analyte Peak Area: 151624
Analyte Ion Transition: 160.953/125.000 Da

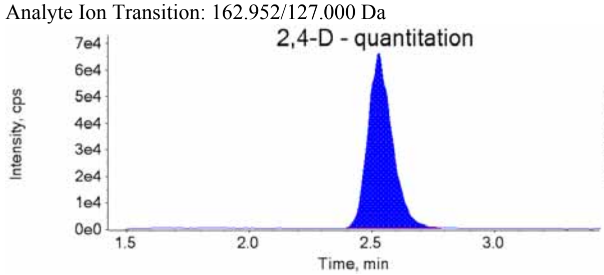
Acquired: 8/27/2010 11:40:31 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 20707
IS Ion Transition: 167.000/131.000 Da

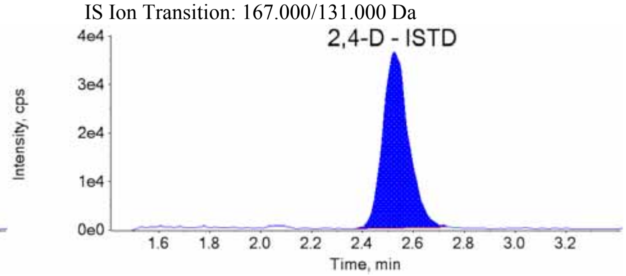




Sample Name: ARA-09-15-10-013-0002A2
Data File: ARA-09-15-10 D27.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 49111
Analyte Ion Transition: 162.952/127.000 Da

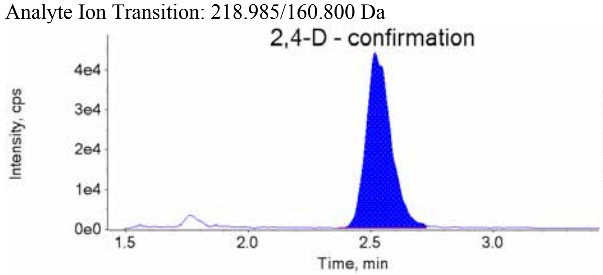
Acquired: 8/27/2010 11:40:31 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 20707
IS Ion Transition: 167.000/131.000 Da

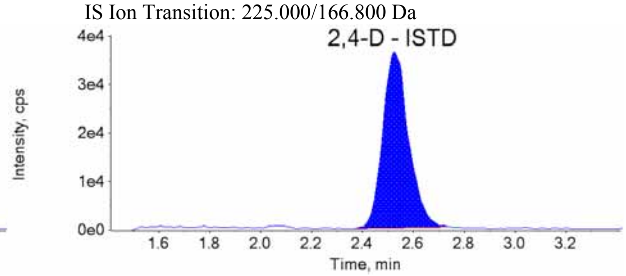




Sample Name: ARA-09-15-10-013-0002A2
Data File: ARA-09-15-10 D27.wiff
Analyte Integration Type: Base To Base
Analyte Peak Area: 450845
Analyte Ion Transition: 218.985/160.800 Da

Acquired: 8/27/2010 11:40:31 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 256420
IS Ion Transition: 225.000/166.800 Da

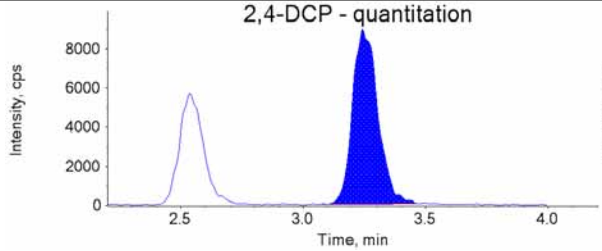


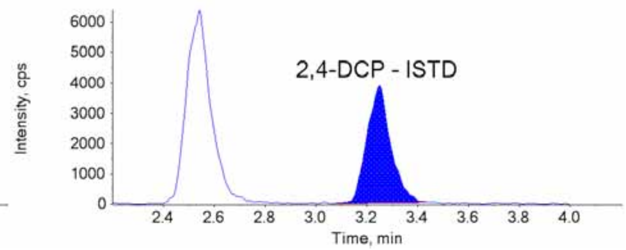


Sample Name: ARA-09-15-10-013-0002A2
Data File: ARA-09-15-10 D27.wiff
Analyte Integration Type: Exponential Skim
Analyte Peak Area: 311858
Analyte Ion Transition: 220.968/163.000 Da

Acquired: 8/27/2010 11:40:31 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 256420
IS Ion Transition: 225.000/166.800 Da

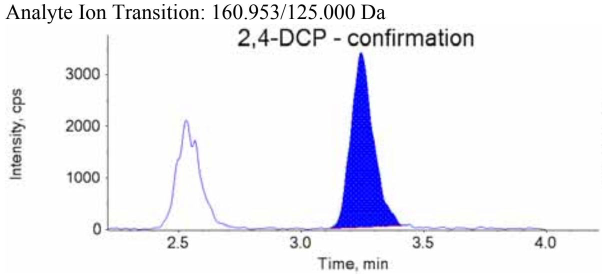
Figure 45. Typical Chromatogram of 2,4-D Analysis, Treated Corn Stover (013-0002)

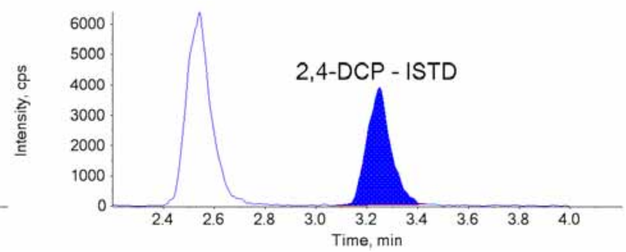




Sample Name: ARA-09-15-10-013-0003A2
Data File: ARA-09-15-10 D27.wiff
Analyte Integration Type: Base To Base
Analyte Peak Area: 64832
Analyte Ion Transition: 160.953/125.000 Da

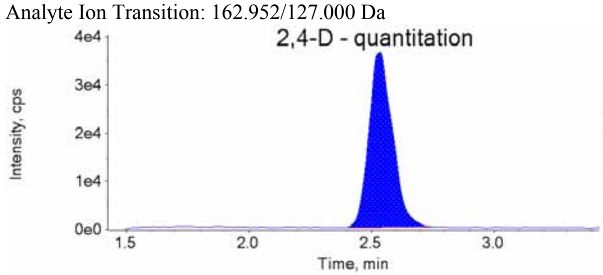
Acquired: 8/27/2010 11:21:05 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 25211
IS Ion Transition: 167.000/131.000 Da

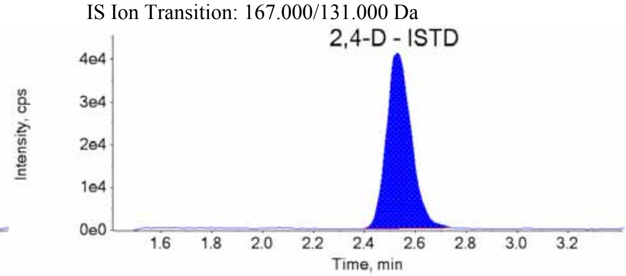




Sample Name: ARA-09-15-10-013-0003A2
Data File: ARA-09-15-10 D27.wiff
Analyte Integration Type: Base To Base
Analyte Peak Area: 21503
Analyte Ion Transition: 162.952/127.000 Da

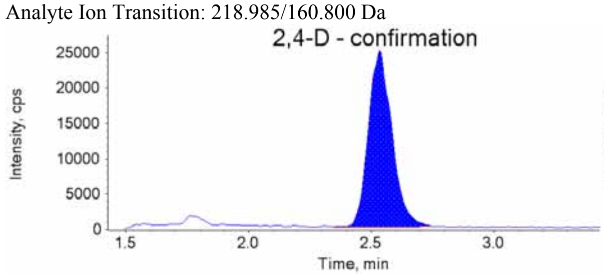
Acquired: 8/27/2010 11:21:05 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 25211
IS Ion Transition: 167.000/131.000 Da

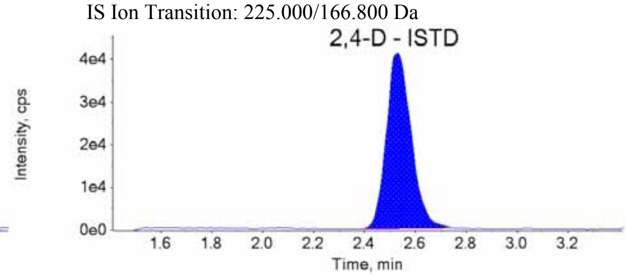




Sample Name: ARA-09-15-10-013-0003A2
Data File: ARA-09-15-10 D27.wiff
Analyte Integration Type: Base To Base
Analyte Peak Area: 245160
Analyte Ion Transition: 218.985/160.800 Da

Acquired: 8/27/2010 11:21:05 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 277832
IS Ion Transition: 225.000/166.800 Da

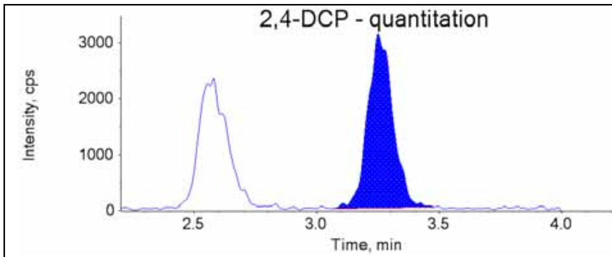


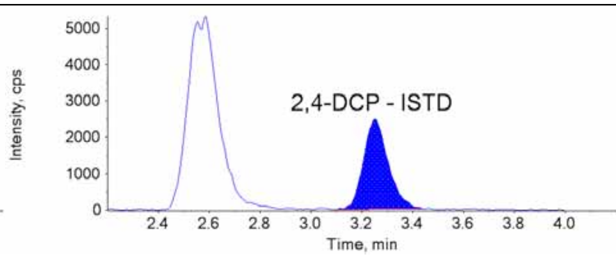


Sample Name: ARA-09-15-10-013-0003A2
Data File: ARA-09-15-10 D27.wiff
Analyte Integration Type: Valley
Analyte Peak Area: 166543
Analyte Ion Transition: 220.968/163.000 Da

Acquired: 8/27/2010 11:21:05 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 277832
IS Ion Transition: 225.000/166.800 Da

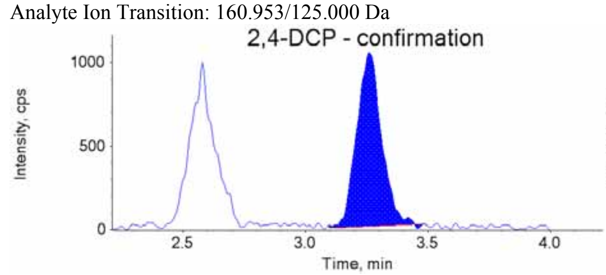
Figure 46. Typical Chromatogram of 2,4-D Analysis, Treated Corn Stover (013-0003)

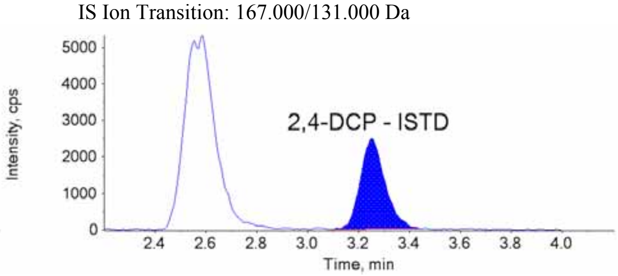




Sample Name: ARA-09-15-10-023-0002A2
Data File: ARA-09-15-10 D28b.wiff
Analyte Integration Type: Base To Base
Analyte Peak Area: 22260
Analyte Ion Transition: 160.953/125.000 Da

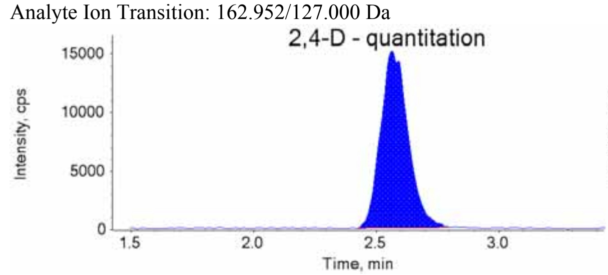
Acquired: 8/31/2010 11:27:12 AM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 16636
IS Ion Transition: 167.000/131.000 Da

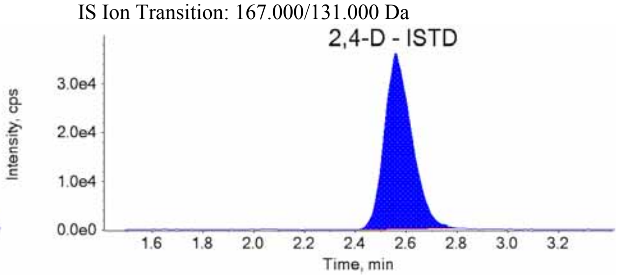




Sample Name: ARA-09-15-10-023-0002A2
Data File: ARA-09-15-10 D28b.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 7377
Analyte Ion Transition: 162.952/127.000 Da

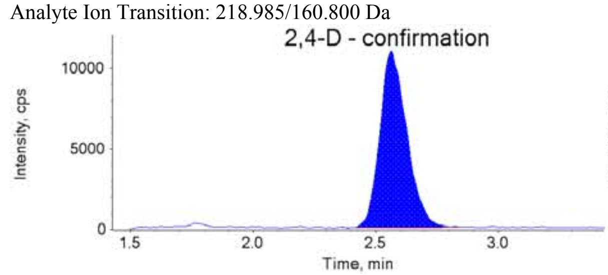
Acquired: 8/31/2010 11:27:12 AM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 16636
IS Ion Transition: 167.000/131.000 Da

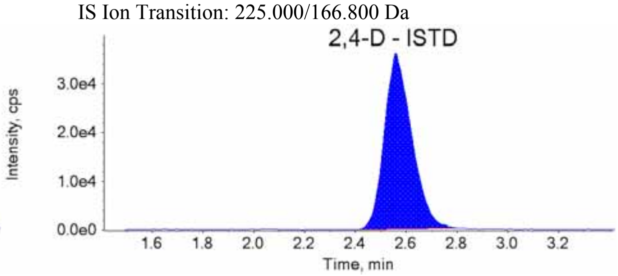




Sample Name: ARA-09-15-10-023-0002A2
Data File: ARA-09-15-10 D28b.wiff
Analyte Integration Type: Base To Base
Analyte Peak Area: 117911
Analyte Ion Transition: 218.985/160.800 Da

Acquired: 8/31/2010 11:27:12 AM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 268678
IS Ion Transition: 225.000/166.800 Da

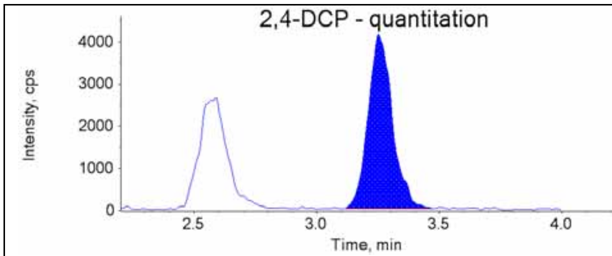


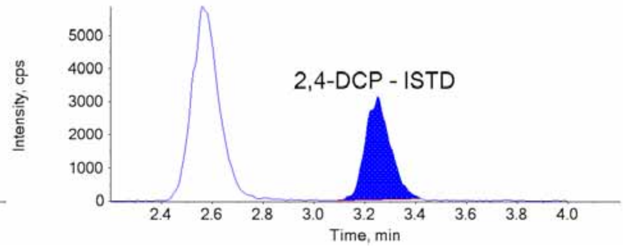


Sample Name: ARA-09-15-10-023-0002A2
Data File: ARA-09-15-10 D28b.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 83956
Analyte Ion Transition: 220.968/163.000 Da

Acquired: 8/31/2010 11:27:12 AM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 268678
IS Ion Transition: 225.000/166.800 Da

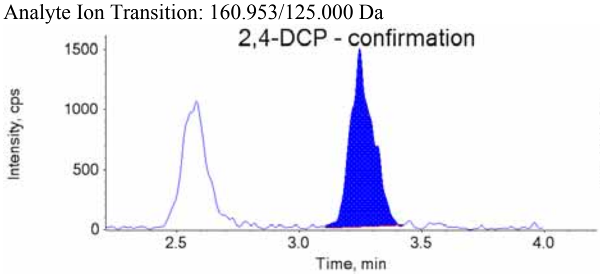
Figure 47. Typical Chromatogram of 2,4-D Analysis, Treated Corn Stover (023-0002)

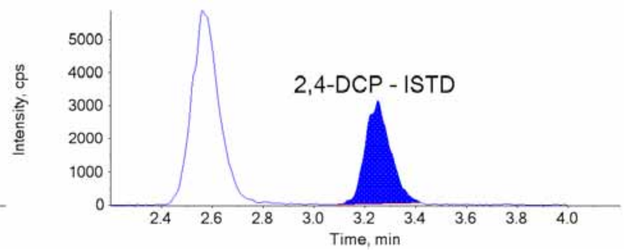




Sample Name: ARA-09-15-10-023-0003A2
Data File: ARA-09-15-10 D28b.wiff
Analyte Integration Type: Base To Base
Analyte Peak Area: 28677
Analyte Ion Transition: 160.953/125.000 Da

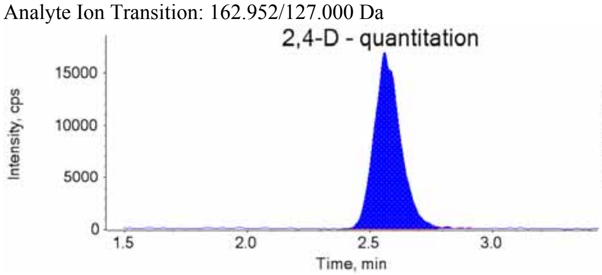
Acquired: 8/31/2010 11:53:07 AM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 21267
IS Ion Transition: 167.000/131.000 Da

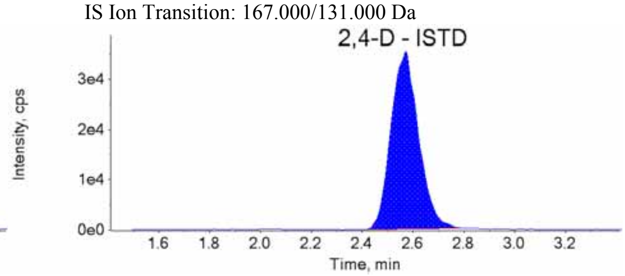




Sample Name: ARA-09-15-10-023-0003A2
Data File: ARA-09-15-10 D28b.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 9387
Analyte Ion Transition: 162.952/127.000 Da

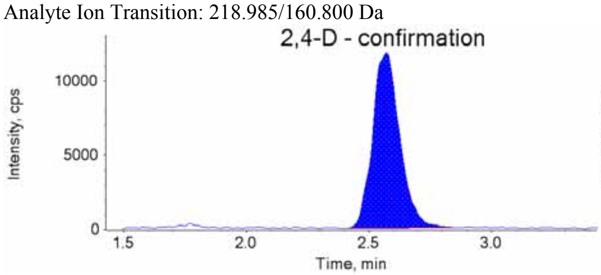
Acquired: 8/31/2010 11:53:07 AM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 21267
IS Ion Transition: 167.000/131.000 Da

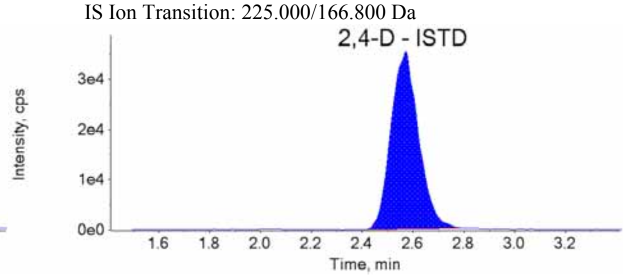




Sample Name: ARA-09-15-10-023-0003A2
Data File: ARA-09-15-10 D28b.wiff
Analyte Integration Type: Base To Base
Analyte Peak Area: 127443
Analyte Ion Transition: 218.985/160.800 Da

Acquired: 8/31/2010 11:53:07 AM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 264982
IS Ion Transition: 225.000/166.800 Da

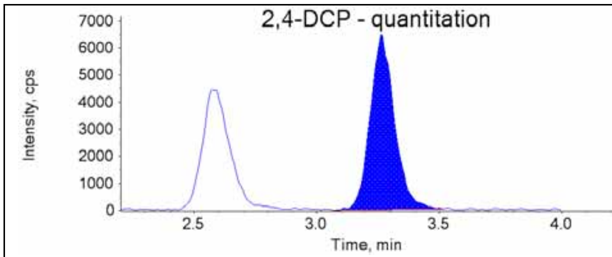


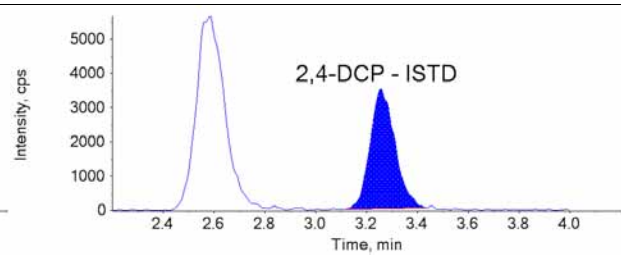


Sample Name: ARA-09-15-10-023-0003A2
Data File: ARA-09-15-10 D28b.wiff
Analyte Integration Type: Valley
Analyte Peak Area: 91255
Analyte Ion Transition: 220.968/163.000 Da

Acquired: 8/31/2010 11:53:07 AM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 264982
IS Ion Transition: 225.000/166.800 Da

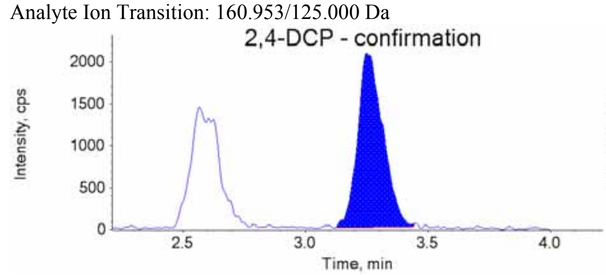
Figure 48. Typical Chromatogram of 2,4-D Analysis, Treated Corn Stover (023-0003)

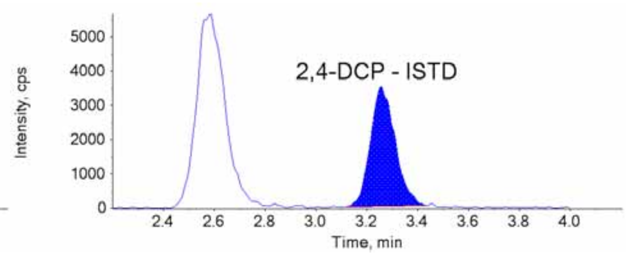




Sample Name: ARA-09-15-10-025-0002A2
Data File: ARA-09-15-10 D28b.wiff
Analyte Integration Type: Base To Base
Analyte Peak Area: 44602
Analyte Ion Transition: 160.953/125.000 Da

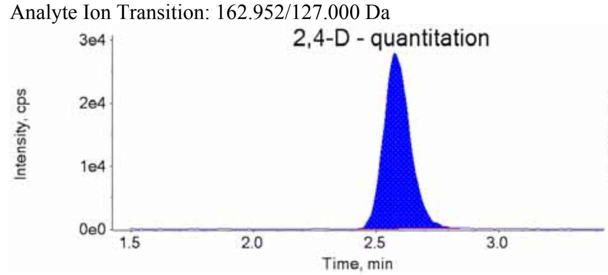
Acquired: 8/31/2010 12:12:30 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 23640
IS Ion Transition: 167.000/131.000 Da

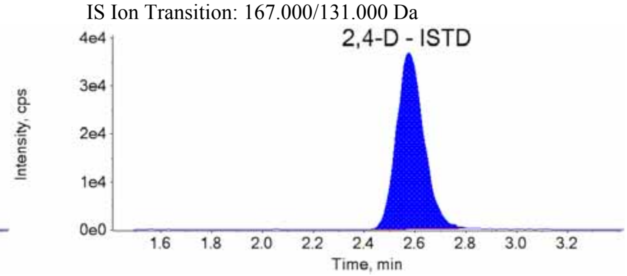




Sample Name: ARA-09-15-10-025-0002A2
Data File: ARA-09-15-10 D28b.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 15067
Analyte Ion Transition: 162.952/127.000 Da

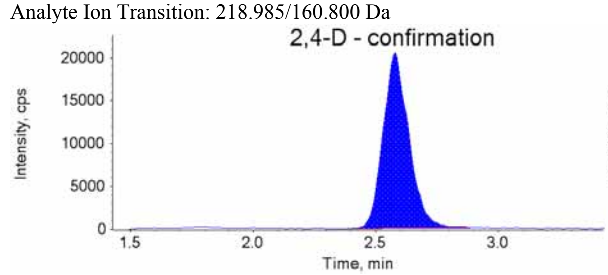
Acquired: 8/31/2010 12:12:30 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 23640
IS Ion Transition: 167.000/131.000 Da

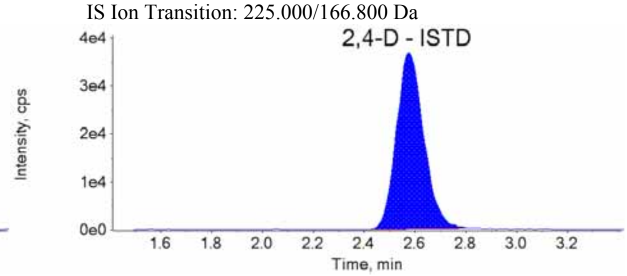




Sample Name: ARA-09-15-10-025-0002A2
Data File: ARA-09-15-10 D28b.wiff
Analyte Integration Type: Base To Base
Analyte Peak Area: 209448
Analyte Ion Transition: 218.985/160.800 Da

Acquired: 8/31/2010 12:12:30 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 274698
IS Ion Transition: 225.000/166.800 Da

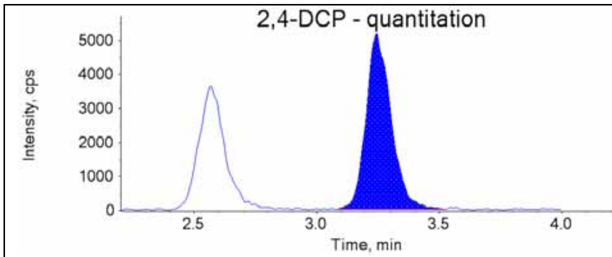


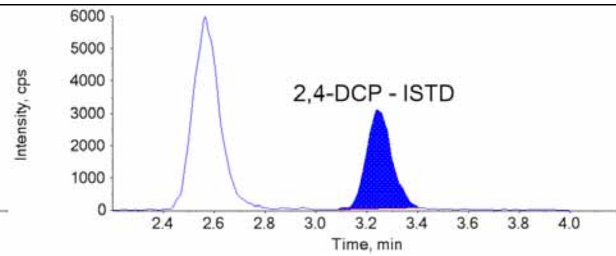


Sample Name: ARA-09-15-10-025-0002A2
Data File: ARA-09-15-10 D28b.wiff
Analyte Integration Type: Base To Base
Analyte Peak Area: 148766
Analyte Ion Transition: 220.968/163.000 Da

Acquired: 8/31/2010 12:12:30 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 274698
IS Ion Transition: 225.000/166.800 Da

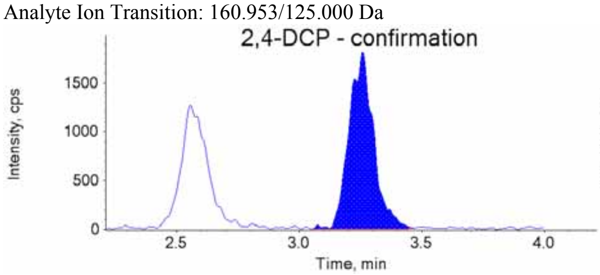
Figure 49. Typical Chromatogram of 2,4-D Analysis, Treated Corn Stover (025-0002)

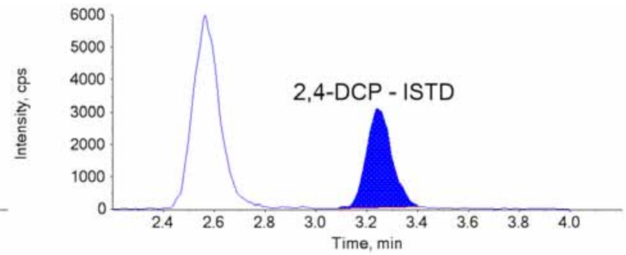




Sample Name: ARA-09-15-10-025-0003A2
Data File: ARA-09-15-10 D28b.wiff
Analyte Integration Type: Base To Base
Analyte Peak Area: 36931
Analyte Ion Transition: 160.953/125.000 Da

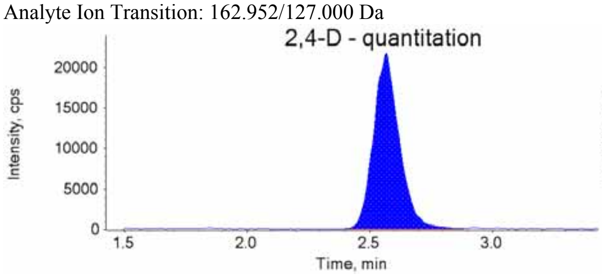
Acquired: 8/31/2010 12:44:56 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 21400
IS Ion Transition: 167.000/131.000 Da

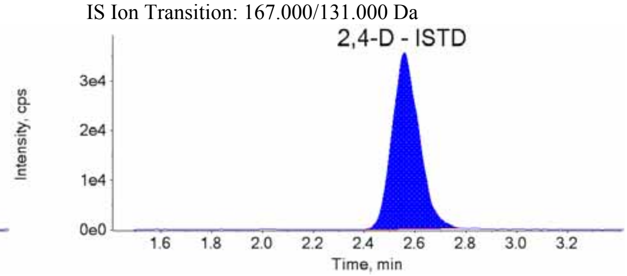




Sample Name: ARA-09-15-10-025-0003A2
Data File: ARA-09-15-10 D28b.wiff
Analyte Integration Type: Valley
Analyte Peak Area: 12733
Analyte Ion Transition: 162.952/127.000 Da

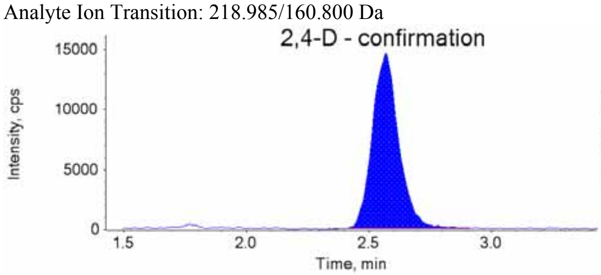
Acquired: 8/31/2010 12:44:56 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 21400
IS Ion Transition: 167.000/131.000 Da

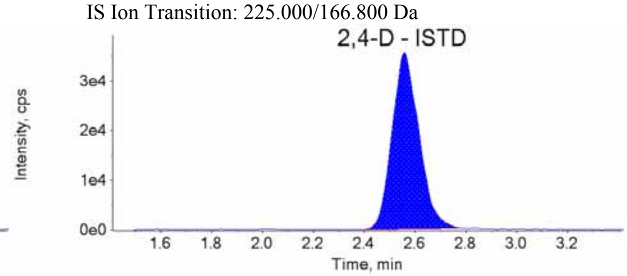




Sample Name: ARA-09-15-10-025-0003A2
Data File: ARA-09-15-10 D28b.wiff
Analyte Integration Type: Base To Base
Analyte Peak Area: 156877
Analyte Ion Transition: 218.985/160.800 Da

Acquired: 8/31/2010 12:44:56 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 260392
IS Ion Transition: 225.000/166.800 Da

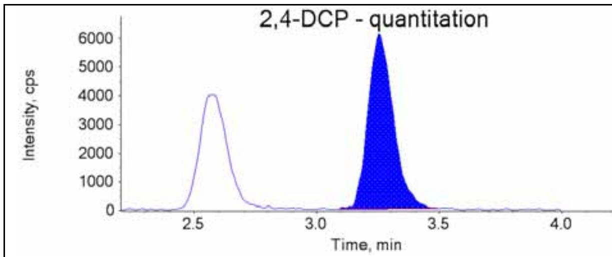


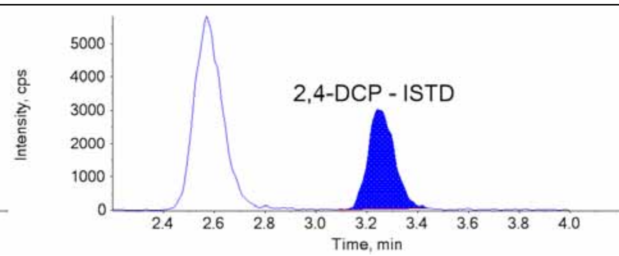


Sample Name: ARA-09-15-10-025-0003A2
Data File: ARA-09-15-10 D28b.wiff
Analyte Integration Type: Base To Base
Analyte Peak Area: 108814
Analyte Ion Transition: 220.968/163.000 Da

Acquired: 8/31/2010 12:44:56 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 260392
IS Ion Transition: 225.000/166.800 Da

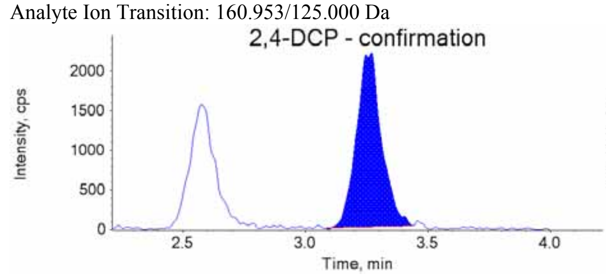
Figure 50. Typical Chromatogram of 2,4-D Analysis, Treated Corn Stover (025-0003)

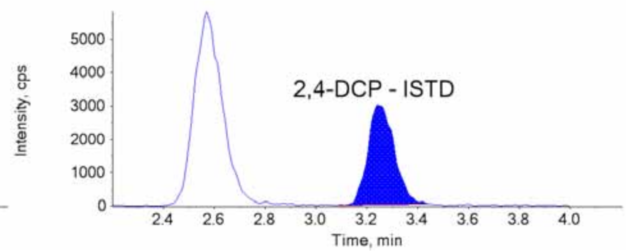




Sample Name: ARA-09-15-10-027-0002A2
Data File: ARA-09-15-10 D28b.wiff
Analyte Integration Type: Base To Base
Analyte Peak Area: 43762
Analyte Ion Transition: 160.953/125.000 Da

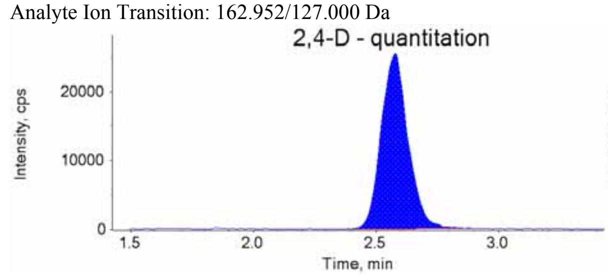
Acquired: 8/31/2010 12:31:58 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 21873
IS Ion Transition: 167.000/131.000 Da

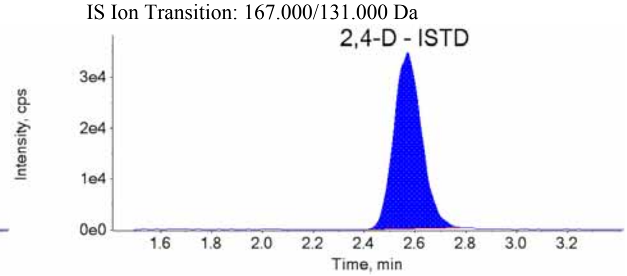




Sample Name: ARA-09-15-10-027-0002A2
Data File: ARA-09-15-10 D28b.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 15728
Analyte Ion Transition: 162.952/127.000 Da

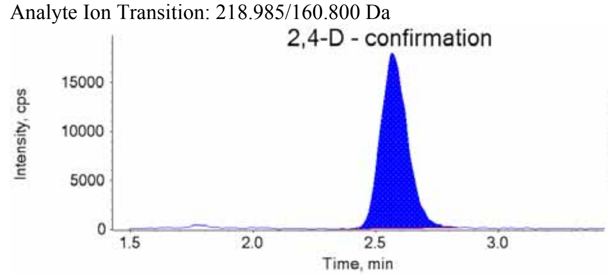
Acquired: 8/31/2010 12:31:58 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 21873
IS Ion Transition: 167.000/131.000 Da

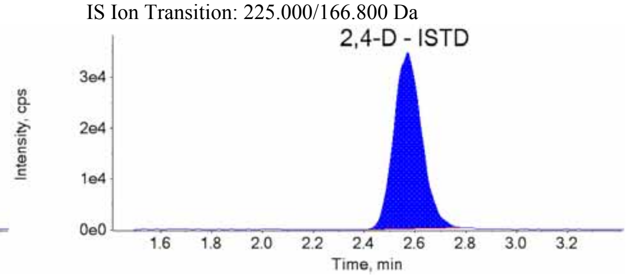




Sample Name: ARA-09-15-10-027-0002A2
Data File: ARA-09-15-10 D28b.wiff
Analyte Integration Type: Base To Base
Analyte Peak Area: 193218
Analyte Ion Transition: 218.985/160.800 Da

Acquired: 8/31/2010 12:31:58 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 262015
IS Ion Transition: 225.000/166.800 Da

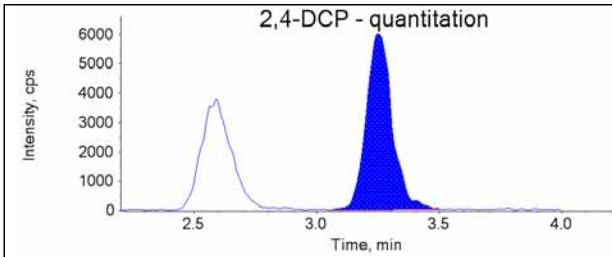


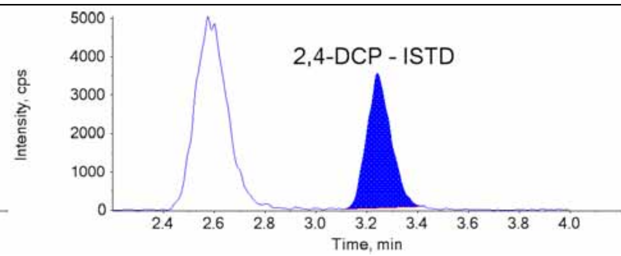


Sample Name: ARA-09-15-10-027-0002A2
Data File: ARA-09-15-10 D28b.wiff
Analyte Integration Type: Base To Base
Analyte Peak Area: 136542
Analyte Ion Transition: 220.968/163.000 Da

Acquired: 8/31/2010 12:31:58 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 262015
IS Ion Transition: 225.000/166.800 Da

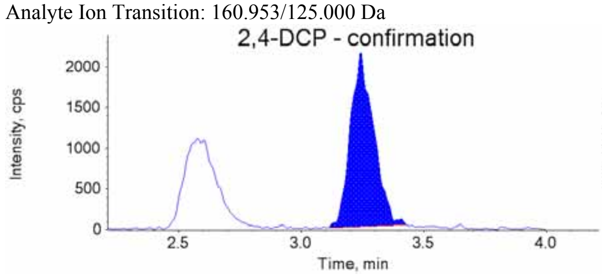
Figure 51. Typical Chromatogram of 2,4-D Analysis, Treated Corn Stover (027-0002)

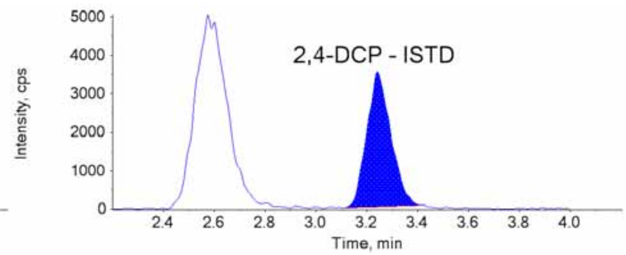




Sample Name: ARA-09-15-10-027-0003A2
Data File: ARA-09-15-10 D28b.wiff
Analyte Integration Type: Base To Base
Analyte Peak Area: 44620
Analyte Ion Transition: 160.953/125.000 Da

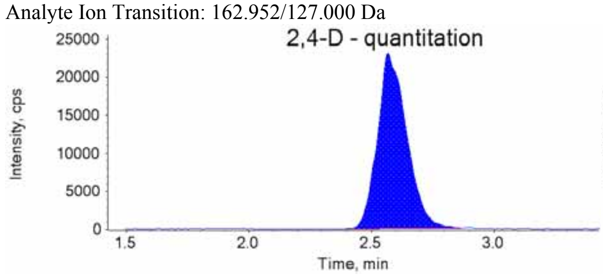
Acquired: 8/31/2010 12:18:59 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 23013
IS Ion Transition: 167.000/131.000 Da

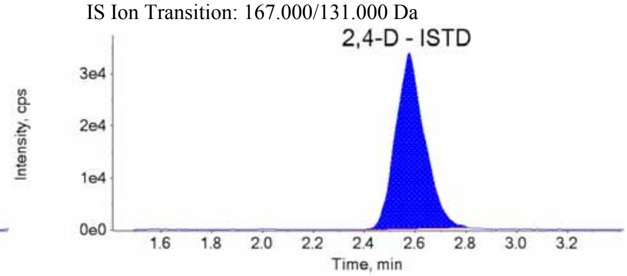




Sample Name: ARA-09-15-10-027-0003A2
Data File: ARA-09-15-10 D28b.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 14234
Analyte Ion Transition: 162.952/127.000 Da

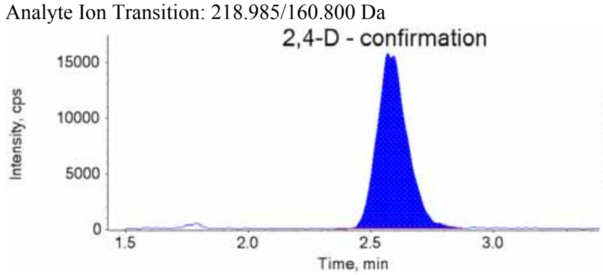
Acquired: 8/31/2010 12:18:59 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 23013
IS Ion Transition: 167.000/131.000 Da

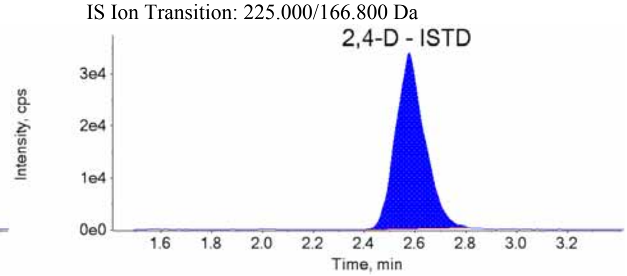




Sample Name: ARA-09-15-10-027-0003A2
Data File: ARA-09-15-10 D28b.wiff
Analyte Integration Type: Base To Base
Analyte Peak Area: 190729
Analyte Ion Transition: 218.985/160.800 Da

Acquired: 8/31/2010 12:18:59 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 265325
IS Ion Transition: 225.000/166.800 Da

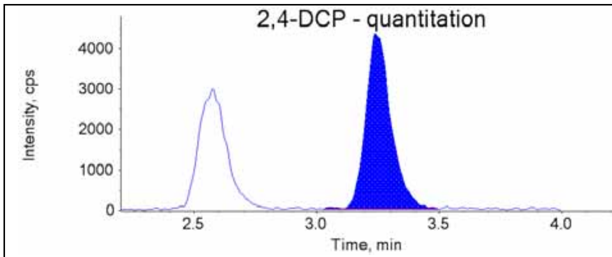


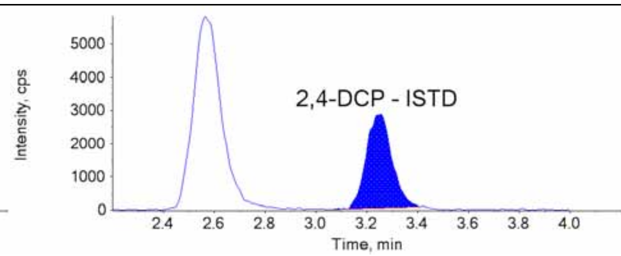


Sample Name: ARA-09-15-10-027-0003A2
Data File: ARA-09-15-10 D28b.wiff
Analyte Integration Type: Base To Base
Analyte Peak Area: 135205
Analyte Ion Transition: 220.968/163.000 Da

Acquired: 8/31/2010 12:18:59 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 265325
IS Ion Transition: 225.000/166.800 Da

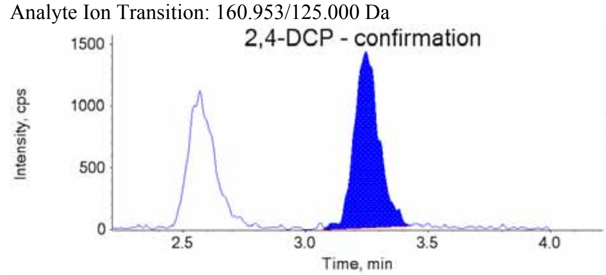
Figure 52. Typical Chromatogram of 2,4-D Analysis, Treated Corn Stover (027-0003)

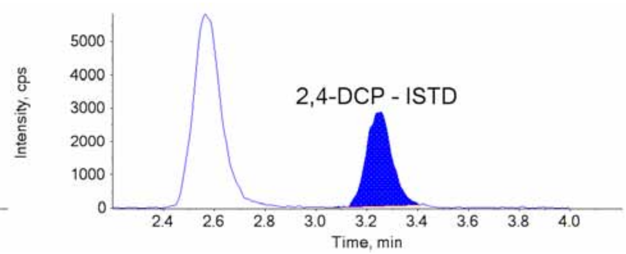




Sample Name: ARA-09-15-10-029-0002A2
Data File: ARA-09-15-10 D28b.wiff
Analyte Integration Type: Base To Base
Analyte Peak Area: 31279
Analyte Ion Transition: 160.953/125.000 Da

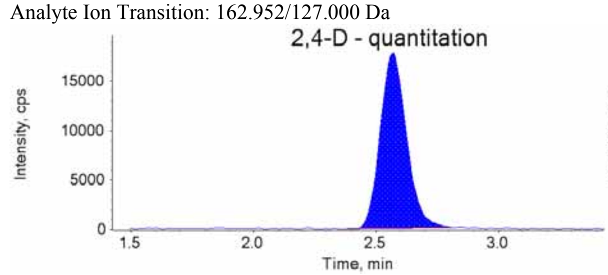
Acquired: 8/31/2010 12:06:02 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 19928
IS Ion Transition: 167.000/131.000 Da

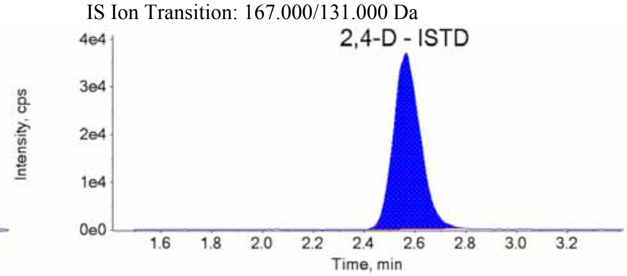




Sample Name: ARA-09-15-10-029-0002A2
Data File: ARA-09-15-10 D28b.wiff
Analyte Integration Type: Valley
Analyte Peak Area: 10126
Analyte Ion Transition: 162.952/127.000 Da

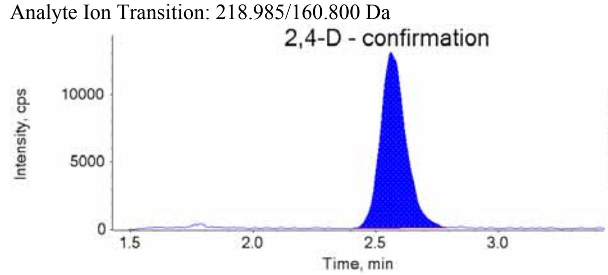
Acquired: 8/31/2010 12:06:02 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 19928
IS Ion Transition: 167.000/131.000 Da

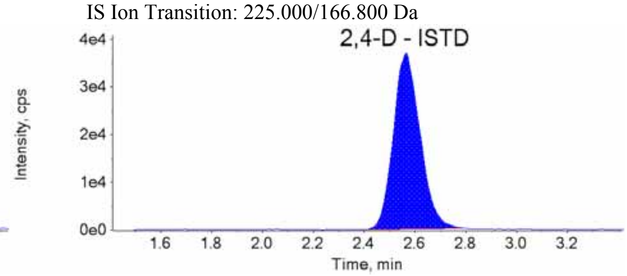




Sample Name: ARA-09-15-10-029-0002A2
Data File: ARA-09-15-10 D28b.wiff
Analyte Integration Type: Base To Base
Analyte Peak Area: 131465
Analyte Ion Transition: 218.985/160.800 Da

Acquired: 8/31/2010 12:06:02 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 268485
IS Ion Transition: 225.000/166.800 Da

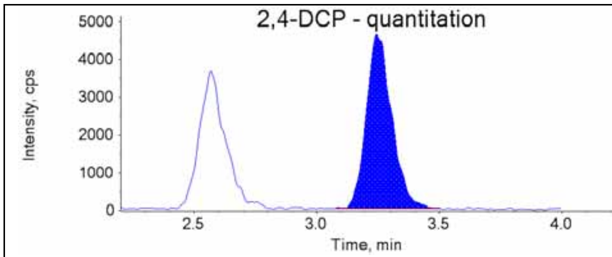


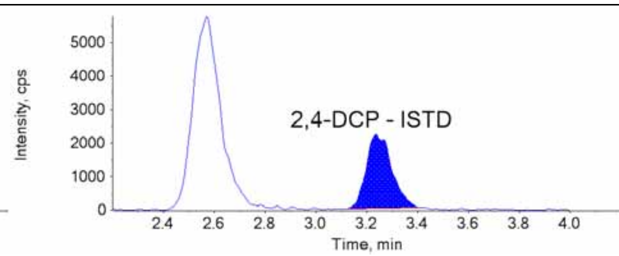


Sample Name: ARA-09-15-10-029-0002A2
Data File: ARA-09-15-10 D28b.wiff
Analyte Integration Type: Base To Base
Analyte Peak Area: 93897
Analyte Ion Transition: 220.968/163.000 Da

Acquired: 8/31/2010 12:06:02 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 268485
IS Ion Transition: 225.000/166.800 Da

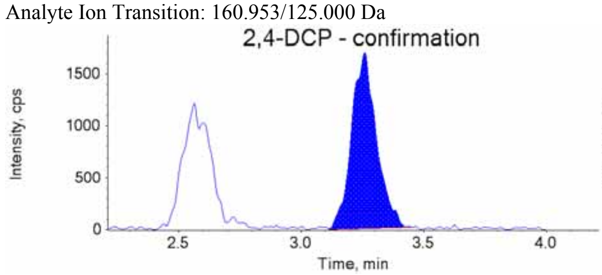
Figure 53. Typical Chromatogram of 2,4-D Analysis, Treated Corn Stover (029-0002)

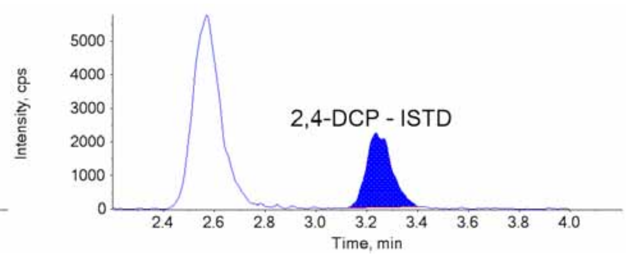




Sample Name: ARA-09-15-10-029-0003A2
Data File: ARA-09-15-10 D28b.wiff
Analyte Integration Type: Base To Base
Analyte Peak Area: 33546
Analyte Ion Transition: 160.953/125.000 Da

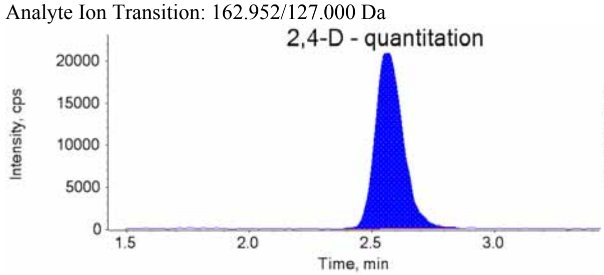
Acquired: 8/31/2010 11:46:37 AM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 15552
IS Ion Transition: 167.000/131.000 Da

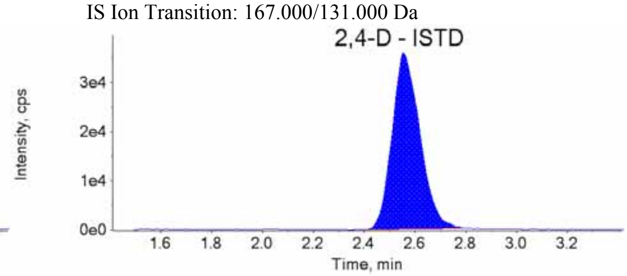




Sample Name: ARA-09-15-10-029-0003A2
Data File: ARA-09-15-10 D28b.wiff
Analyte Integration Type: Valley
Analyte Peak Area: 11935
Analyte Ion Transition: 162.952/127.000 Da

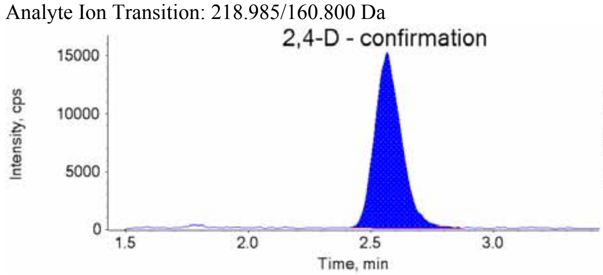
Acquired: 8/31/2010 11:46:37 AM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 15552
IS Ion Transition: 167.000/131.000 Da

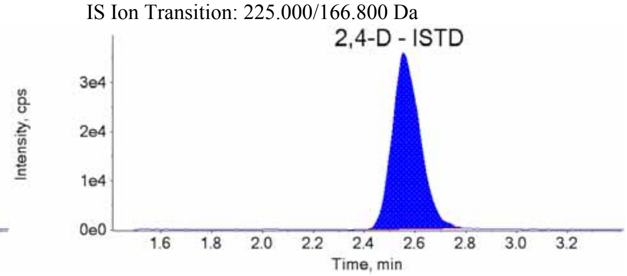




Sample Name: ARA-09-15-10-029-0003A2
Data File: ARA-09-15-10 D28b.wiff
Analyte Integration Type: Base To Base
Analyte Peak Area: 165221
Analyte Ion Transition: 218.985/160.800 Da

Acquired: 8/31/2010 11:46:37 AM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 271102
IS Ion Transition: 225.000/166.800 Da

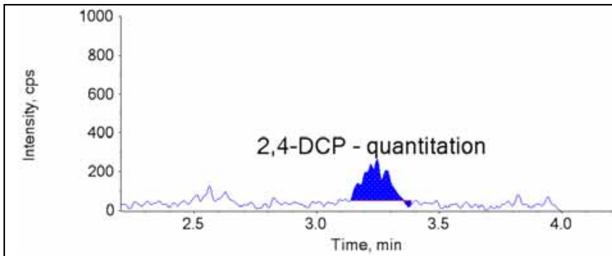


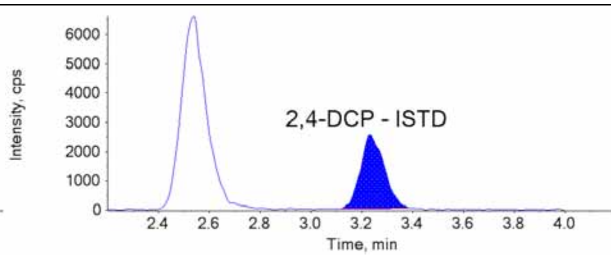


Sample Name: ARA-09-15-10-029-0003A2
Data File: ARA-09-15-10 D28b.wiff
Analyte Integration Type: Base To Base
Analyte Peak Area: 113811
Analyte Ion Transition: 220.968/163.000 Da

Acquired: 8/31/2010 11:46:37 AM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 271102
IS Ion Transition: 225.000/166.800 Da

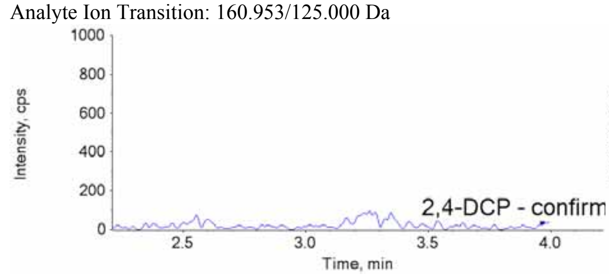
Figure 54. Typical Chromatogram of 2,4-D Analysis, Treated Corn Stover (029-0003)

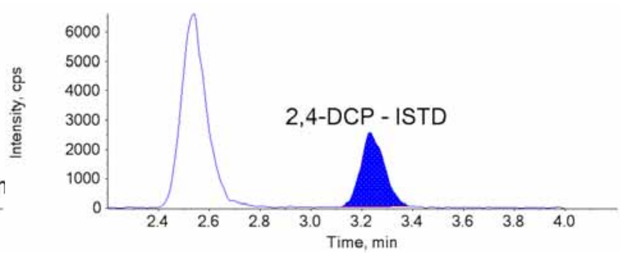




Sample Name: ARA-09-15-10-141-0001A2
Data File: ARA-09-15-10 D19c.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 1393
Analyte Ion Transition: 160.953/125.000 Da

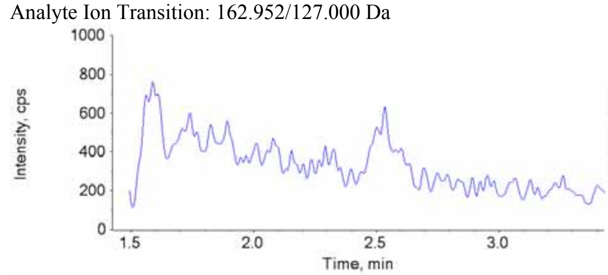
Acquired: 8/18/2010 4:23:38 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 16399
IS Ion Transition: 167.000/131.000 Da

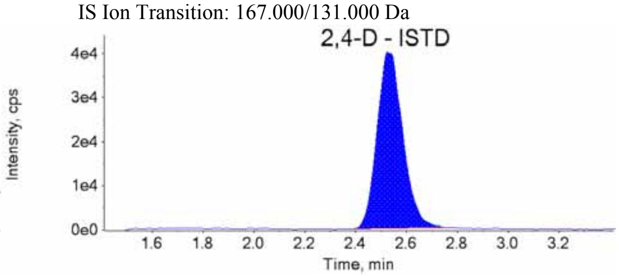




Sample Name: ARA-09-15-10-141-0001A2
Data File: ARA-09-15-10 D19c.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 0
Analyte Ion Transition: 162.952/127.000 Da

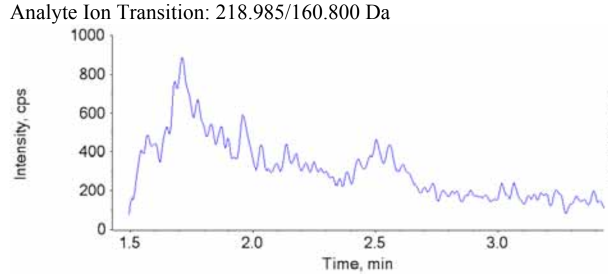
Acquired: 8/18/2010 4:23:38 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 16399
IS Ion Transition: 167.000/131.000 Da

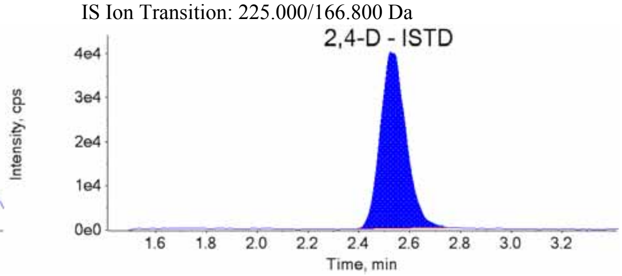




Sample Name: ARA-09-15-10-141-0001A2
Data File: ARA-09-15-10 D19c.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 0
Analyte Ion Transition: 218.985/160.800 Da

Acquired: 8/18/2010 4:23:38 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 279413
IS Ion Transition: 225.000/166.800 Da

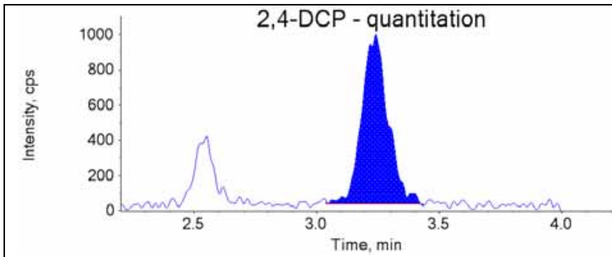


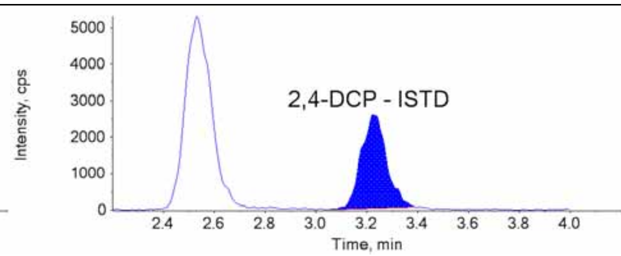


Sample Name: ARA-09-15-10-141-0001A2
Data File: ARA-09-15-10 D19c.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 0
Analyte Ion Transition: 220.968/163.000 Da

Acquired: 8/18/2010 4:23:38 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 279413
IS Ion Transition: 225.000/166.800 Da

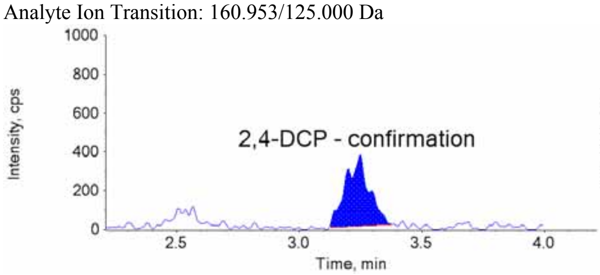
Figure 55. Typical Chromatogram of 2,4-D Analysis, Control Corn Aspirated Grain Fractions (141-0001)

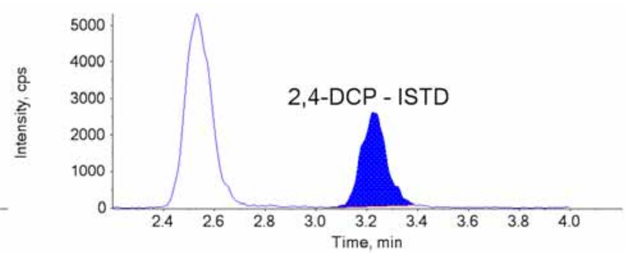




Sample Name: ARA-09-15-10-141-0001A4 + 0.01 µg/g
Data File: ARA-09-15-10 D19c.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 7184
Analyte Ion Transition: 160.953/125.000 Da

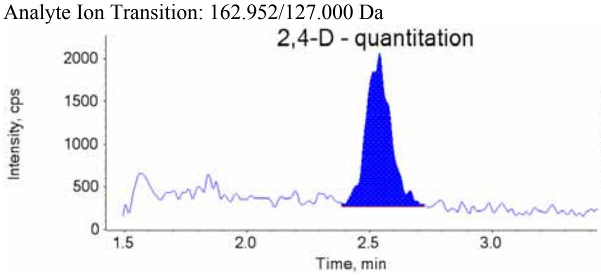
Acquired: 8/18/2010 9:14:51 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 17902
IS Ion Transition: 167.000/131.000 Da

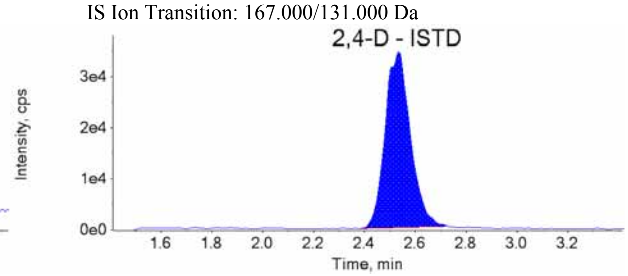




Sample Name: ARA-09-15-10-141-0001A4 + 0.01 µg/g
Data File: ARA-09-15-10 D19c.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 2444
Analyte Ion Transition: 162.952/127.000 Da

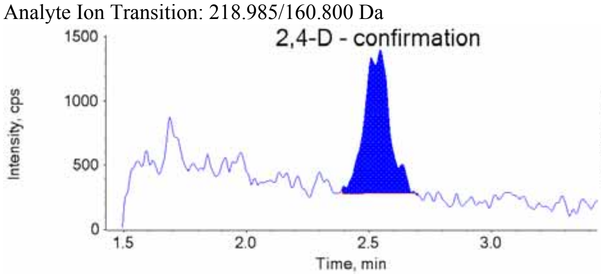
Acquired: 8/18/2010 9:14:51 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 17902
IS Ion Transition: 167.000/131.000 Da

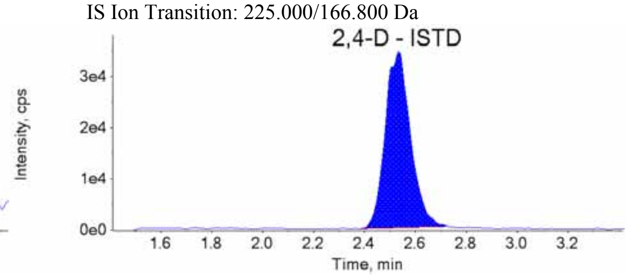




Sample Name: ARA-09-15-10-141-0001A4 + 0.01 µg/g
Data File: ARA-09-15-10 D19c.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 12467
Analyte Ion Transition: 218.985/160.800 Da

Acquired: 8/18/2010 9:14:51 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 237781
IS Ion Transition: 225.000/166.800 Da

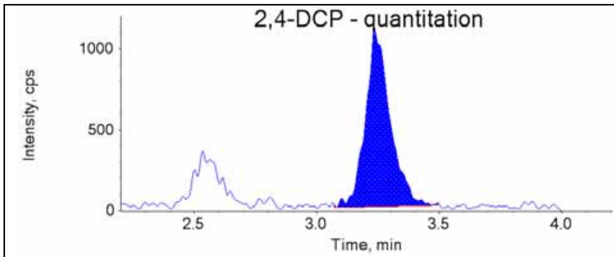


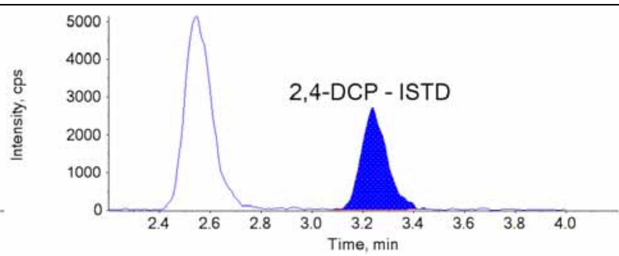


Sample Name: ARA-09-15-10-141-0001A4 + 0.01 µg/g
Data File: ARA-09-15-10 D19c.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 8105
Analyte Ion Transition: 220.968/163.000 Da

Acquired: 8/18/2010 9:14:51 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 237781
IS Ion Transition: 225.000/166.800 Da

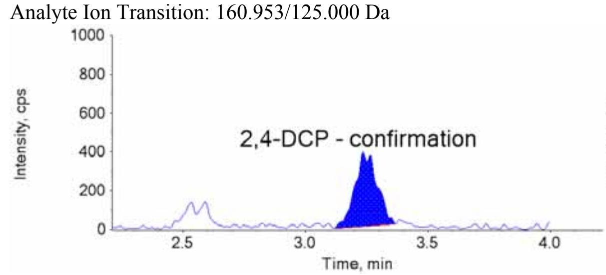
Figure 56. Typical Chromatogram of 2,4-D Analysis, Corn Aspirated Grain Fractions 0.01 µg/g Recovery

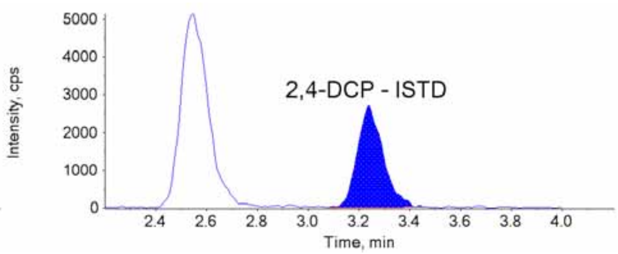




Sample Name: ARA-09-15-10-141-0001A5 + 0.01 µg/g
Data File: ARA-09-15-10 D19c.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 7671
Analyte Ion Transition: 160.953/125.000 Da

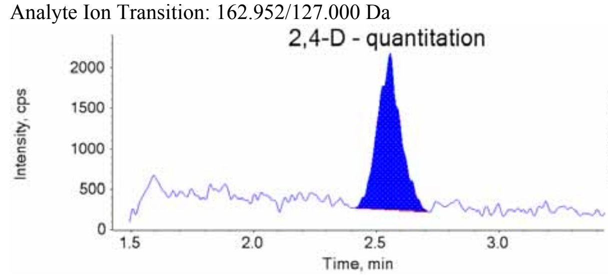
Acquired: 8/18/2010 9:21:19 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 18837
IS Ion Transition: 167.000/131.000 Da

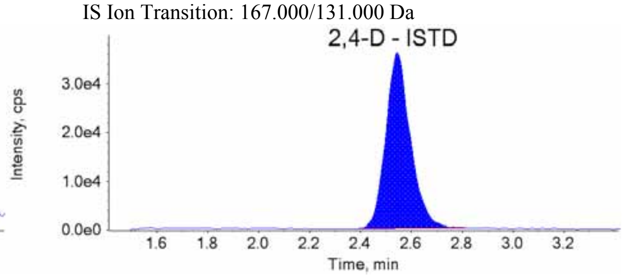




Sample Name: ARA-09-15-10-141-0001A5 + 0.01 µg/g
Data File: ARA-09-15-10 D19c.wiff
Analyte Integration Type: Base To Base
Analyte Peak Area: 2450
Analyte Ion Transition: 162.952/127.000 Da

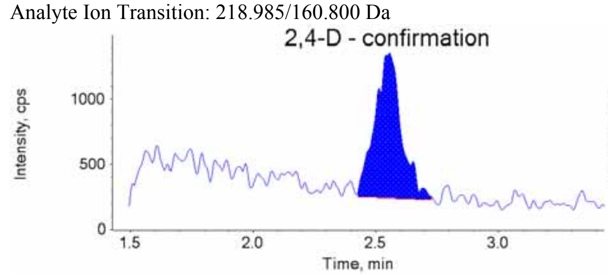
Acquired: 8/18/2010 9:21:19 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 18837
IS Ion Transition: 167.000/131.000 Da

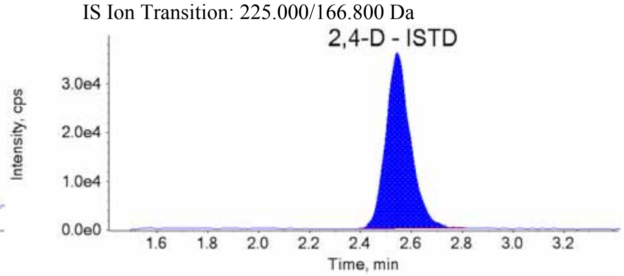




Sample Name: ARA-09-15-10-141-0001A5 + 0.01 µg/g
Data File: ARA-09-15-10 D19c.wiff
Analyte Integration Type: Base To Base
Analyte Peak Area: 12557
Analyte Ion Transition: 218.985/160.800 Da

Acquired: 8/18/2010 9:21:19 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 242006
IS Ion Transition: 225.000/166.800 Da

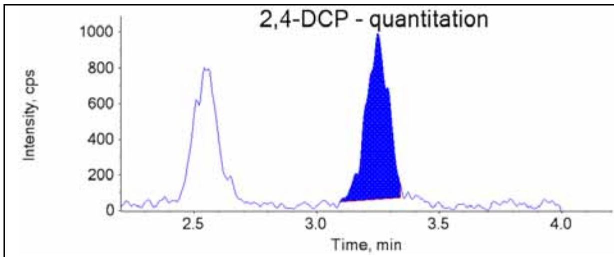


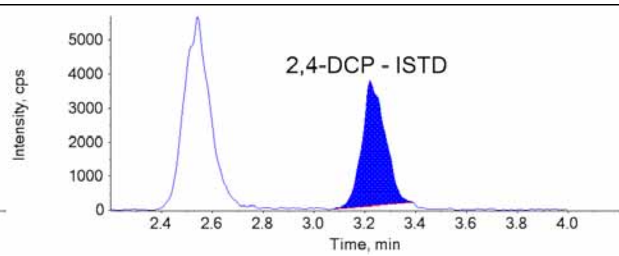


Sample Name: ARA-09-15-10-141-0001A5 + 0.01 µg/g
Data File: ARA-09-15-10 D19c.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 8453
Analyte Ion Transition: 220.968/163.000 Da

Acquired: 8/18/2010 9:21:19 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 242006
IS Ion Transition: 225.000/166.800 Da

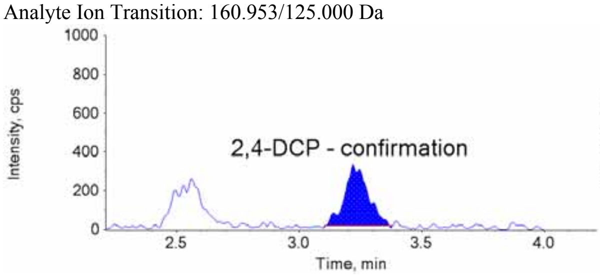
Figure 57. Typical Chromatogram of 2,4-D Analysis, Corn Aspirated Grain Fractions 0.01 µg/g Recovery

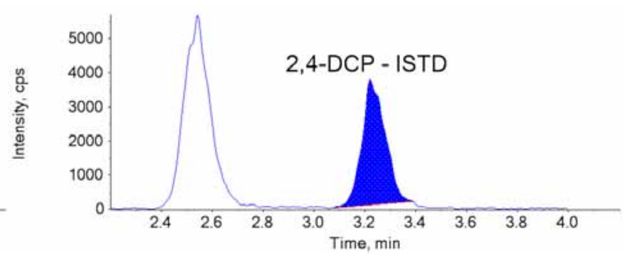




Sample Name: ARA-09-15-10-141-0002A2
Data File: ARA-09-15-10 D19c.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 5890
Analyte Ion Transition: 160.953/125.000 Da

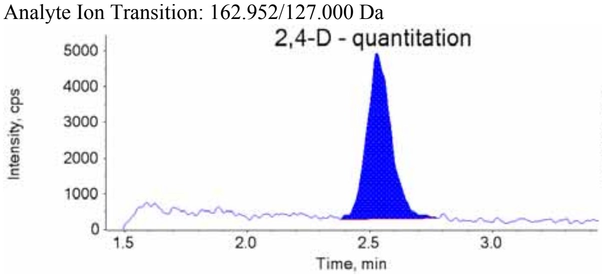
Acquired: 8/18/2010 4:30:09 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 23316
IS Ion Transition: 167.000/131.000 Da

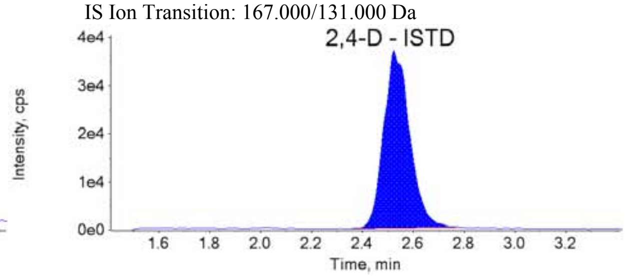




Sample Name: ARA-09-15-10-141-0002A2
Data File: ARA-09-15-10 D19c.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 2060
Analyte Ion Transition: 162.952/127.000 Da

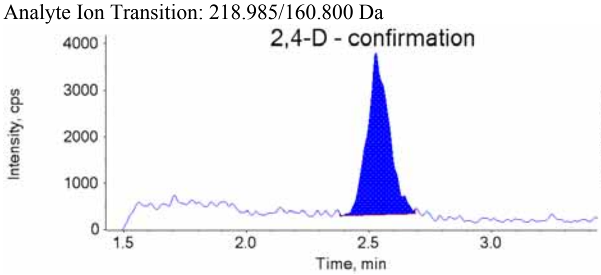
Acquired: 8/18/2010 4:30:09 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 23316
IS Ion Transition: 167.000/131.000 Da

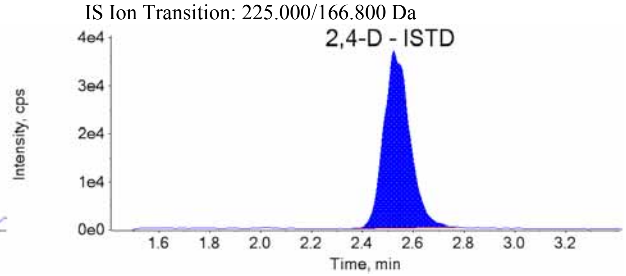




Sample Name: ARA-09-15-10-141-0002A2
Data File: ARA-09-15-10 D19c.wiff
Analyte Integration Type: Base To Base
Analyte Peak Area: 32324
Analyte Ion Transition: 218.985/160.800 Da

Acquired: 8/18/2010 4:30:09 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 262099
IS Ion Transition: 225.000/166.800 Da

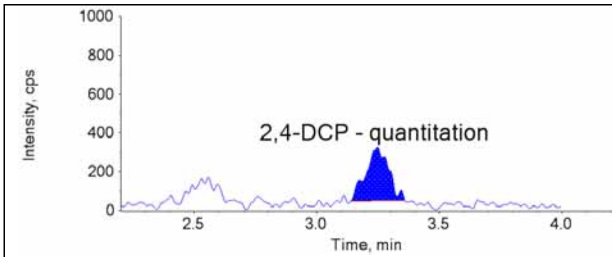


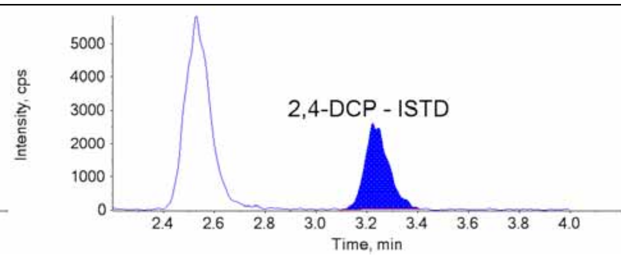


Sample Name: ARA-09-15-10-141-0002A2
Data File: ARA-09-15-10 D19c.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 21580
Analyte Ion Transition: 220.968/163.000 Da

Acquired: 8/18/2010 4:30:09 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 262099
IS Ion Transition: 225.000/166.800 Da

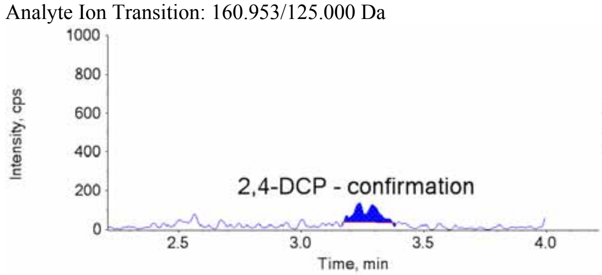
Figure 58. Typical Chromatogram of 2,4-D Analysis, Treated Corn Aspirated Grain Fractions (141-0002)

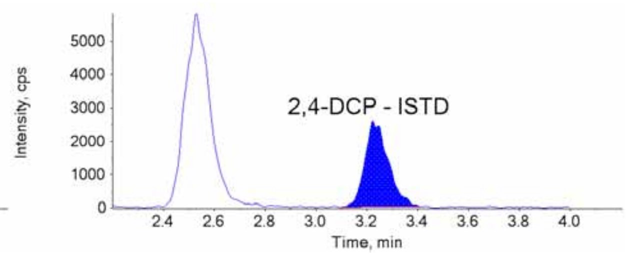




Sample Name: ARA-09-15-10-150-0002A2
Data File: ARA-09-15-10 D19c.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 1783
Analyte Ion Transition: 160.953/125.000 Da

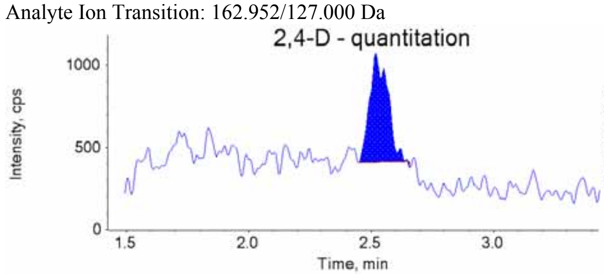
Acquired: 8/18/2010 4:43:04 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 16581
IS Ion Transition: 167.000/131.000 Da

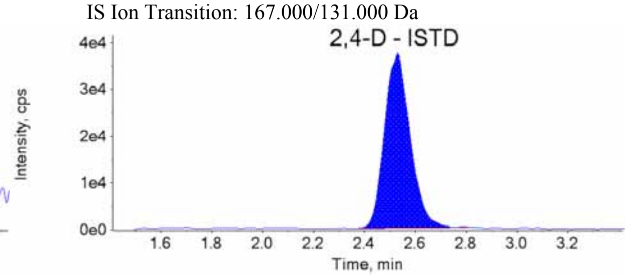




Sample Name: ARA-09-15-10-150-0002A2
Data File: ARA-09-15-10 D19c.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 608
Analyte Ion Transition: 162.952/127.000 Da

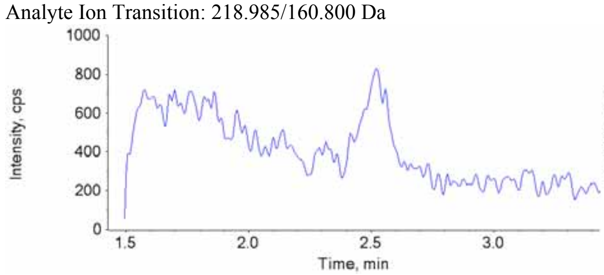
Acquired: 8/18/2010 4:43:04 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 16581
IS Ion Transition: 167.000/131.000 Da

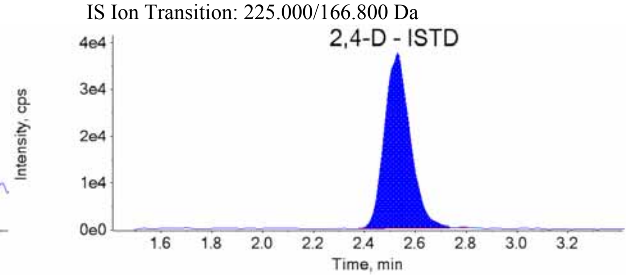




Sample Name: ARA-09-15-10-150-0002A2
Data File: ARA-09-15-10 D19c.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 3404
Analyte Ion Transition: 218.985/160.800 Da

Acquired: 8/18/2010 4:43:04 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 261343
IS Ion Transition: 225.000/166.800 Da

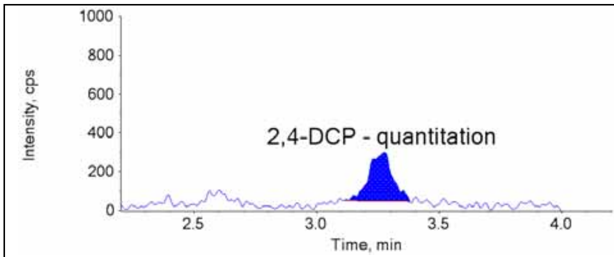


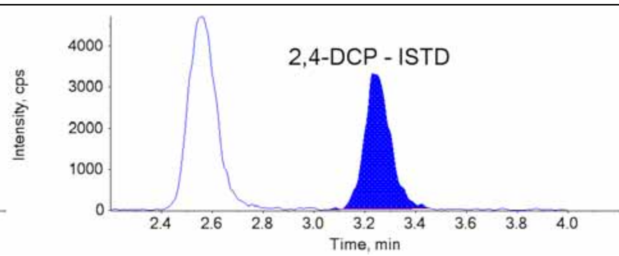


Sample Name: ARA-09-15-10-150-0002A2
Data File: ARA-09-15-10 D19c.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 0
Analyte Ion Transition: 220.968/163.000 Da

Acquired: 8/18/2010 4:43:04 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 261343
IS Ion Transition: 225.000/166.800 Da

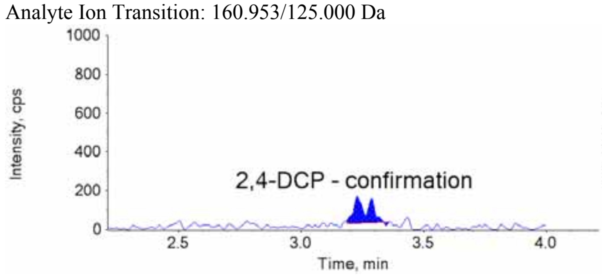
Figure 59. Typical Chromatogram of 2,4-D Analysis, Treated Corn Aspirated Grain Fractions (150-0002)

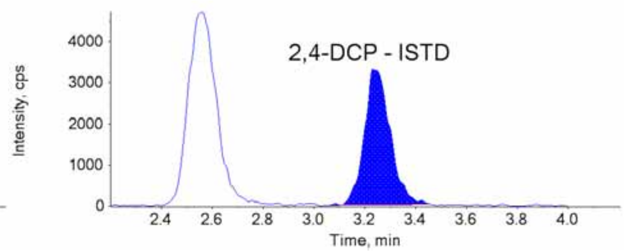




Sample Name: ARA-09-15-10-145-0001A22
Data File: ARA-09-15-10 D19c.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 1723
Analyte Ion Transition: 160.953/125.000 Da

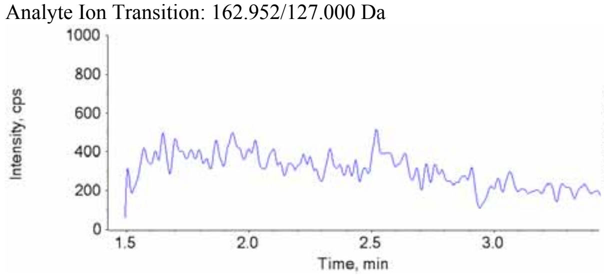
Acquired: 8/18/2010 5:47:56 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 23418
IS Ion Transition: 167.000/131.000 Da

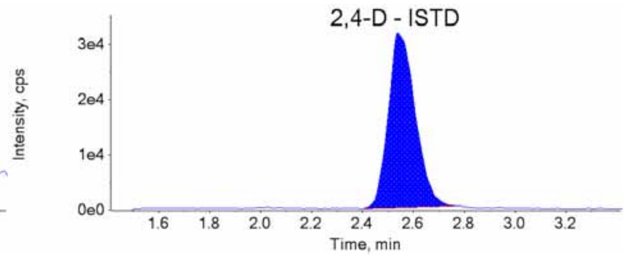




Sample Name: ARA-09-15-10-145-0001A22
Data File: ARA-09-15-10 D19c.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 602
Analyte Ion Transition: 162.952/127.000 Da

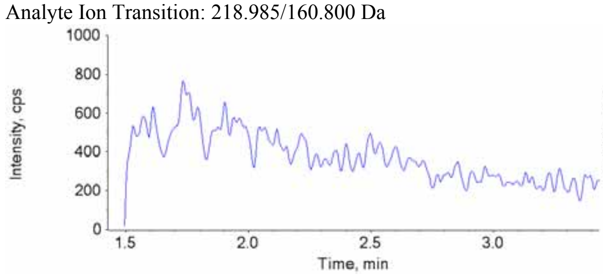
Acquired: 8/18/2010 5:47:56 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 23418
IS Ion Transition: 167.000/131.000 Da

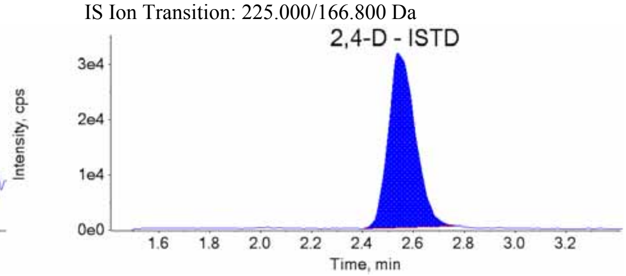




Sample Name: ARA-09-15-10-145-0001A22
Data File: ARA-09-15-10 D19c.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 0
Analyte Ion Transition: 218.985/160.800 Da

Acquired: 8/18/2010 5:47:56 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 233741
IS Ion Transition: 225.000/166.800 Da

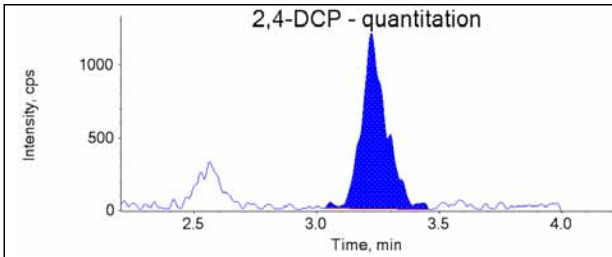


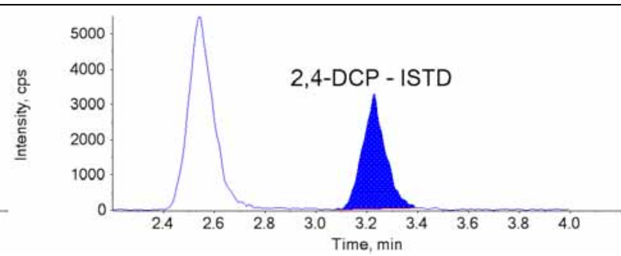


Sample Name: ARA-09-15-10-145-0001A22
Data File: ARA-09-15-10 D19c.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 0
Analyte Ion Transition: 220.968/163.000 Da

Acquired: 8/18/2010 5:47:56 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 233741
IS Ion Transition: 225.000/166.800 Da

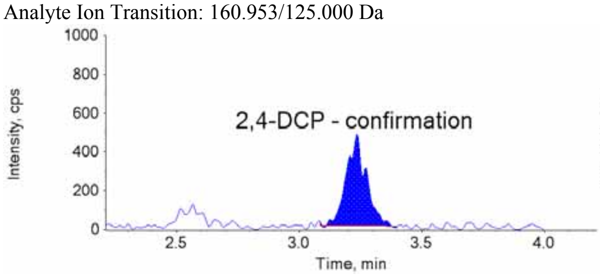
Figure 60. Typical Chromatogram of 2,4-D Analysis, Control Corn Grits (145-0001)

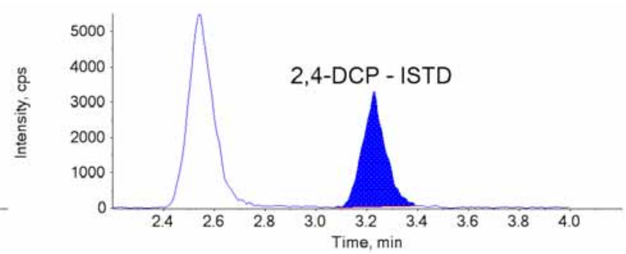




Sample Name: ARA-09-15-10-145-0001A24 + 0.01 µg/g
Data File: ARA-09-15-10 D19c.wiff
Analyte Integration Type: Base To Base
Analyte Peak Area: 8381
Analyte Ion Transition: 160.953/125.000 Da

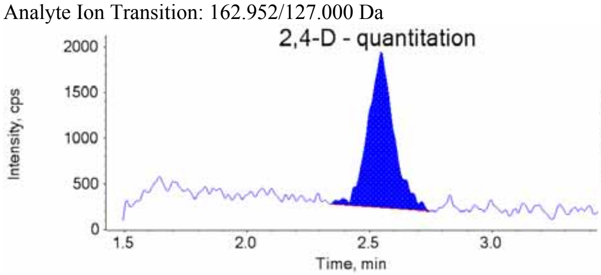
Acquired: 8/18/2010 9:47:10 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 20063
IS Ion Transition: 167.000/131.000 Da

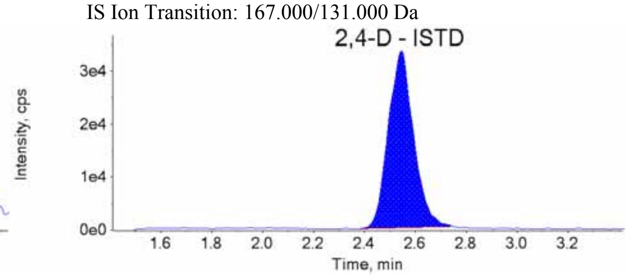




Sample Name: ARA-09-15-10-145-0001A24 + 0.01 µg/g
Data File: ARA-09-15-10 D19c.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 2657
Analyte Ion Transition: 162.952/127.000 Da

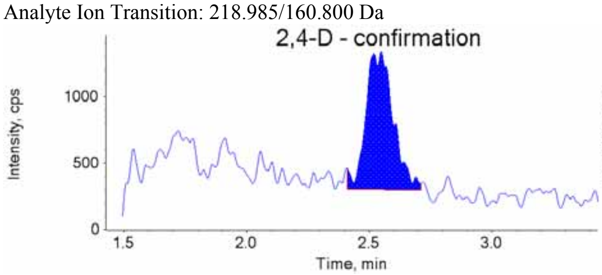
Acquired: 8/18/2010 9:47:10 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 20063
IS Ion Transition: 167.000/131.000 Da

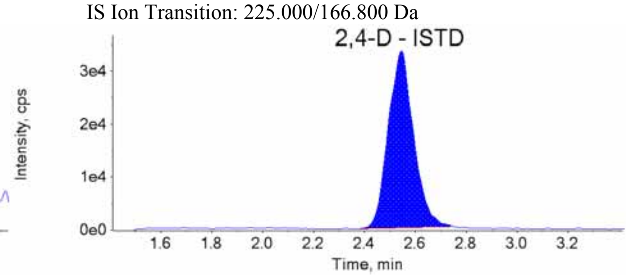




Sample Name: ARA-09-15-10-145-0001A24 + 0.01 µg/g
Data File: ARA-09-15-10 D19c.wiff
Analyte Integration Type: Base To Base
Analyte Peak Area: 12482
Analyte Ion Transition: 218.985/160.800 Da

Acquired: 8/18/2010 9:47:10 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 230486
IS Ion Transition: 225.000/166.800 Da

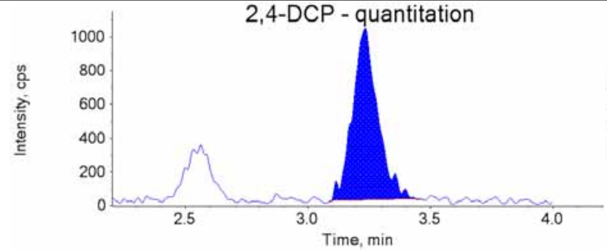


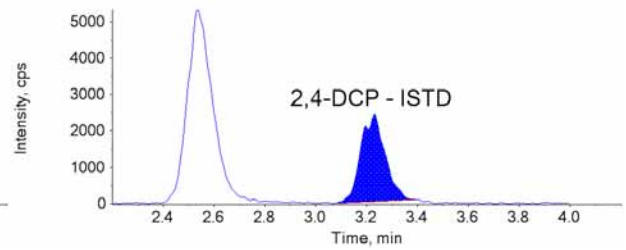


Sample Name: ARA-09-15-10-145-0001A24 + 0.01 µg/g
Data File: ARA-09-15-10 D19c.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 8142
Analyte Ion Transition: 220.968/163.000 Da

Acquired: 8/18/2010 9:47:10 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 230486
IS Ion Transition: 225.000/166.800 Da

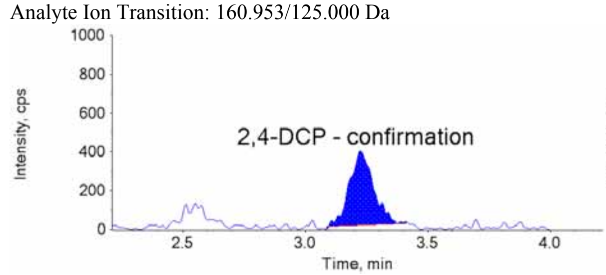
Figure 61. Typical Chromatogram of 2,4-D Analysis, Corn Grits 0.01 µg/g Recovery

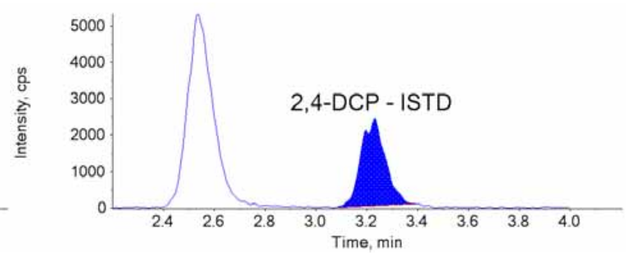




Sample Name: ARA-09-15-10-145-0001A25 + 0.01 µg/g
Data File: ARA-09-15-10 D19c.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 7305
Analyte Ion Transition: 160.953/125.000 Da

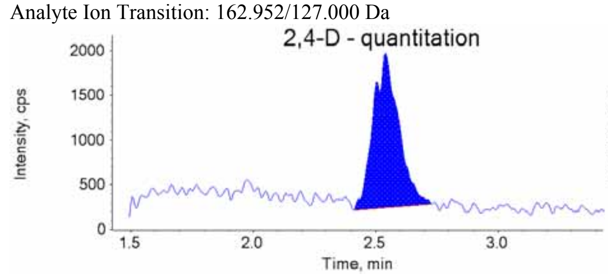
Acquired: 8/18/2010 9:53:37 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 16009
IS Ion Transition: 167.000/131.000 Da

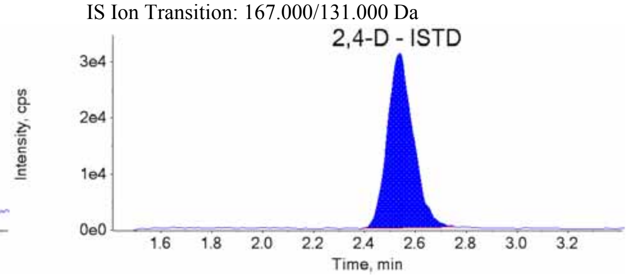




Sample Name: ARA-09-15-10-145-0001A25 + 0.01 µg/g
Data File: ARA-09-15-10 D19c.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 2626
Analyte Ion Transition: 162.952/127.000 Da

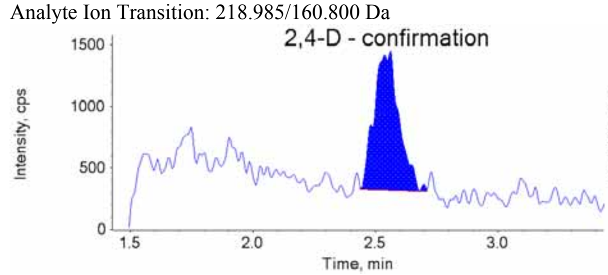
Acquired: 8/18/2010 9:53:37 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 16009
IS Ion Transition: 167.000/131.000 Da

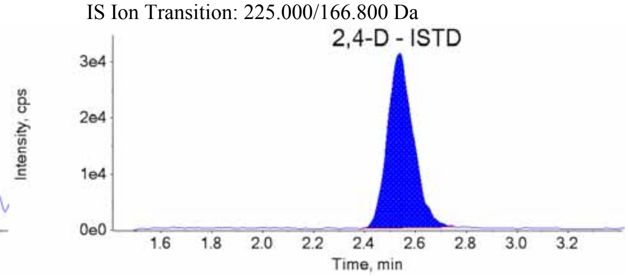




Sample Name: ARA-09-15-10-145-0001A25 + 0.01 µg/g
Data File: ARA-09-15-10 D19c.wiff
Analyte Integration Type: Base To Base
Analyte Peak Area: 12603
Analyte Ion Transition: 218.985/160.800 Da

Acquired: 8/18/2010 9:53:37 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 219091
IS Ion Transition: 225.000/166.800 Da

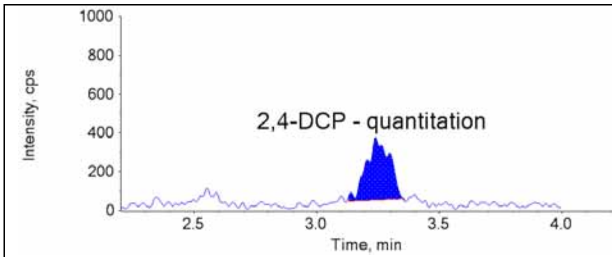


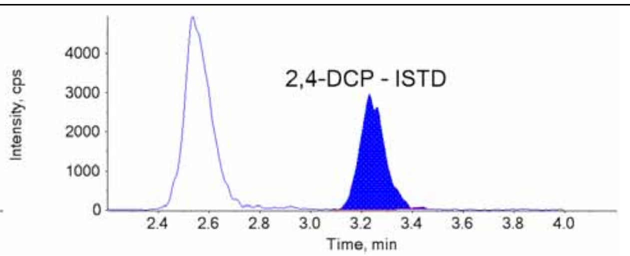


Sample Name: ARA-09-15-10-145-0001A25 + 0.01 µg/g
Data File: ARA-09-15-10 D19c.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 8044
Analyte Ion Transition: 220.968/163.000 Da

Acquired: 8/18/2010 9:53:37 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 219091
IS Ion Transition: 225.000/166.800 Da

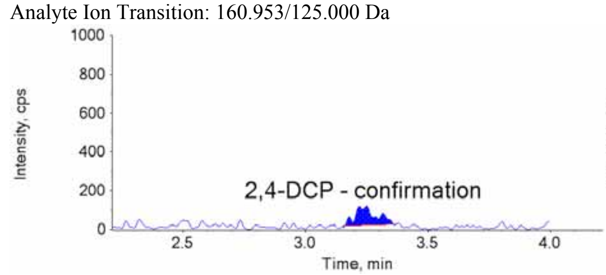
Figure 62. Typical Chromatogram of 2,4-D Analysis, Corn Grits 0.01 µg/g Recovery

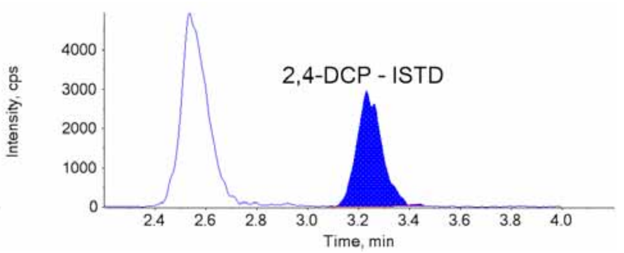




Sample Name: ARA-09-15-10-145-0003A2
Data File: ARA-09-15-10 D19c.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 2128
Analyte Ion Transition: 160.953/125.000 Da

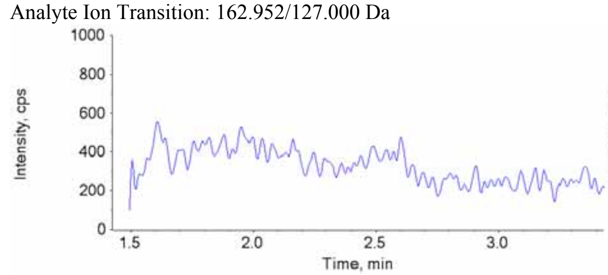
Acquired: 8/18/2010 6:00:52 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 20103
IS Ion Transition: 167.000/131.000 Da

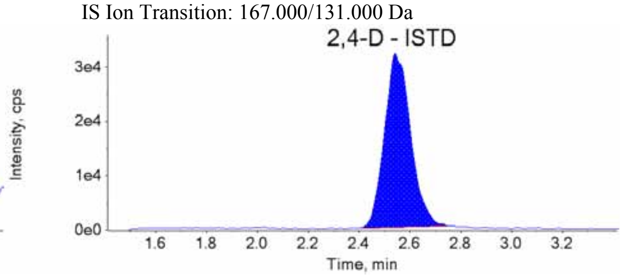




Sample Name: ARA-09-15-10-145-0003A2
Data File: ARA-09-15-10 D19c.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 582
Analyte Ion Transition: 162.952/127.000 Da

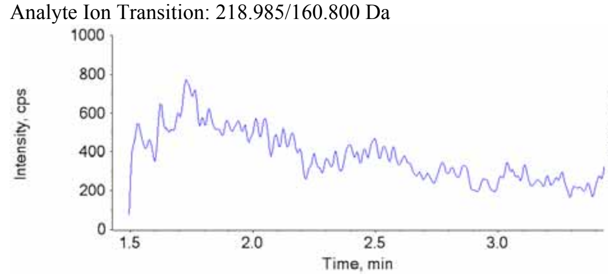
Acquired: 8/18/2010 6:00:52 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 20103
IS Ion Transition: 167.000/131.000 Da

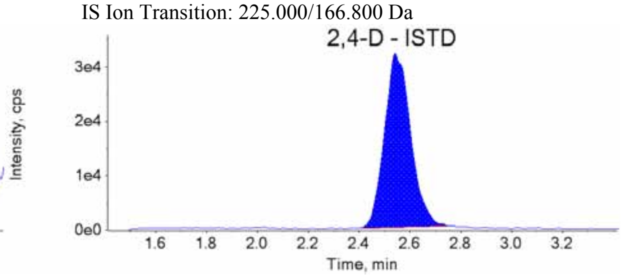




Sample Name: ARA-09-15-10-145-0003A2
Data File: ARA-09-15-10 D19c.wiff
Analyte Integration Type: No Peak
Analyte Peak Area: 0
Analyte Ion Transition: 218.985/160.800 Da

Acquired: 8/18/2010 6:00:52 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 229432
IS Ion Transition: 225.000/166.800 Da

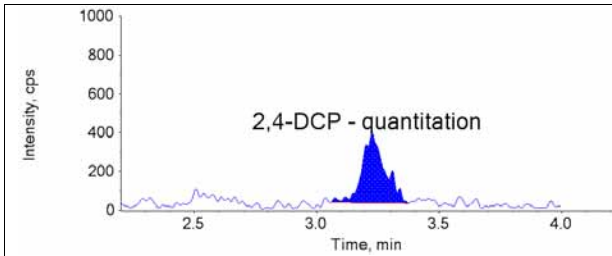


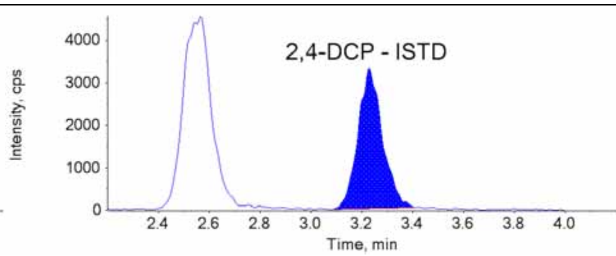


Sample Name: ARA-09-15-10-145-0003A2
Data File: ARA-09-15-10 D19c.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 0
Analyte Ion Transition: 220.968/163.000 Da

Acquired: 8/18/2010 6:00:52 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 229432
IS Ion Transition: 225.000/166.800 Da

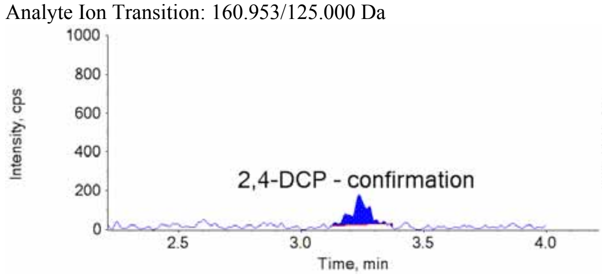
Figure 63. Typical Chromatogram of 2,4-D Analysis, Treated Corn Grits (145-0003)

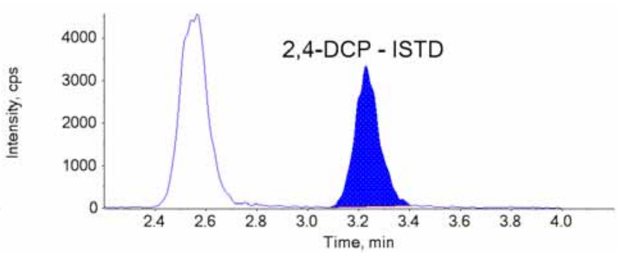




Sample Name: ARA-09-15-10-145-0004A2
Data File: ARA-09-15-10 D19c.wiff
Analyte Integration Type: Base To Base
Analyte Peak Area: 2320
Analyte Ion Transition: 160.953/125.000 Da

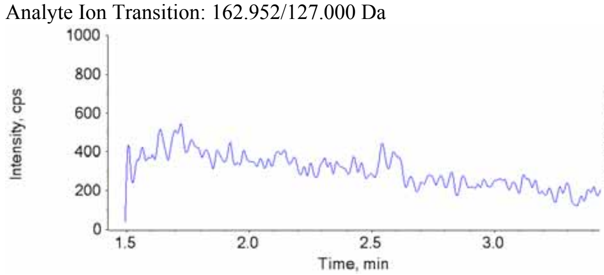
Acquired: 8/18/2010 6:07:21 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 22037
IS Ion Transition: 167.000/131.000 Da

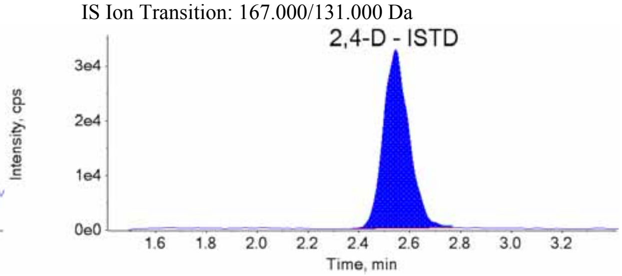




Sample Name: ARA-09-15-10-145-0004A2
Data File: ARA-09-15-10 D19c.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 696
Analyte Ion Transition: 162.952/127.000 Da

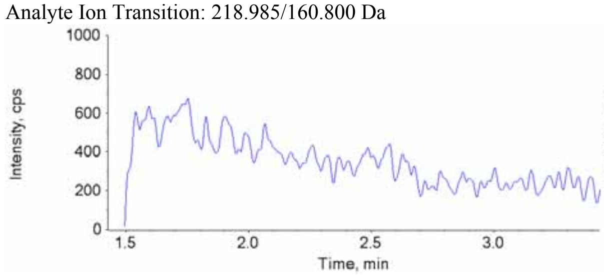
Acquired: 8/18/2010 6:07:21 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 22037
IS Ion Transition: 167.000/131.000 Da

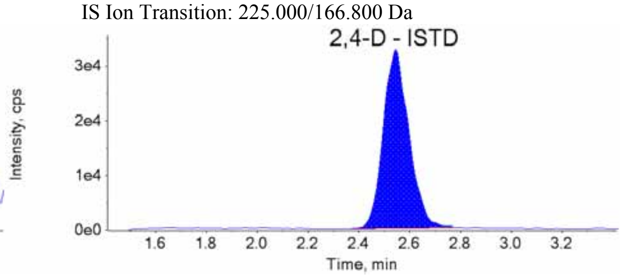




Sample Name: ARA-09-15-10-145-0004A2
Data File: ARA-09-15-10 D19c.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 0
Analyte Ion Transition: 218.985/160.800 Da

Acquired: 8/18/2010 6:07:21 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 229652
IS Ion Transition: 225.000/166.800 Da

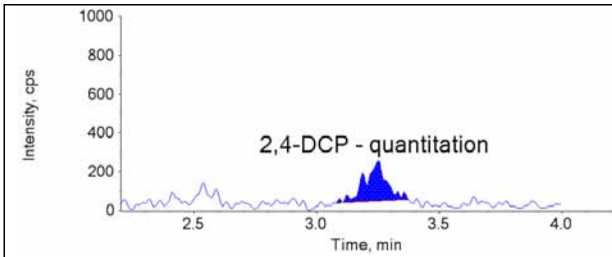


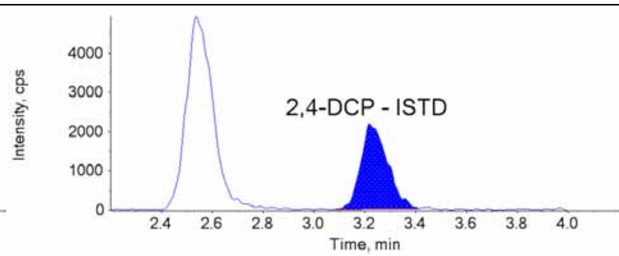


Sample Name: ARA-09-15-10-145-0004A2
Data File: ARA-09-15-10 D19c.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 0
Analyte Ion Transition: 220.968/163.000 Da

Acquired: 8/18/2010 6:07:21 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 229652
IS Ion Transition: 225.000/166.800 Da

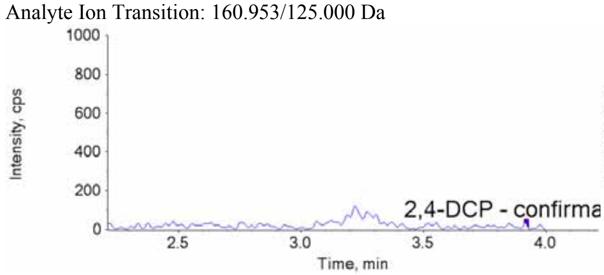
Figure 64. Typical Chromatogram of 2,4-D Analysis, Treated Corn Grits (145-0004)

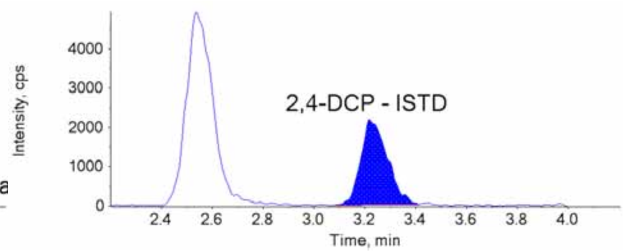




Sample Name: ARA-09-15-10-144-0001A22
Data File: ARA-09-15-10 D19c.wiff
Analyte Integration Type: Base To Base
Analyte Peak Area: 1292
Analyte Ion Transition: 160.953/125.000 Da

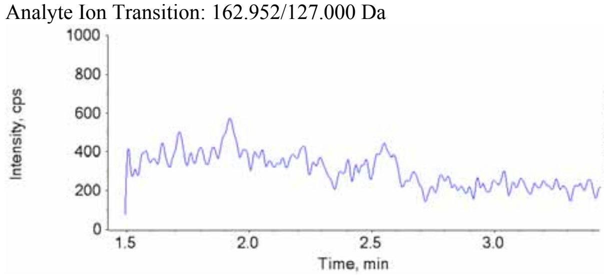
Acquired: 8/18/2010 6:46:11 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 15916
IS Ion Transition: 167.000/131.000 Da

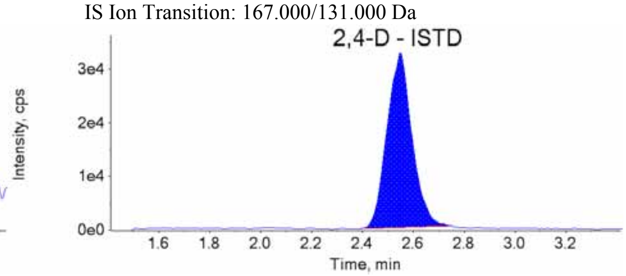




Sample Name: ARA-09-15-10-144-0001A22
Data File: ARA-09-15-10 D19c.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 0
Analyte Ion Transition: 162.952/127.000 Da

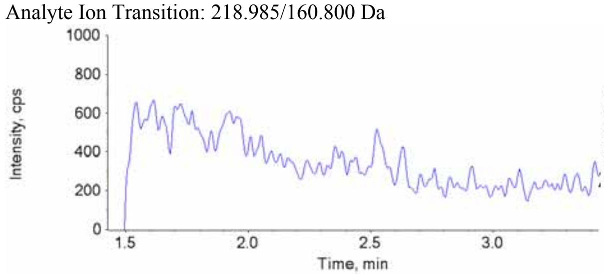
Acquired: 8/18/2010 6:46:11 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 15916
IS Ion Transition: 167.000/131.000 Da

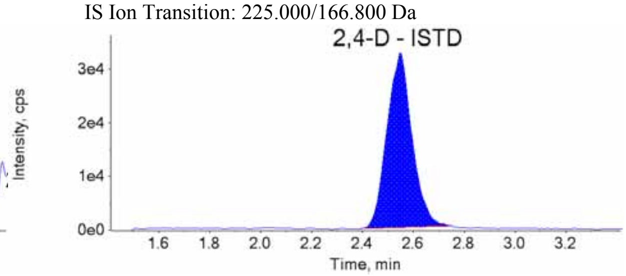




Sample Name: ARA-09-15-10-144-0001A22
Data File: ARA-09-15-10 D19c.wiff
Analyte Integration Type: No Peak
Analyte Peak Area: 0
Analyte Ion Transition: 218.985/160.800 Da

Acquired: 8/18/2010 6:46:11 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 221073
IS Ion Transition: 225.000/166.800 Da

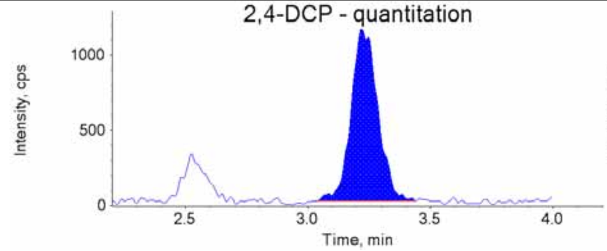


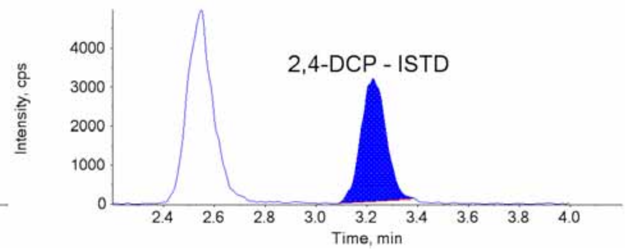


Sample Name: ARA-09-15-10-144-0001A22
Data File: ARA-09-15-10 D19c.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 0
Analyte Ion Transition: 220.968/163.000 Da

Acquired: 8/18/2010 6:46:11 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 221073
IS Ion Transition: 225.000/166.800 Da

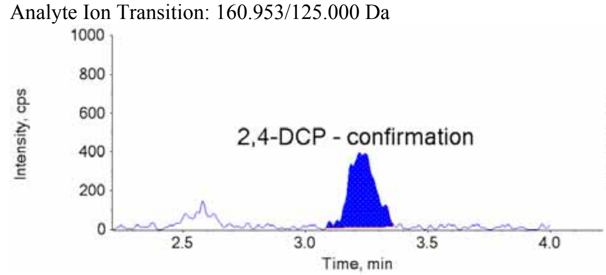
Figure 65. Typical Chromatogram of 2,4-D Analysis, Control Corn Meal (144-0001)

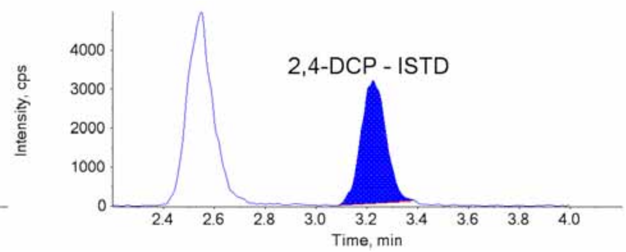




Sample Name: ARA-09-15-10-144-0001A24 + 0.01 µg/g
Data File: ARA-09-15-10 D19c.wiff
Analyte Integration Type: Base To Base
Analyte Peak Area: 8765
Analyte Ion Transition: 160.953/125.000 Da

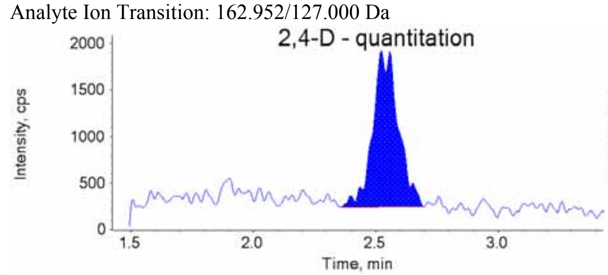
Acquired: 8/18/2010 10:00:05 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 21665
IS Ion Transition: 167.000/131.000 Da

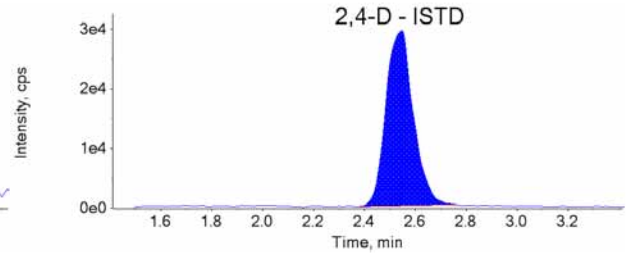




Sample Name: ARA-09-15-10-144-0001A24 + 0.01 µg/g
Data File: ARA-09-15-10 D19c.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 2940
Analyte Ion Transition: 162.952/127.000 Da

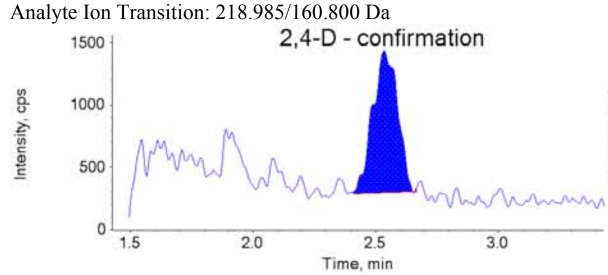
Acquired: 8/18/2010 10:00:05 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 21665
IS Ion Transition: 167.000/131.000 Da

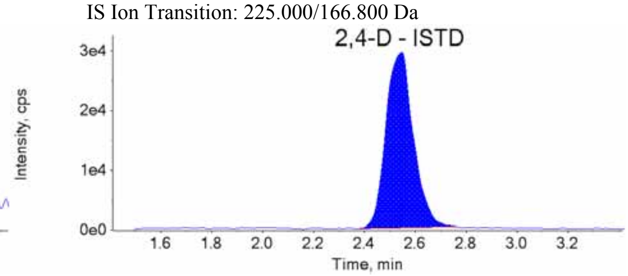




Sample Name: ARA-09-15-10-144-0001A24 + 0.01 µg/g
Data File: ARA-09-15-10 D19c.wiff
Analyte Integration Type: Base To Base
Analyte Peak Area: 12333
Analyte Ion Transition: 218.985/160.800 Da

Acquired: 8/18/2010 10:00:05 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 220343
IS Ion Transition: 225.000/166.800 Da

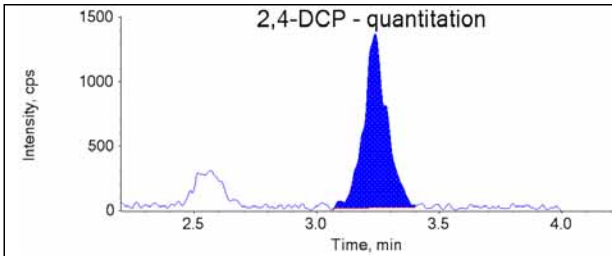


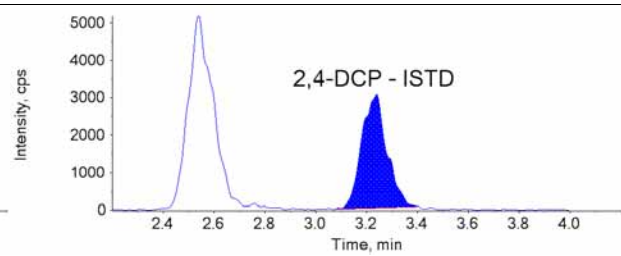


Sample Name: ARA-09-15-10-144-0001A24 + 0.01 µg/g
Data File: ARA-09-15-10 D19c.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 8154
Analyte Ion Transition: 220.968/163.000 Da

Acquired: 8/18/2010 10:00:05 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 220343
IS Ion Transition: 225.000/166.800 Da

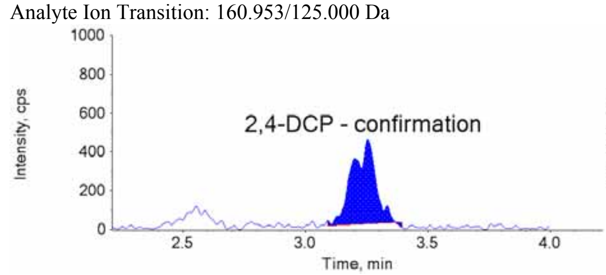
Figure 66. Typical Chromatogram of 2,4-D Analysis, Corn Meal 0.01 µg/g Recovery

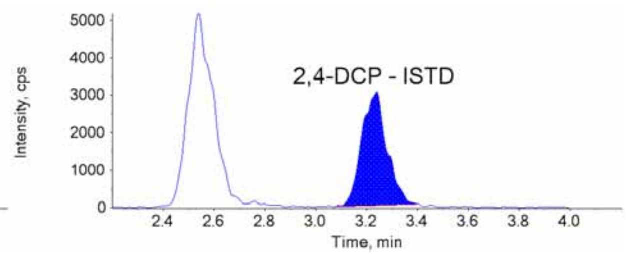




Sample Name: ARA-09-15-10-144-0001A25 + 0.01 µg/g
Data File: ARA-09-15-10 D19c.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 8942
Analyte Ion Transition: 160.953/125.000 Da

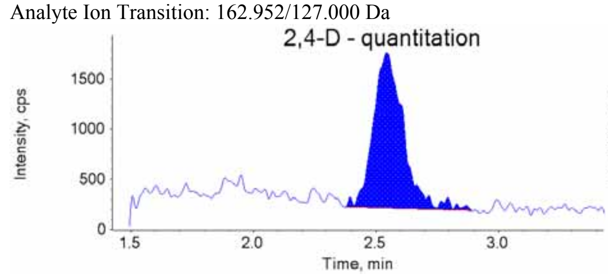
Acquired: 8/18/2010 10:06:33 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 20939
IS Ion Transition: 167.000/131.000 Da

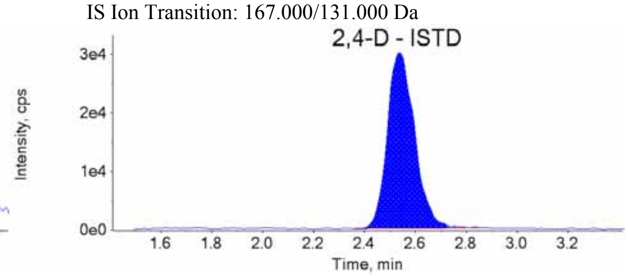




Sample Name: ARA-09-15-10-144-0001A25 + 0.01 µg/g
Data File: ARA-09-15-10 D19c.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 2946
Analyte Ion Transition: 162.952/127.000 Da

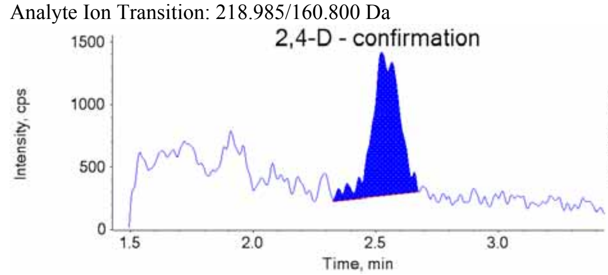
Acquired: 8/18/2010 10:06:33 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 20939
IS Ion Transition: 167.000/131.000 Da

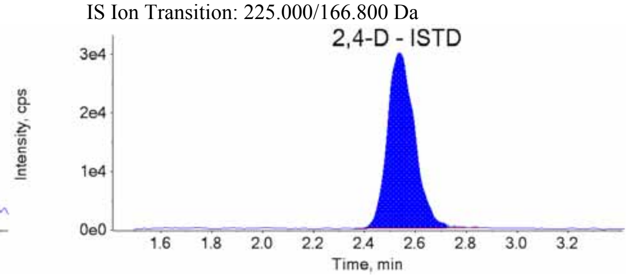




Sample Name: ARA-09-15-10-144-0001A25 + 0.01 µg/g
Data File: ARA-09-15-10 D19c.wiff
Analyte Integration Type: Base To Base
Analyte Peak Area: 13379
Analyte Ion Transition: 218.985/160.800 Da

Acquired: 8/18/2010 10:06:33 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 223536
IS Ion Transition: 225.000/166.800 Da

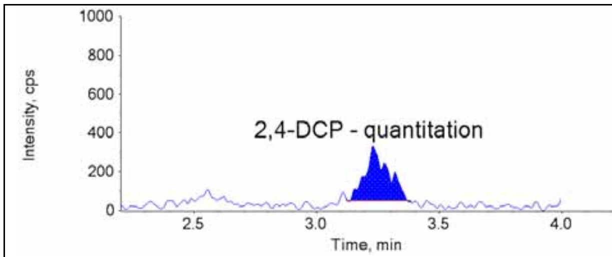


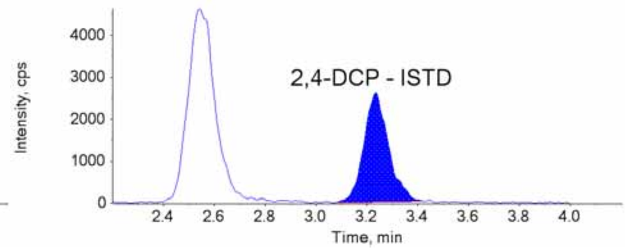


Sample Name: ARA-09-15-10-144-0001A25 + 0.01 µg/g
Data File: ARA-09-15-10 D19c.wiff
Analyte Integration Type: Valley
Analyte Peak Area: 8977
Analyte Ion Transition: 220.968/163.000 Da

Acquired: 8/18/2010 10:06:33 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 223536
IS Ion Transition: 225.000/166.800 Da

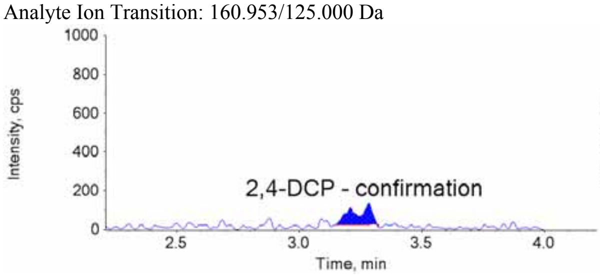
Figure 67. Typical Chromatogram of 2,4-D Analysis, Corn Meal 0.01 µg/g Recovery

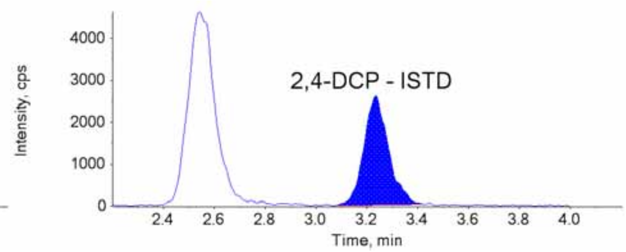




Sample Name: ARA-09-15-10-144-0003A2
Data File: ARA-09-15-10 D19c.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 1813
Analyte Ion Transition: 160.953/125.000 Da

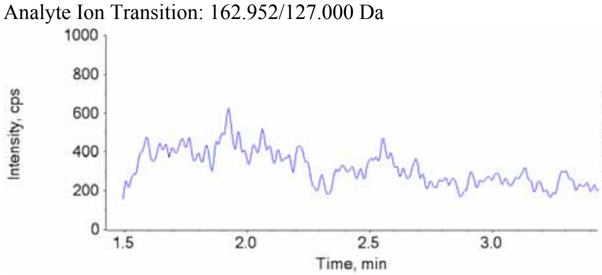
Acquired: 8/18/2010 6:59:07 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 17571
IS Ion Transition: 167.000/131.000 Da

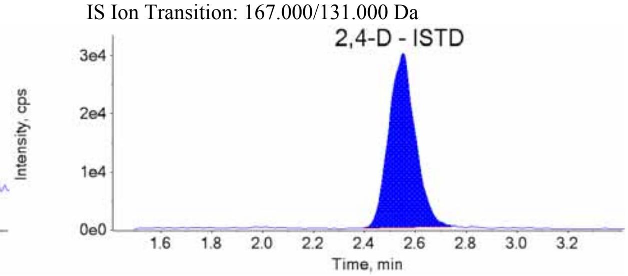




Sample Name: ARA-09-15-10-144-0003A2
Data File: ARA-09-15-10 D19c.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 567
Analyte Ion Transition: 162.952/127.000 Da

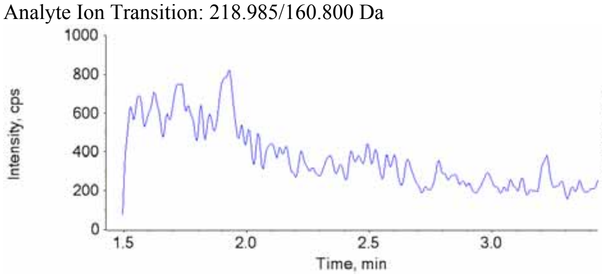
Acquired: 8/18/2010 6:59:07 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 17571
IS Ion Transition: 167.000/131.000 Da

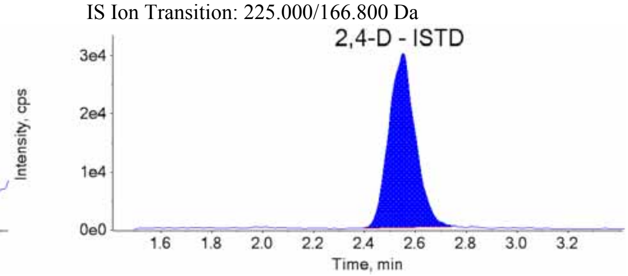




Sample Name: ARA-09-15-10-144-0003A2
Data File: ARA-09-15-10 D19c.wiff
Analyte Integration Type: No Peak
Analyte Peak Area: 0
Analyte Ion Transition: 218.985/160.800 Da

Acquired: 8/18/2010 6:59:07 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 216864
IS Ion Transition: 225.000/166.800 Da

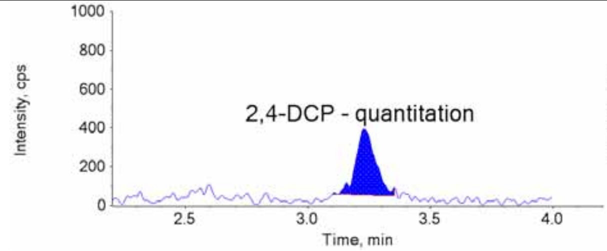


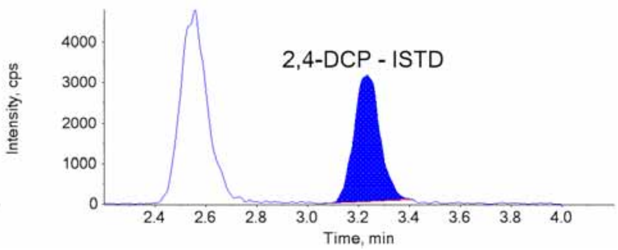


Sample Name: ARA-09-15-10-144-0003A2
Data File: ARA-09-15-10 D19c.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 0
Analyte Ion Transition: 220.968/163.000 Da

Acquired: 8/18/2010 6:59:07 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 216864
IS Ion Transition: 225.000/166.800 Da

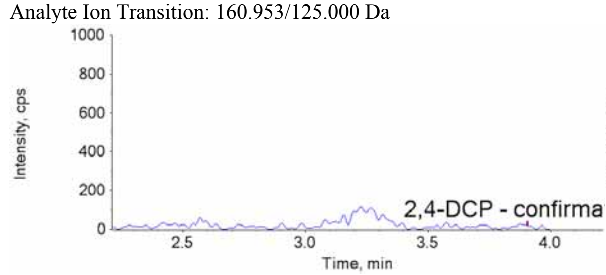
Figure 68. Typical Chromatogram of 2,4-D Analysis, Treated Corn Meal (144-0003)

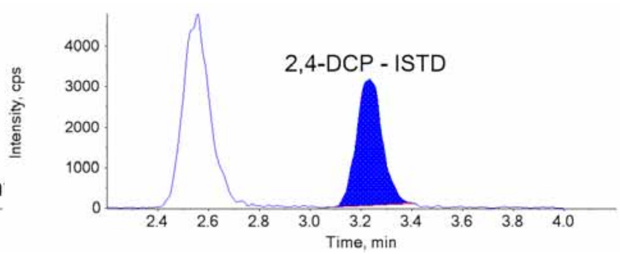




Sample Name: ARA-09-15-10-144-0004A2
Data File: ARA-09-15-10 D19c.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 1894
Analyte Ion Transition: 160.953/125.000 Da

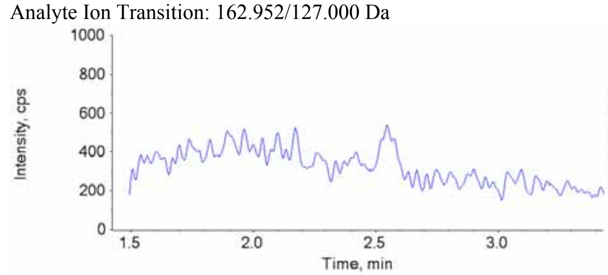
Acquired: 8/18/2010 7:05:35 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 22253
IS Ion Transition: 167.000/131.000 Da

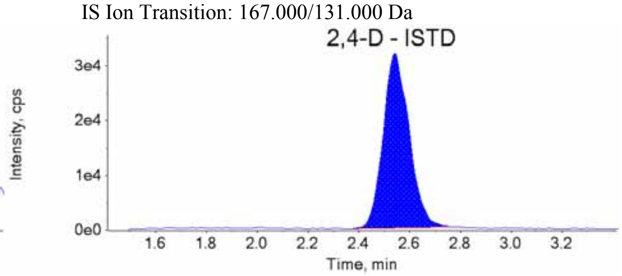




Sample Name: ARA-09-15-10-144-0004A2
Data File: ARA-09-15-10 D19c.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 0
Analyte Ion Transition: 162.952/127.000 Da

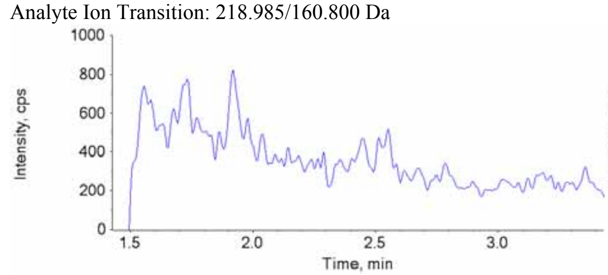
Acquired: 8/18/2010 7:05:35 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 22253
IS Ion Transition: 167.000/131.000 Da

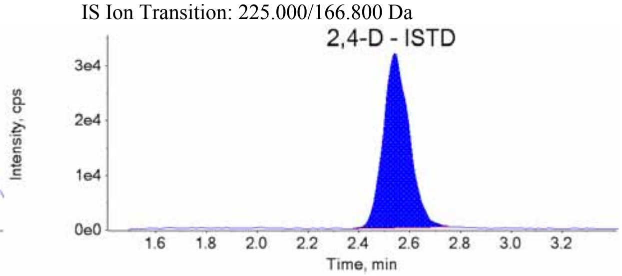




Sample Name: ARA-09-15-10-144-0004A2
Data File: ARA-09-15-10 D19c.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 0
Analyte Ion Transition: 218.985/160.800 Da

Acquired: 8/18/2010 7:05:35 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 226225
IS Ion Transition: 225.000/166.800 Da

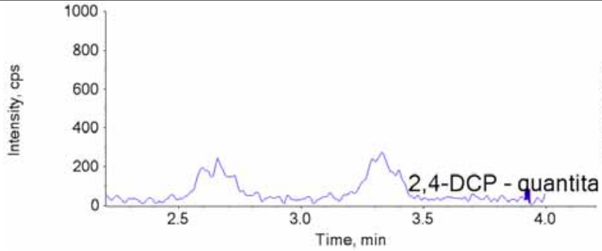


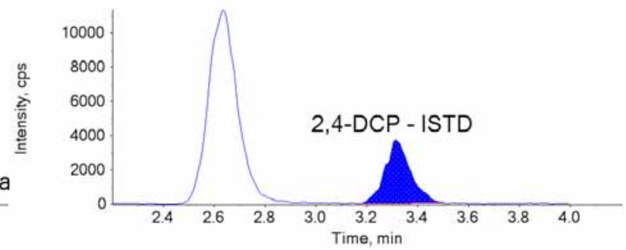


Sample Name: ARA-09-15-10-144-0004A2
Data File: ARA-09-15-10 D19c.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 0
Analyte Ion Transition: 220.968/163.000 Da

Acquired: 8/18/2010 7:05:35 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 226225
IS Ion Transition: 225.000/166.800 Da

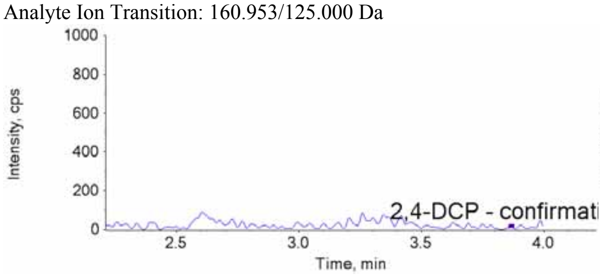
Figure 69. Typical Chromatogram of 2,4-D Analysis, Treated Corn Meal (144-0004)

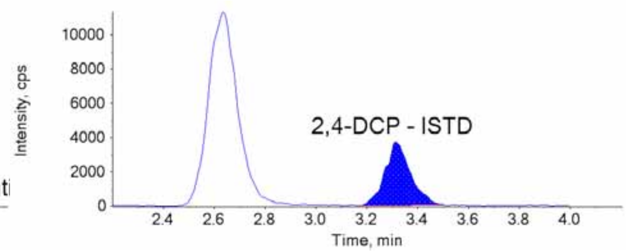




Sample Name: ARA-09-15-10-143-0001A30
Data File: ARA-09-15-10 D20.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 0
Analyte Ion Transition: 160.953/125.000 Da

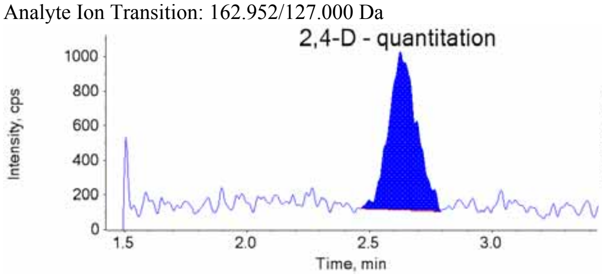
Acquired: 8/13/2010 5:19:37 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 25816
IS Ion Transition: 167.000/131.000 Da

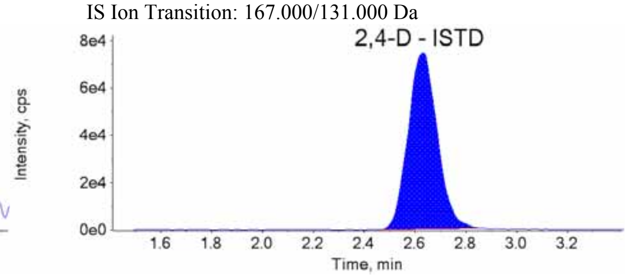




Sample Name: ARA-09-15-10-143-0001A30
Data File: ARA-09-15-10 D20.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 0
Analyte Ion Transition: 162.952/127.000 Da

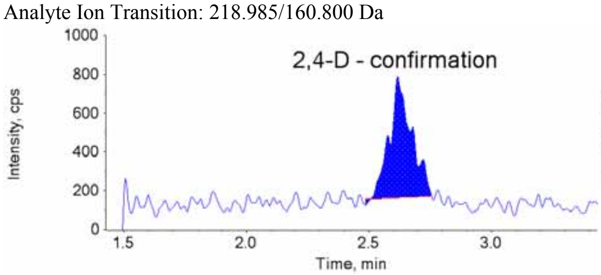
Acquired: 8/13/2010 5:19:37 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 25816
IS Ion Transition: 167.000/131.000 Da

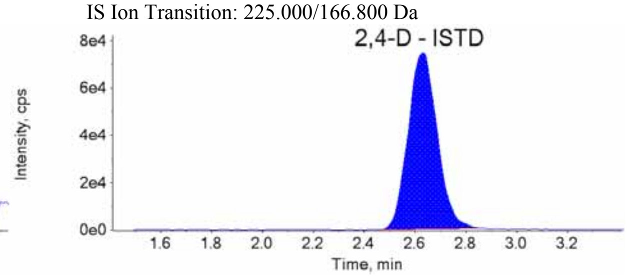




Sample Name: ARA-09-15-10-143-0001A30
Data File: ARA-09-15-10 D20.wiff
Analyte Integration Type: Base To Base
Analyte Peak Area: 7301
Analyte Ion Transition: 218.985/160.800 Da

Acquired: 8/13/2010 5:19:37 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 591836
IS Ion Transition: 225.000/166.800 Da

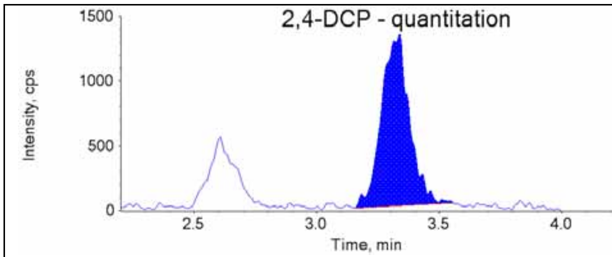


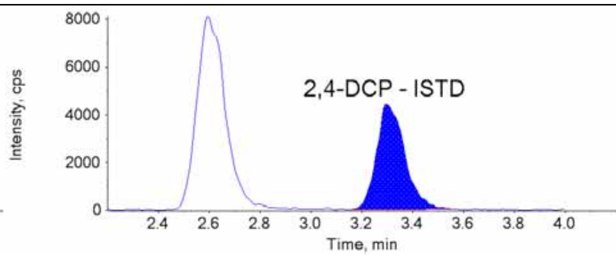


Sample Name: ARA-09-15-10-143-0001A30
Data File: ARA-09-15-10 D20.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 3894
Analyte Ion Transition: 220.968/163.000 Da

Acquired: 8/13/2010 5:19:37 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 591836
IS Ion Transition: 225.000/166.800 Da

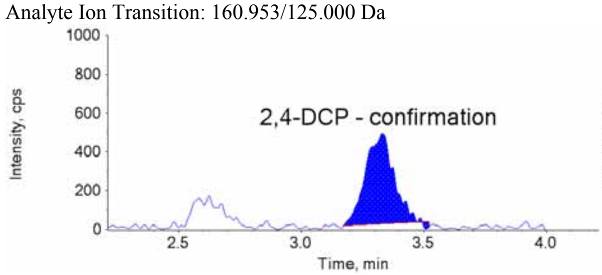
Figure 70. Typical Chromatogram of 2,4-D Analysis, Control Corn Oil, Wet Processed (143-0001)

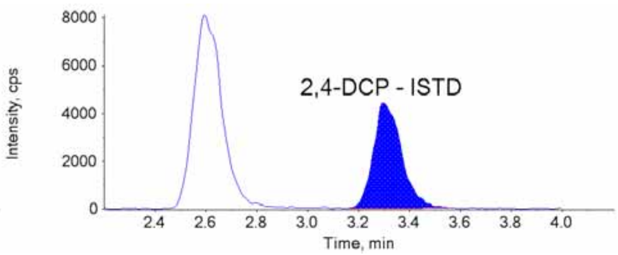




Sample Name: ARA-09-15-10-143-0001A32 + 0.01 µg/g
Data File: ARA-09-15-10 D20.wiff
Analyte Integration Type: Base To Base
Analyte Peak Area: 10582
Analyte Ion Transition: 160.953/125.000 Da

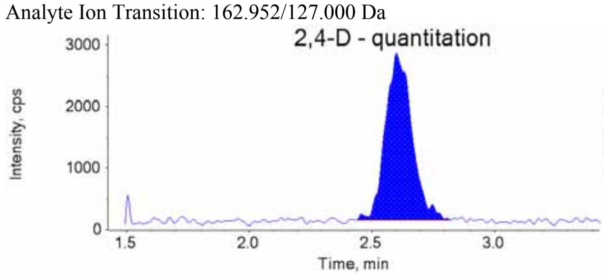
Acquired: 8/13/2010 7:42:07 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 34121
IS Ion Transition: 167.000/131.000 Da

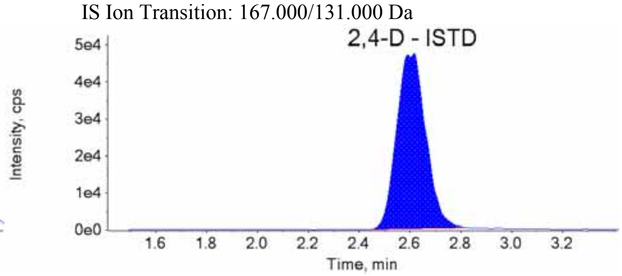




Sample Name: ARA-09-15-10-143-0001A32 + 0.01 µg/g
Data File: ARA-09-15-10 D20.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 3797
Analyte Ion Transition: 162.952/127.000 Da

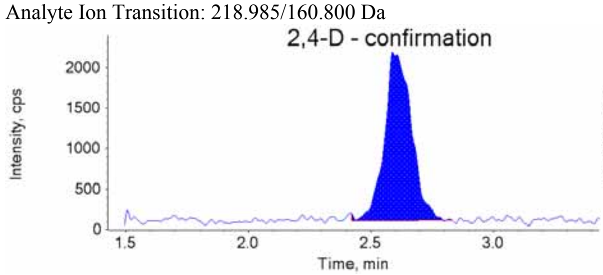
Acquired: 8/13/2010 7:42:07 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 34121
IS Ion Transition: 167.000/131.000 Da

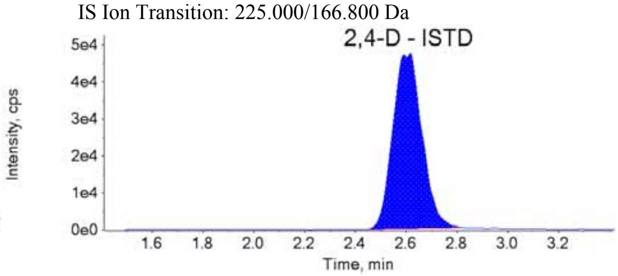




Sample Name: ARA-09-15-10-143-0001A32 + 0.01 µg/g
Data File: ARA-09-15-10 D20.wiff
Analyte Integration Type: Base To Base
Analyte Peak Area: 20554
Analyte Ion Transition: 218.985/160.800 Da

Acquired: 8/13/2010 7:42:07 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 377955
IS Ion Transition: 225.000/166.800 Da

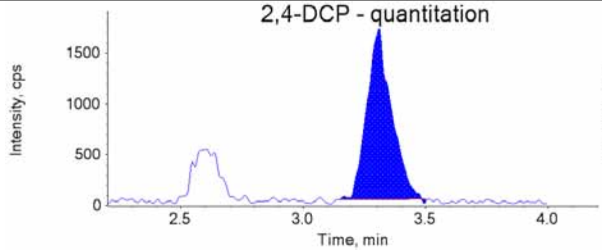


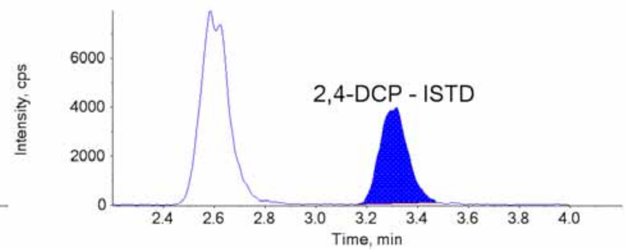


Sample Name: ARA-09-15-10-143-0001A32 + 0.01 µg/g
Data File: ARA-09-15-10 D20.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 15989
Analyte Ion Transition: 220.968/163.000 Da

Acquired: 8/13/2010 7:42:07 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 377955
IS Ion Transition: 225.000/166.800 Da

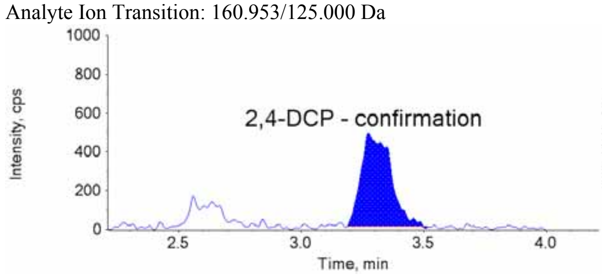
Figure 71. Typical Chromatogram of 2,4-D Analysis, Corn Oil (Wet Processed) 0.01 µg/g Recovery

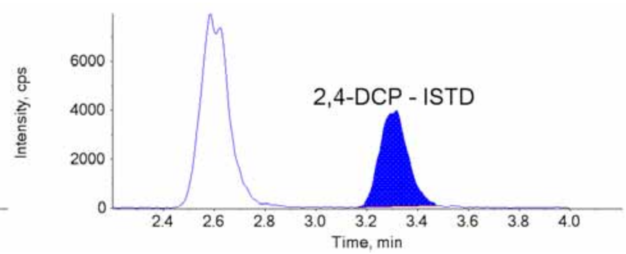




Sample Name: ARA-09-15-10-143-0001A33 + 0.01 µg/g
Data File: ARA-09-15-10 D20.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 11848
Analyte Ion Transition: 160.953/125.000 Da

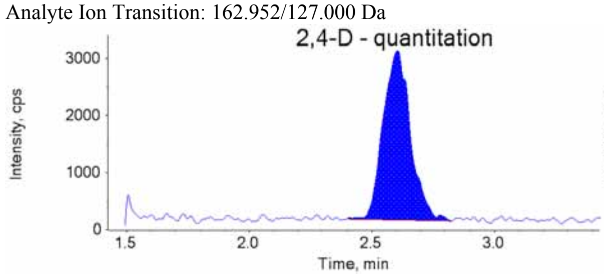
Acquired: 8/13/2010 7:55:06 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 30434
IS Ion Transition: 167.000/131.000 Da

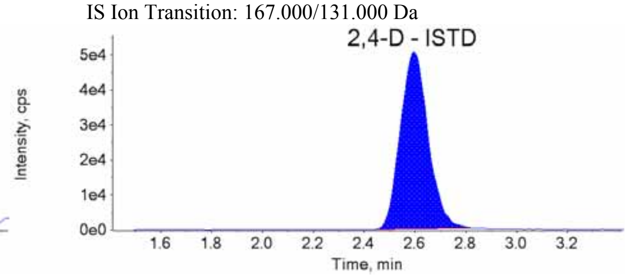




Sample Name: ARA-09-15-10-143-0001A33 + 0.01 µg/g
Data File: ARA-09-15-10 D20.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 4074
Analyte Ion Transition: 162.952/127.000 Da

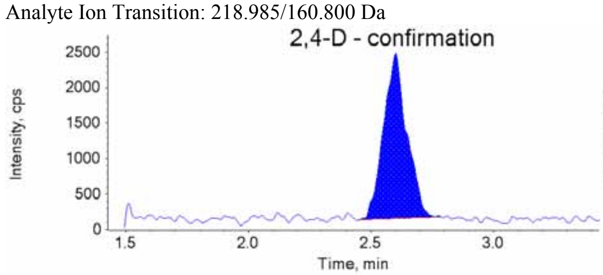
Acquired: 8/13/2010 7:55:06 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 30434
IS Ion Transition: 167.000/131.000 Da

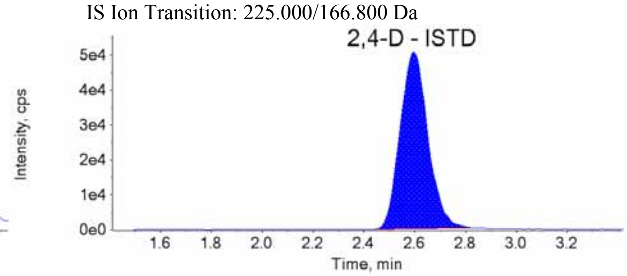




Sample Name: ARA-09-15-10-143-0001A33 + 0.01 µg/g
Data File: ARA-09-15-10 D20.wiff
Analyte Integration Type: Base To Base
Analyte Peak Area: 23310
Analyte Ion Transition: 218.985/160.800 Da

Acquired: 8/13/2010 7:55:06 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 383130
IS Ion Transition: 225.000/166.800 Da

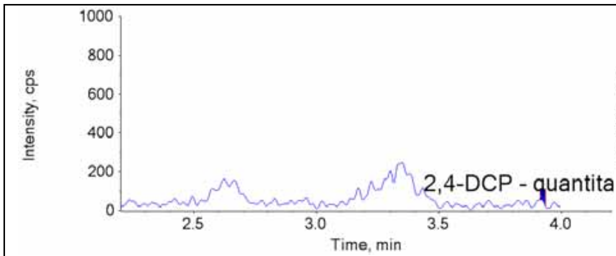


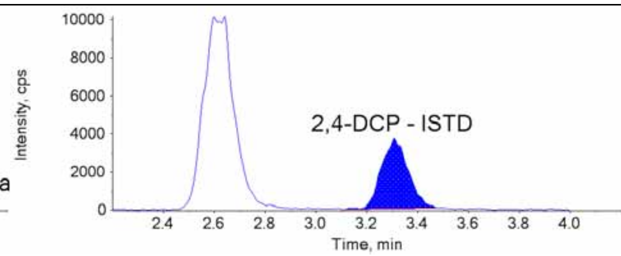


Sample Name: ARA-09-15-10-143-0001A33 + 0.01 µg/g
Data File: ARA-09-15-10 D20.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 15399
Analyte Ion Transition: 220.968/163.000 Da

Acquired: 8/13/2010 7:55:06 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 383130
IS Ion Transition: 225.000/166.800 Da

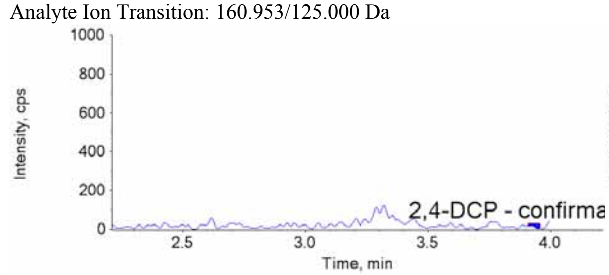
Figure 72. Typical Chromatogram of 2,4-D Analysis, Corn Oil (Wet Processed) 0.01 µg/g Recovery

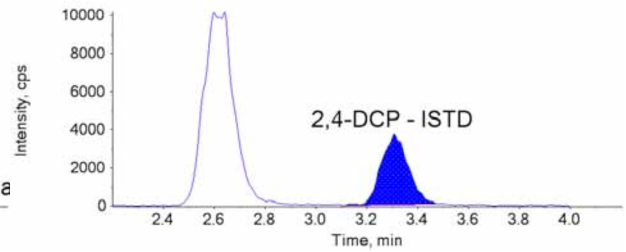




Sample Name: ARA-09-15-10-143-0003A3
Data File: ARA-09-15-10 D20.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 0
Analyte Ion Transition: 160.953/125.000 Da

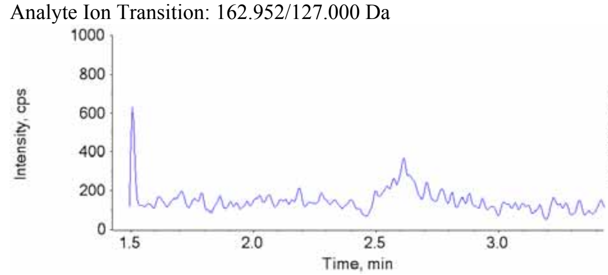
Acquired: 8/13/2010 5:32:31 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 27392
IS Ion Transition: 167.000/131.000 Da

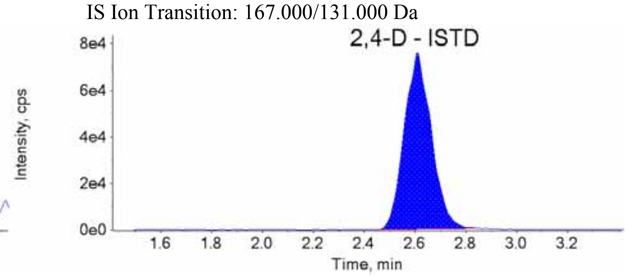




Sample Name: ARA-09-15-10-143-0003A3
Data File: ARA-09-15-10 D20.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 0
Analyte Ion Transition: 162.952/127.000 Da

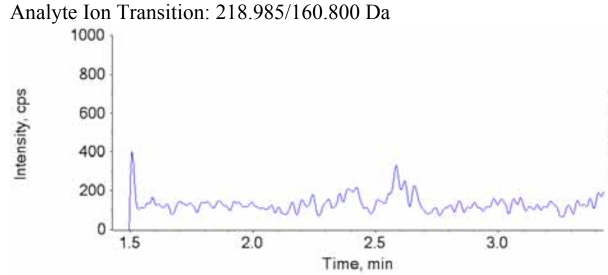
Acquired: 8/13/2010 5:32:31 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 27392
IS Ion Transition: 167.000/131.000 Da

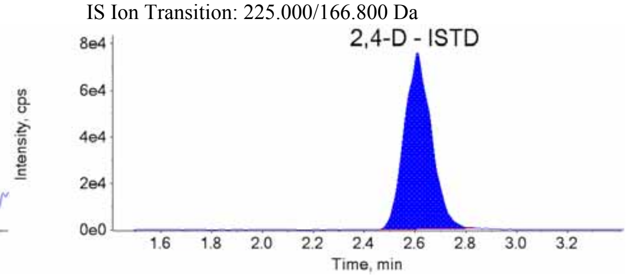




Sample Name: ARA-09-15-10-143-0003A3
Data File: ARA-09-15-10 D20.wiff
Analyte Integration Type: No Peak
Analyte Peak Area: 0
Analyte Ion Transition: 218.985/160.800 Da

Acquired: 8/13/2010 5:32:31 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 559797
IS Ion Transition: 225.000/166.800 Da

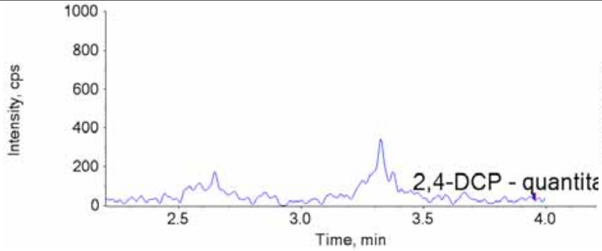


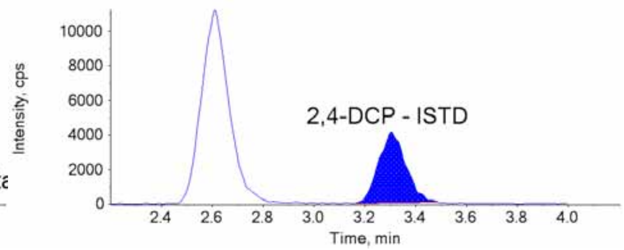


Sample Name: ARA-09-15-10-143-0003A3
Data File: ARA-09-15-10 D20.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 0
Analyte Ion Transition: 220.968/163.000 Da

Acquired: 8/13/2010 5:32:31 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 559797
IS Ion Transition: 225.000/166.800 Da

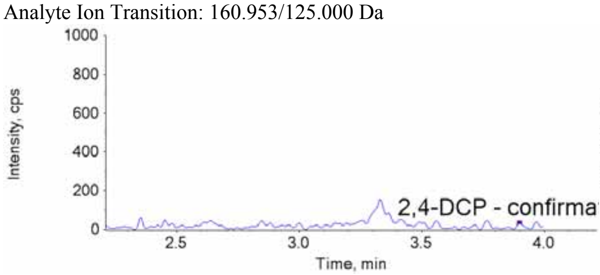
Figure 73. Typical Chromatogram of 2,4-D Analysis, Treated Corn Oil, Wet Processed (143-0003)

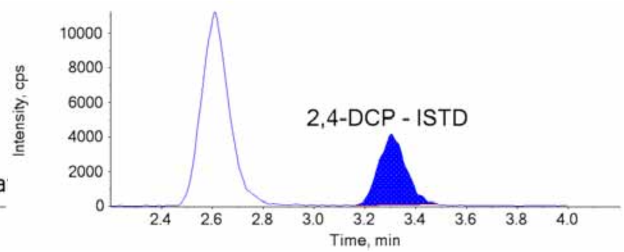




Sample Name: ARA-09-15-10-143-0004A3
Data File: ARA-09-15-10 D20.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 0
Analyte Ion Transition: 160.953/125.000 Da

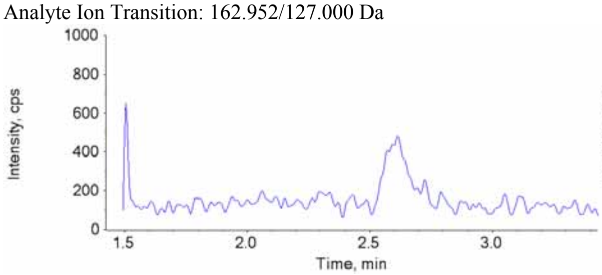
Acquired: 8/13/2010 5:45:27 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 30726
IS Ion Transition: 167.000/131.000 Da

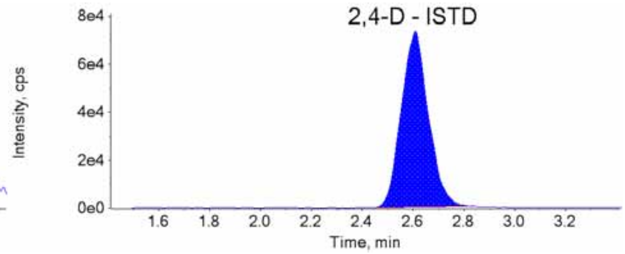




Sample Name: ARA-09-15-10-143-0004A3
Data File: ARA-09-15-10 D20.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 0
Analyte Ion Transition: 162.952/127.000 Da

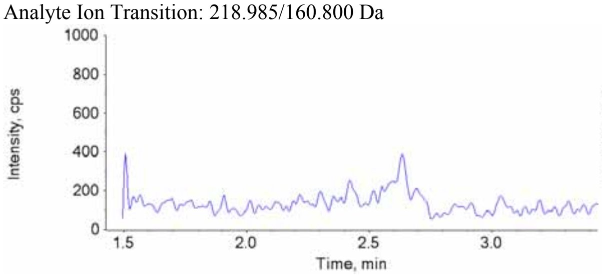
Acquired: 8/13/2010 5:45:27 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 30726
IS Ion Transition: 167.000/131.000 Da

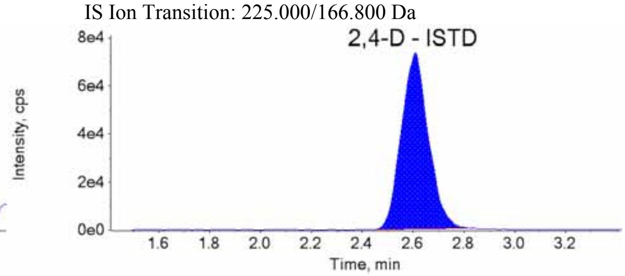




Sample Name: ARA-09-15-10-143-0004A3
Data File: ARA-09-15-10 D20.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 0
Analyte Ion Transition: 218.985/160.800 Da

Acquired: 8/13/2010 5:45:27 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 539523
IS Ion Transition: 225.000/166.800 Da

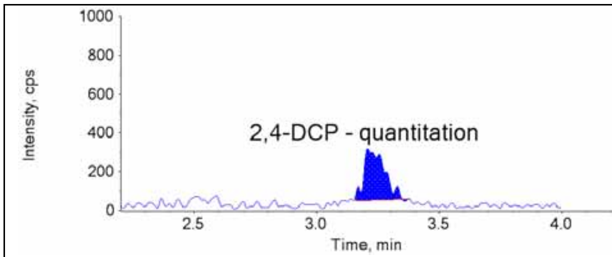


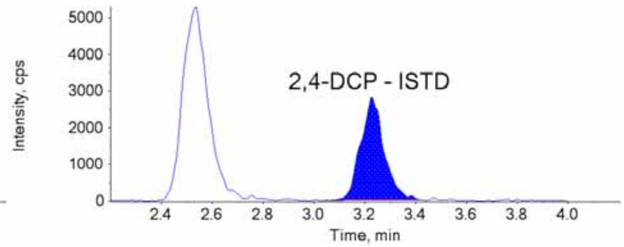


Sample Name: ARA-09-15-10-143-0004A3
Data File: ARA-09-15-10 D20.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 0
Analyte Ion Transition: 220.968/163.000 Da

Acquired: 8/13/2010 5:45:27 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 539523
IS Ion Transition: 225.000/166.800 Da

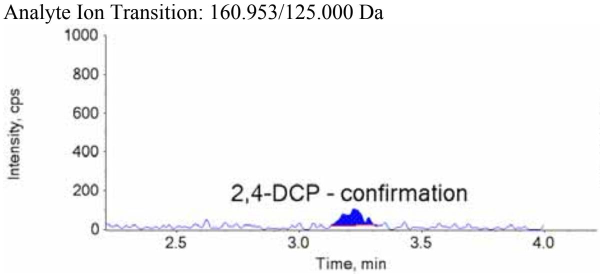
Figure 74. Typical Chromatogram of 2,4-D Analysis, Treated Corn Oil, Wet Processed (143-0004)

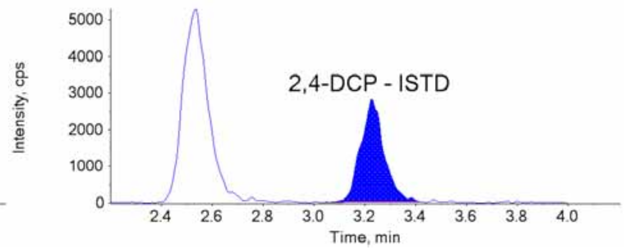




Sample Name: ARA-09-15-10-142-0001A30
Data File: ARA-09-15-10 D19c.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 1562
Analyte Ion Transition: 160.953/125.000 Da

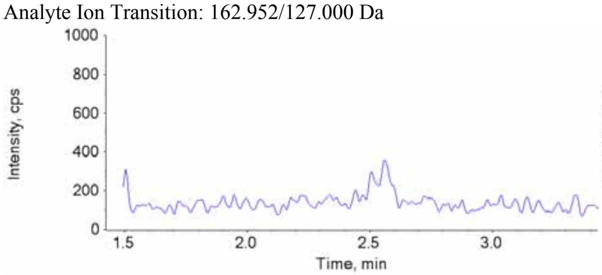
Acquired: 8/18/2010 7:44:21 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 17827
IS Ion Transition: 167.000/131.000 Da

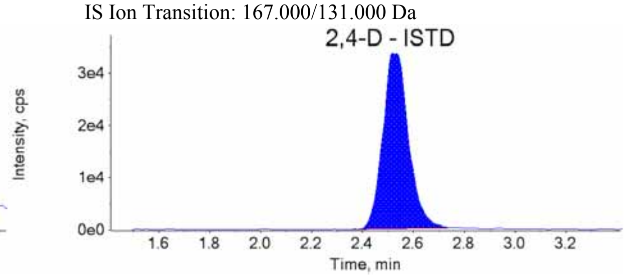




Sample Name: ARA-09-15-10-142-0001A30
Data File: ARA-09-15-10 D19c.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 456
Analyte Ion Transition: 162.952/127.000 Da

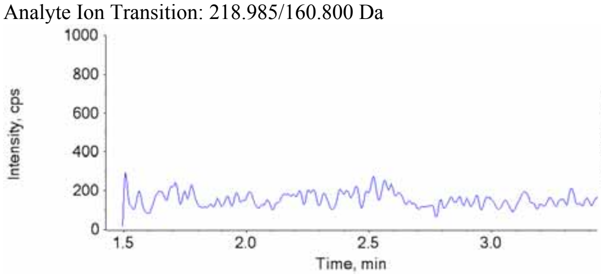
Acquired: 8/18/2010 7:44:21 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 17827
IS Ion Transition: 167.000/131.000 Da

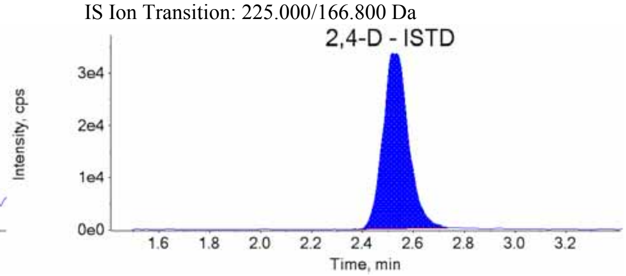




Sample Name: ARA-09-15-10-142-0001A30
Data File: ARA-09-15-10 D19c.wiff
Analyte Integration Type: No Peak
Analyte Peak Area: 0
Analyte Ion Transition: 218.985/160.800 Da

Acquired: 8/18/2010 7:44:21 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 235788
IS Ion Transition: 225.000/166.800 Da

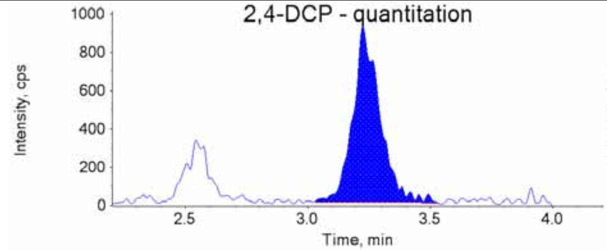


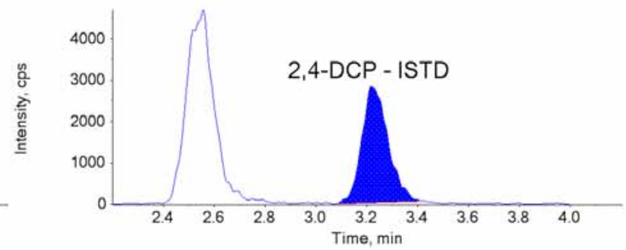


Sample Name: ARA-09-15-10-142-0001A30
Data File: ARA-09-15-10 D19c.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 0
Analyte Ion Transition: 220.968/163.000 Da

Acquired: 8/18/2010 7:44:21 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 235788
IS Ion Transition: 225.000/166.800 Da

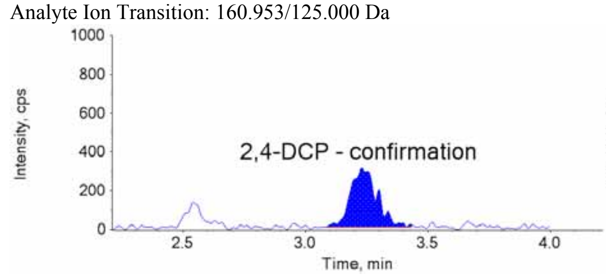
Figure 75. Typical Chromatogram of 2,4-D Analysis, Control Corn Starch (142-0001)

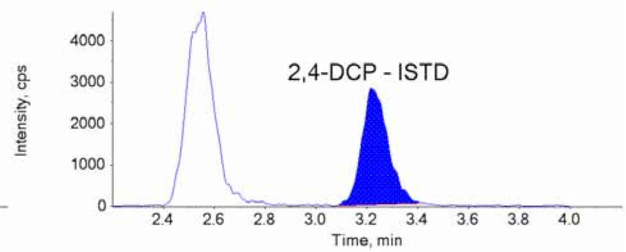




Sample Name: ARA-09-15-10-142-0001A32 + 0.01 µg/g
Data File: ARA-09-15-10 D19c.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 7108
Analyte Ion Transition: 160.953/125.000 Da

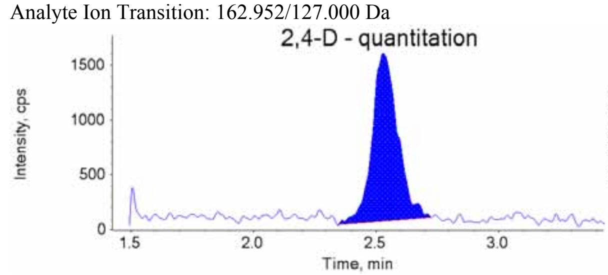
Acquired: 8/18/2010 10:13:03 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 20297
IS Ion Transition: 167.000/131.000 Da

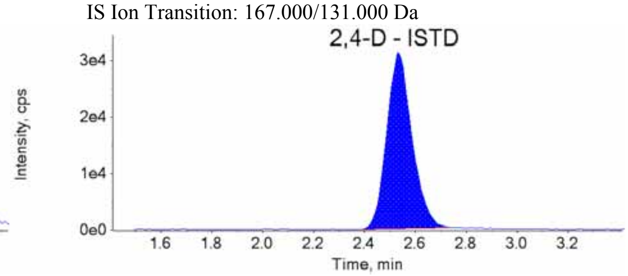




Sample Name: ARA-09-15-10-142-0001A32 + 0.01 µg/g
Data File: ARA-09-15-10 D19c.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 2288
Analyte Ion Transition: 162.952/127.000 Da

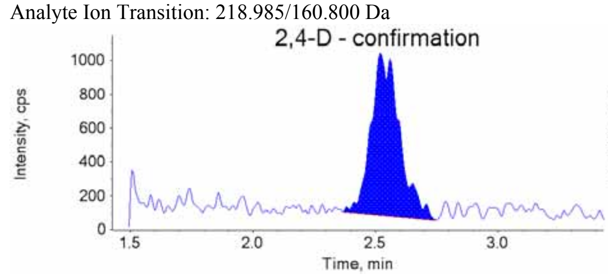
Acquired: 8/18/2010 10:13:03 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 20297
IS Ion Transition: 167.000/131.000 Da

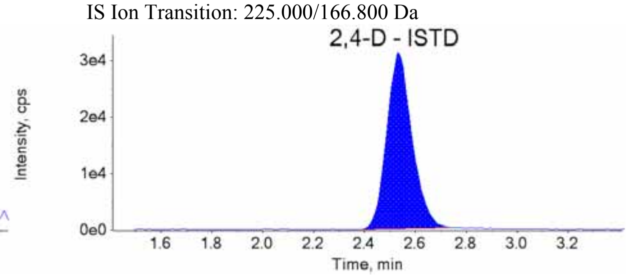




Sample Name: ARA-09-15-10-142-0001A32 + 0.01 µg/g
Data File: ARA-09-15-10 D19c.wiff
Analyte Integration Type: Base To Base
Analyte Peak Area: 11275
Analyte Ion Transition: 218.985/160.800 Da

Acquired: 8/18/2010 10:13:03 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 213529
IS Ion Transition: 225.000/166.800 Da

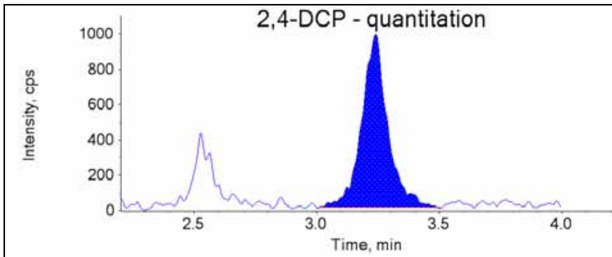


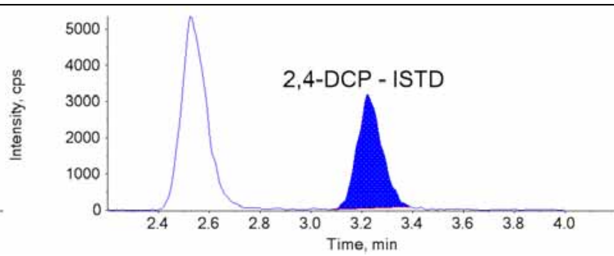


Sample Name: ARA-09-15-10-142-0001A32 + 0.01 µg/g
Data File: ARA-09-15-10 D19c.wiff
Analyte Integration Type: Base To Base
Analyte Peak Area: 7844
Analyte Ion Transition: 220.968/163.000 Da

Acquired: 8/18/2010 10:13:03 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 213529
IS Ion Transition: 225.000/166.800 Da

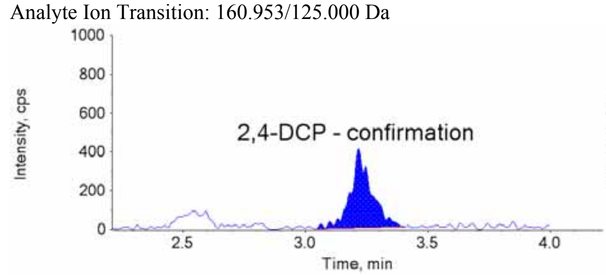
Figure 76. Typical Chromatogram of 2,4-D Analysis, Corn Starch 0.01 µg/g Recovery

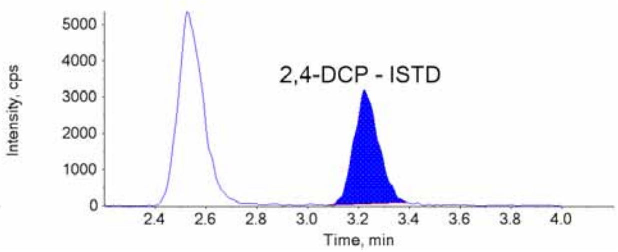




Sample Name: ARA-09-15-10-142-0001A33 + 0.01 µg/g
Data File: ARA-09-15-10 D19c.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 7262
Analyte Ion Transition: 160.953/125.000 Da

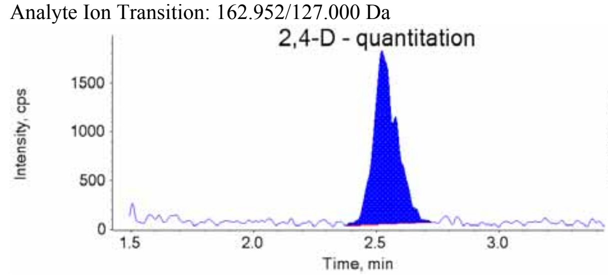
Acquired: 8/18/2010 10:19:32 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 21160
IS Ion Transition: 167.000/131.000 Da

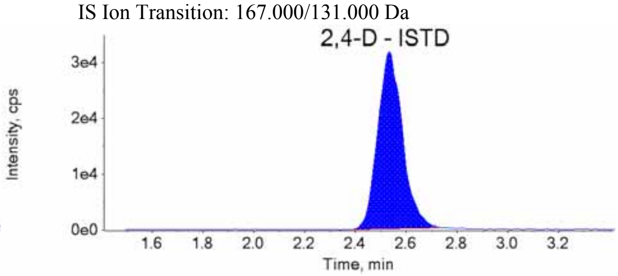




Sample Name: ARA-09-15-10-142-0001A33 + 0.01 µg/g
Data File: ARA-09-15-10 D19c.wiff
Analyte Integration Type: Base To Base
Analyte Peak Area: 2379
Analyte Ion Transition: 162.952/127.000 Da

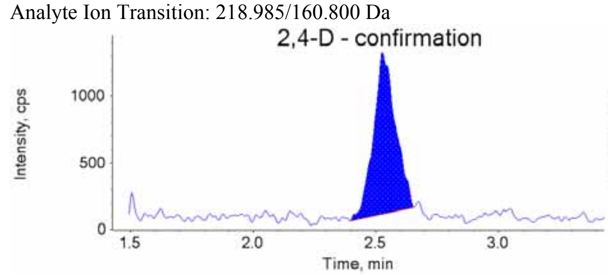
Acquired: 8/18/2010 10:19:32 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 21160
IS Ion Transition: 167.000/131.000 Da

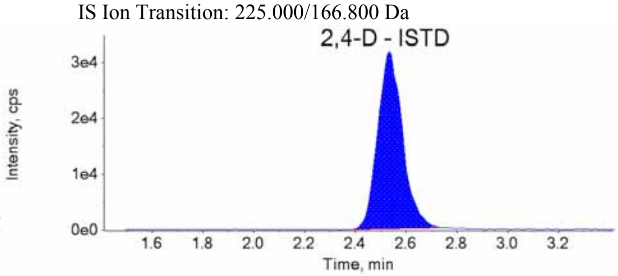




Sample Name: ARA-09-15-10-142-0001A33 + 0.01 µg/g
Data File: ARA-09-15-10 D19c.wiff
Analyte Integration Type: Base To Base
Analyte Peak Area: 11766
Analyte Ion Transition: 218.985/160.800 Da

Acquired: 8/18/2010 10:19:32 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 214223
IS Ion Transition: 225.000/166.800 Da

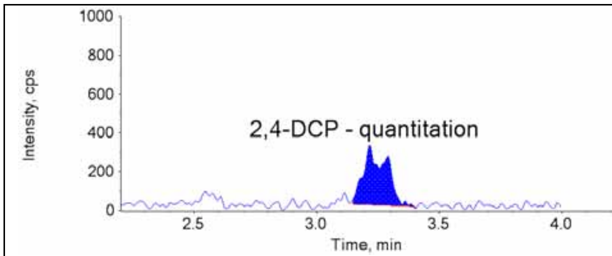


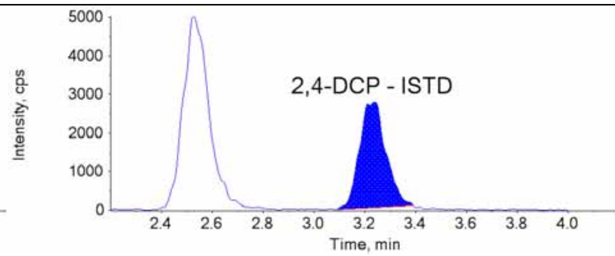


Sample Name: ARA-09-15-10-142-0001A33 + 0.01 µg/g
Data File: ARA-09-15-10 D19c.wiff
Analyte Integration Type: Base To Base
Analyte Peak Area: 7565
Analyte Ion Transition: 220.968/163.000 Da

Acquired: 8/18/2010 10:19:32 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 214223
IS Ion Transition: 225.000/166.800 Da

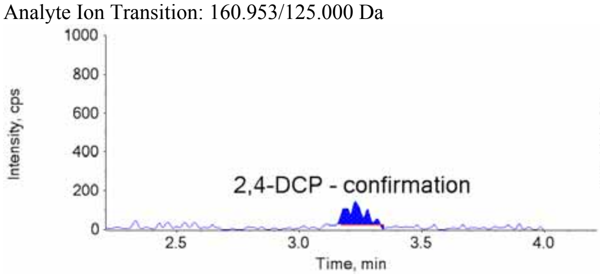
Figure 77. Typical Chromatogram of 2,4-D Analysis, Corn Starch 0.01 µg/g Recovery

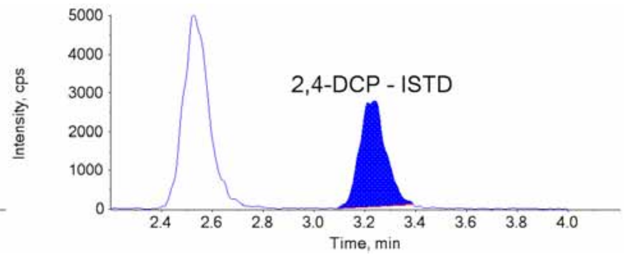




Sample Name: ARA-09-15-10-142-0003A3
Data File: ARA-09-15-10 D19c.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 1994
Analyte Ion Transition: 160.953/125.000 Da

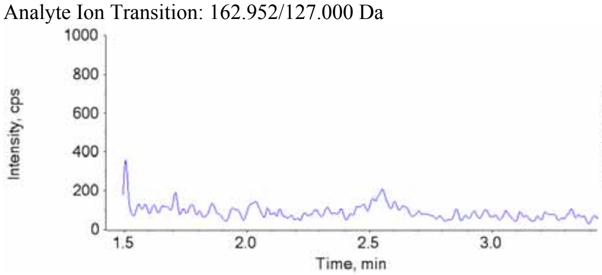
Acquired: 8/18/2010 7:57:16 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 19865
IS Ion Transition: 167.000/131.000 Da

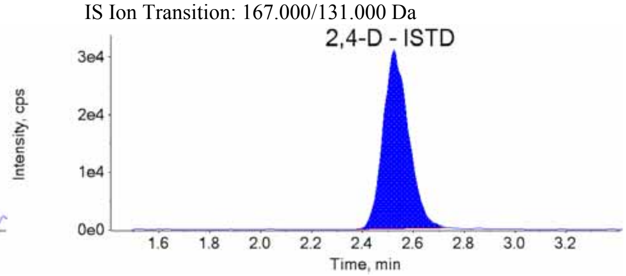




Sample Name: ARA-09-15-10-142-0003A3
Data File: ARA-09-15-10 D19c.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 617
Analyte Ion Transition: 162.952/127.000 Da

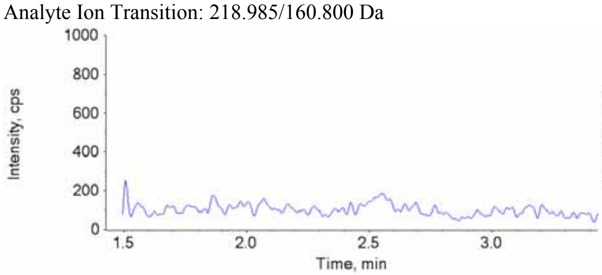
Acquired: 8/18/2010 7:57:16 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 19865
IS Ion Transition: 167.000/131.000 Da

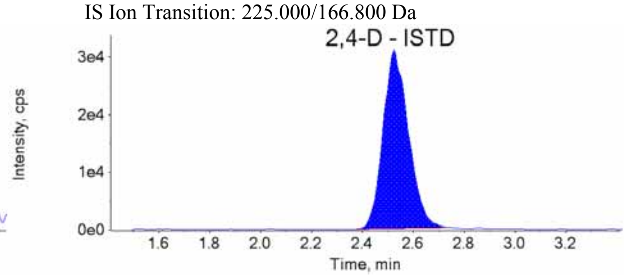




Sample Name: ARA-09-15-10-142-0003A3
Data File: ARA-09-15-10 D19c.wiff
Analyte Integration Type: No Peak
Analyte Peak Area: 0
Analyte Ion Transition: 218.985/160.800 Da

Acquired: 8/18/2010 7:57:16 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 214185
IS Ion Transition: 225.000/166.800 Da

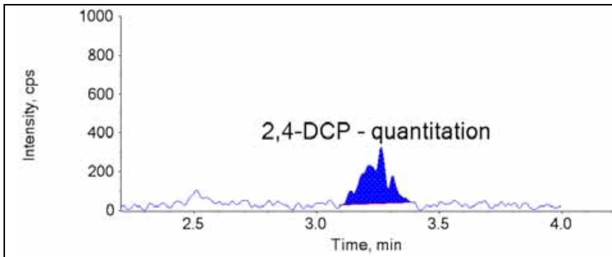


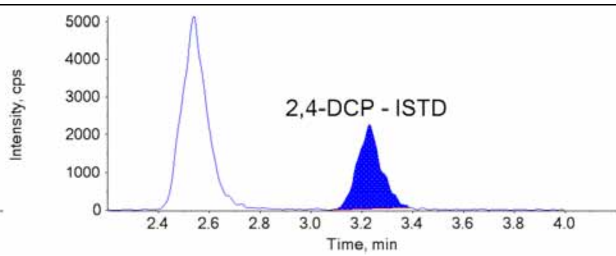


Sample Name: ARA-09-15-10-142-0003A3
Data File: ARA-09-15-10 D19c.wiff
Analyte Integration Type: No Peak
Analyte Peak Area: 0
Analyte Ion Transition: 220.968/163.000 Da

Acquired: 8/18/2010 7:57:16 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 214185
IS Ion Transition: 225.000/166.800 Da

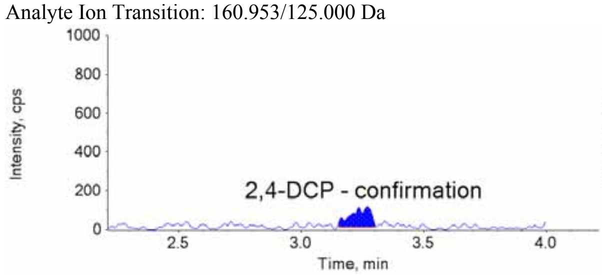
Figure 78. Typical Chromatogram of 2,4-D Analysis, Treated Corn Starch (142-0003)

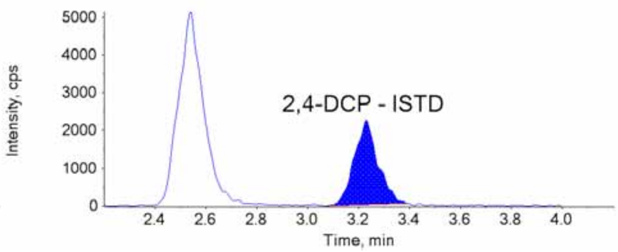




Sample Name: ARA-09-15-10-142-0004A4
Data File: ARA-09-15-10 D19c.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 1880
Analyte Ion Transition: 160.953/125.000 Da

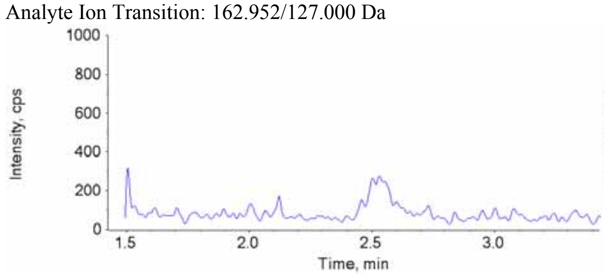
Acquired: 8/18/2010 8:03:43 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 14382
IS Ion Transition: 167.000/131.000 Da

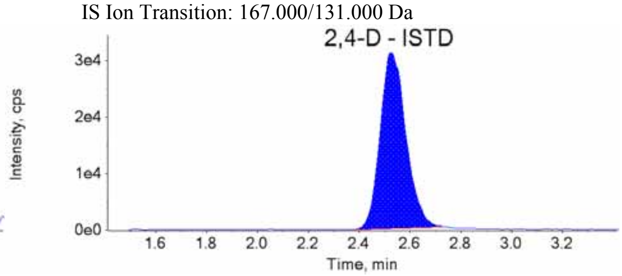




Sample Name: ARA-09-15-10-142-0004A4
Data File: ARA-09-15-10 D19c.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 589
Analyte Ion Transition: 162.952/127.000 Da

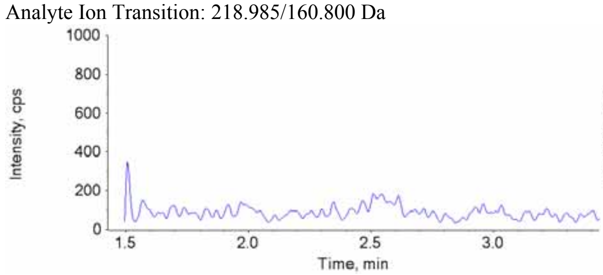
Acquired: 8/18/2010 8:03:43 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 14382
IS Ion Transition: 167.000/131.000 Da

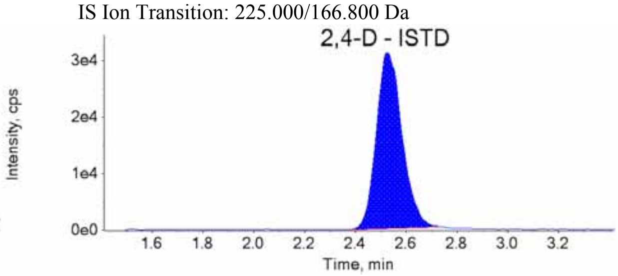




Sample Name: ARA-09-15-10-142-0004A4
Data File: ARA-09-15-10 D19c.wiff
Analyte Integration Type: Manual
Analyte Peak Area: 0
Analyte Ion Transition: 218.985/160.800 Da

Acquired: 8/18/2010 8:03:43 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 218707
IS Ion Transition: 225.000/166.800 Da



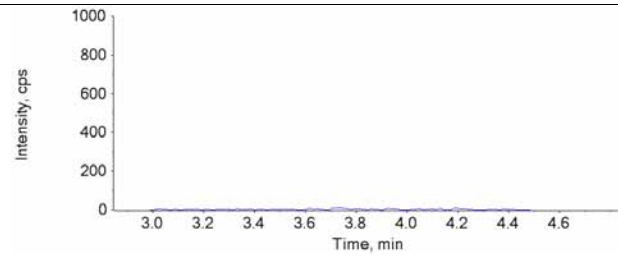


Sample Name: ARA-09-15-10-142-0004A4
Data File: ARA-09-15-10 D19c.wiff
Analyte Integration Type: No Peak
Analyte Peak Area: 0
Analyte Ion Transition: 220.968/163.000 Da

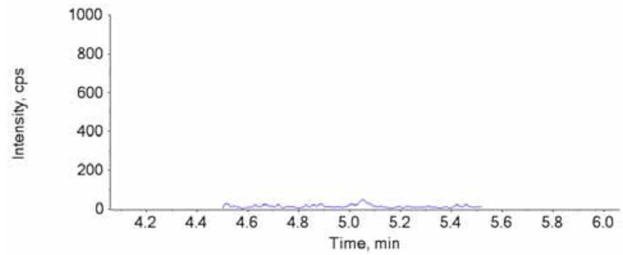
Acquired: 8/18/2010 8:03:43 PM
Injection Volume (µL): 30
IS Integration Type: Base To Base
IS Peak Area: 218707
IS Ion Transition: 225.000/166.800 Da

Figure 79. Typical Chromatogram of 2,4-D Analysis, Treated Corn Starch (142-0004)

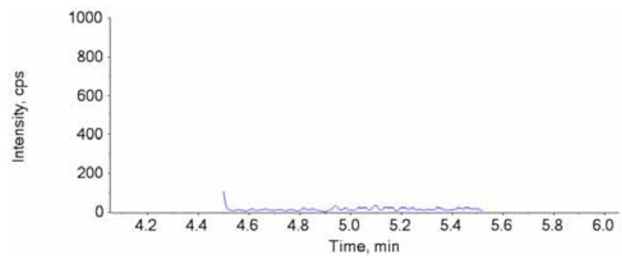
Sample Name: Reagent Blank
Data File: ARA-09-15-10 Q02.wiff
Sample Index: 1
Acquired: 5/20/2010 12:34:57 AM
Analyte Peak Area: 0
Analyte Ion Transition: 343.1/108.1 amu



Sample Name: Reagent Blank
Data File: ARA-09-15-10 Q02.wiff
Sample Index: 1
Acquired: 5/20/2010 12:34:57 AM
Analyte Peak Area: 0
Analyte Ion Transition: 373.2/299.0 amu



Sample Name: Reagent Blank
Data File: ARA-09-15-10 Q02.wiff
Sample Index: 1
Acquired: 5/20/2010 12:34:57 AM
Analyte Peak Area: 0
Analyte Ion Transition: 373.2/91.0 amu



Sample Name: Reagent Blank
Data File: ARA-09-15-10 Q02.wiff
Sample Index: 1
Acquired: 5/20/2010 12:34:57 AM
Analyte Peak Area: 0
Analyte Ion Transition: 343.1/270.9 amu

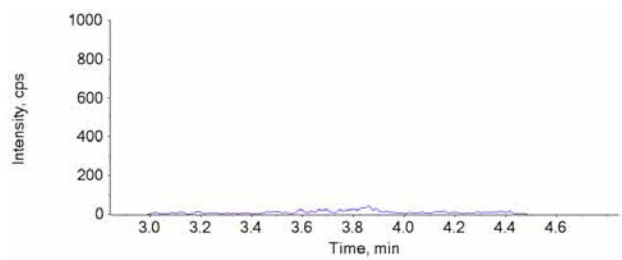
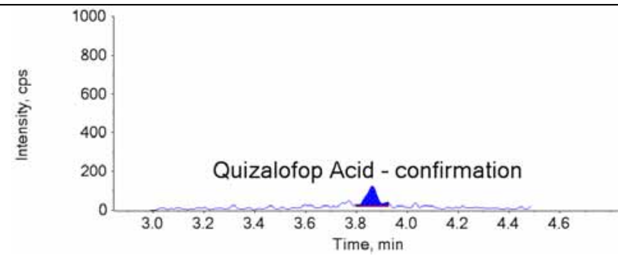
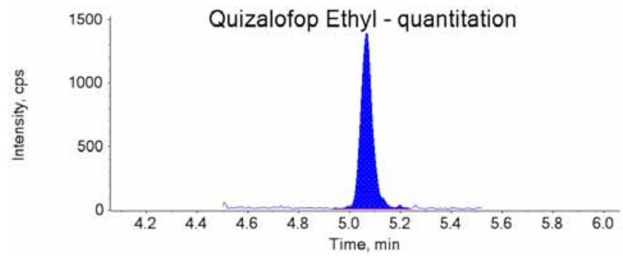


Figure 80. Typical Chromatogram of Quizalofop Analysis, Reagent Blank

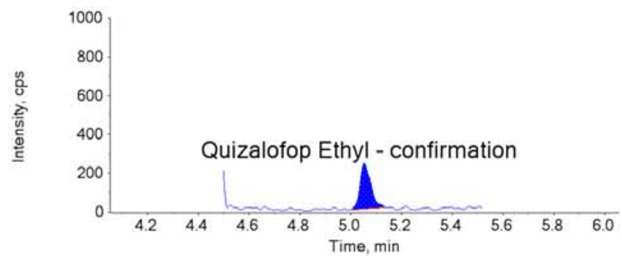
Sample Name: 0.25 std
Data File: ARA-09-15-10 Q02.wiff
Sample Index: 8
Acquired: 5/20/2010 2:02:18 AM
Analyte Peak Area: 323
Analyte Ion Transition: 343.1/108.1 amu



Sample Name: 0.25 std
Data File: ARA-09-15-10 Q02.wiff
Sample Index: 8
Acquired: 5/20/2010 2:02:18 AM
Analyte Peak Area: 4529
Analyte Ion Transition: 373.2/299.0 amu



Sample Name: 0.25 std
Data File: ARA-09-15-10 Q02.wiff
Sample Index: 8
Acquired: 5/20/2010 2:02:18 AM
Analyte Peak Area: 691
Analyte Ion Transition: 373.2/91.0 amu



Sample Name: 0.25 std
Data File: ARA-09-15-10 Q02.wiff
Sample Index: 8
Acquired: 5/20/2010 2:02:18 AM
Analyte Peak Area: 5520
Analyte Ion Transition: 343.1/270.9 amu

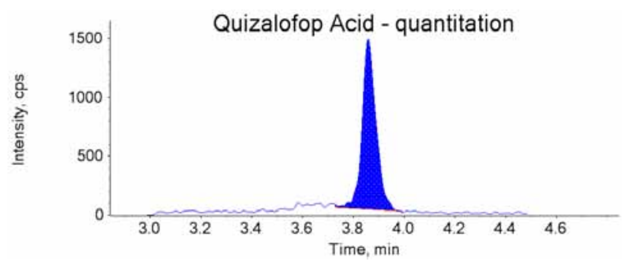
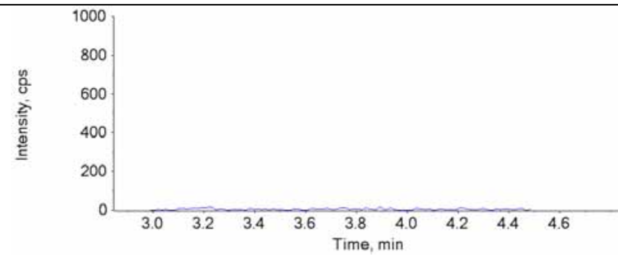
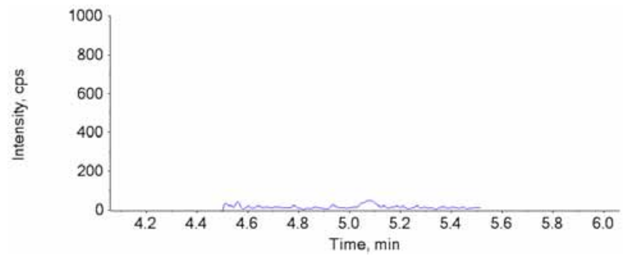


Figure 81. Typical Chromatogram of Quizalofop Analysis, 0.25 ng/mL Calibration Standard

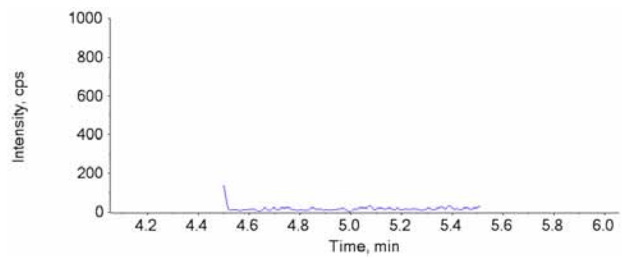
Sample Name: ARA-09-15-10-020-0001A8
Data File: ARA-09-15-10 Q02.wiff
Sample Index: 2
Acquired: 5/20/2010 12:47:25 AM
Analyte Peak Area: 0
Analyte Ion Transition: 343.1/108.1 amu



Sample Name: ARA-09-15-10-020-0001A8
Data File: ARA-09-15-10 Q02.wiff
Sample Index: 2
Acquired: 5/20/2010 12:47:25 AM
Analyte Peak Area: 0
Analyte Ion Transition: 373.2/299.0 amu



Sample Name: ARA-09-15-10-020-0001A8
Data File: ARA-09-15-10 Q02.wiff
Sample Index: 2
Acquired: 5/20/2010 12:47:25 AM
Analyte Peak Area: 0
Analyte Ion Transition: 373.2/91.0 amu



Sample Name: ARA-09-15-10-020-0001A8
Data File: ARA-09-15-10 Q02.wiff
Sample Index: 2
Acquired: 5/20/2010 12:47:25 AM
Analyte Peak Area: 0
Analyte Ion Transition: 343.1/270.9 amu

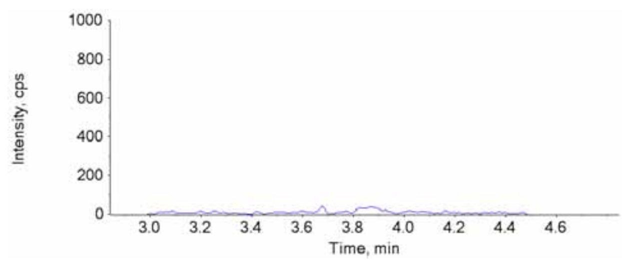
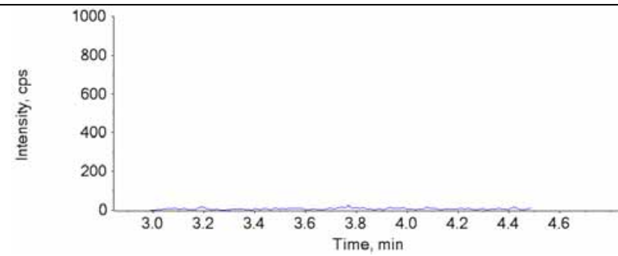
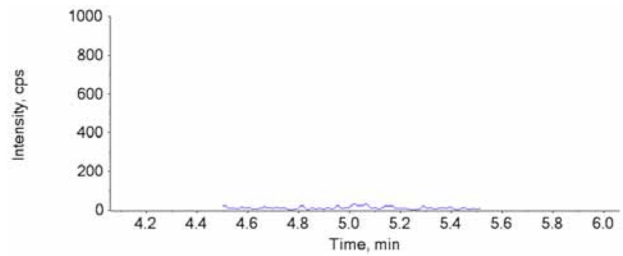


Figure 82. Typical Chromatogram of Quizalofop Analysis, Control Corn Grain (020-0001)

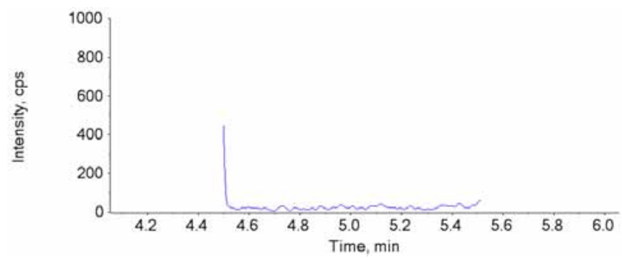
Sample Name: ARA-09-15-10-022-0001A1
Data File: ARA-09-15-10 Q01.wiff
Sample Index: 12
Acquired: 5/18/2010 9:21:50 PM
Analyte Peak Area: 0
Analyte Ion Transition: 343.1/108.1 amu



Sample Name: ARA-09-15-10-022-0001A1
Data File: ARA-09-15-10 Q01.wiff
Sample Index: 12
Acquired: 5/18/2010 9:21:50 PM
Analyte Peak Area: 0
Analyte Ion Transition: 373.2/299.0 amu



Sample Name: ARA-09-15-10-022-0001A1
Data File: ARA-09-15-10 Q01.wiff
Sample Index: 12
Acquired: 5/18/2010 9:21:50 PM
Analyte Peak Area: 0
Analyte Ion Transition: 373.2/91.0 amu



Sample Name: ARA-09-15-10-022-0001A1
Data File: ARA-09-15-10 Q01.wiff
Sample Index: 12
Acquired: 5/18/2010 9:21:50 PM
Analyte Peak Area: 0
Analyte Ion Transition: 343.1/270.9 amu

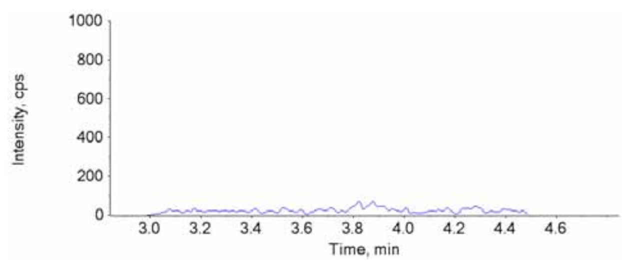
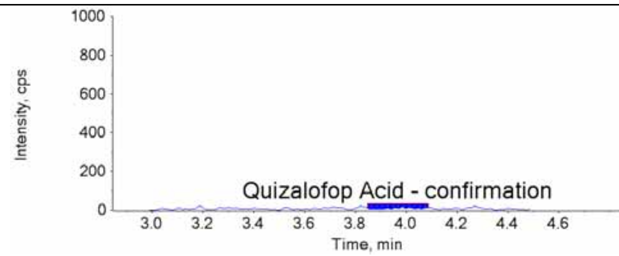
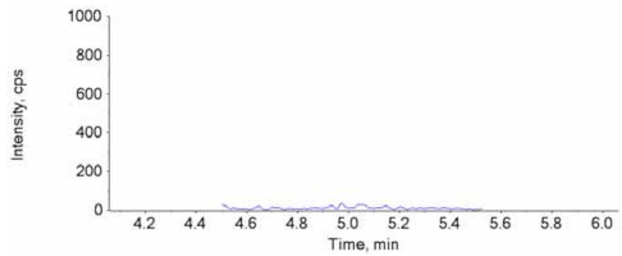


Figure 83. Typical Chromatogram of Quizalofop Analysis, Control Corn Grain (022-0001)

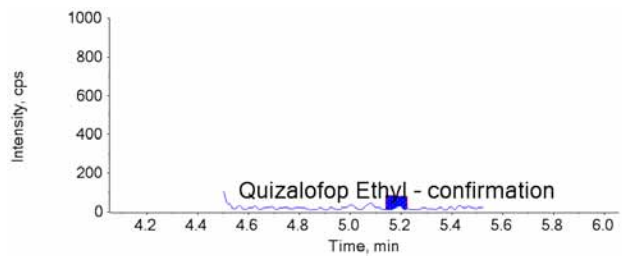
Sample Name: ARA-09-15-10-024-0001A1
Data File: ARA-09-15-10 Q01.wiff
Sample Index: 13
Acquired: 5/18/2010 9:34:19 PM
Analyte Peak Area: 0
Analyte Ion Transition: 343.1/108.1 amu



Sample Name: ARA-09-15-10-024-0001A1
Data File: ARA-09-15-10 Q01.wiff
Sample Index: 13
Acquired: 5/18/2010 9:34:19 PM
Analyte Peak Area: 0
Analyte Ion Transition: 373.2/299.0 amu



Sample Name: ARA-09-15-10-024-0001A1
Data File: ARA-09-15-10 Q01.wiff
Sample Index: 13
Acquired: 5/18/2010 9:34:19 PM
Analyte Peak Area: 0
Analyte Ion Transition: 373.2/91.0 amu



Sample Name: ARA-09-15-10-024-0001A1
Data File: ARA-09-15-10 Q01.wiff
Sample Index: 13
Acquired: 5/18/2010 9:34:19 PM
Analyte Peak Area: 0
Analyte Ion Transition: 343.1/270.9 amu

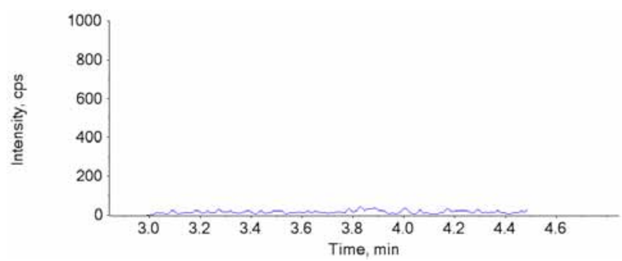
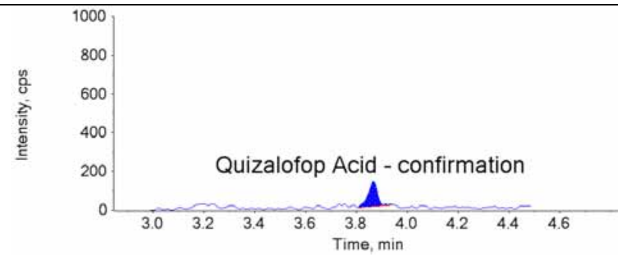
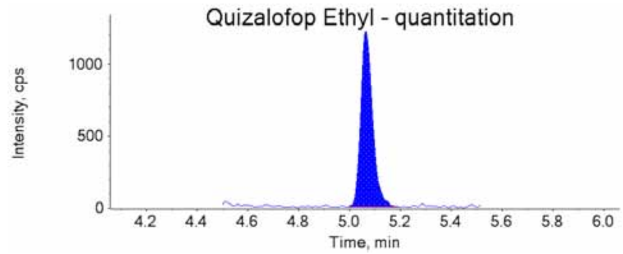


Figure 84. Typical Chromatogram of Quizalofop Analysis, Control Corn Grain (024-0001)

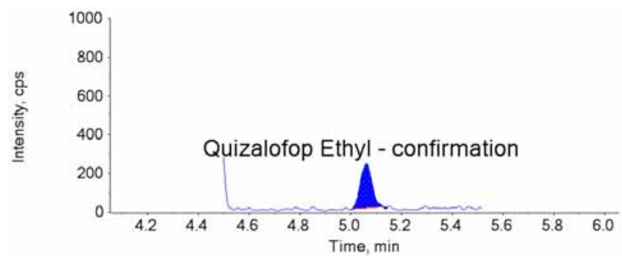
Sample Name: ARA-09-15-10-020-0001A10 + 0.01 µg/g
Data File: ARA-09-15-10 Q02.wiff
Sample Index: 4
Acquired: 5/20/2010 1:12:22 AM
Analyte Peak Area: 336
Analyte Ion Transition: 343.1/108.1 amu



Sample Name: ARA-09-15-10-020-0001A10 + 0.01 µg/g
Data File: ARA-09-15-10 Q02.wiff
Sample Index: 4
Acquired: 5/20/2010 1:12:22 AM
Analyte Peak Area: 3964
Analyte Ion Transition: 373.2/299.0 amu



Sample Name: ARA-09-15-10-020-0001A10 + 0.01 µg/g
Data File: ARA-09-15-10 Q02.wiff
Sample Index: 4
Acquired: 5/20/2010 1:12:22 AM
Analyte Peak Area: 732
Analyte Ion Transition: 373.2/91.0 amu



Sample Name: ARA-09-15-10-020-0001A10 + 0.01 µg/g
Data File: ARA-09-15-10 Q02.wiff
Sample Index: 4
Acquired: 5/20/2010 1:12:22 AM
Analyte Peak Area: 5172
Analyte Ion Transition: 343.1/270.9 amu

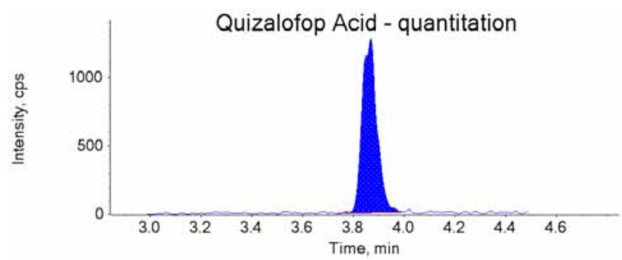
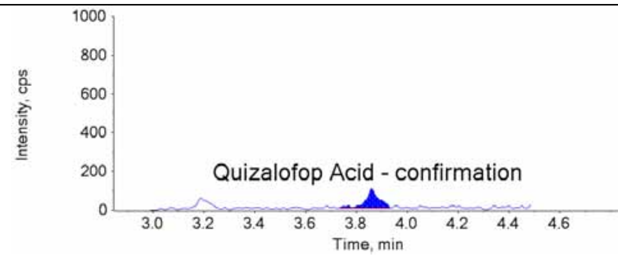
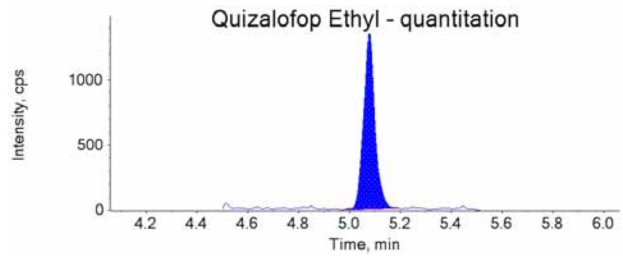


Figure 85. Typical Chromatogram of Quizalofop Analysis, Corn Grain 0.01 µg/g Recovery

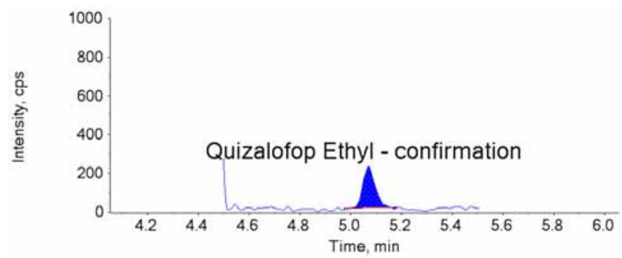
Sample Name: ARA-09-15-10-020-0001A11 + 0.01 µg/g
Data File: ARA-09-15-10 Q02.wiff
Sample Index: 5
Acquired: 5/20/2010 1:24:51 AM
Analyte Peak Area: 388
Analyte Ion Transition: 343.1/108.1 amu



Sample Name: ARA-09-15-10-020-0001A11 + 0.01 µg/g
Data File: ARA-09-15-10 Q02.wiff
Sample Index: 5
Acquired: 5/20/2010 1:24:51 AM
Analyte Peak Area: 4179
Analyte Ion Transition: 373.2/299.0 amu



Sample Name: ARA-09-15-10-020-0001A11 + 0.01 µg/g
Data File: ARA-09-15-10 Q02.wiff
Sample Index: 5
Acquired: 5/20/2010 1:24:51 AM
Analyte Peak Area: 703
Analyte Ion Transition: 373.2/91.0 amu



Sample Name: ARA-09-15-10-020-0001A11 + 0.01 µg/g
Data File: ARA-09-15-10 Q02.wiff
Sample Index: 5
Acquired: 5/20/2010 1:24:51 AM
Analyte Peak Area: 5258
Analyte Ion Transition: 343.1/270.9 amu

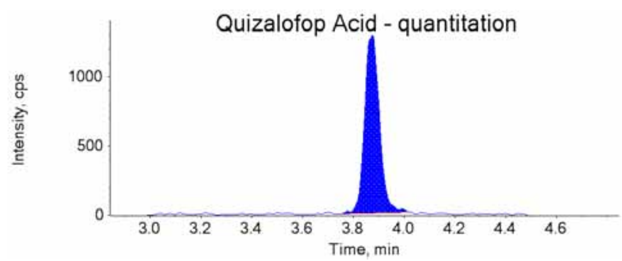
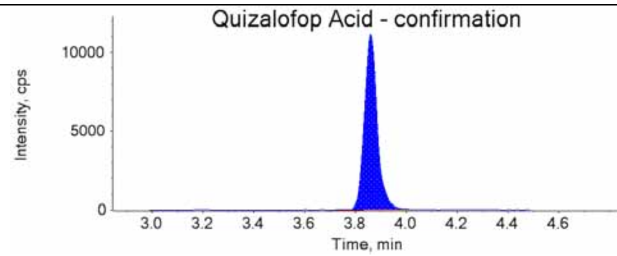
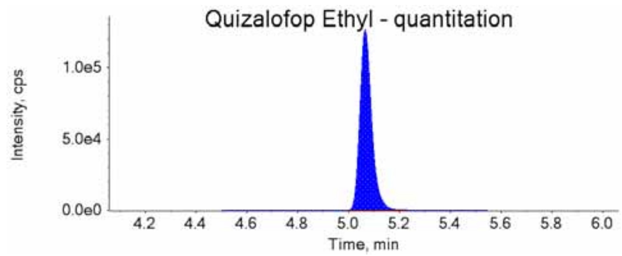


Figure 86. Typical Chromatogram of Quizalofop Analysis, Corn Grain 0.01 µg/g Recovery

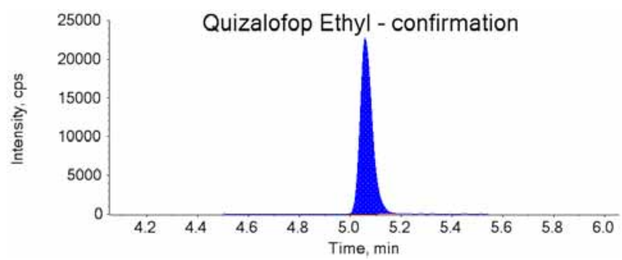
Sample Name: ARA-09-15-10-020-0001A12 + 1.0 µg/g
Data File: ARA-09-15-10 Q02.wiff
Sample Index: 6
Acquired: 5/20/2010 1:37:19 AM
Analyte Peak Area: 39383
Analyte Ion Transition: 343.1/108.1 amu



Sample Name: ARA-09-15-10-020-0001A12 + 1.0 µg/g
Data File: ARA-09-15-10 Q02.wiff
Sample Index: 6
Acquired: 5/20/2010 1:37:19 AM
Analyte Peak Area: 406851
Analyte Ion Transition: 373.2/299.0 amu



Sample Name: ARA-09-15-10-020-0001A12 + 1.0 µg/g
Data File: ARA-09-15-10 Q02.wiff
Sample Index: 6
Acquired: 5/20/2010 1:37:19 AM
Analyte Peak Area: 74486
Analyte Ion Transition: 373.2/91.0 amu



Sample Name: ARA-09-15-10-020-0001A12 + 1.0 µg/g
Data File: ARA-09-15-10 Q02.wiff
Sample Index: 6
Acquired: 5/20/2010 1:37:19 AM
Analyte Peak Area: 508658
Analyte Ion Transition: 343.1/270.9 amu

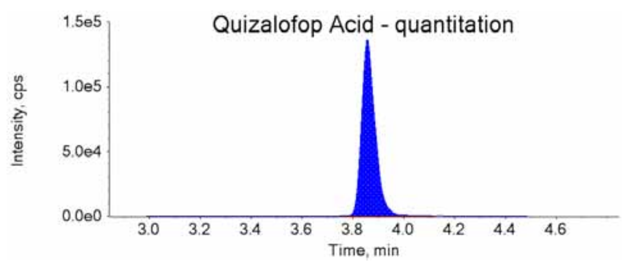
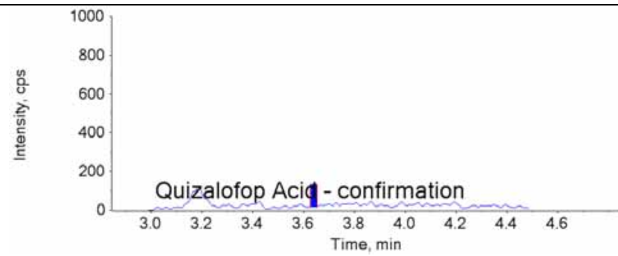
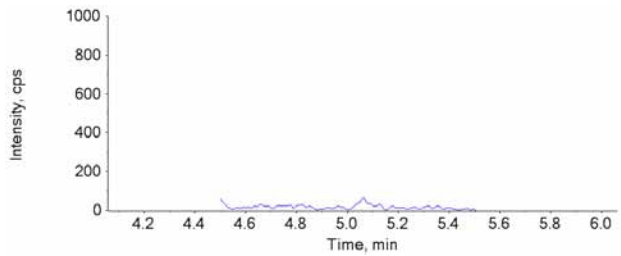


Figure 87. Typical Chromatogram of Quizalofop Analysis, Corn Grain 1.00 µg/g Recovery

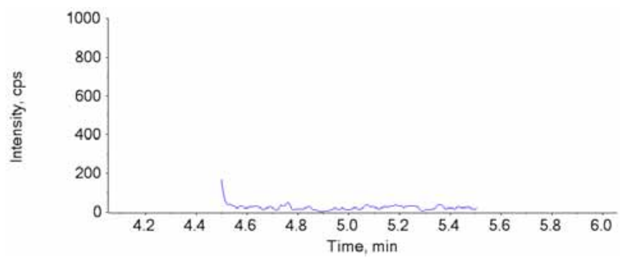
Sample Name: ARA-09-15-10-148-0002A1
Data File: ARA-09-15-10 Q02.wiff
Sample Index: 13
Acquired: 5/20/2010 3:04:39 AM
Analyte Peak Area: 0
Analyte Ion Transition: 343.1/108.1 amu



Sample Name: ARA-09-15-10-148-0002A1
Data File: ARA-09-15-10 Q02.wiff
Sample Index: 13
Acquired: 5/20/2010 3:04:39 AM
Analyte Peak Area: 0
Analyte Ion Transition: 373.2/299.0 amu



Sample Name: ARA-09-15-10-148-0002A1
Data File: ARA-09-15-10 Q02.wiff
Sample Index: 13
Acquired: 5/20/2010 3:04:39 AM
Analyte Peak Area: 0
Analyte Ion Transition: 373.2/91.0 amu



Sample Name: ARA-09-15-10-148-0002A1
Data File: ARA-09-15-10 Q02.wiff
Sample Index: 13
Acquired: 5/20/2010 3:04:39 AM
Analyte Peak Area: 0
Analyte Ion Transition: 343.1/270.9 amu

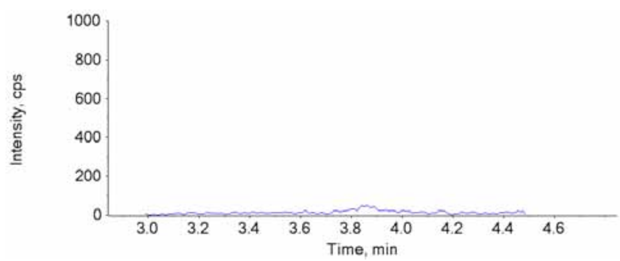
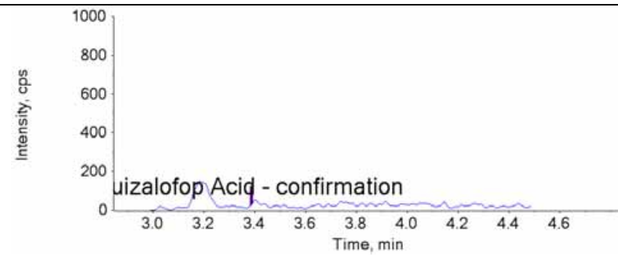
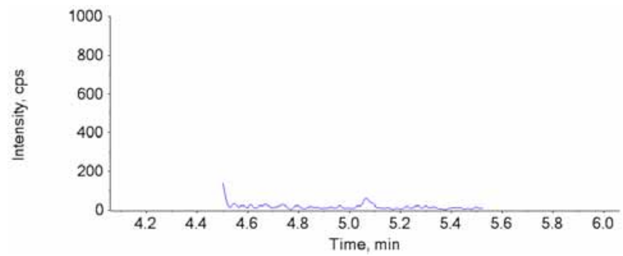


Figure 88. Typical Chromatogram of Quizalofop Analysis, Treated Corn Grain (148-0002)

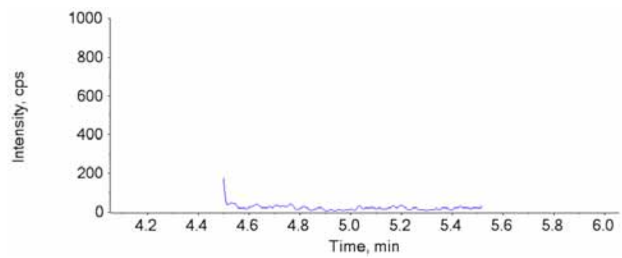
Sample Name: ARA-09-15-10-148-0003A1
Data File: ARA-09-15-10 Q02.wiff
Sample Index: 14
Acquired: 5/20/2010 3:17:09 AM
Analyte Peak Area: 0
Analyte Ion Transition: 343.1/108.1 amu



Sample Name: ARA-09-15-10-148-0003A1
Data File: ARA-09-15-10 Q02.wiff
Sample Index: 14
Acquired: 5/20/2010 3:17:09 AM
Analyte Peak Area: 0
Analyte Ion Transition: 373.2/299.0 amu



Sample Name: ARA-09-15-10-148-0003A1
Data File: ARA-09-15-10 Q02.wiff
Sample Index: 14
Acquired: 5/20/2010 3:17:09 AM
Analyte Peak Area: 0
Analyte Ion Transition: 373.2/91.0 amu



Sample Name: ARA-09-15-10-148-0003A1
Data File: ARA-09-15-10 Q02.wiff
Sample Index: 14
Acquired: 5/20/2010 3:17:09 AM
Analyte Peak Area: 0
Analyte Ion Transition: 343.1/270.9 amu

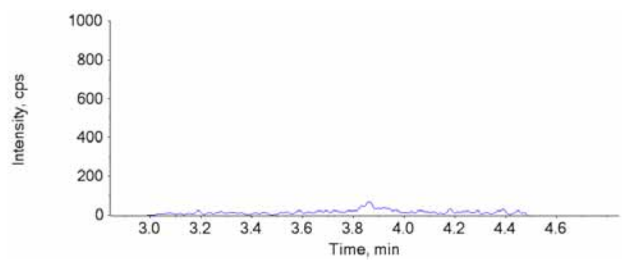
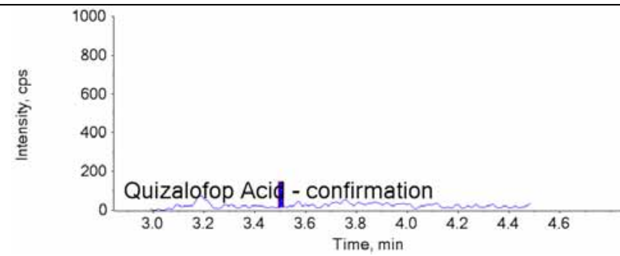
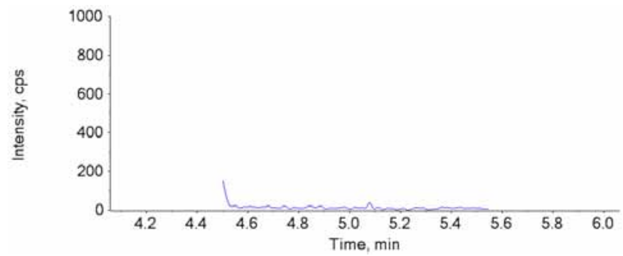


Figure 89. Typical Chromatogram of Quizalofop Analysis, Treated Corn Grain (148-0003)

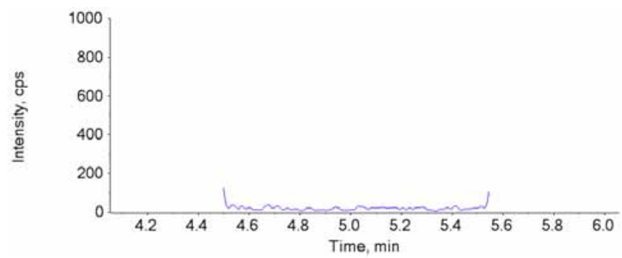
Sample Name: ARA-09-15-10-020-0002A1
Data File: ARA-09-15-10 Q02.wiff
Sample Index: 26
Acquired: 5/20/2010 5:46:45 AM
Analyte Peak Area: 0
Analyte Ion Transition: 343.1/108.1 amu



Sample Name: ARA-09-15-10-020-0002A1
Data File: ARA-09-15-10 Q02.wiff
Sample Index: 26
Acquired: 5/20/2010 5:46:45 AM
Analyte Peak Area: 0
Analyte Ion Transition: 373.2/299.0 amu



Sample Name: ARA-09-15-10-020-0002A1
Data File: ARA-09-15-10 Q02.wiff
Sample Index: 26
Acquired: 5/20/2010 5:46:45 AM
Analyte Peak Area: 0
Analyte Ion Transition: 373.2/91.0 amu



Sample Name: ARA-09-15-10-020-0002A1
Data File: ARA-09-15-10 Q02.wiff
Sample Index: 26
Acquired: 5/20/2010 5:46:45 AM
Analyte Peak Area: 0
Analyte Ion Transition: 343.1/270.9 amu

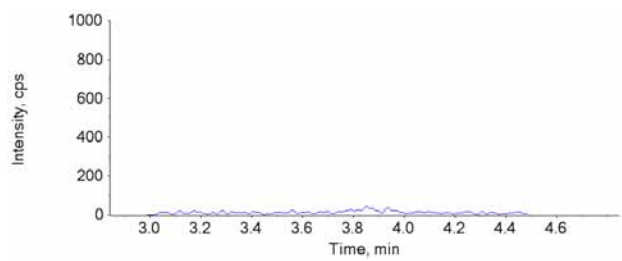
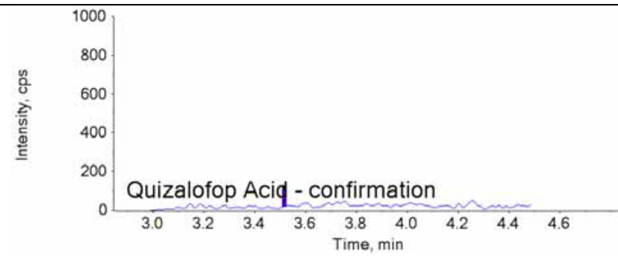
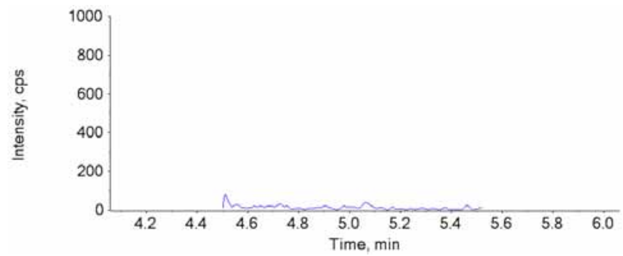


Figure 90. Typical Chromatogram of Quizalofop Analysis, Treated Corn Grain (020-0002)

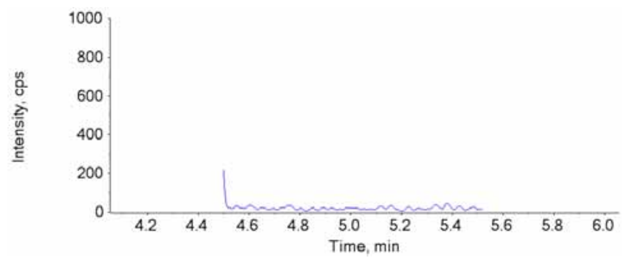
Sample Name: ARA-09-15-10-020-0003A1
Data File: ARA-09-15-10 Q02.wiff
Sample Index: 27
Acquired: 5/20/2010 5:59:11 AM
Analyte Peak Area: 0
Analyte Ion Transition: 343.1/108.1 amu



Sample Name: ARA-09-15-10-020-0003A1
Data File: ARA-09-15-10 Q02.wiff
Sample Index: 27
Acquired: 5/20/2010 5:59:11 AM
Analyte Peak Area: 0
Analyte Ion Transition: 373.2/299.0 amu



Sample Name: ARA-09-15-10-020-0003A1
Data File: ARA-09-15-10 Q02.wiff
Sample Index: 27
Acquired: 5/20/2010 5:59:11 AM
Analyte Peak Area: 0
Analyte Ion Transition: 373.2/91.0 amu



Sample Name: ARA-09-15-10-020-0003A1
Data File: ARA-09-15-10 Q02.wiff
Sample Index: 27
Acquired: 5/20/2010 5:59:11 AM
Analyte Peak Area: 0
Analyte Ion Transition: 343.1/270.9 amu

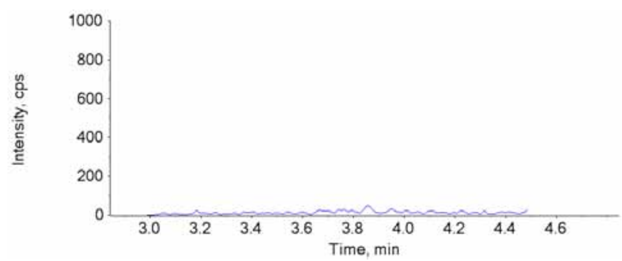
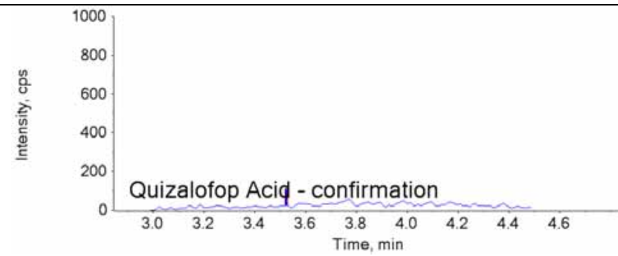
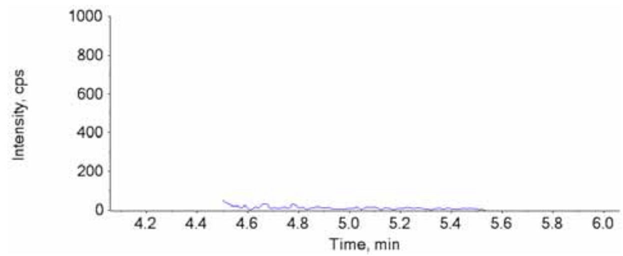


Figure 91. Typical Chromatogram of Quizalofop Analysis, Treated Corn Grain (020-0003)

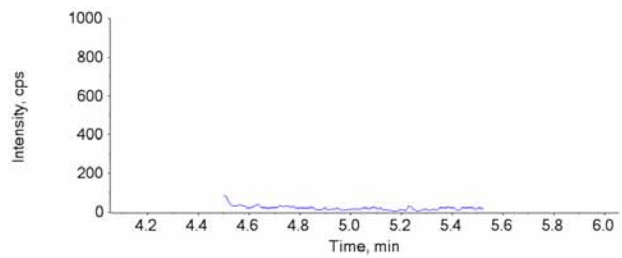
Sample Name: ARA-09-15-10-022-0002A1
Data File: ARA-09-15-10 Q02.wiff
Sample Index: 28
Acquired: 5/20/2010 6:11:39 AM
Analyte Peak Area: 0
Analyte Ion Transition: 343.1/108.1 amu



Sample Name: ARA-09-15-10-022-0002A1
Data File: ARA-09-15-10 Q02.wiff
Sample Index: 28
Acquired: 5/20/2010 6:11:39 AM
Analyte Peak Area: 0
Analyte Ion Transition: 373.2/299.0 amu



Sample Name: ARA-09-15-10-022-0002A1
Data File: ARA-09-15-10 Q02.wiff
Sample Index: 28
Acquired: 5/20/2010 6:11:39 AM
Analyte Peak Area: 0
Analyte Ion Transition: 373.2/91.0 amu



Sample Name: ARA-09-15-10-022-0002A1
Data File: ARA-09-15-10 Q02.wiff
Sample Index: 28
Acquired: 5/20/2010 6:11:39 AM
Analyte Peak Area: 0
Analyte Ion Transition: 343.1/270.9 amu

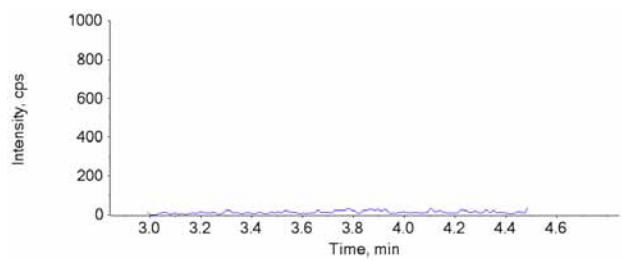
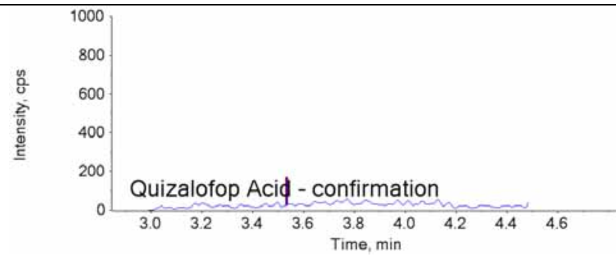
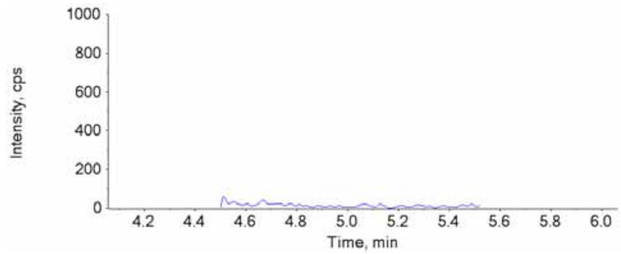


Figure 92. Typical Chromatogram of Quizalofop Analysis, Treated Corn Grain (022-0002)

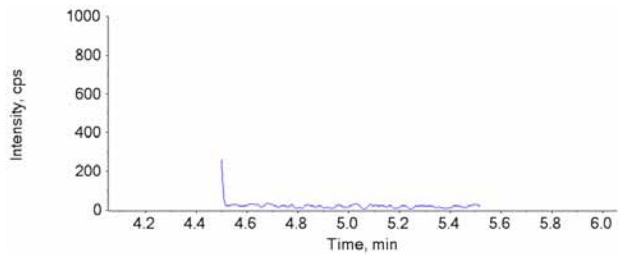
Sample Name: ARA-09-15-10-022-0003A1
Data File: ARA-09-15-10 Q02.wiff
Sample Index: 29
Acquired: 5/20/2010 6:24:09 AM
Analyte Peak Area: 0
Analyte Ion Transition: 343.1/108.1 amu



Sample Name: ARA-09-15-10-022-0003A1
Data File: ARA-09-15-10 Q02.wiff
Sample Index: 29
Acquired: 5/20/2010 6:24:09 AM
Analyte Peak Area: 0
Analyte Ion Transition: 373.2/299.0 amu



Sample Name: ARA-09-15-10-022-0003A1
Data File: ARA-09-15-10 Q02.wiff
Sample Index: 29
Acquired: 5/20/2010 6:24:09 AM
Analyte Peak Area: 0
Analyte Ion Transition: 373.2/91.0 amu



Sample Name: ARA-09-15-10-022-0003A1
Data File: ARA-09-15-10 Q02.wiff
Sample Index: 29
Acquired: 5/20/2010 6:24:09 AM
Analyte Peak Area: 0
Analyte Ion Transition: 343.1/270.9 amu

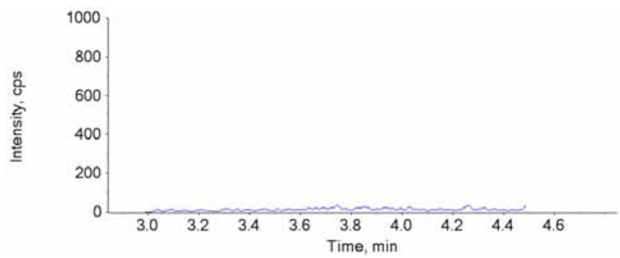
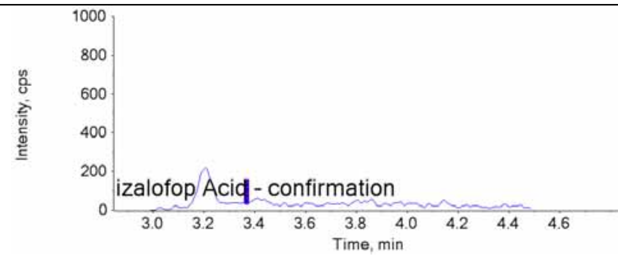
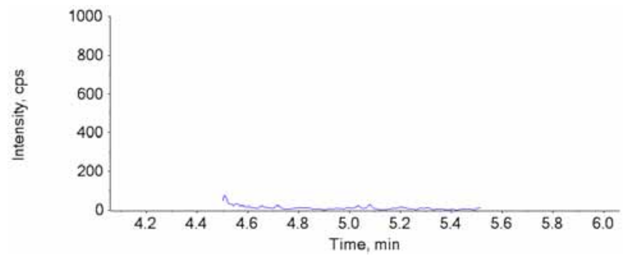


Figure 93. Typical Chromatogram of Quizalofop Analysis, Treated Corn Grain (022-0003)

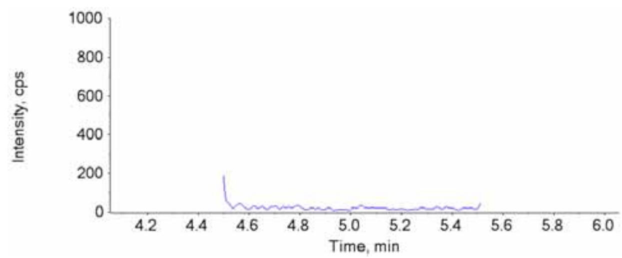
Sample Name: ARA-09-15-10-032-0002A1
Data File: ARA-09-15-10 Q02.wiff
Sample Index: 37
Acquired: 5/20/2010 8:04:00 AM
Analyte Peak Area: 0
Analyte Ion Transition: 343.1/108.1 amu



Sample Name: ARA-09-15-10-032-0002A1
Data File: ARA-09-15-10 Q02.wiff
Sample Index: 37
Acquired: 5/20/2010 8:04:00 AM
Analyte Peak Area: 0
Analyte Ion Transition: 373.2/299.0 amu



Sample Name: ARA-09-15-10-032-0002A1
Data File: ARA-09-15-10 Q02.wiff
Sample Index: 37
Acquired: 5/20/2010 8:04:00 AM
Analyte Peak Area: 0
Analyte Ion Transition: 373.2/91.0 amu



Sample Name: ARA-09-15-10-032-0002A1
Data File: ARA-09-15-10 Q02.wiff
Sample Index: 37
Acquired: 5/20/2010 8:04:00 AM
Analyte Peak Area: 0
Analyte Ion Transition: 343.1/270.9 amu

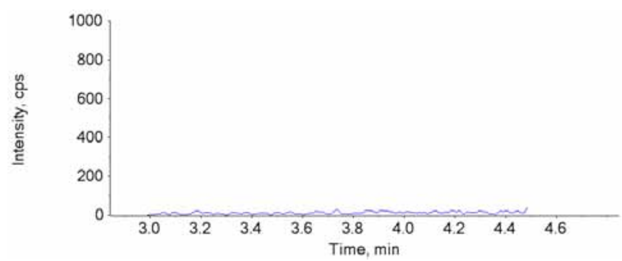
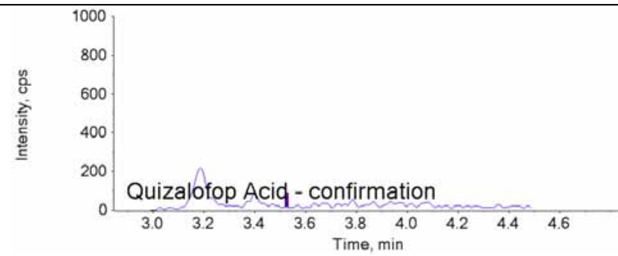
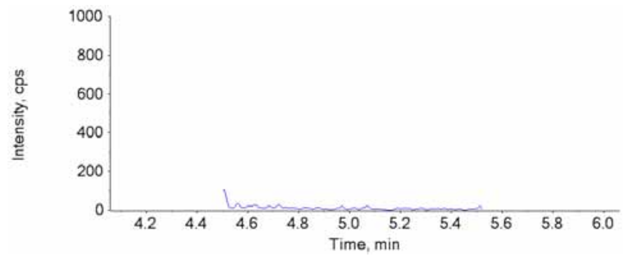


Figure 94. Typical Chromatogram of Quizalofop Analysis, Treated Corn Grain (032-0002)

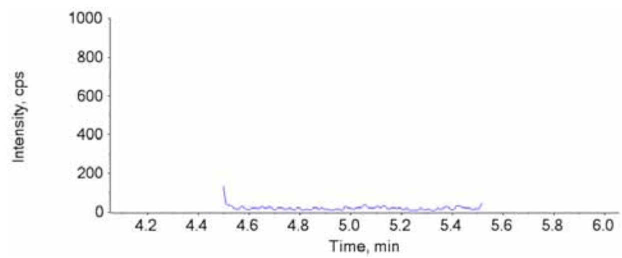
Sample Name: ARA-09-15-10-032-0003A1
Data File: ARA-09-15-10 Q02.wiff
Sample Index: 38
Acquired: 5/20/2010 8:16:28 AM
Analyte Peak Area: 0
Analyte Ion Transition: 343.1/108.1 amu



Sample Name: ARA-09-15-10-032-0003A1
Data File: ARA-09-15-10 Q02.wiff
Sample Index: 38
Acquired: 5/20/2010 8:16:28 AM
Analyte Peak Area: 0
Analyte Ion Transition: 373.2/299.0 amu



Sample Name: ARA-09-15-10-032-0003A1
Data File: ARA-09-15-10 Q02.wiff
Sample Index: 38
Acquired: 5/20/2010 8:16:28 AM
Analyte Peak Area: 0
Analyte Ion Transition: 373.2/91.0 amu



Sample Name: ARA-09-15-10-032-0003A1
Data File: ARA-09-15-10 Q02.wiff
Sample Index: 38
Acquired: 5/20/2010 8:16:28 AM
Analyte Peak Area: 0
Analyte Ion Transition: 343.1/270.9 amu

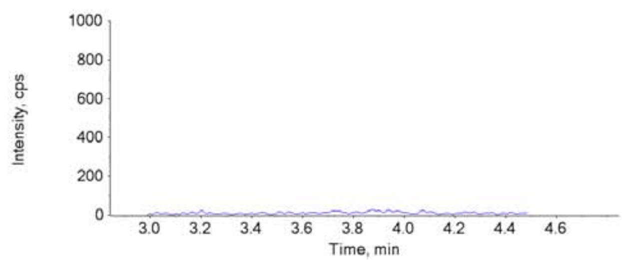
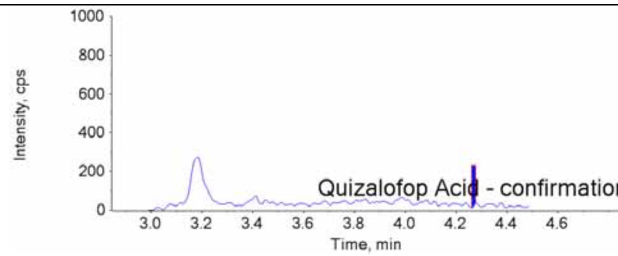
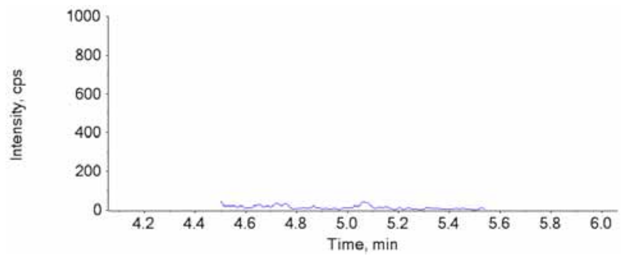


Figure 95. Typical Chromatogram of Quizalofop Analysis, Treated Corn Grain (032-0003)

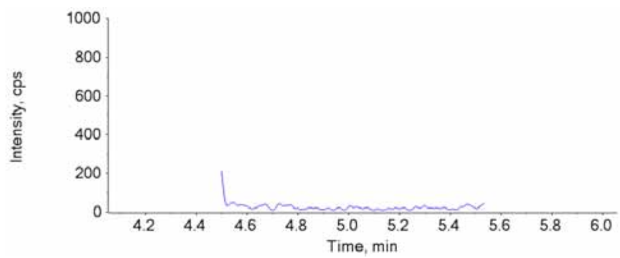
Sample Name: ARA-09-15-10-060-0002A1
Data File: ARA-09-15-10 Q02.wiff
Sample Index: 51
Acquired: 5/20/2010 10:58:35 AM
Analyte Peak Area: 0
Analyte Ion Transition: 343.1/108.1 amu



Sample Name: ARA-09-15-10-060-0002A1
Data File: ARA-09-15-10 Q02.wiff
Sample Index: 51
Acquired: 5/20/2010 10:58:35 AM
Analyte Peak Area: 0
Analyte Ion Transition: 373.2/299.0 amu



Sample Name: ARA-09-15-10-060-0002A1
Data File: ARA-09-15-10 Q02.wiff
Sample Index: 51
Acquired: 5/20/2010 10:58:35 AM
Analyte Peak Area: 0
Analyte Ion Transition: 373.2/91.0 amu



Sample Name: ARA-09-15-10-060-0002A1
Data File: ARA-09-15-10 Q02.wiff
Sample Index: 51
Acquired: 5/20/2010 10:58:35 AM
Analyte Peak Area: 0
Analyte Ion Transition: 343.1/270.9 amu

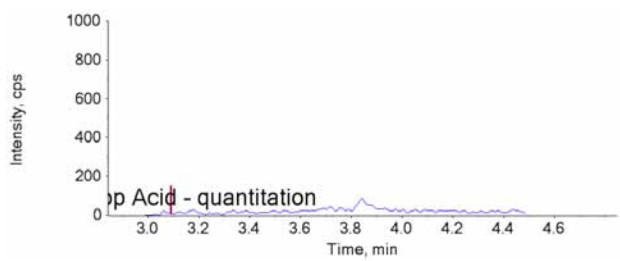
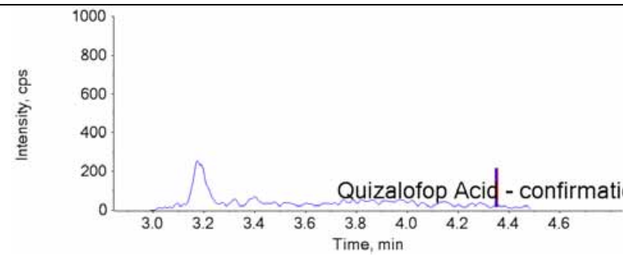
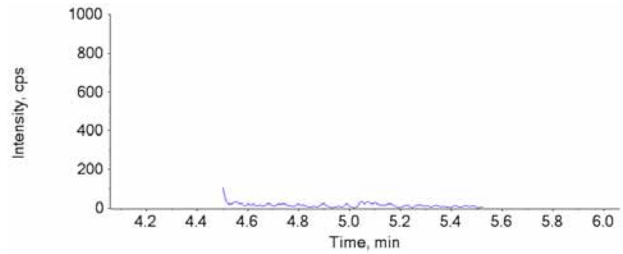


Figure 96. Typical Chromatogram of Quizalofop Analysis, Treated Corn Grain (060-0002)

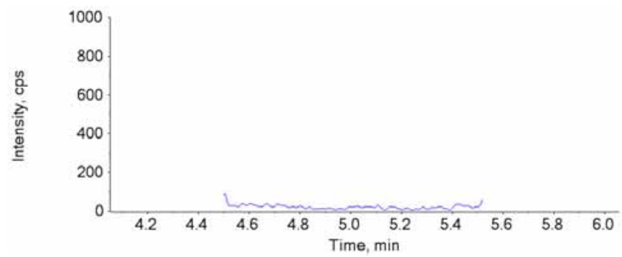
Sample Name: ARA-09-15-10-060-0003A1
Data File: ARA-09-15-10 Q02.wiff
Sample Index: 52
Acquired: 5/20/2010 11:11:02 AM
Analyte Peak Area: 0
Analyte Ion Transition: 343.1/108.1 amu



Sample Name: ARA-09-15-10-060-0003A1
Data File: ARA-09-15-10 Q02.wiff
Sample Index: 52
Acquired: 5/20/2010 11:11:02 AM
Analyte Peak Area: 0
Analyte Ion Transition: 373.2/299.0 amu



Sample Name: ARA-09-15-10-060-0003A1
Data File: ARA-09-15-10 Q02.wiff
Sample Index: 52
Acquired: 5/20/2010 11:11:02 AM
Analyte Peak Area: 0
Analyte Ion Transition: 373.2/91.0 amu



Sample Name: ARA-09-15-10-060-0003A1
Data File: ARA-09-15-10 Q02.wiff
Sample Index: 52
Acquired: 5/20/2010 11:11:02 AM
Analyte Peak Area: 0
Analyte Ion Transition: 343.1/270.9 amu

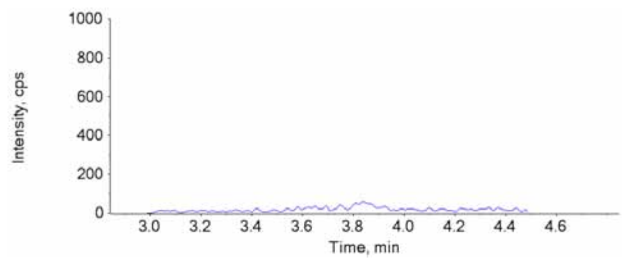
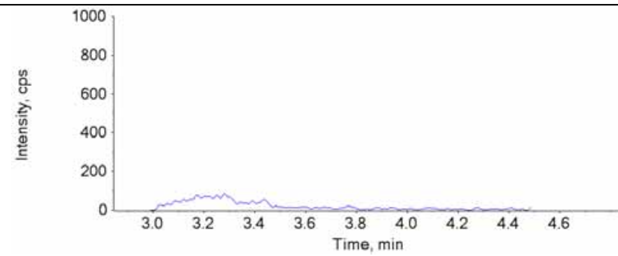
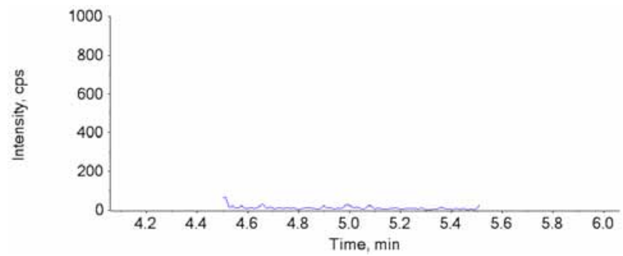


Figure 97. Typical Chromatogram of Quizalofop Analysis, Treated Corn Grain (060-0003)

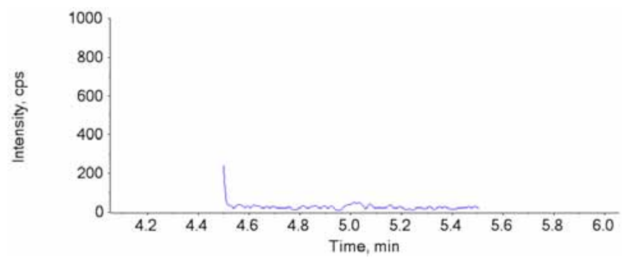
Sample Name: ARA-09-15-10-157-0001A7
Data File: ARA-09-15-10 Q05c.wiff
Sample Index: 2
Acquired: 6/1/2010 4:02:10 PM
Analyte Peak Area: 0
Analyte Ion Transition: 343.1/108.1 amu



Sample Name: ARA-09-15-10-157-0001A7
Data File: ARA-09-15-10 Q05c.wiff
Sample Index: 2
Acquired: 6/1/2010 4:02:10 PM
Analyte Peak Area: 0
Analyte Ion Transition: 373.2/299.0 amu



Sample Name: ARA-09-15-10-157-0001A7
Data File: ARA-09-15-10 Q05c.wiff
Sample Index: 2
Acquired: 6/1/2010 4:02:10 PM
Analyte Peak Area: 0
Analyte Ion Transition: 373.2/91.0 amu



Sample Name: ARA-09-15-10-157-0001A7
Data File: ARA-09-15-10 Q05c.wiff
Sample Index: 2
Acquired: 6/1/2010 4:02:10 PM
Analyte Peak Area: 0
Analyte Ion Transition: 343.1/270.9 amu

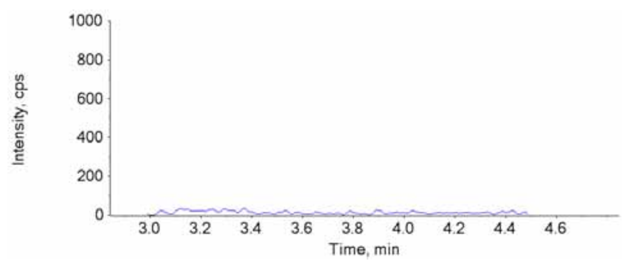
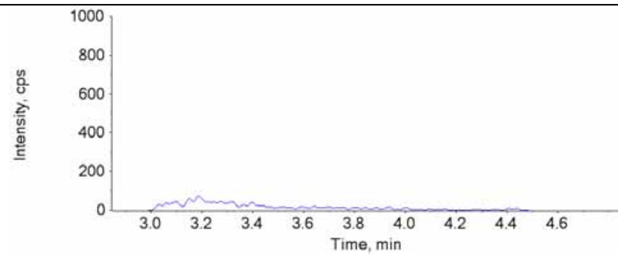
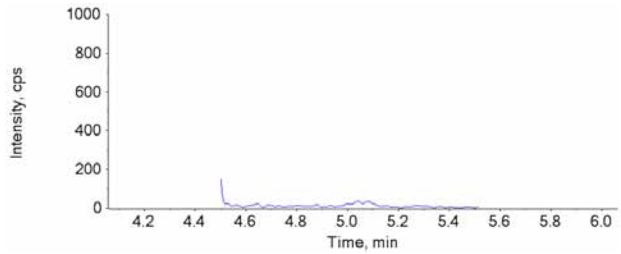


Figure 98. Typical Chromatogram of Quizalofop Analysis, Control Corn Forage (157-0001)

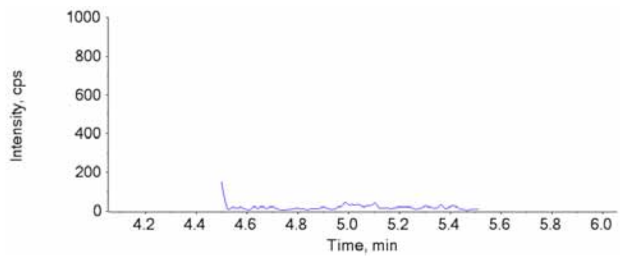
Sample Name: ARA-09-15-10-095-0001A1
Data File: ARA-09-15-10 Q05c.wiff
Sample Index: 9
Acquired: 6/1/2010 5:29:31 PM
Analyte Peak Area: 0
Analyte Ion Transition: 343.1/108.1 amu



Sample Name: ARA-09-15-10-095-0001A1
Data File: ARA-09-15-10 Q05c.wiff
Sample Index: 9
Acquired: 6/1/2010 5:29:31 PM
Analyte Peak Area: 0
Analyte Ion Transition: 373.2/299.0 amu



Sample Name: ARA-09-15-10-095-0001A1
Data File: ARA-09-15-10 Q05c.wiff
Sample Index: 9
Acquired: 6/1/2010 5:29:31 PM
Analyte Peak Area: 0
Analyte Ion Transition: 373.2/91.0 amu



Sample Name: ARA-09-15-10-095-0001A1
Data File: ARA-09-15-10 Q05c.wiff
Sample Index: 9
Acquired: 6/1/2010 5:29:31 PM
Analyte Peak Area: 0
Analyte Ion Transition: 343.1/270.9 amu

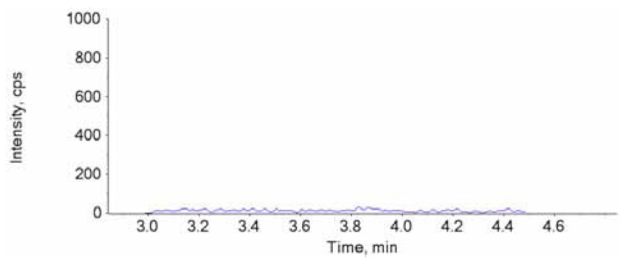
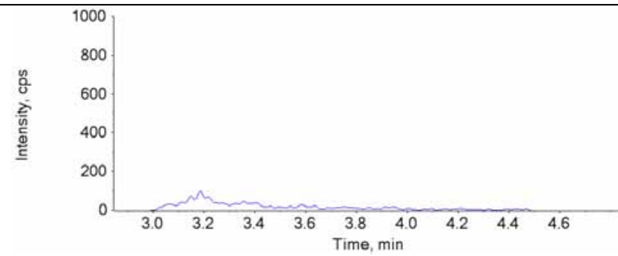
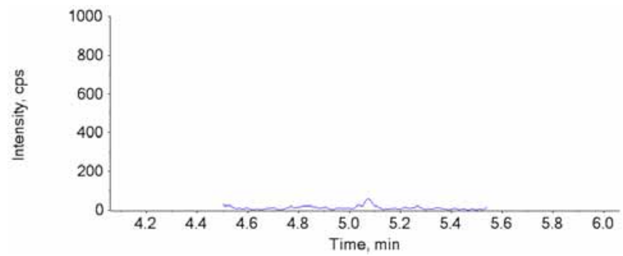


Figure 99. Typical Chromatogram of Quizalofop Analysis, Control Corn Forage (095-0001)

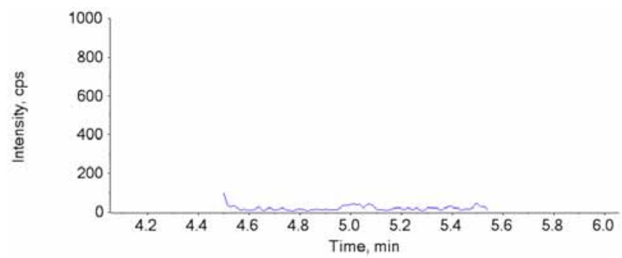
Sample Name: ARA-09-15-10-096-0001A1
Data File: ARA-09-15-10 Q05c.wiff
Sample Index: 10
Acquired: 6/1/2010 5:42:02 PM
Analyte Peak Area: 0
Analyte Ion Transition: 343.1/108.1 amu



Sample Name: ARA-09-15-10-096-0001A1
Data File: ARA-09-15-10 Q05c.wiff
Sample Index: 10
Acquired: 6/1/2010 5:42:02 PM
Analyte Peak Area: 0
Analyte Ion Transition: 373.2/299.0 amu



Sample Name: ARA-09-15-10-096-0001A1
Data File: ARA-09-15-10 Q05c.wiff
Sample Index: 10
Acquired: 6/1/2010 5:42:02 PM
Analyte Peak Area: 0
Analyte Ion Transition: 373.2/91.0 amu



Sample Name: ARA-09-15-10-096-0001A1
Data File: ARA-09-15-10 Q05c.wiff
Sample Index: 10
Acquired: 6/1/2010 5:42:02 PM
Analyte Peak Area: 0
Analyte Ion Transition: 343.1/270.9 amu

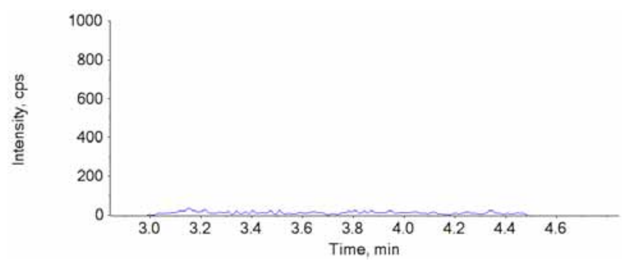
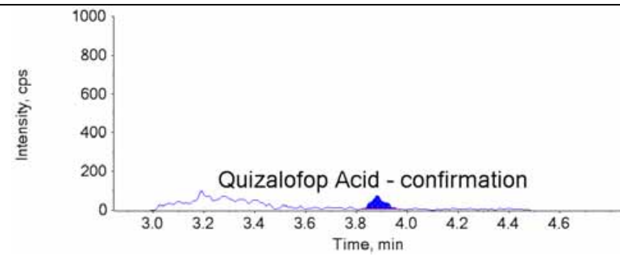
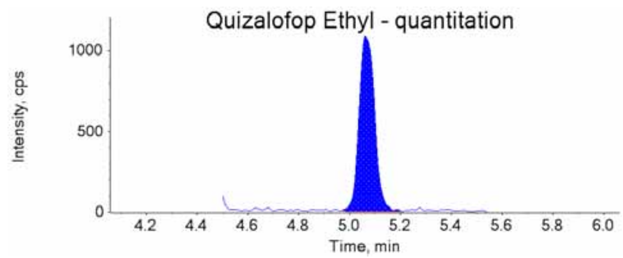


Figure 100. Typical Chromatogram of Quizalofop Analysis, Control Corn Forage (096-0001)

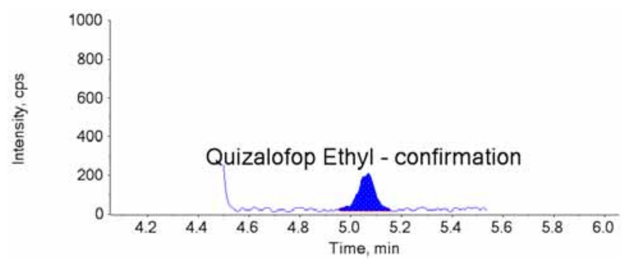
Sample Name: ARA-09-15-10-157-0001A9 + 0.01 µg/g
Data File: ARA-09-15-10 Q05c.wiff
Sample Index: 4
Acquired: 6/1/2010 4:27:09 PM
Analyte Peak Area: 237
Analyte Ion Transition: 343.1/108.1 amu



Sample Name: ARA-09-15-10-157-0001A9 + 0.01 µg/g
Data File: ARA-09-15-10 Q05c.wiff
Sample Index: 4
Acquired: 6/1/2010 4:27:09 PM
Analyte Peak Area: 4646
Analyte Ion Transition: 373.2/299.0 amu



Sample Name: ARA-09-15-10-157-0001A9 + 0.01 µg/g
Data File: ARA-09-15-10 Q05c.wiff
Sample Index: 4
Acquired: 6/1/2010 4:27:09 PM
Analyte Peak Area: 926
Analyte Ion Transition: 373.2/91.0 amu



Sample Name: ARA-09-15-10-157-0001A9 + 0.01 µg/g
Data File: ARA-09-15-10 Q05c.wiff
Sample Index: 4
Acquired: 6/1/2010 4:27:09 PM
Analyte Peak Area: 3092
Analyte Ion Transition: 343.1/270.9 amu

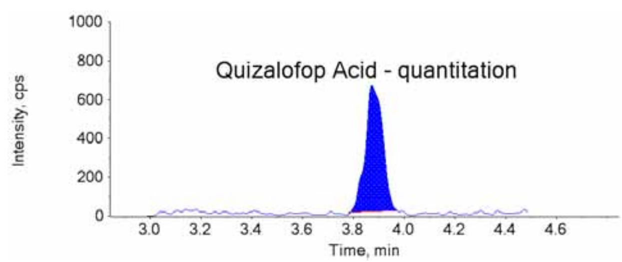
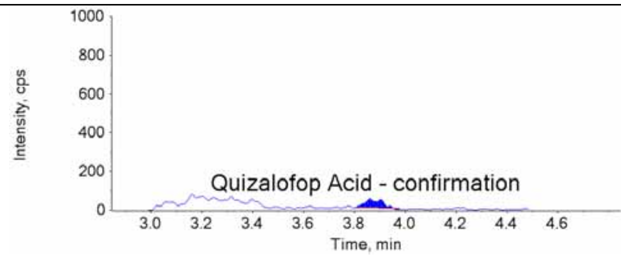
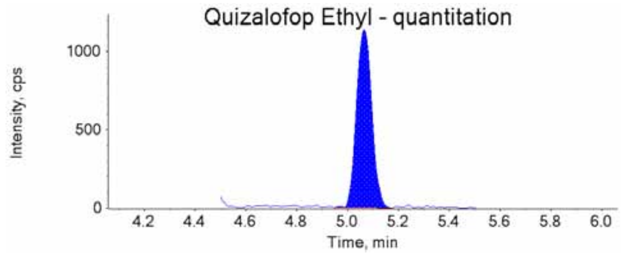


Figure 101. Typical Chromatogram of Quizalofop Analysis, Corn Forage 0.01 µg/g Recovery

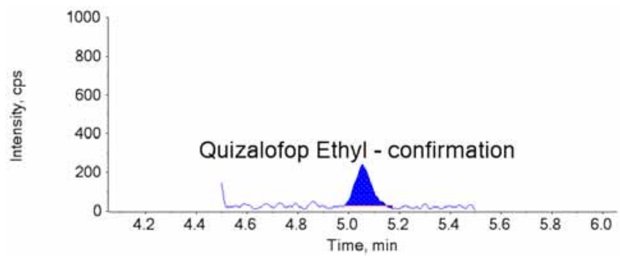
Sample Name: ARA-09-15-10-157-0001A10 + 0.01 µg/g
Data File: ARA-09-15-10 Q05c.wiff
Sample Index: 5
Acquired: 6/1/2010 4:39:37 PM
Analyte Peak Area: 214
Analyte Ion Transition: 343.1/108.1 amu



Sample Name: ARA-09-15-10-157-0001A10 + 0.01 µg/g
Data File: ARA-09-15-10 Q05c.wiff
Sample Index: 5
Acquired: 6/1/2010 4:39:37 PM
Analyte Peak Area: 4758
Analyte Ion Transition: 373.2/299.0 amu



Sample Name: ARA-09-15-10-157-0001A10 + 0.01 µg/g
Data File: ARA-09-15-10 Q05c.wiff
Sample Index: 5
Acquired: 6/1/2010 4:39:37 PM
Analyte Peak Area: 924
Analyte Ion Transition: 373.2/91.0 amu



Sample Name: ARA-09-15-10-157-0001A10 + 0.01 µg/g
Data File: ARA-09-15-10 Q05c.wiff
Sample Index: 5
Acquired: 6/1/2010 4:39:37 PM
Analyte Peak Area: 3071
Analyte Ion Transition: 343.1/270.9 amu

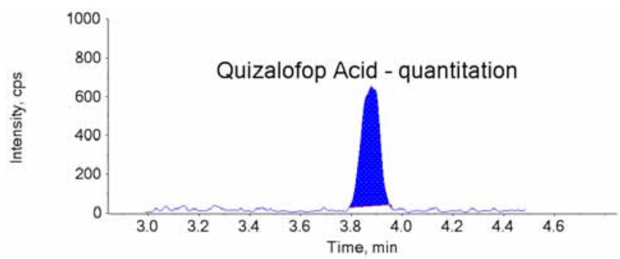
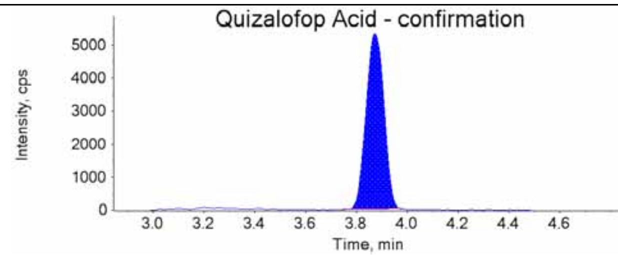
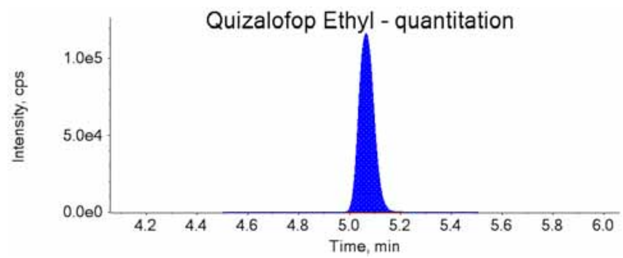


Figure 102. Typical Chromatogram of Quizalofop Analysis, Corn Forage 0.01 µg/g Recovery

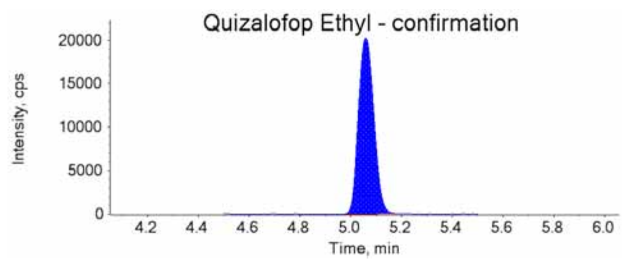
Sample Name: ARA-09-15-10-157-0001A11 + 1.0 µg/g
Data File: ARA-09-15-10 Q05c.wiff
Sample Index: 6
Acquired: 6/1/2010 4:52:06 PM
Analyte Peak Area: 25982
Analyte Ion Transition: 343.1/108.1 amu



Sample Name: ARA-09-15-10-157-0001A11 + 1.0 µg/g
Data File: ARA-09-15-10 Q05c.wiff
Sample Index: 6
Acquired: 6/1/2010 4:52:06 PM
Analyte Peak Area: 489698
Analyte Ion Transition: 373.2/299.0 amu



Sample Name: ARA-09-15-10-157-0001A11 + 1.0 µg/g
Data File: ARA-09-15-10 Q05c.wiff
Sample Index: 6
Acquired: 6/1/2010 4:52:06 PM
Analyte Peak Area: 85705
Analyte Ion Transition: 373.2/91.0 amu



Sample Name: ARA-09-15-10-157-0001A11 + 1.0 µg/g
Data File: ARA-09-15-10 Q05c.wiff
Sample Index: 6
Acquired: 6/1/2010 4:52:06 PM
Analyte Peak Area: 344059
Analyte Ion Transition: 343.1/270.9 amu

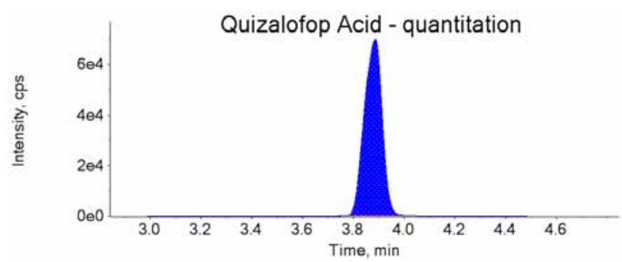
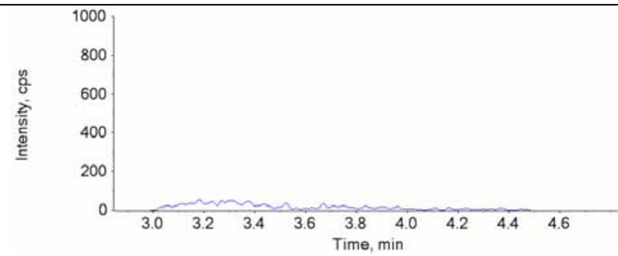
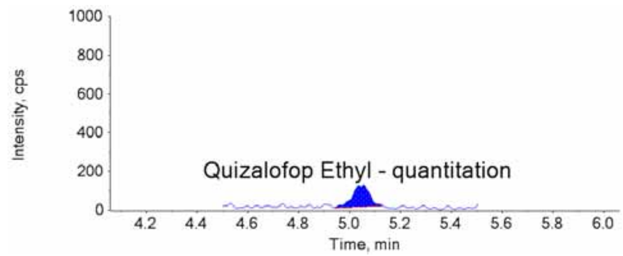


Figure 103. Typical Chromatogram of Quizalofop Analysis, Corn Forage 1.00 µg/g Recovery

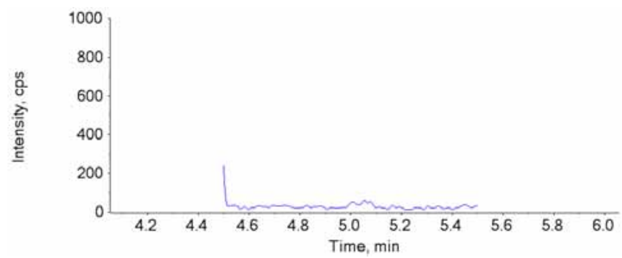
Sample Name: ARA-09-15-10-009-0002A1
Data File: ARA-09-15-10 Q05c.wiff
Sample Index: 41
Acquired: 6/2/2010 12:09:04 AM
Analyte Peak Area: 0
Analyte Ion Transition: 343.1/108.1 amu



Sample Name: ARA-09-15-10-009-0002A1
Data File: ARA-09-15-10 Q05c.wiff
Sample Index: 41
Acquired: 6/2/2010 12:09:04 AM
Analyte Peak Area: 531
Analyte Ion Transition: 373.2/299.0 amu



Sample Name: ARA-09-15-10-009-0002A1
Data File: ARA-09-15-10 Q05c.wiff
Sample Index: 41
Acquired: 6/2/2010 12:09:04 AM
Analyte Peak Area: 0
Analyte Ion Transition: 373.2/91.0 amu



Sample Name: ARA-09-15-10-009-0002A1
Data File: ARA-09-15-10 Q05c.wiff
Sample Index: 41
Acquired: 6/2/2010 12:09:04 AM
Analyte Peak Area: 0
Analyte Ion Transition: 343.1/270.9 amu

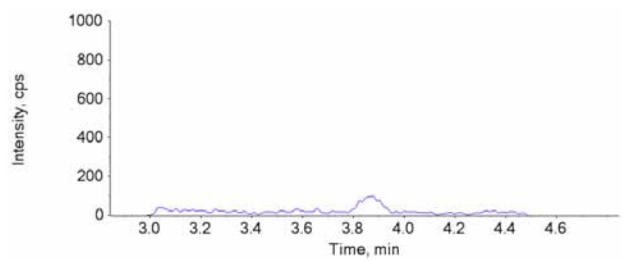
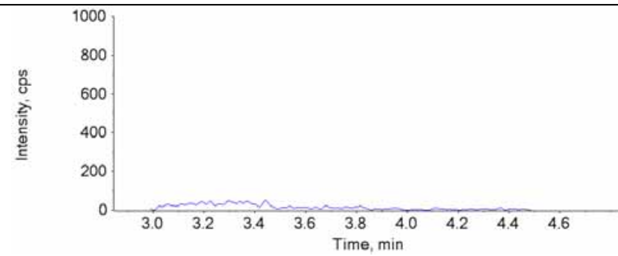
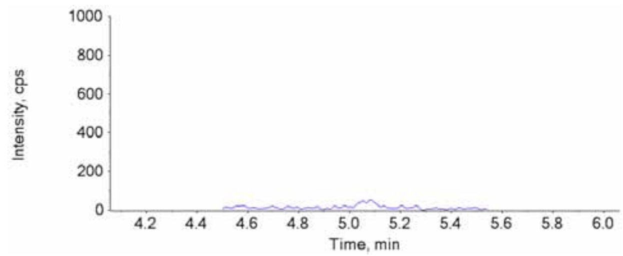


Figure 104. Typical Chromatogram of Quizalofop Analysis, Treated Corn Forage (009-0002)

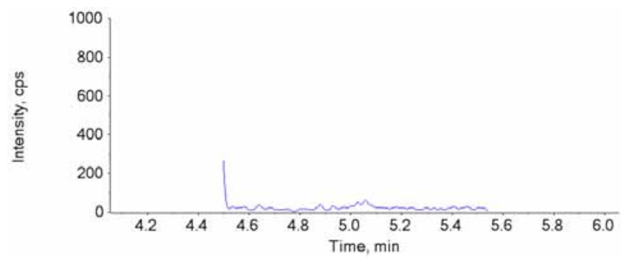
Sample Name: ARA-09-15-10-009-0003A1
Data File: ARA-09-15-10 Q05c.wiff
Sample Index: 42
Acquired: 6/2/2010 12:21:33 AM
Analyte Peak Area: 0
Analyte Ion Transition: 343.1/108.1 amu



Sample Name: ARA-09-15-10-009-0003A1
Data File: ARA-09-15-10 Q05c.wiff
Sample Index: 42
Acquired: 6/2/2010 12:21:33 AM
Analyte Peak Area: 0
Analyte Ion Transition: 373.2/299.0 amu



Sample Name: ARA-09-15-10-009-0003A1
Data File: ARA-09-15-10 Q05c.wiff
Sample Index: 42
Acquired: 6/2/2010 12:21:33 AM
Analyte Peak Area: 0
Analyte Ion Transition: 373.2/91.0 amu



Sample Name: ARA-09-15-10-009-0003A1
Data File: ARA-09-15-10 Q05c.wiff
Sample Index: 42
Acquired: 6/2/2010 12:21:33 AM
Analyte Peak Area: 0
Analyte Ion Transition: 343.1/270.9 amu

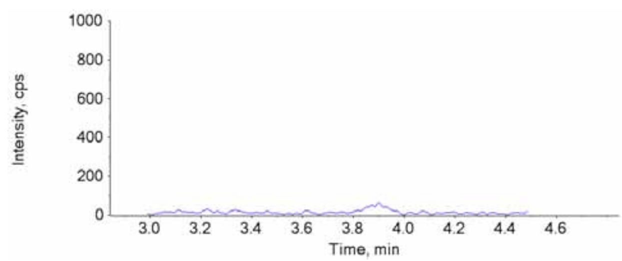
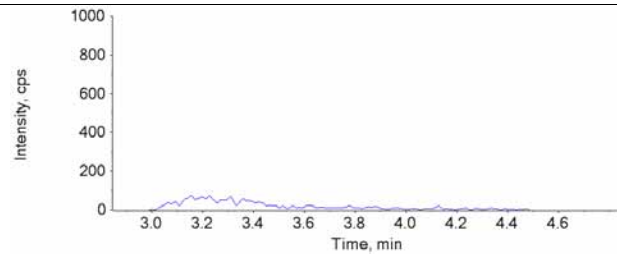
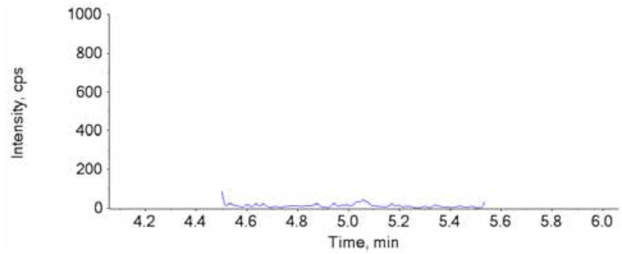


Figure 105. Typical Chromatogram of Quizalofop Analysis, Treated Corn Forage (009-0003)

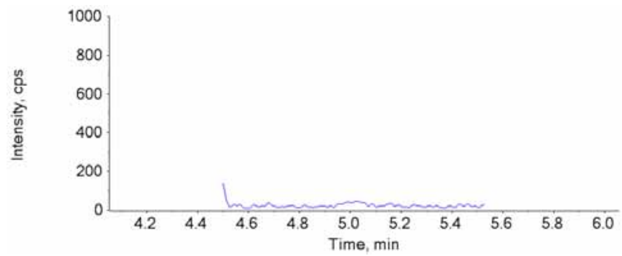
Sample Name: ARA-09-15-10-010-0002A1
Data File: ARA-09-15-10 Q05c.wiff
Sample Index: 43
Acquired: 6/2/2010 12:34:04 AM
Analyte Peak Area: 0
Analyte Ion Transition: 343.1/108.1 amu



Sample Name: ARA-09-15-10-010-0002A1
Data File: ARA-09-15-10 Q05c.wiff
Sample Index: 43
Acquired: 6/2/2010 12:34:04 AM
Analyte Peak Area: 0
Analyte Ion Transition: 373.2/299.0 amu



Sample Name: ARA-09-15-10-010-0002A1
Data File: ARA-09-15-10 Q05c.wiff
Sample Index: 43
Acquired: 6/2/2010 12:34:04 AM
Analyte Peak Area: 0
Analyte Ion Transition: 373.2/91.0 amu



Sample Name: ARA-09-15-10-010-0002A1
Data File: ARA-09-15-10 Q05c.wiff
Sample Index: 43
Acquired: 6/2/2010 12:34:04 AM
Analyte Peak Area: 0
Analyte Ion Transition: 343.1/270.9 amu

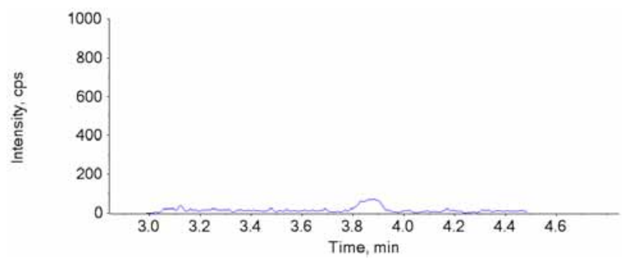
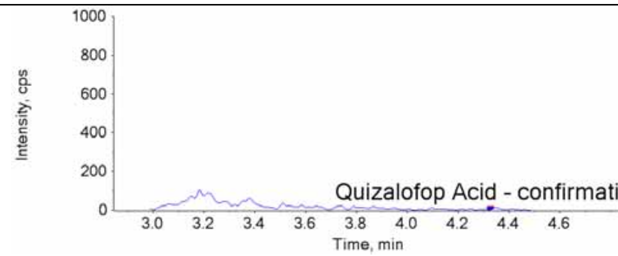
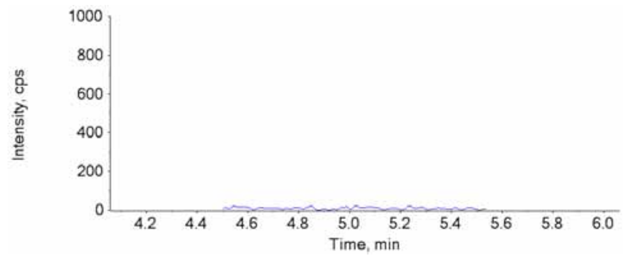


Figure 106. Typical Chromatogram of Quizalofop Analysis, Treated Corn Forage (010-0002)

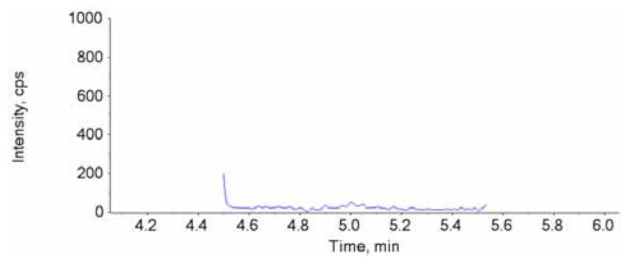
Sample Name: ARA-09-15-10-010-0003A1
Data File: ARA-09-15-10 Q05c.wiff
Sample Index: 44
Acquired: 6/2/2010 12:46:33 AM
Analyte Peak Area: 0
Analyte Ion Transition: 343.1/108.1 amu



Sample Name: ARA-09-15-10-010-0003A1
Data File: ARA-09-15-10 Q05c.wiff
Sample Index: 44
Acquired: 6/2/2010 12:46:33 AM
Analyte Peak Area: 0
Analyte Ion Transition: 373.2/299.0 amu



Sample Name: ARA-09-15-10-010-0003A1
Data File: ARA-09-15-10 Q05c.wiff
Sample Index: 44
Acquired: 6/2/2010 12:46:33 AM
Analyte Peak Area: 0
Analyte Ion Transition: 373.2/91.0 amu



Sample Name: ARA-09-15-10-010-0003A1
Data File: ARA-09-15-10 Q05c.wiff
Sample Index: 44
Acquired: 6/2/2010 12:46:33 AM
Analyte Peak Area: 0
Analyte Ion Transition: 343.1/270.9 amu

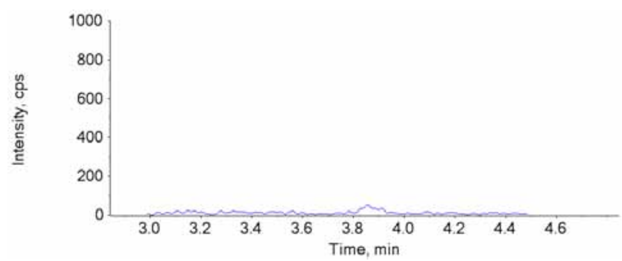
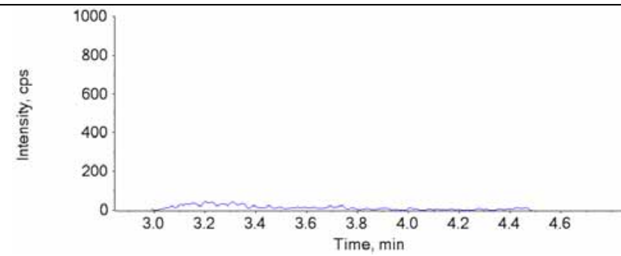
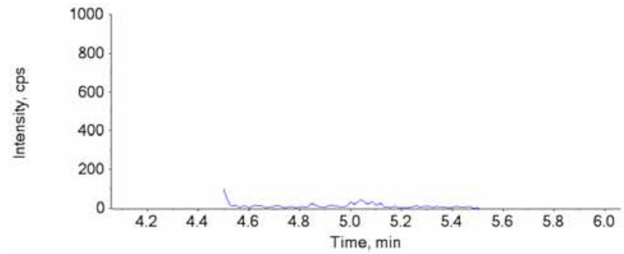


Figure 107. Typical Chromatogram of Quizalofop Analysis, Treated Corn Forage (010-0003)

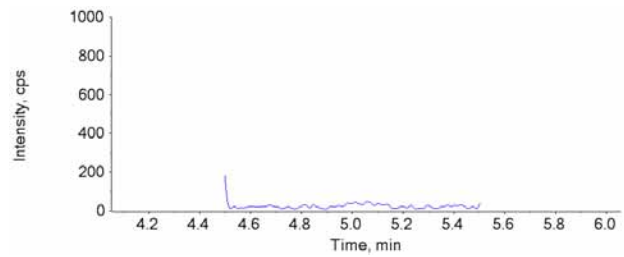
Sample Name: ARA-09-15-10-014-0002A1
Data File: ARA-09-15-10 Q05c.wiff
Sample Index: 45
Acquired: 6/2/2010 12:59:00 AM
Analyte Peak Area: 0
Analyte Ion Transition: 343.1/108.1 amu



Sample Name: ARA-09-15-10-014-0002A1
Data File: ARA-09-15-10 Q05c.wiff
Sample Index: 45
Acquired: 6/2/2010 12:59:00 AM
Analyte Peak Area: 0
Analyte Ion Transition: 373.2/299.0 amu



Sample Name: ARA-09-15-10-014-0002A1
Data File: ARA-09-15-10 Q05c.wiff
Sample Index: 45
Acquired: 6/2/2010 12:59:00 AM
Analyte Peak Area: 0
Analyte Ion Transition: 373.2/91.0 amu



Sample Name: ARA-09-15-10-014-0002A1
Data File: ARA-09-15-10 Q05c.wiff
Sample Index: 45
Acquired: 6/2/2010 12:59:00 AM
Analyte Peak Area: 0
Analyte Ion Transition: 343.1/270.9 amu

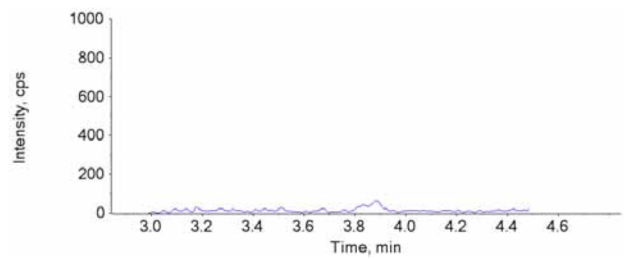
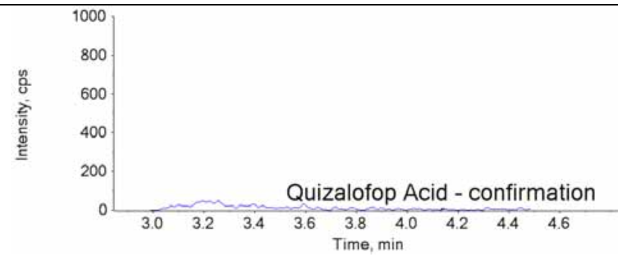
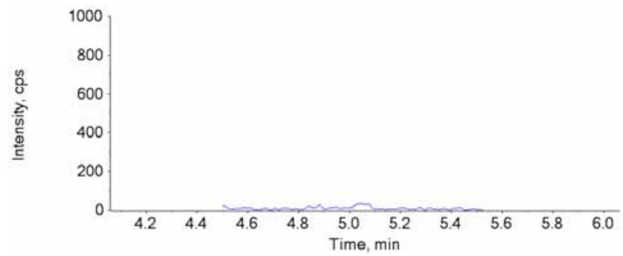


Figure 108. Typical Chromatogram of Quizalofop Analysis, Treated Corn Forage (014-0002)

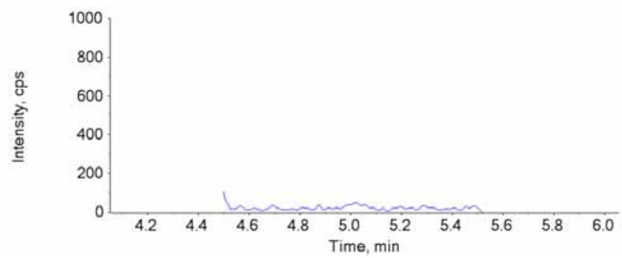
Sample Name: ARA-09-15-10-014-0003A1
Data File: ARA-09-15-10 Q05c.wiff
Sample Index: 46
Acquired: 6/2/2010 1:11:29 AM
Analyte Peak Area: 0
Analyte Ion Transition: 343.1/108.1 amu



Sample Name: ARA-09-15-10-014-0003A1
Data File: ARA-09-15-10 Q05c.wiff
Sample Index: 46
Acquired: 6/2/2010 1:11:29 AM
Analyte Peak Area: 0
Analyte Ion Transition: 373.2/299.0 amu



Sample Name: ARA-09-15-10-014-0003A1
Data File: ARA-09-15-10 Q05c.wiff
Sample Index: 46
Acquired: 6/2/2010 1:11:29 AM
Analyte Peak Area: 0
Analyte Ion Transition: 373.2/91.0 amu



Sample Name: ARA-09-15-10-014-0003A1
Data File: ARA-09-15-10 Q05c.wiff
Sample Index: 46
Acquired: 6/2/2010 1:11:29 AM
Analyte Peak Area: 0
Analyte Ion Transition: 343.1/270.9 amu

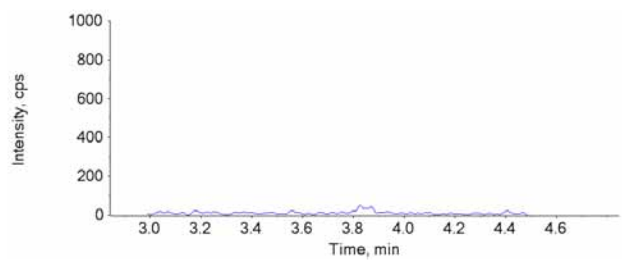
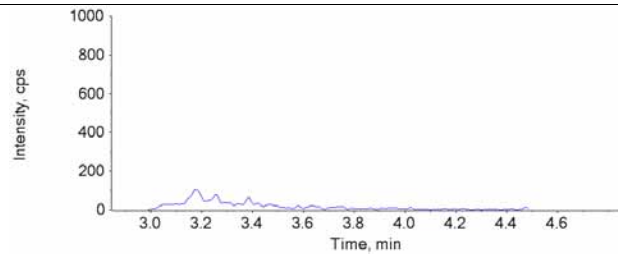
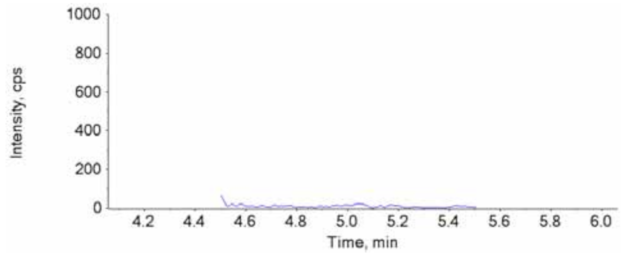


Figure 109. Typical Chromatogram of Quizalofop Analysis, Treated Corn Forage (014-0003)

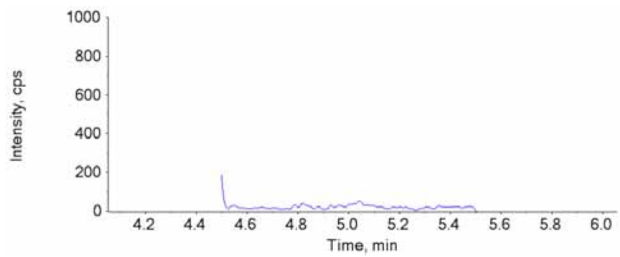
Sample Name: ARA-09-15-10-015-0002A1
Data File: ARA-09-15-10 Q05c.wiff
Sample Index: 47
Acquired: 6/2/2010 1:23:56 AM
Analyte Peak Area: 0
Analyte Ion Transition: 343.1/108.1 amu



Sample Name: ARA-09-15-10-015-0002A1
Data File: ARA-09-15-10 Q05c.wiff
Sample Index: 47
Acquired: 6/2/2010 1:23:56 AM
Analyte Peak Area: 0
Analyte Ion Transition: 373.2/299.0 amu



Sample Name: ARA-09-15-10-015-0002A1
Data File: ARA-09-15-10 Q05c.wiff
Sample Index: 47
Acquired: 6/2/2010 1:23:56 AM
Analyte Peak Area: 0
Analyte Ion Transition: 373.2/91.0 amu



Sample Name: ARA-09-15-10-015-0002A1
Data File: ARA-09-15-10 Q05c.wiff
Sample Index: 47
Acquired: 6/2/2010 1:23:56 AM
Analyte Peak Area: 0
Analyte Ion Transition: 343.1/270.9 amu

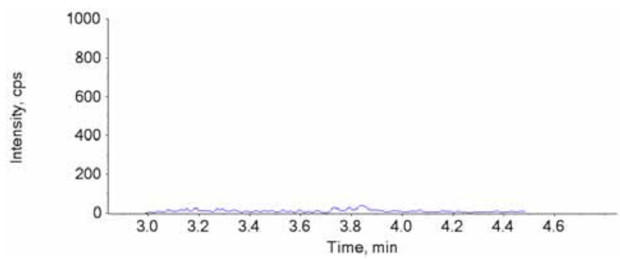
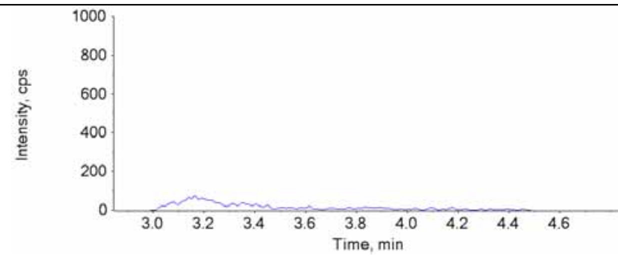
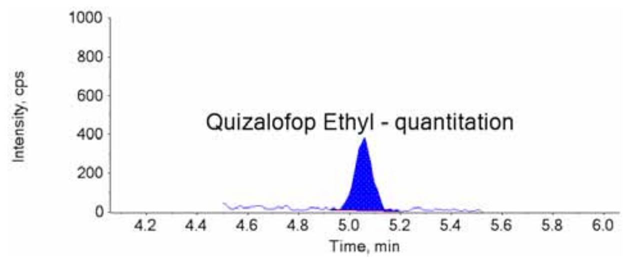


Figure 110. Typical Chromatogram of Quizalofop Analysis, Treated Corn Forage (015-0002)

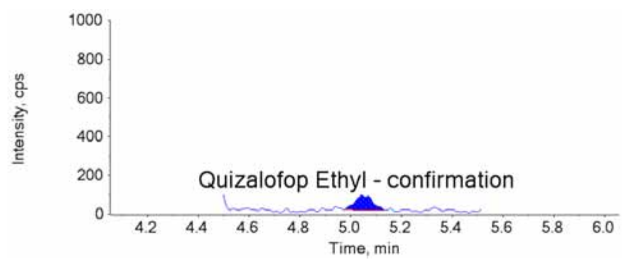
Sample Name: ARA-09-15-10-015-0003A1
Data File: ARA-09-15-10 Q05c.wiff
Sample Index: 49
Acquired: 6/2/2010 1:48:52 AM
Analyte Peak Area: 0
Analyte Ion Transition: 343.1/108.1 amu



Sample Name: ARA-09-15-10-015-0003A1
Data File: ARA-09-15-10 Q05c.wiff
Sample Index: 49
Acquired: 6/2/2010 1:48:52 AM
Analyte Peak Area: 1824
Analyte Ion Transition: 373.2/299.0 amu



Sample Name: ARA-09-15-10-015-0003A1
Data File: ARA-09-15-10 Q05c.wiff
Sample Index: 49
Acquired: 6/2/2010 1:48:52 AM
Analyte Peak Area: 375
Analyte Ion Transition: 373.2/91.0 amu



Sample Name: ARA-09-15-10-015-0003A1
Data File: ARA-09-15-10 Q05c.wiff
Sample Index: 49
Acquired: 6/2/2010 1:48:52 AM
Analyte Peak Area: 517
Analyte Ion Transition: 343.1/270.9 amu

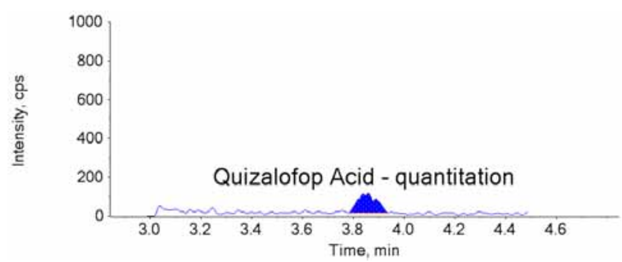
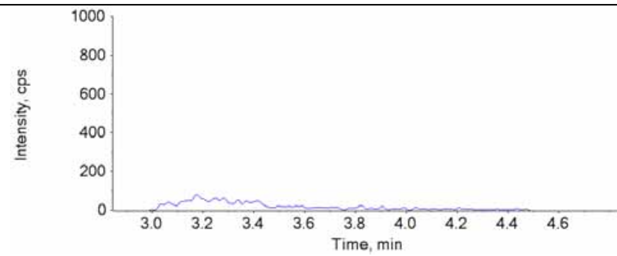
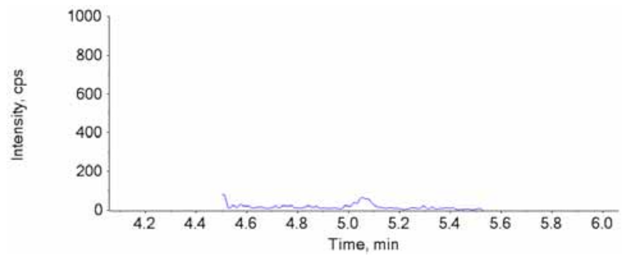


Figure 111. Typical Chromatogram of Quizalofop Analysis, Treated Corn Forage (015-0003)

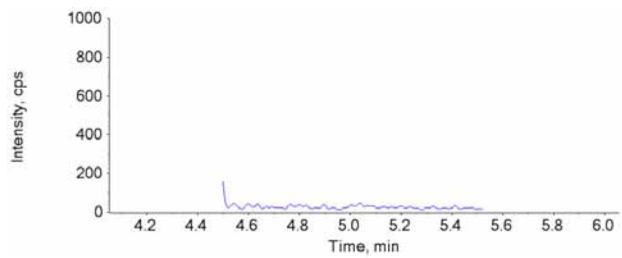
Sample Name: ARA-09-15-10-016-0002A1
Data File: ARA-09-15-10 Q05c.wiff
Sample Index: 50
Acquired: 6/2/2010 2:01:20 AM
Analyte Peak Area: 0
Analyte Ion Transition: 343.1/108.1 amu



Sample Name: ARA-09-15-10-016-0002A1
Data File: ARA-09-15-10 Q05c.wiff
Sample Index: 50
Acquired: 6/2/2010 2:01:20 AM
Analyte Peak Area: 0
Analyte Ion Transition: 373.2/299.0 amu



Sample Name: ARA-09-15-10-016-0002A1
Data File: ARA-09-15-10 Q05c.wiff
Sample Index: 50
Acquired: 6/2/2010 2:01:20 AM
Analyte Peak Area: 0
Analyte Ion Transition: 373.2/91.0 amu



Sample Name: ARA-09-15-10-016-0002A1
Data File: ARA-09-15-10 Q05c.wiff
Sample Index: 50
Acquired: 6/2/2010 2:01:20 AM
Analyte Peak Area: 0
Analyte Ion Transition: 343.1/270.9 amu

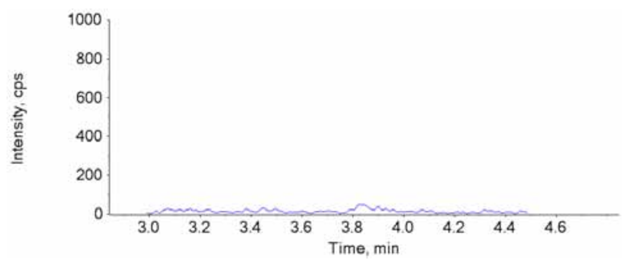
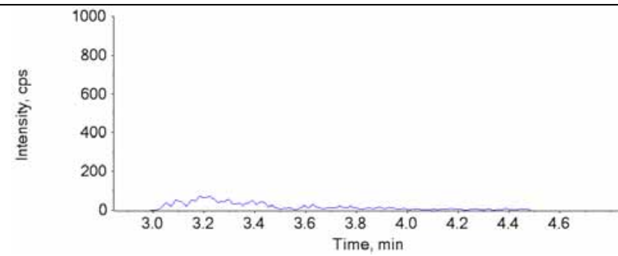
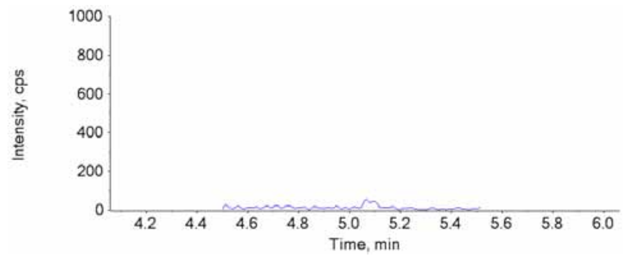


Figure 112. Typical Chromatogram of Quizalofop Analysis, Treated Corn Forage (016-0002)

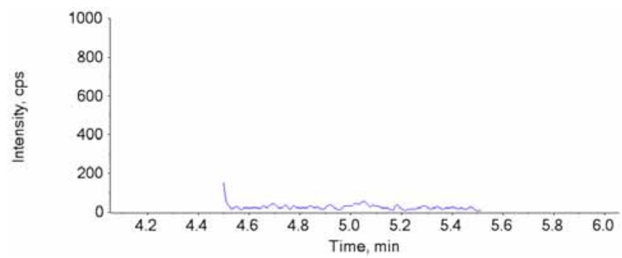
Sample Name: ARA-09-15-10-016-0003A1
Data File: ARA-09-15-10 Q05c.wiff
Sample Index: 51
Acquired: 6/2/2010 2:13:49 AM
Analyte Peak Area: 0
Analyte Ion Transition: 343.1/108.1 amu



Sample Name: ARA-09-15-10-016-0003A1
Data File: ARA-09-15-10 Q05c.wiff
Sample Index: 51
Acquired: 6/2/2010 2:13:49 AM
Analyte Peak Area: 0
Analyte Ion Transition: 373.2/299.0 amu



Sample Name: ARA-09-15-10-016-0003A1
Data File: ARA-09-15-10 Q05c.wiff
Sample Index: 51
Acquired: 6/2/2010 2:13:49 AM
Analyte Peak Area: 0
Analyte Ion Transition: 373.2/91.0 amu



Sample Name: ARA-09-15-10-016-0003A1
Data File: ARA-09-15-10 Q05c.wiff
Sample Index: 51
Acquired: 6/2/2010 2:13:49 AM
Analyte Peak Area: 0
Analyte Ion Transition: 343.1/270.9 amu

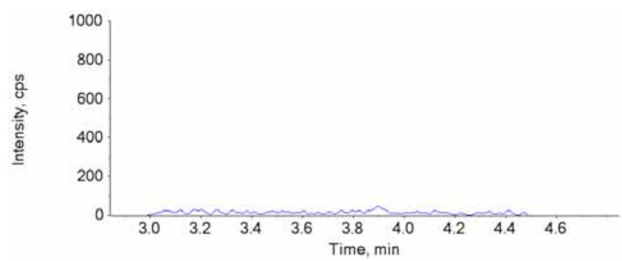
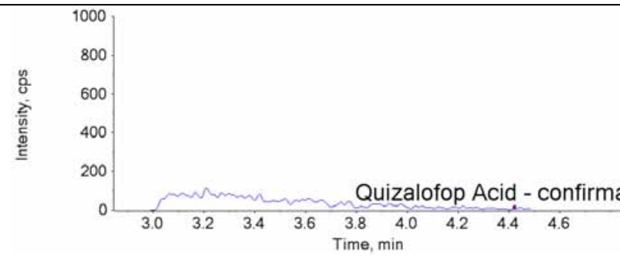
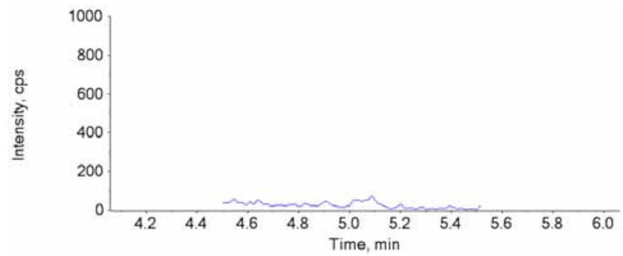


Figure 113. Typical Chromatogram of Quizalofop Analysis, Treated Corn Forage (016-0003)

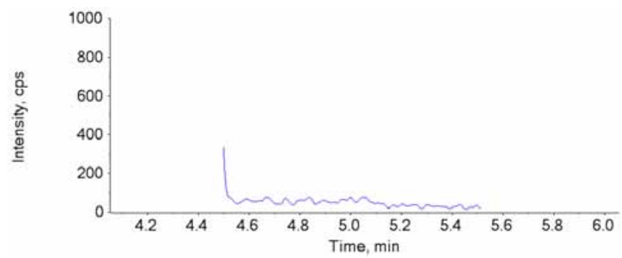
Sample Name: ARA-09-15-10-158-0001A1
Data File: ARA-09-15-10 Q09b.wiff
Sample Index: 2
Acquired: 6/4/2010 2:41:04 PM
Analyte Peak Area: 0
Analyte Ion Transition: 343.1/108.1 amu



Sample Name: ARA-09-15-10-158-0001A1
Data File: ARA-09-15-10 Q09b.wiff
Sample Index: 2
Acquired: 6/4/2010 2:41:04 PM
Analyte Peak Area: 0
Analyte Ion Transition: 373.2/299.0 amu



Sample Name: ARA-09-15-10-158-0001A1
Data File: ARA-09-15-10 Q09b.wiff
Sample Index: 2
Acquired: 6/4/2010 2:41:04 PM
Analyte Peak Area: 0
Analyte Ion Transition: 373.2/91.0 amu



Sample Name: ARA-09-15-10-158-0001A1
Data File: ARA-09-15-10 Q09b.wiff
Sample Index: 2
Acquired: 6/4/2010 2:41:04 PM
Analyte Peak Area: 0
Analyte Ion Transition: 343.1/270.9 amu

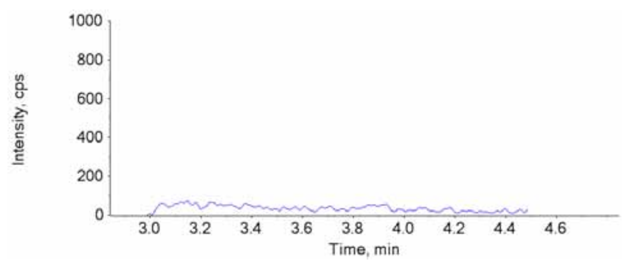
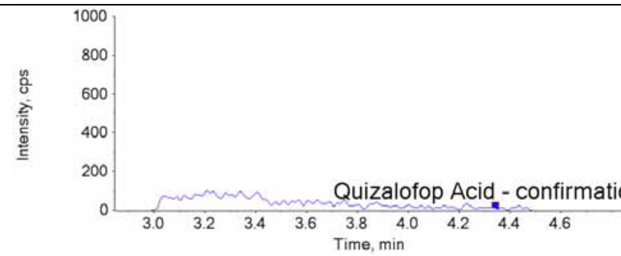
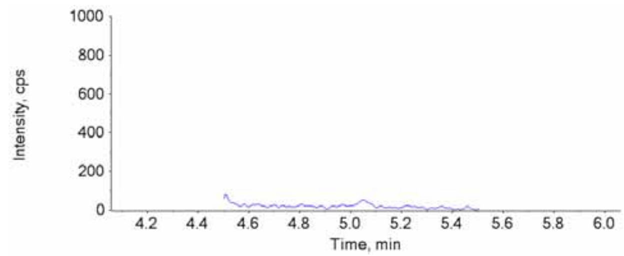


Figure 114. Typical Chromatogram of Quizalofop Analysis, Control Corn Stover (158-0001)

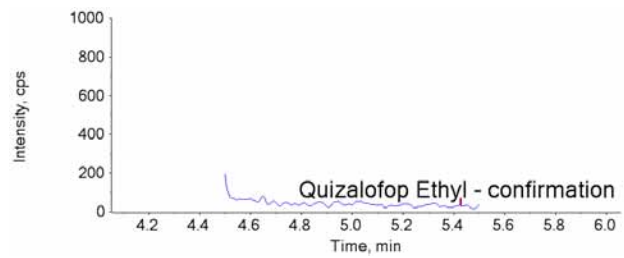
Sample Name: ARA-09-15-10-008-0001A1
Data File: ARA-09-15-10 Q09b.wiff
Sample Index: 10
Acquired: 6/4/2010 4:20:59 PM
Analyte Peak Area: 0
Analyte Ion Transition: 343.1/108.1 amu



Sample Name: ARA-09-15-10-008-0001A1
Data File: ARA-09-15-10 Q09b.wiff
Sample Index: 10
Acquired: 6/4/2010 4:20:59 PM
Analyte Peak Area: 0
Analyte Ion Transition: 373.2/299.0 amu



Sample Name: ARA-09-15-10-008-0001A1
Data File: ARA-09-15-10 Q09b.wiff
Sample Index: 10
Acquired: 6/4/2010 4:20:59 PM
Analyte Peak Area: 0
Analyte Ion Transition: 373.2/91.0 amu



Sample Name: ARA-09-15-10-008-0001A1
Data File: ARA-09-15-10 Q09b.wiff
Sample Index: 10
Acquired: 6/4/2010 4:20:59 PM
Analyte Peak Area: 0
Analyte Ion Transition: 343.1/270.9 amu

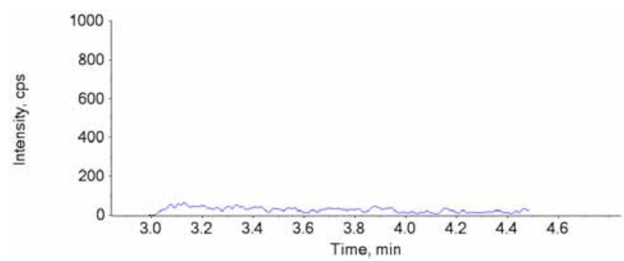
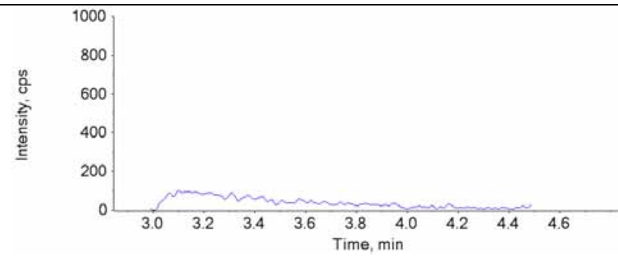
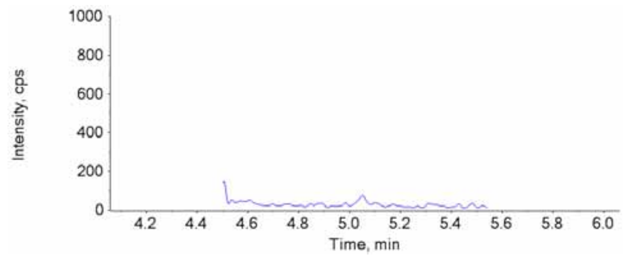


Figure 115. Typical Chromatogram of Quizalofop Analysis, Control Corn Stover (008-0001)

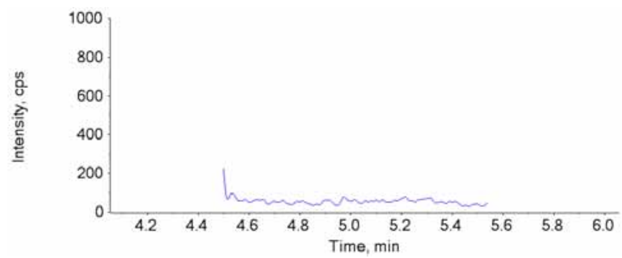
Sample Name: ARA-09-15-10-013-0001A1
Data File: ARA-09-15-10 Q09b.wiff
Sample Index: 11
Acquired: 6/4/2010 4:33:27 PM
Analyte Peak Area: 0
Analyte Ion Transition: 343.1/108.1 amu



Sample Name: ARA-09-15-10-013-0001A1
Data File: ARA-09-15-10 Q09b.wiff
Sample Index: 11
Acquired: 6/4/2010 4:33:27 PM
Analyte Peak Area: 0
Analyte Ion Transition: 373.2/299.0 amu



Sample Name: ARA-09-15-10-013-0001A1
Data File: ARA-09-15-10 Q09b.wiff
Sample Index: 11
Acquired: 6/4/2010 4:33:27 PM
Analyte Peak Area: 0
Analyte Ion Transition: 373.2/91.0 amu



Sample Name: ARA-09-15-10-013-0001A1
Data File: ARA-09-15-10 Q09b.wiff
Sample Index: 11
Acquired: 6/4/2010 4:33:27 PM
Analyte Peak Area: 0
Analyte Ion Transition: 343.1/270.9 amu

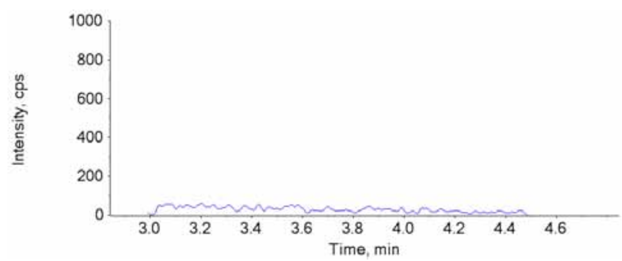
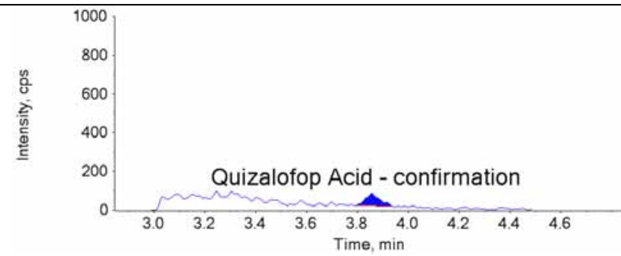
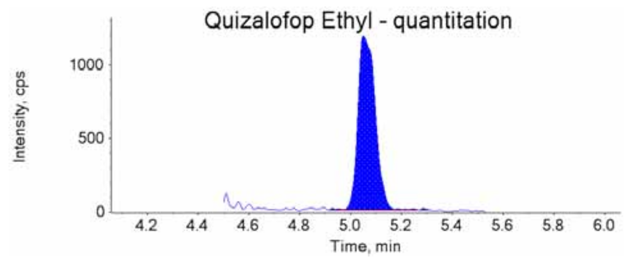


Figure 116. Typical Chromatogram of Quizalofop Analysis, Control Corn Stover (013-0001)

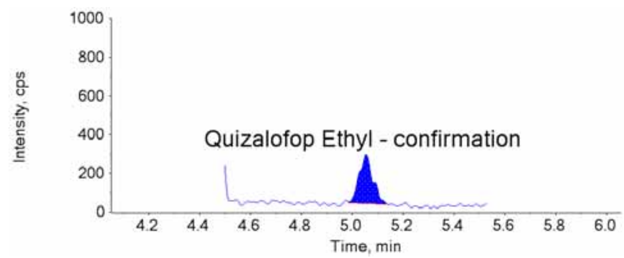
Sample Name: ARA-09-15-10-158-0001A3 + 0.01 µg/g
Data File: ARA-09-15-10 Q09b.wiff
Sample Index: 4
Acquired: 6/4/2010 3:06:04 PM
Analyte Peak Area: 245
Analyte Ion Transition: 343.1/108.1 amu



Sample Name: ARA-09-15-10-158-0001A3 + 0.01 µg/g
Data File: ARA-09-15-10 Q09b.wiff
Sample Index: 4
Acquired: 6/4/2010 3:06:04 PM
Analyte Peak Area: 5667
Analyte Ion Transition: 373.2/299.0 amu



Sample Name: ARA-09-15-10-158-0001A3 + 0.01 µg/g
Data File: ARA-09-15-10 Q09b.wiff
Sample Index: 4
Acquired: 6/4/2010 3:06:04 PM
Analyte Peak Area: 944
Analyte Ion Transition: 373.2/91.0 amu



Sample Name: ARA-09-15-10-158-0001A3 + 0.01 µg/g
Data File: ARA-09-15-10 Q09b.wiff
Sample Index: 4
Acquired: 6/4/2010 3:06:04 PM
Analyte Peak Area: 3488
Analyte Ion Transition: 343.1/270.9 amu

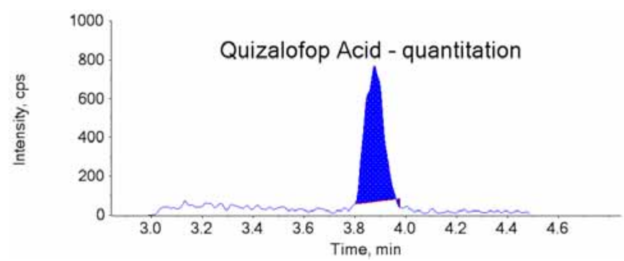
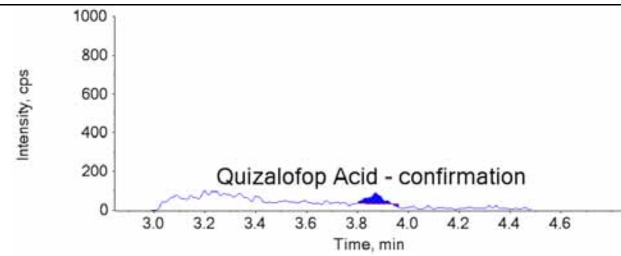
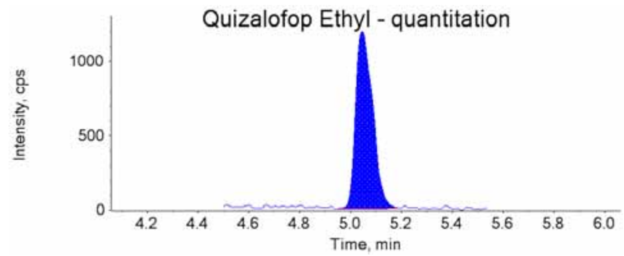


Figure 117. Typical Chromatogram of Quizalofop Analysis, Corn Stover 0.01 µg/g Recovery

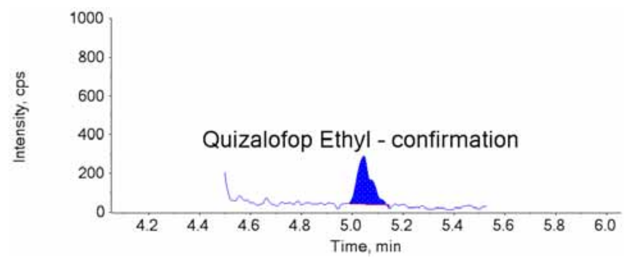
Sample Name: ARA-09-15-10-158-0001A4 + 0.01 µg/g
Data File: ARA-09-15-10 Q09b.wiff
Sample Index: 5
Acquired: 6/4/2010 3:18:33 PM
Analyte Peak Area: 233
Analyte Ion Transition: 343.1/108.1 amu



Sample Name: ARA-09-15-10-158-0001A4 + 0.01 µg/g
Data File: ARA-09-15-10 Q09b.wiff
Sample Index: 5
Acquired: 6/4/2010 3:18:33 PM
Analyte Peak Area: 5591
Analyte Ion Transition: 373.2/299.0 amu



Sample Name: ARA-09-15-10-158-0001A4 + 0.01 µg/g
Data File: ARA-09-15-10 Q09b.wiff
Sample Index: 5
Acquired: 6/4/2010 3:18:33 PM
Analyte Peak Area: 956
Analyte Ion Transition: 373.2/91.0 amu



Sample Name: ARA-09-15-10-158-0001A4 + 0.01 µg/g
Data File: ARA-09-15-10 Q09b.wiff
Sample Index: 5
Acquired: 6/4/2010 3:18:33 PM
Analyte Peak Area: 3452
Analyte Ion Transition: 343.1/270.9 amu

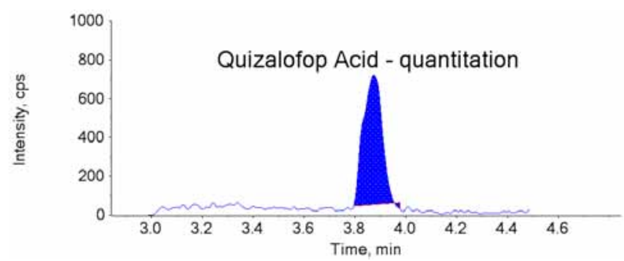
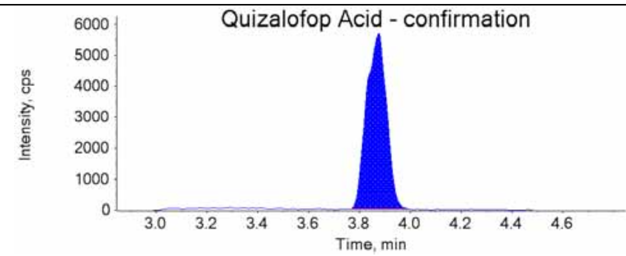
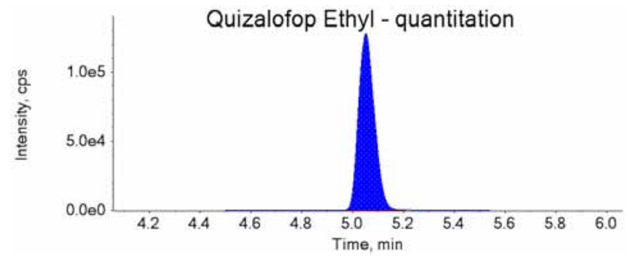


Figure 118. Typical Chromatogram of Quizalofop Analysis, Corn Stover 0.01 µg/g Recovery

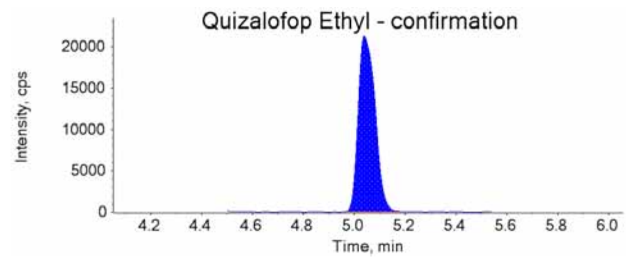
Sample Name: ARA-09-15-10-158-0001A5 + 1.0 µg/g
Data File: ARA-09-15-10 Q09b.wiff
Sample Index: 6
Acquired: 6/4/2010 3:30:59 PM
Analyte Peak Area: 31020
Analyte Ion Transition: 343.1/108.1 amu



Sample Name: ARA-09-15-10-158-0001A5 + 1.0 µg/g
Data File: ARA-09-15-10 Q09b.wiff
Sample Index: 6
Acquired: 6/4/2010 3:30:59 PM
Analyte Peak Area: 554123
Analyte Ion Transition: 373.2/299.0 amu



Sample Name: ARA-09-15-10-158-0001A5 + 1.0 µg/g
Data File: ARA-09-15-10 Q09b.wiff
Sample Index: 6
Acquired: 6/4/2010 3:30:59 PM
Analyte Peak Area: 97954
Analyte Ion Transition: 373.2/91.0 amu



Sample Name: ARA-09-15-10-158-0001A5 + 1.0 µg/g
Data File: ARA-09-15-10 Q09b.wiff
Sample Index: 6
Acquired: 6/4/2010 3:30:59 PM
Analyte Peak Area: 387200
Analyte Ion Transition: 343.1/270.9 amu

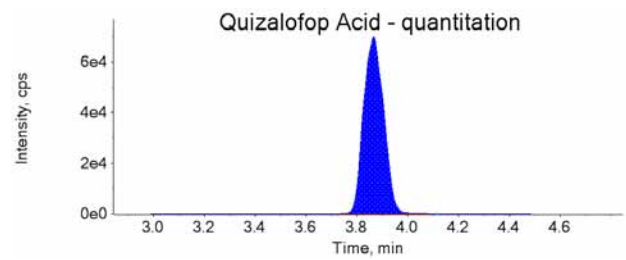
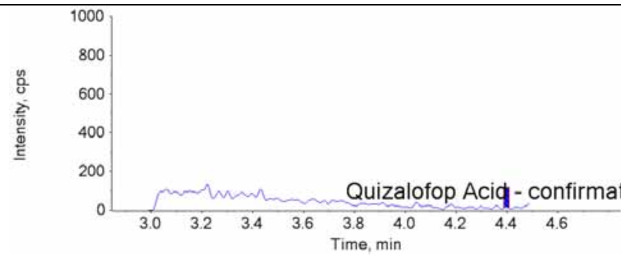
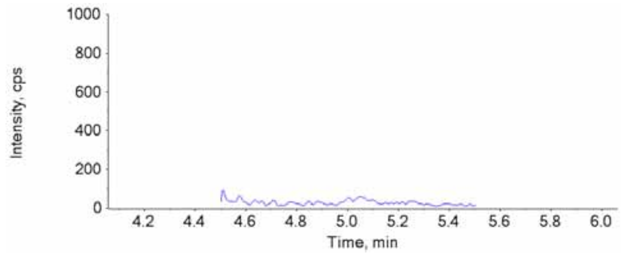


Figure 119. Typical Chromatogram of Quizalofop Analysis, Corn Stover 1.00 µg/g Recovery

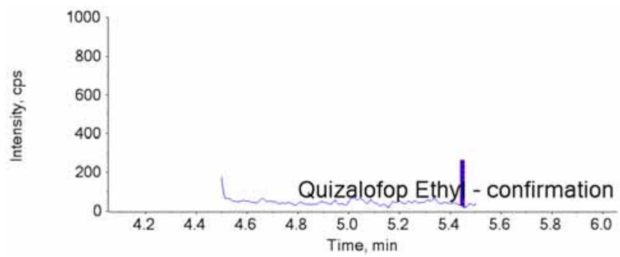
Sample Name: ARA-09-15-10-013-0002A1
Data File: ARA-09-15-10 Q10b.wiff
Sample Index: 9
Acquired: 6/5/2010 3:49:44 AM
Analyte Peak Area: 0
Analyte Ion Transition: 343.1/108.1 amu



Sample Name: ARA-09-15-10-013-0002A1
Data File: ARA-09-15-10 Q10b.wiff
Sample Index: 9
Acquired: 6/5/2010 3:49:44 AM
Analyte Peak Area: 0
Analyte Ion Transition: 373.2/299.0 amu



Sample Name: ARA-09-15-10-013-0002A1
Data File: ARA-09-15-10 Q10b.wiff
Sample Index: 9
Acquired: 6/5/2010 3:49:44 AM
Analyte Peak Area: 0
Analyte Ion Transition: 373.2/91.0 amu



Sample Name: ARA-09-15-10-013-0002A1
Data File: ARA-09-15-10 Q10b.wiff
Sample Index: 9
Acquired: 6/5/2010 3:49:44 AM
Analyte Peak Area: 547
Analyte Ion Transition: 343.1/270.9 amu

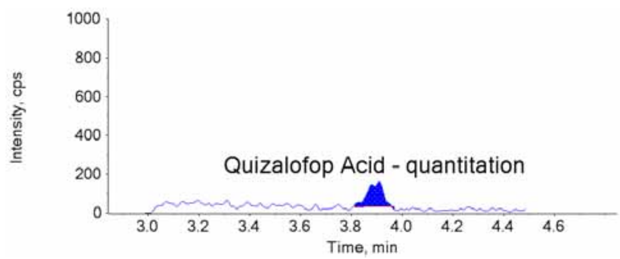
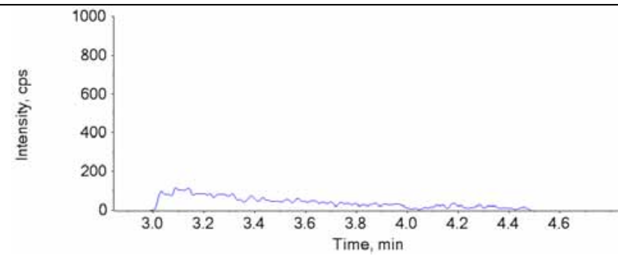
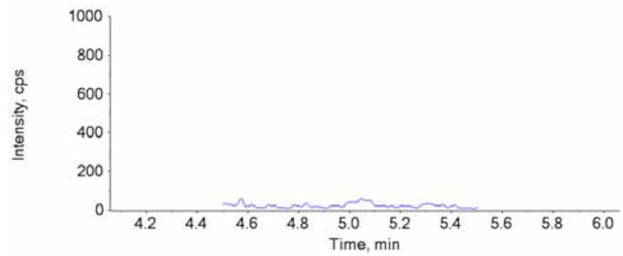


Figure 120. Typical Chromatogram of Quizalofop Analysis, Treated Corn Stover (013-0002)

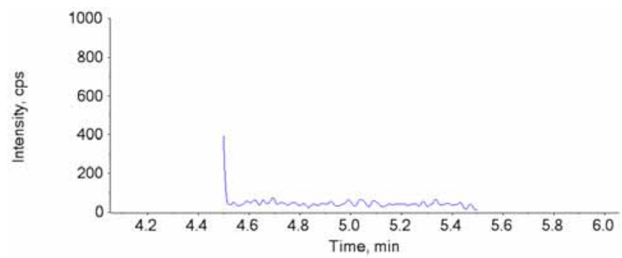
Sample Name: ARA-09-15-10-013-0003A1
Data File: ARA-09-15-10 Q10b.wiff
Sample Index: 10
Acquired: 6/5/2010 4:02:14 AM
Analyte Peak Area: 0
Analyte Ion Transition: 343.1/108.1 amu



Sample Name: ARA-09-15-10-013-0003A1
Data File: ARA-09-15-10 Q10b.wiff
Sample Index: 10
Acquired: 6/5/2010 4:02:14 AM
Analyte Peak Area: 0
Analyte Ion Transition: 373.2/299.0 amu



Sample Name: ARA-09-15-10-013-0003A1
Data File: ARA-09-15-10 Q10b.wiff
Sample Index: 10
Acquired: 6/5/2010 4:02:14 AM
Analyte Peak Area: 0
Analyte Ion Transition: 373.2/91.0 amu



Sample Name: ARA-09-15-10-013-0003A1
Data File: ARA-09-15-10 Q10b.wiff
Sample Index: 10
Acquired: 6/5/2010 4:02:14 AM
Analyte Peak Area: 446
Analyte Ion Transition: 343.1/270.9 amu

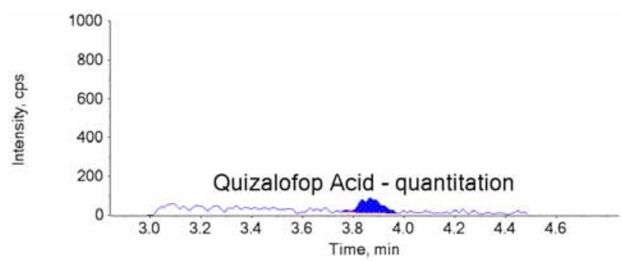
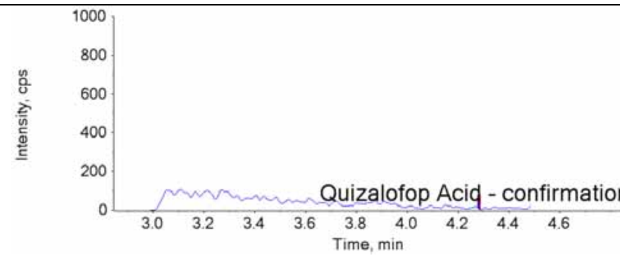
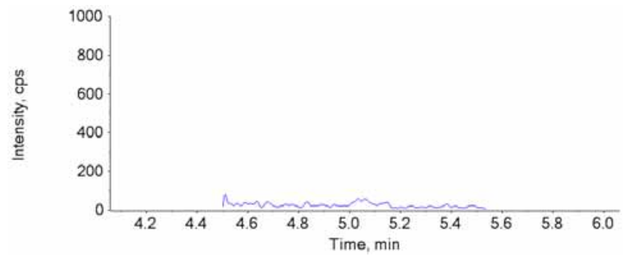


Figure 121. Typical Chromatogram of Quizalofop Analysis, Treated Corn Stover (013-0003)

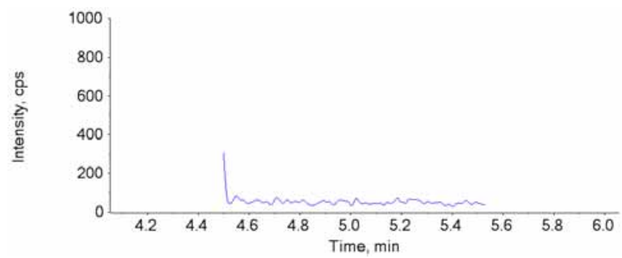
Sample Name: ARA-09-15-10-023-0002A1
Data File: ARA-09-15-10 Q10b.wiff
Sample Index: 13
Acquired: 6/5/2010 4:39:38 AM
Analyte Peak Area: 0
Analyte Ion Transition: 343.1/108.1 amu



Sample Name: ARA-09-15-10-023-0002A1
Data File: ARA-09-15-10 Q10b.wiff
Sample Index: 13
Acquired: 6/5/2010 4:39:38 AM
Analyte Peak Area: 0
Analyte Ion Transition: 373.2/299.0 amu



Sample Name: ARA-09-15-10-023-0002A1
Data File: ARA-09-15-10 Q10b.wiff
Sample Index: 13
Acquired: 6/5/2010 4:39:38 AM
Analyte Peak Area: 0
Analyte Ion Transition: 373.2/91.0 amu



Sample Name: ARA-09-15-10-023-0002A1
Data File: ARA-09-15-10 Q10b.wiff
Sample Index: 13
Acquired: 6/5/2010 4:39:38 AM
Analyte Peak Area: 741
Analyte Ion Transition: 343.1/270.9 amu

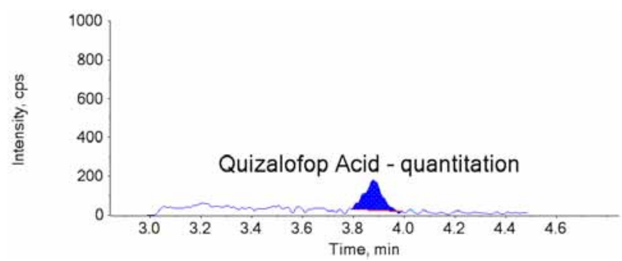
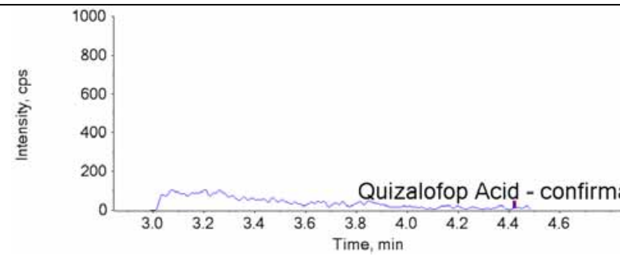
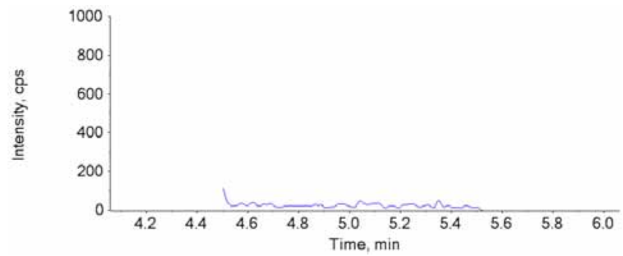


Figure 122. Typical Chromatogram of Quizalofop Analysis, Treated Corn Stover (023-0002)

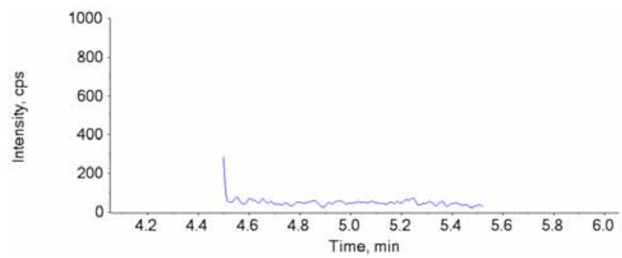
Sample Name: ARA-09-15-10-023-0003A1
Data File: ARA-09-15-10 Q10b.wiff
Sample Index: 14
Acquired: 6/5/2010 4:52:06 AM
Analyte Peak Area: 0
Analyte Ion Transition: 343.1/108.1 amu



Sample Name: ARA-09-15-10-023-0003A1
Data File: ARA-09-15-10 Q10b.wiff
Sample Index: 14
Acquired: 6/5/2010 4:52:06 AM
Analyte Peak Area: 0
Analyte Ion Transition: 373.2/299.0 amu



Sample Name: ARA-09-15-10-023-0003A1
Data File: ARA-09-15-10 Q10b.wiff
Sample Index: 14
Acquired: 6/5/2010 4:52:06 AM
Analyte Peak Area: 0
Analyte Ion Transition: 373.2/91.0 amu



Sample Name: ARA-09-15-10-023-0003A1
Data File: ARA-09-15-10 Q10b.wiff
Sample Index: 14
Acquired: 6/5/2010 4:52:06 AM
Analyte Peak Area: 927
Analyte Ion Transition: 343.1/270.9 amu

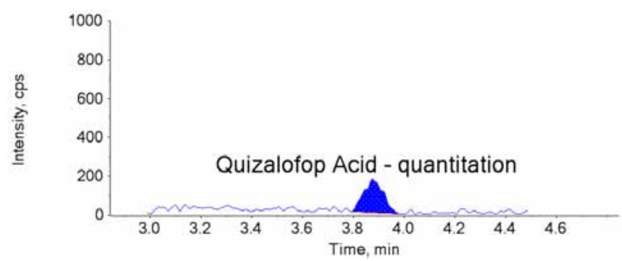
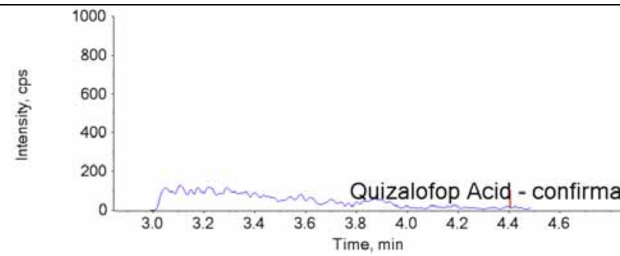
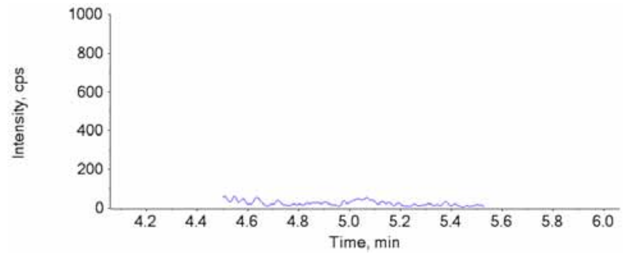


Figure 123. Typical Chromatogram of Quizalofop Analysis, Treated Corn Stover (023-0003)

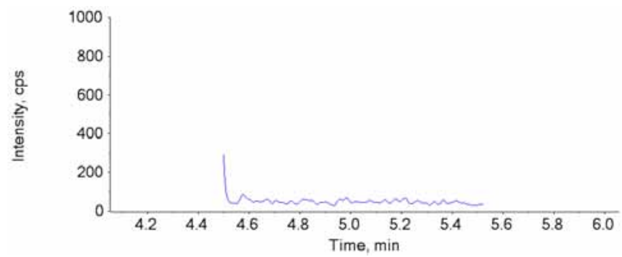
Sample Name: ARA-09-15-10-025-0002A1
Data File: ARA-09-15-10 Q10b.wiff
Sample Index: 15
Acquired: 6/5/2010 5:04:33 AM
Analyte Peak Area: 0
Analyte Ion Transition: 343.1/108.1 amu



Sample Name: ARA-09-15-10-025-0002A1
Data File: ARA-09-15-10 Q10b.wiff
Sample Index: 15
Acquired: 6/5/2010 5:04:33 AM
Analyte Peak Area: 0
Analyte Ion Transition: 373.2/299.0 amu



Sample Name: ARA-09-15-10-025-0002A1
Data File: ARA-09-15-10 Q10b.wiff
Sample Index: 15
Acquired: 6/5/2010 5:04:33 AM
Analyte Peak Area: 0
Analyte Ion Transition: 373.2/91.0 amu



Sample Name: ARA-09-15-10-025-0002A1
Data File: ARA-09-15-10 Q10b.wiff
Sample Index: 15
Acquired: 6/5/2010 5:04:33 AM
Analyte Peak Area: 1152
Analyte Ion Transition: 343.1/270.9 amu

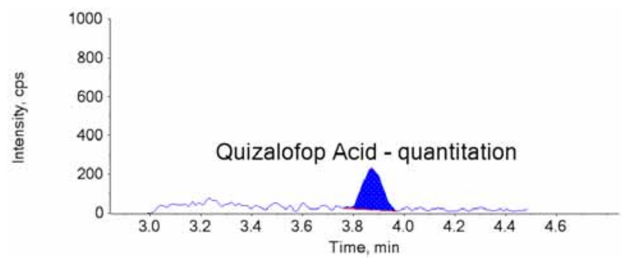
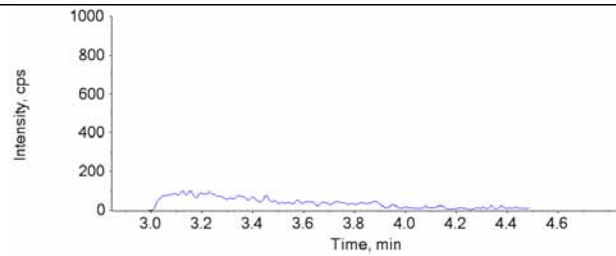
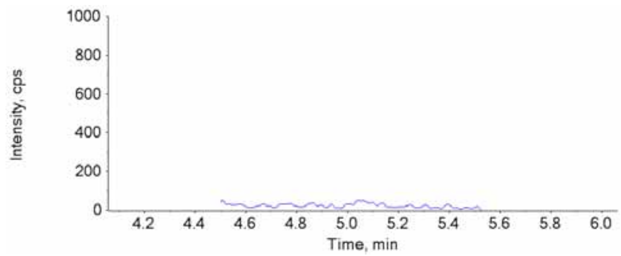


Figure 124. Typical Chromatogram of Quizalofop Analysis, Treated Corn Stover (025-0002)

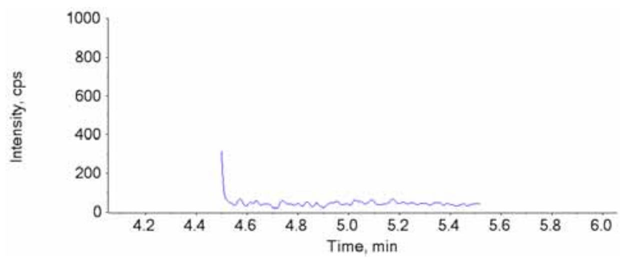
Sample Name: ARA-09-15-10-025-0003A1
Data File: ARA-09-15-10 Q10b.wiff
Sample Index: 17
Acquired: 6/5/2010 5:29:31 AM
Analyte Peak Area: 0
Analyte Ion Transition: 343.1/108.1 amu



Sample Name: ARA-09-15-10-025-0003A1
Data File: ARA-09-15-10 Q10b.wiff
Sample Index: 17
Acquired: 6/5/2010 5:29:31 AM
Analyte Peak Area: 0
Analyte Ion Transition: 373.2/299.0 amu



Sample Name: ARA-09-15-10-025-0003A1
Data File: ARA-09-15-10 Q10b.wiff
Sample Index: 17
Acquired: 6/5/2010 5:29:31 AM
Analyte Peak Area: 0
Analyte Ion Transition: 373.2/91.0 amu



Sample Name: ARA-09-15-10-025-0003A1
Data File: ARA-09-15-10 Q10b.wiff
Sample Index: 17
Acquired: 6/5/2010 5:29:31 AM
Analyte Peak Area: 582
Analyte Ion Transition: 343.1/270.9 amu

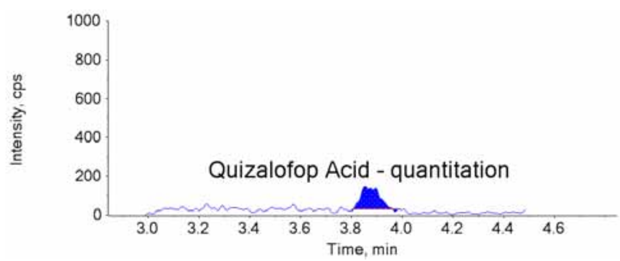
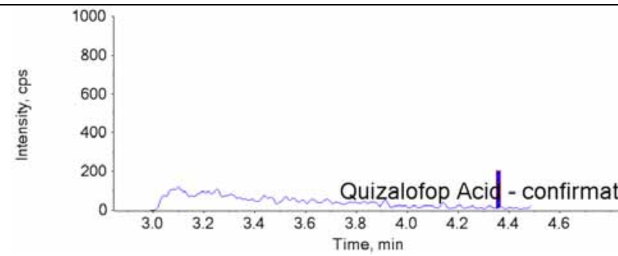
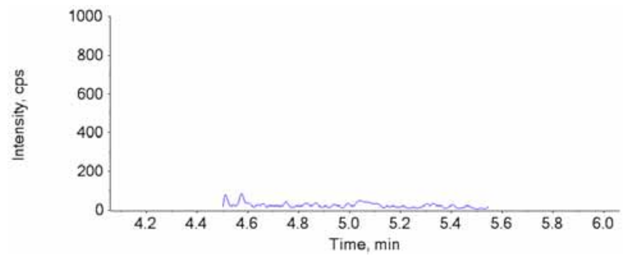


Figure 125. Typical Chromatogram of Quizalofop Analysis, Treated Corn Stover (025-0003)

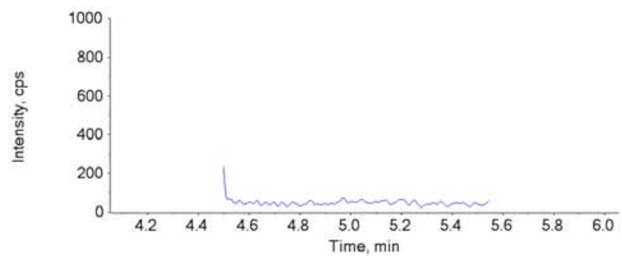
Sample Name: ARA-09-15-10-027-0002A1
Data File: ARA-09-15-10 Q10b.wiff
Sample Index: 18
Acquired: 6/5/2010 5:41:58 AM
Analyte Peak Area: 0
Analyte Ion Transition: 343.1/108.1 amu



Sample Name: ARA-09-15-10-027-0002A1
Data File: ARA-09-15-10 Q10b.wiff
Sample Index: 18
Acquired: 6/5/2010 5:41:58 AM
Analyte Peak Area: 0
Analyte Ion Transition: 373.2/299.0 amu



Sample Name: ARA-09-15-10-027-0002A1
Data File: ARA-09-15-10 Q10b.wiff
Sample Index: 18
Acquired: 6/5/2010 5:41:58 AM
Analyte Peak Area: 0
Analyte Ion Transition: 373.2/91.0 amu



Sample Name: ARA-09-15-10-027-0002A1
Data File: ARA-09-15-10 Q10b.wiff
Sample Index: 18
Acquired: 6/5/2010 5:41:58 AM
Analyte Peak Area: 825
Analyte Ion Transition: 343.1/270.9 amu

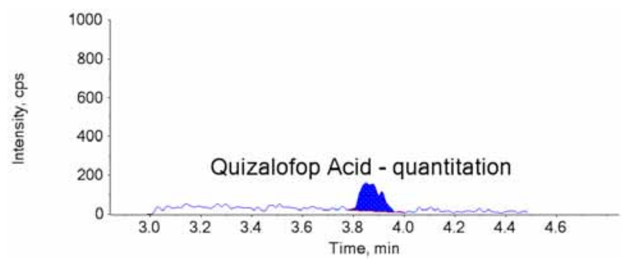
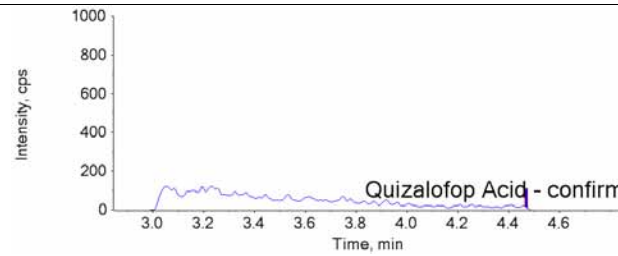
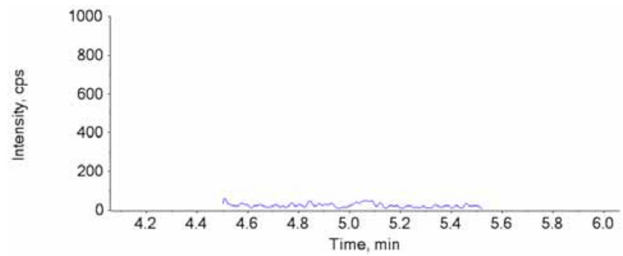


Figure 126. Typical Chromatogram of Quizalofop Analysis, Treated Corn Stover (027-0002)

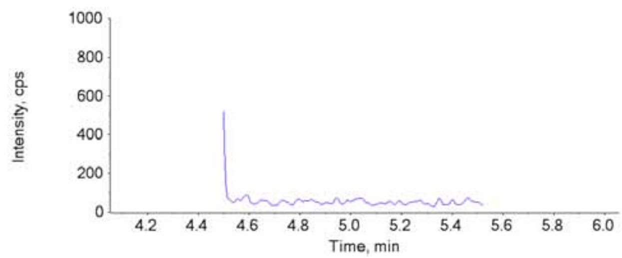
Sample Name: ARA-09-15-10-027-0003A1
Data File: ARA-09-15-10 Q10b.wiff
Sample Index: 19
Acquired: 6/5/2010 5:54:26 AM
Analyte Peak Area: 0
Analyte Ion Transition: 343.1/108.1 amu



Sample Name: ARA-09-15-10-027-0003A1
Data File: ARA-09-15-10 Q10b.wiff
Sample Index: 19
Acquired: 6/5/2010 5:54:26 AM
Analyte Peak Area: 0
Analyte Ion Transition: 373.2/299.0 amu



Sample Name: ARA-09-15-10-027-0003A1
Data File: ARA-09-15-10 Q10b.wiff
Sample Index: 19
Acquired: 6/5/2010 5:54:26 AM
Analyte Peak Area: 0
Analyte Ion Transition: 373.2/91.0 amu



Sample Name: ARA-09-15-10-027-0003A1
Data File: ARA-09-15-10 Q10b.wiff
Sample Index: 19
Acquired: 6/5/2010 5:54:26 AM
Analyte Peak Area: 673
Analyte Ion Transition: 343.1/270.9 amu

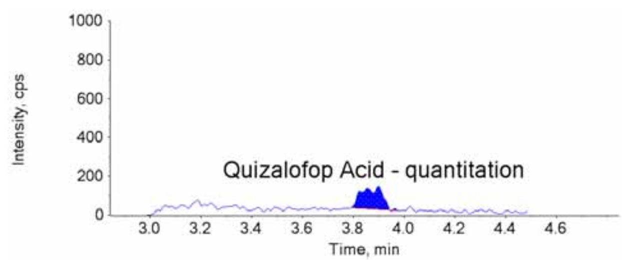
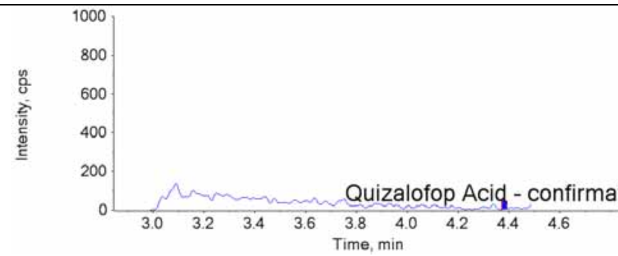
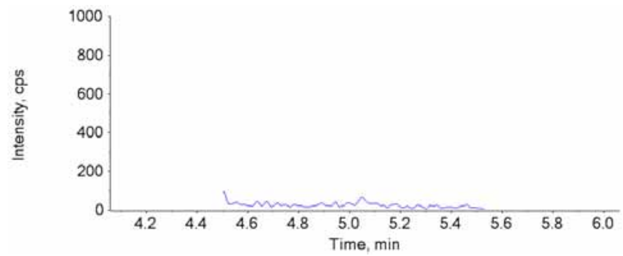


Figure 127. Typical Chromatogram of Quizalofop Analysis, Treated Corn Stover (027-0003)

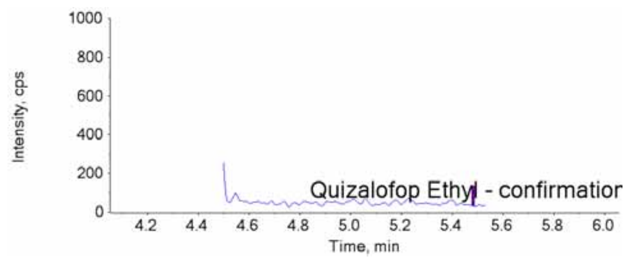
Sample Name: ARA-09-15-10-029-0002A1
Data File: ARA-09-15-10 Q10b.wiff
Sample Index: 20
Acquired: 6/5/2010 6:06:56 AM
Analyte Peak Area: 0
Analyte Ion Transition: 343.1/108.1 amu



Sample Name: ARA-09-15-10-029-0002A1
Data File: ARA-09-15-10 Q10b.wiff
Sample Index: 20
Acquired: 6/5/2010 6:06:56 AM
Analyte Peak Area: 0
Analyte Ion Transition: 373.2/299.0 amu



Sample Name: ARA-09-15-10-029-0002A1
Data File: ARA-09-15-10 Q10b.wiff
Sample Index: 20
Acquired: 6/5/2010 6:06:56 AM
Analyte Peak Area: 0
Analyte Ion Transition: 373.2/91.0 amu



Sample Name: ARA-09-15-10-029-0002A1
Data File: ARA-09-15-10 Q10b.wiff
Sample Index: 20
Acquired: 6/5/2010 6:06:56 AM
Analyte Peak Area: 539
Analyte Ion Transition: 343.1/270.9 amu

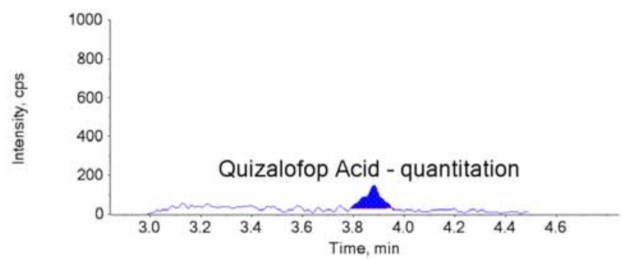
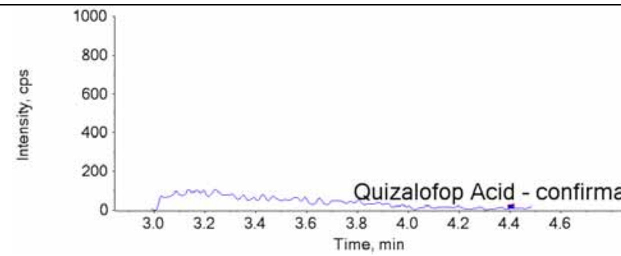
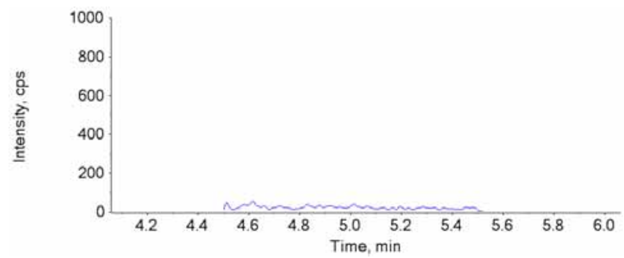


Figure 128. Typical Chromatogram of Quizalofop Analysis, Treated Corn Stover (029-0002)

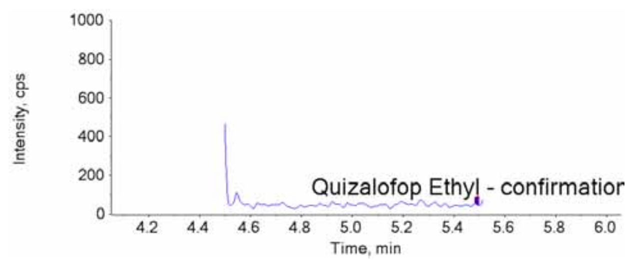
Sample Name: ARA-09-15-10-029-0003A1
Data File: ARA-09-15-10 Q10b.wiff
Sample Index: 21
Acquired: 6/5/2010 6:19:25 AM
Analyte Peak Area: 0
Analyte Ion Transition: 343.1/108.1 amu



Sample Name: ARA-09-15-10-029-0003A1
Data File: ARA-09-15-10 Q10b.wiff
Sample Index: 21
Acquired: 6/5/2010 6:19:25 AM
Analyte Peak Area: 0
Analyte Ion Transition: 373.2/299.0 amu



Sample Name: ARA-09-15-10-029-0003A1
Data File: ARA-09-15-10 Q10b.wiff
Sample Index: 21
Acquired: 6/5/2010 6:19:25 AM
Analyte Peak Area: 0
Analyte Ion Transition: 373.2/91.0 amu



Sample Name: ARA-09-15-10-029-0003A1
Data File: ARA-09-15-10 Q10b.wiff
Sample Index: 21
Acquired: 6/5/2010 6:19:25 AM
Analyte Peak Area: 0
Analyte Ion Transition: 343.1/270.9 amu

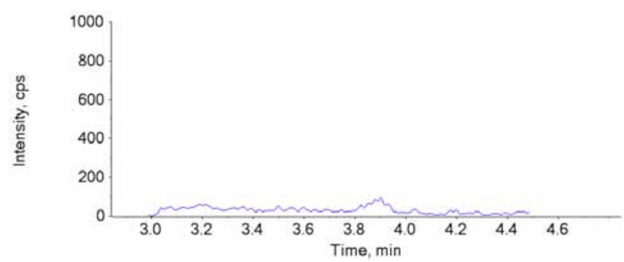
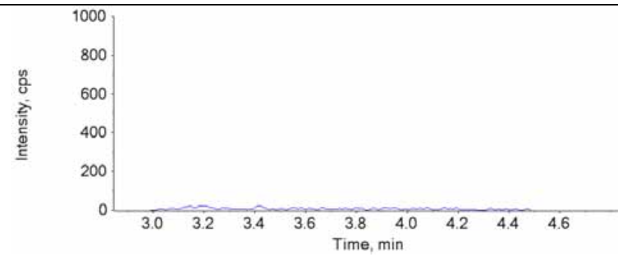
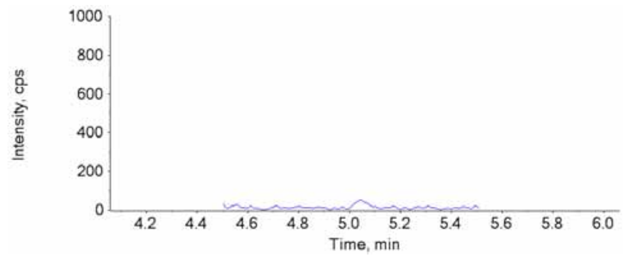


Figure 129. Typical Chromatogram of Quizalofop Analysis, Treated Corn Stover (029-0003)

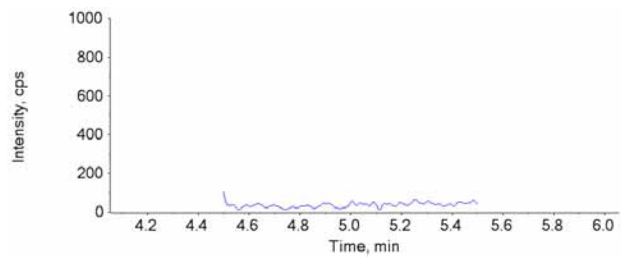
Sample Name: ARA-09-15-10-150-0001A1
Data File: ARA-09-15-10 Q12.wiff
Sample Index: 2
Acquired: 6/8/2010 6:10:39 PM
Analyte Peak Area: 0
Analyte Ion Transition: 343.1/108.1 amu



Sample Name: ARA-09-15-10-150-0001A1
Data File: ARA-09-15-10 Q12.wiff
Sample Index: 2
Acquired: 6/8/2010 6:10:39 PM
Analyte Peak Area: 0
Analyte Ion Transition: 373.2/299.0 amu



Sample Name: ARA-09-15-10-150-0001A1
Data File: ARA-09-15-10 Q12.wiff
Sample Index: 2
Acquired: 6/8/2010 6:10:39 PM
Analyte Peak Area: 0
Analyte Ion Transition: 373.2/91.0 amu



Sample Name: ARA-09-15-10-150-0001A1
Data File: ARA-09-15-10 Q12.wiff
Sample Index: 2
Acquired: 6/8/2010 6:10:39 PM
Analyte Peak Area: 0
Analyte Ion Transition: 343.1/270.9 amu

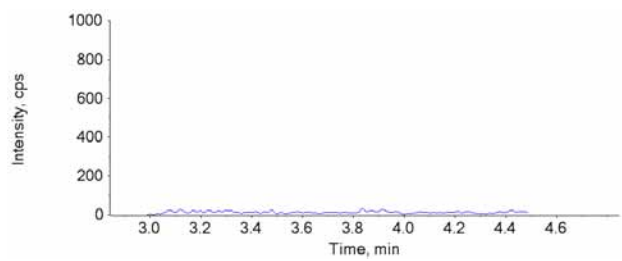
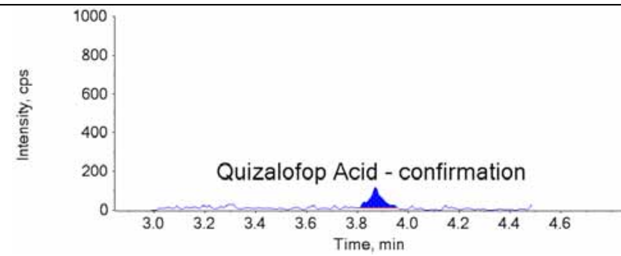
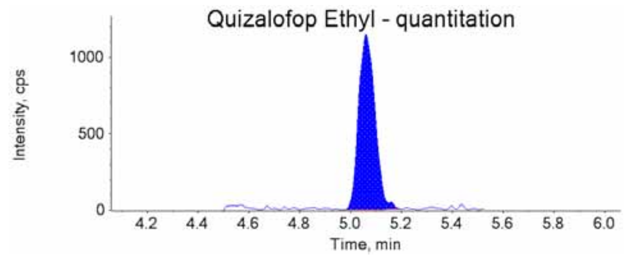


Figure 130. Typical Chromatogram of Quizalofop Analysis, Control Corn Aspirated Grain Fractions (150-0001)

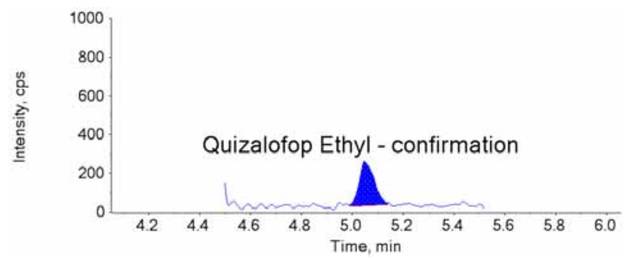
Sample Name: ARA-09-15-10-150-0001A3 + 0.01 µg/g
Data File: ARA-09-15-10 Q12.wiff
Sample Index: 11
Acquired: 6/8/2010 8:02:55 PM
Analyte Peak Area: 338
Analyte Ion Transition: 343.1/108.1 amu



Sample Name: ARA-09-15-10-150-0001A3 + 0.01 µg/g
Data File: ARA-09-15-10 Q12.wiff
Sample Index: 11
Acquired: 6/8/2010 8:02:55 PM
Analyte Peak Area: 5130
Analyte Ion Transition: 373.2/299.0 amu



Sample Name: ARA-09-15-10-150-0001A3 + 0.01 µg/g
Data File: ARA-09-15-10 Q12.wiff
Sample Index: 11
Acquired: 6/8/2010 8:02:55 PM
Analyte Peak Area: 940
Analyte Ion Transition: 373.2/91.0 amu



Sample Name: ARA-09-15-10-150-0001A3 + 0.01 µg/g
Data File: ARA-09-15-10 Q12.wiff
Sample Index: 11
Acquired: 6/8/2010 8:02:55 PM
Analyte Peak Area: 4875
Analyte Ion Transition: 343.1/270.9 amu

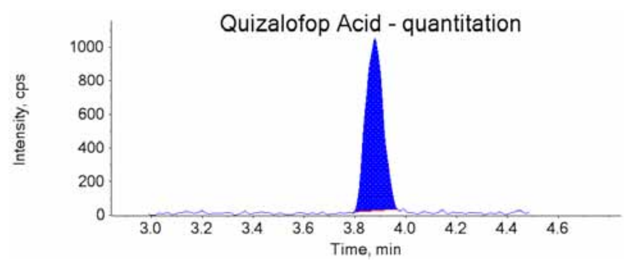
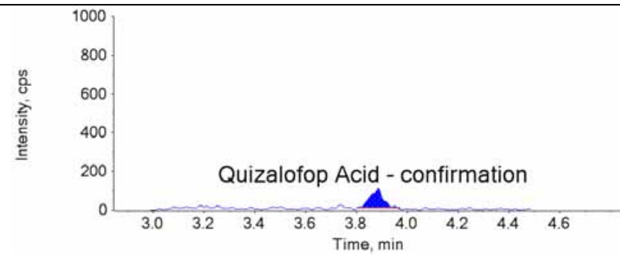
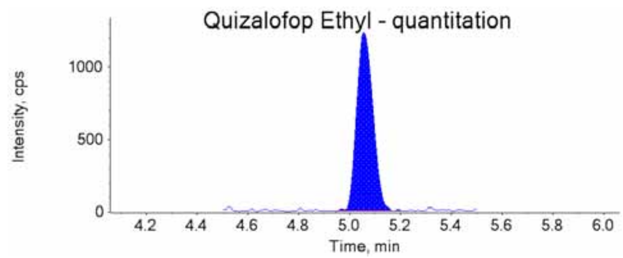


Figure 131. Typical Chromatogram of Quizalofop Analysis, Corn Aspirated Grain Fractions 0.01 µg/g Recovery

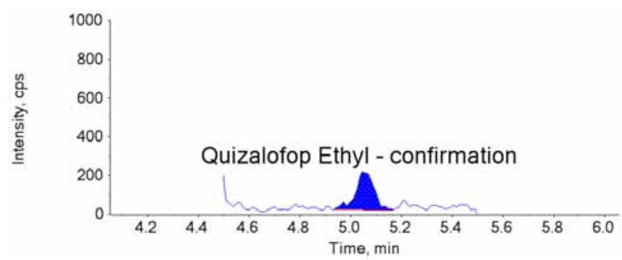
Sample Name: ARA-09-15-10-150-0001A4 + 0.01 µg/g
Data File: ARA-09-15-10 Q12.wiff
Sample Index: 12
Acquired: 6/8/2010 8:15:23 PM
Analyte Peak Area: 340
Analyte Ion Transition: 343.1/108.1 amu



Sample Name: ARA-09-15-10-150-0001A4 + 0.01 µg/g
Data File: ARA-09-15-10 Q12.wiff
Sample Index: 12
Acquired: 6/8/2010 8:15:23 PM
Analyte Peak Area: 5517
Analyte Ion Transition: 373.2/299.0 amu



Sample Name: ARA-09-15-10-150-0001A4 + 0.01 µg/g
Data File: ARA-09-15-10 Q12.wiff
Sample Index: 12
Acquired: 6/8/2010 8:15:23 PM
Analyte Peak Area: 1005
Analyte Ion Transition: 373.2/91.0 amu



Sample Name: ARA-09-15-10-150-0001A4 + 0.01 µg/g
Data File: ARA-09-15-10 Q12.wiff
Sample Index: 12
Acquired: 6/8/2010 8:15:23 PM
Analyte Peak Area: 5075
Analyte Ion Transition: 343.1/270.9 amu

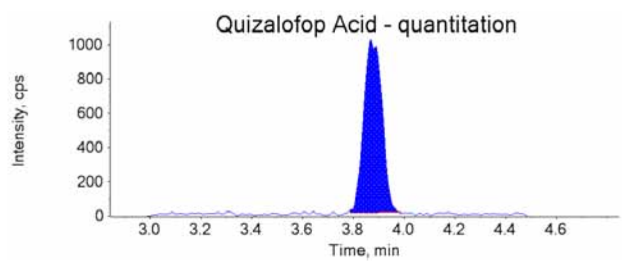
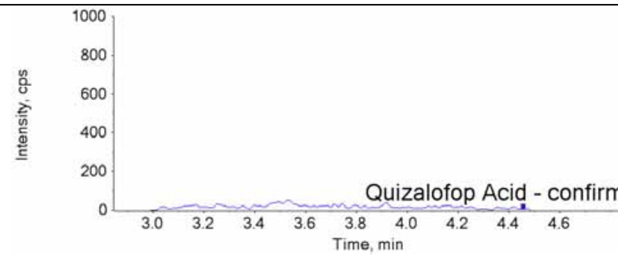
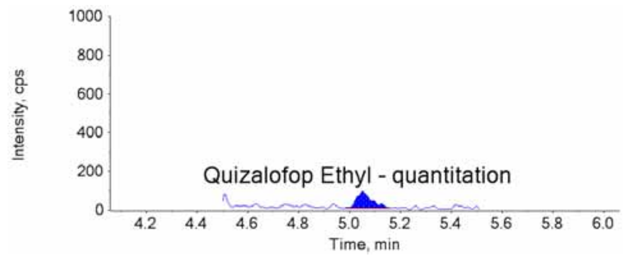


Figure 132. Typical Chromatogram of Quizalofop Analysis, Corn Aspirated Grain Fractions 0.01 µg/g Recovery

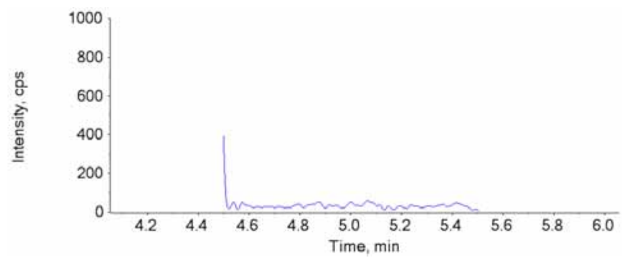
Sample Name: ARA-09-15-10-141-0002A1
Data File: ARA-09-15-10 Q12.wiff
Sample Index: 50
Acquired: 6/9/2010 4:09:22 AM
Analyte Peak Area: 0
Analyte Ion Transition: 343.1/108.1 amu



Sample Name: ARA-09-15-10-141-0002A1
Data File: ARA-09-15-10 Q12.wiff
Sample Index: 50
Acquired: 6/9/2010 4:09:22 AM
Analyte Peak Area: 359
Analyte Ion Transition: 373.2/299.0 amu



Sample Name: ARA-09-15-10-141-0002A1
Data File: ARA-09-15-10 Q12.wiff
Sample Index: 50
Acquired: 6/9/2010 4:09:22 AM
Analyte Peak Area: 0
Analyte Ion Transition: 373.2/91.0 amu



Sample Name: ARA-09-15-10-141-0002A1
Data File: ARA-09-15-10 Q12.wiff
Sample Index: 50
Acquired: 6/9/2010 4:09:22 AM
Analyte Peak Area: 0
Analyte Ion Transition: 343.1/270.9 amu

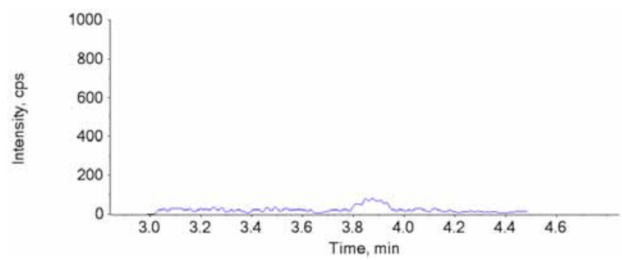
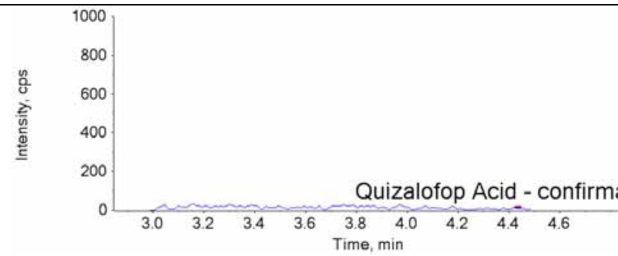
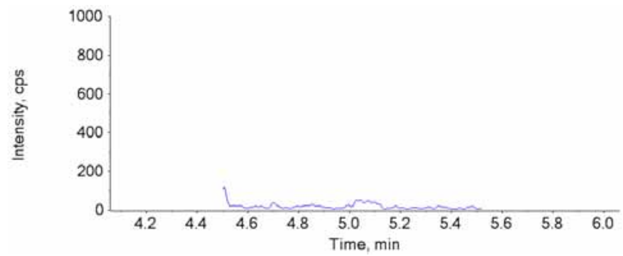


Figure 133. Typical Chromatogram of Quizalofop Analysis, Treated Corn Aspirated Grain Fractions (141-0002)

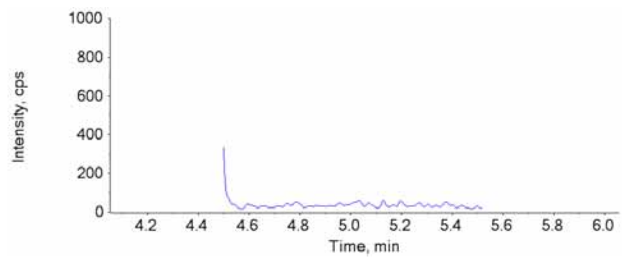
Sample Name: ARA-09-15-10-150-0002A1
Data File: ARA-09-15-10 Q12.wiff
Sample Index: 51
Acquired: 6/9/2010 4:21:51 AM
Analyte Peak Area: 0
Analyte Ion Transition: 343.1/108.1 amu



Sample Name: ARA-09-15-10-150-0002A1
Data File: ARA-09-15-10 Q12.wiff
Sample Index: 51
Acquired: 6/9/2010 4:21:51 AM
Analyte Peak Area: 0
Analyte Ion Transition: 373.2/299.0 amu



Sample Name: ARA-09-15-10-150-0002A1
Data File: ARA-09-15-10 Q12.wiff
Sample Index: 51
Acquired: 6/9/2010 4:21:51 AM
Analyte Peak Area: 0
Analyte Ion Transition: 373.2/91.0 amu



Sample Name: ARA-09-15-10-150-0002A1
Data File: ARA-09-15-10 Q12.wiff
Sample Index: 51
Acquired: 6/9/2010 4:21:51 AM
Analyte Peak Area: 0
Analyte Ion Transition: 343.1/270.9 amu

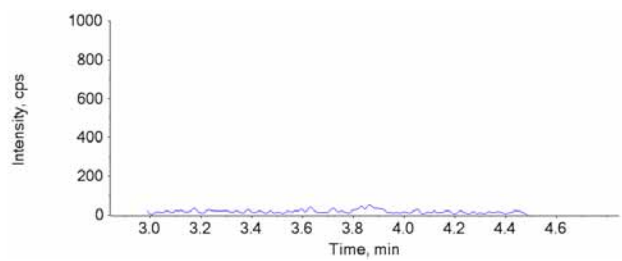
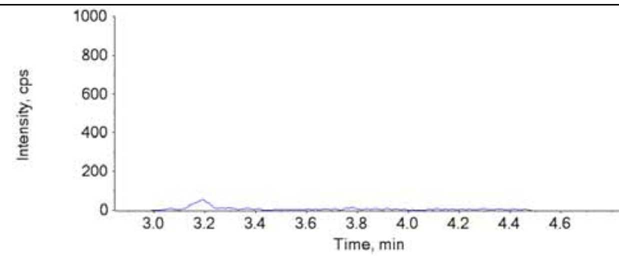
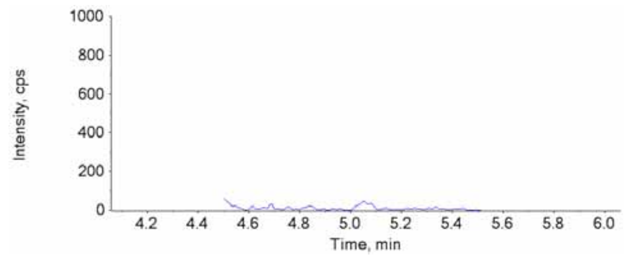


Figure 134. Typical Chromatogram of Quizalofop Analysis, Treated Corn Aspirated Grain Fractions (150-0002)

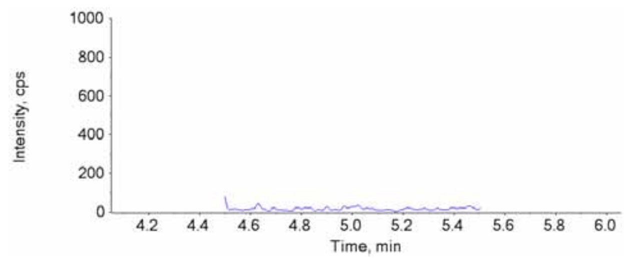
Sample Name: ARA-09-15-10-145-0001A9
Data File: ARA-09-15-10 Q13.wiff
Sample Index: 4
Acquired: 6/10/2010 4:34:14 AM
Analyte Peak Area: 0
Analyte Ion Transition: 343.1/108.1 amu



Sample Name: ARA-09-15-10-145-0001A9
Data File: ARA-09-15-10 Q13.wiff
Sample Index: 4
Acquired: 6/10/2010 4:34:14 AM
Analyte Peak Area: 0
Analyte Ion Transition: 373.2/299.0 amu



Sample Name: ARA-09-15-10-145-0001A9
Data File: ARA-09-15-10 Q13.wiff
Sample Index: 4
Acquired: 6/10/2010 4:34:14 AM
Analyte Peak Area: 0
Analyte Ion Transition: 373.2/91.0 amu



Sample Name: ARA-09-15-10-145-0001A9
Data File: ARA-09-15-10 Q13.wiff
Sample Index: 4
Acquired: 6/10/2010 4:34:14 AM
Analyte Peak Area: 0
Analyte Ion Transition: 343.1/270.9 amu

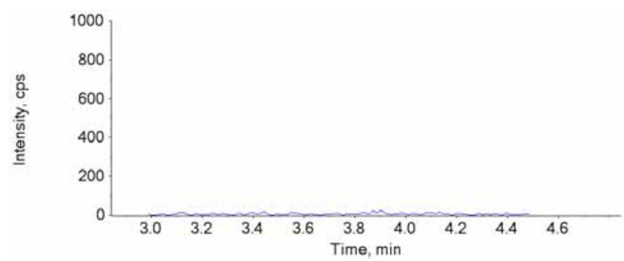
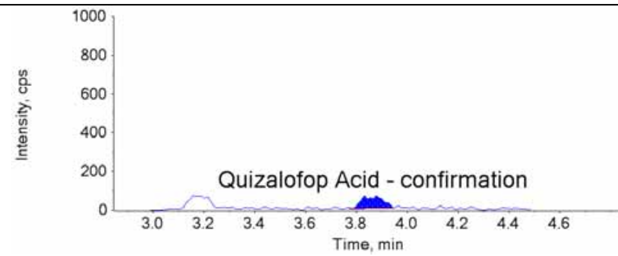
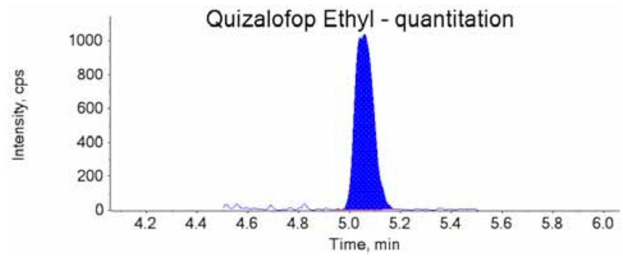


Figure 135. Typical Chromatogram of Quizalofop Analysis, Control Corn Grits (145-0001)

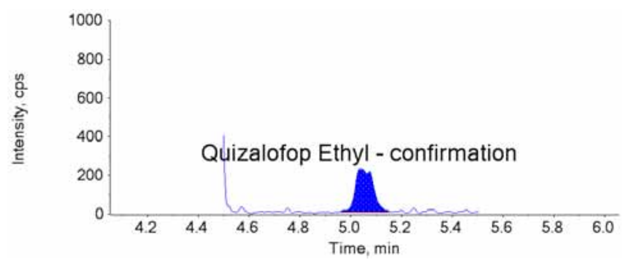
Sample Name: ARA-09-15-10-145-0001A11 + 0.01 µg/g
Data File: ARA-09-15-10 Q13.wiff
Sample Index: 18
Acquired: 6/10/2010 7:29:01 AM
Analyte Peak Area: 383
Analyte Ion Transition: 343.1/108.1 amu



Sample Name: ARA-09-15-10-145-0001A11 + 0.01 µg/g
Data File: ARA-09-15-10 Q13.wiff
Sample Index: 18
Acquired: 6/10/2010 7:29:01 AM
Analyte Peak Area: 5328
Analyte Ion Transition: 373.2/299.0 amu



Sample Name: ARA-09-15-10-145-0001A11 + 0.01 µg/g
Data File: ARA-09-15-10 Q13.wiff
Sample Index: 18
Acquired: 6/10/2010 7:29:01 AM
Analyte Peak Area: 1103
Analyte Ion Transition: 373.2/91.0 amu



Sample Name: ARA-09-15-10-145-0001A11 + 0.01 µg/g
Data File: ARA-09-15-10 Q13.wiff
Sample Index: 18
Acquired: 6/10/2010 7:29:01 AM
Analyte Peak Area: 5013
Analyte Ion Transition: 343.1/270.9 amu

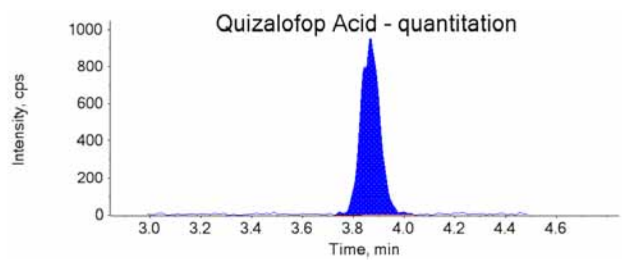
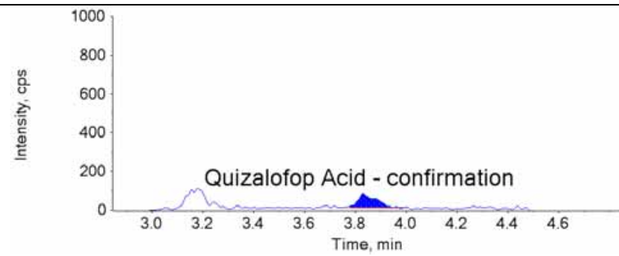
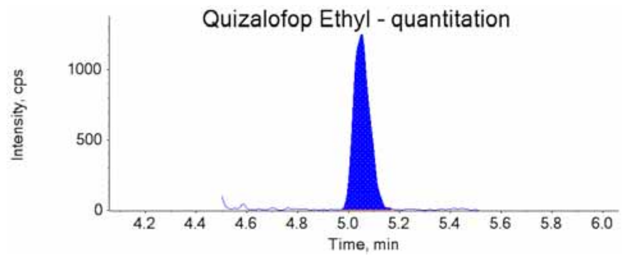


Figure 136. Typical Chromatogram of Quizalofop Analysis, Corn Grits 0.01 µg/g Recovery

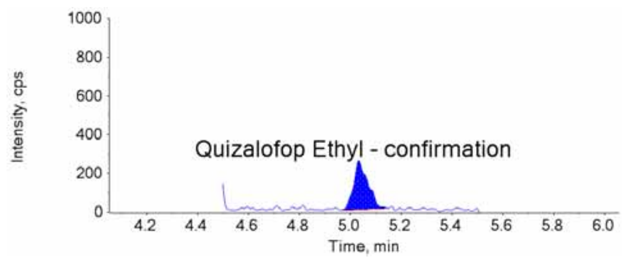
Sample Name: ARA-09-15-10-145-0001A12 + 0.01 µg/g
Data File: ARA-09-15-10 Q13.wiff
Sample Index: 19
Acquired: 6/10/2010 7:41:29 AM
Analyte Peak Area: 342
Analyte Ion Transition: 343.1/108.1 amu



Sample Name: ARA-09-15-10-145-0001A12 + 0.01 µg/g
Data File: ARA-09-15-10 Q13.wiff
Sample Index: 19
Acquired: 6/10/2010 7:41:29 AM
Analyte Peak Area: 5616
Analyte Ion Transition: 373.2/299.0 amu



Sample Name: ARA-09-15-10-145-0001A12 + 0.01 µg/g
Data File: ARA-09-15-10 Q13.wiff
Sample Index: 19
Acquired: 6/10/2010 7:41:29 AM
Analyte Peak Area: 985
Analyte Ion Transition: 373.2/91.0 amu



Sample Name: ARA-09-15-10-145-0001A12 + 0.01 µg/g
Data File: ARA-09-15-10 Q13.wiff
Sample Index: 19
Acquired: 6/10/2010 7:41:29 AM
Analyte Peak Area: 5403
Analyte Ion Transition: 343.1/270.9 amu

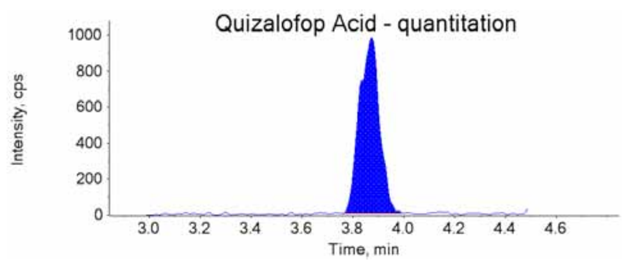
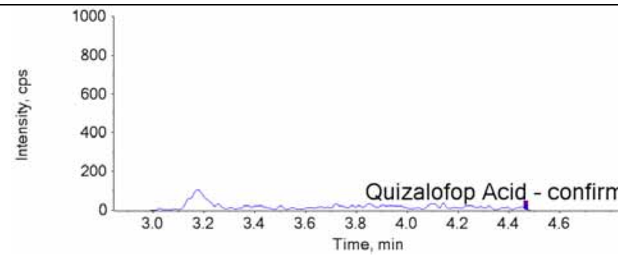
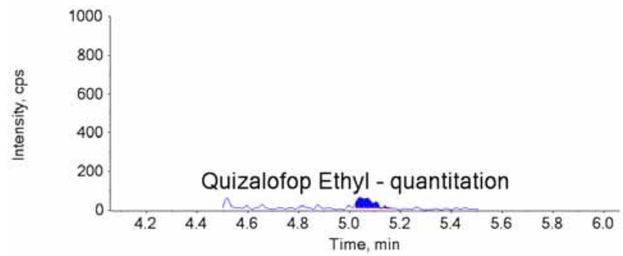


Figure 137. Typical Chromatogram of Quizalofop Analysis, Corn Grits 0.01 µg/g Recovery

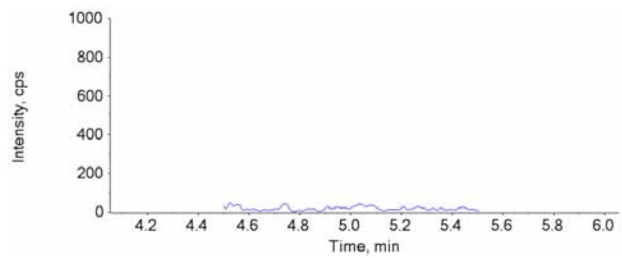
Sample Name: ARA-09-15-10-145-0003A1
Data File: ARA-09-15-10 Q13.wiff
Sample Index: 43
Acquired: 6/10/2010 12:41:01 PM
Analyte Peak Area: 0
Analyte Ion Transition: 343.1/108.1 amu



Sample Name: ARA-09-15-10-145-0003A1
Data File: ARA-09-15-10 Q13.wiff
Sample Index: 43
Acquired: 6/10/2010 12:41:01 PM
Analyte Peak Area: 219
Analyte Ion Transition: 373.2/299.0 amu



Sample Name: ARA-09-15-10-145-0003A1
Data File: ARA-09-15-10 Q13.wiff
Sample Index: 43
Acquired: 6/10/2010 12:41:01 PM
Analyte Peak Area: 0
Analyte Ion Transition: 373.2/91.0 amu



Sample Name: ARA-09-15-10-145-0003A1
Data File: ARA-09-15-10 Q13.wiff
Sample Index: 43
Acquired: 6/10/2010 12:41:01 PM
Analyte Peak Area: 89
Analyte Ion Transition: 343.1/270.9 amu

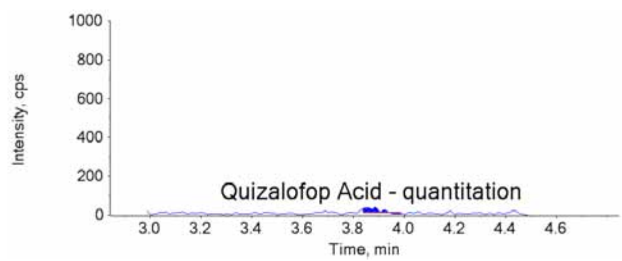
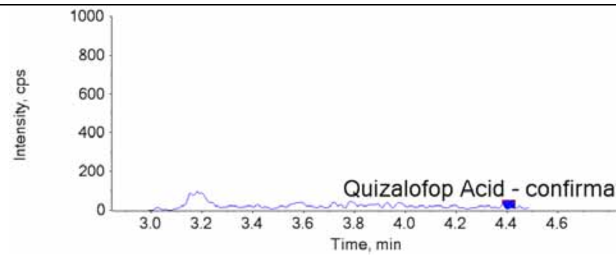
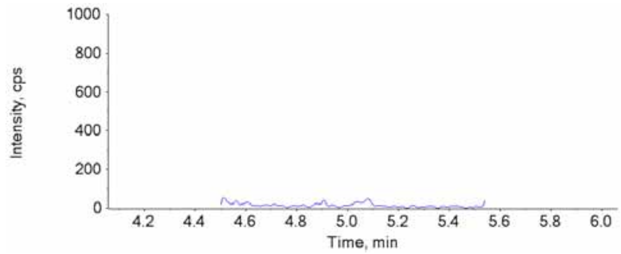


Figure 138. Typical Chromatogram of Quizalofop Analysis, Treated Corn Grits (145-0003)

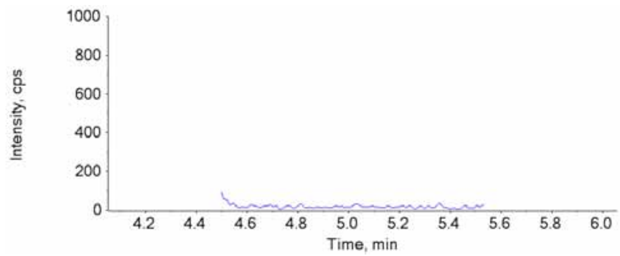
Sample Name: ARA-09-15-10-145-0004A1
Data File: ARA-09-15-10 Q13.wiff
Sample Index: 44
Acquired: 6/10/2010 12:53:29 PM
Analyte Peak Area: 0
Analyte Ion Transition: 343.1/108.1 amu



Sample Name: ARA-09-15-10-145-0004A1
Data File: ARA-09-15-10 Q13.wiff
Sample Index: 44
Acquired: 6/10/2010 12:53:29 PM
Analyte Peak Area: 0
Analyte Ion Transition: 373.2/299.0 amu



Sample Name: ARA-09-15-10-145-0004A1
Data File: ARA-09-15-10 Q13.wiff
Sample Index: 44
Acquired: 6/10/2010 12:53:29 PM
Analyte Peak Area: 0
Analyte Ion Transition: 373.2/91.0 amu



Sample Name: ARA-09-15-10-145-0004A1
Data File: ARA-09-15-10 Q13.wiff
Sample Index: 44
Acquired: 6/10/2010 12:53:29 PM
Analyte Peak Area: 0
Analyte Ion Transition: 343.1/270.9 amu

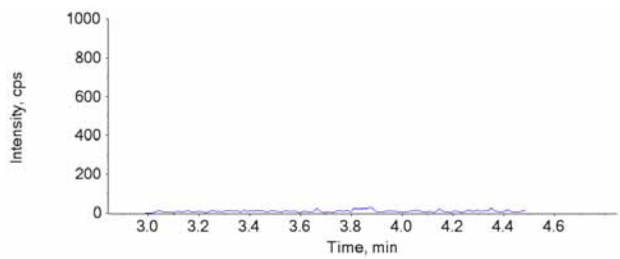
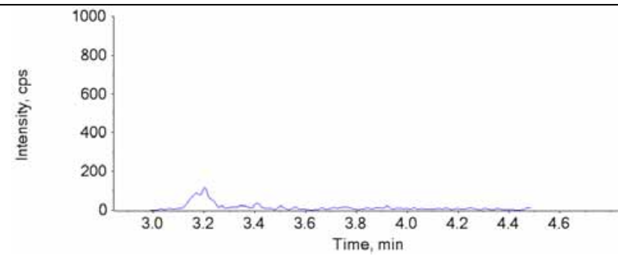
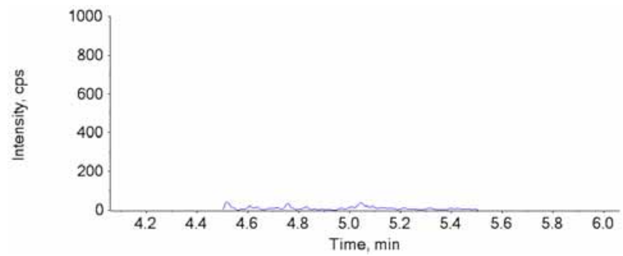


Figure 139. Typical Chromatogram of Quizalofop Analysis, Treated Corn Grits (145-0004)

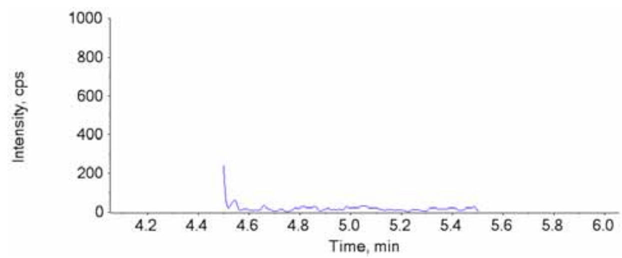
Sample Name: ARA-09-15-10-144-0001A9
Data File: ARA-09-15-10 Q13.wiff
Sample Index: 5
Acquired: 6/10/2010 4:46:43 AM
Analyte Peak Area: 0
Analyte Ion Transition: 343.1/108.1 amu



Sample Name: ARA-09-15-10-144-0001A9
Data File: ARA-09-15-10 Q13.wiff
Sample Index: 5
Acquired: 6/10/2010 4:46:43 AM
Analyte Peak Area: 0
Analyte Ion Transition: 373.2/299.0 amu



Sample Name: ARA-09-15-10-144-0001A9
Data File: ARA-09-15-10 Q13.wiff
Sample Index: 5
Acquired: 6/10/2010 4:46:43 AM
Analyte Peak Area: 0
Analyte Ion Transition: 373.2/91.0 amu



Sample Name: ARA-09-15-10-144-0001A9
Data File: ARA-09-15-10 Q13.wiff
Sample Index: 5
Acquired: 6/10/2010 4:46:43 AM
Analyte Peak Area: 0
Analyte Ion Transition: 343.1/270.9 amu

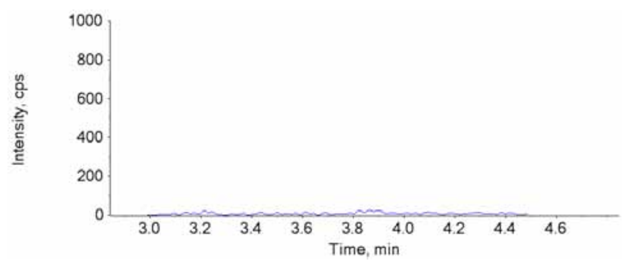
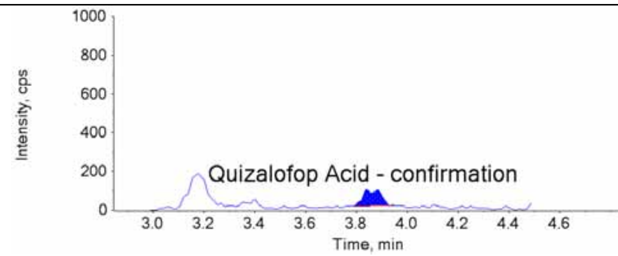
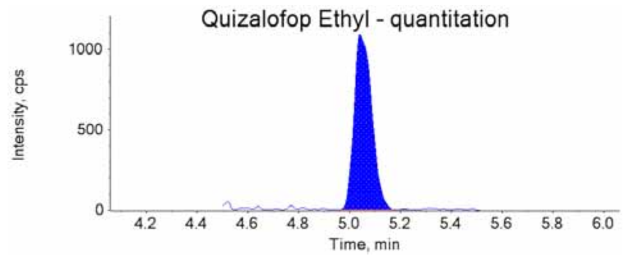


Figure 140. Typical Chromatogram of Quizalofop Analysis, Control Corn Meal (144-0001)

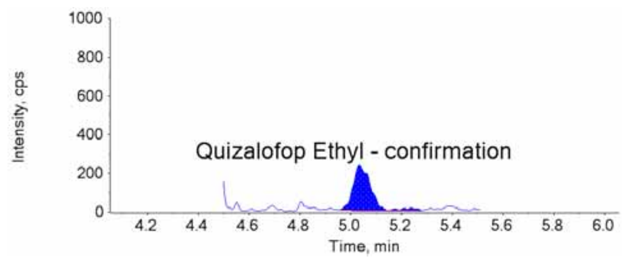
Sample Name: ARA-09-15-10-144-0001A11 + 0.01 µg/g
Data File: ARA-09-15-10 Q13.wiff
Sample Index: 21
Acquired: 6/10/2010 8:06:26 AM
Analyte Peak Area: 415
Analyte Ion Transition: 343.1/108.1 amu



Sample Name: ARA-09-15-10-144-0001A11 + 0.01 µg/g
Data File: ARA-09-15-10 Q13.wiff
Sample Index: 21
Acquired: 6/10/2010 8:06:26 AM
Analyte Peak Area: 5310
Analyte Ion Transition: 373.2/299.0 amu



Sample Name: ARA-09-15-10-144-0001A11 + 0.01 µg/g
Data File: ARA-09-15-10 Q13.wiff
Sample Index: 21
Acquired: 6/10/2010 8:06:26 AM
Analyte Peak Area: 1164
Analyte Ion Transition: 373.2/91.0 amu



Sample Name: ARA-09-15-10-144-0001A11 + 0.01 µg/g
Data File: ARA-09-15-10 Q13.wiff
Sample Index: 21
Acquired: 6/10/2010 8:06:26 AM
Analyte Peak Area: 5503
Analyte Ion Transition: 343.1/270.9 amu

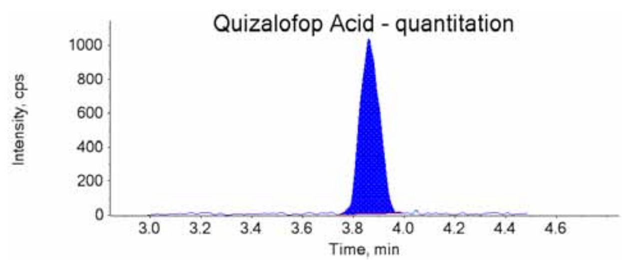
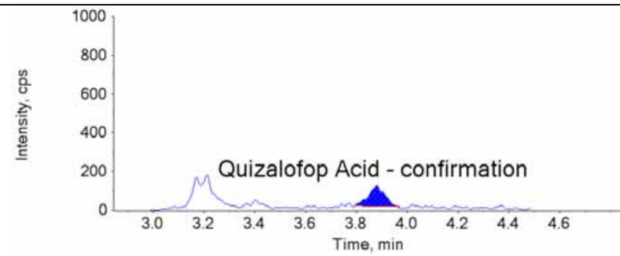
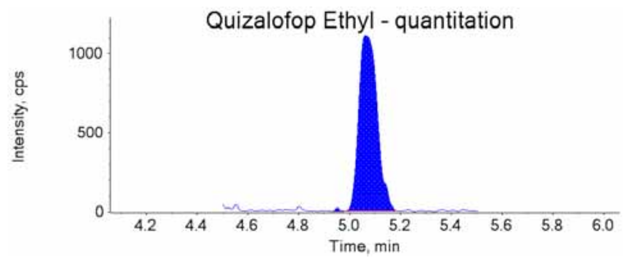


Figure 141. Typical Chromatogram of Quizalofop Analysis, Corn Meal 0.01 µg/g Recovery

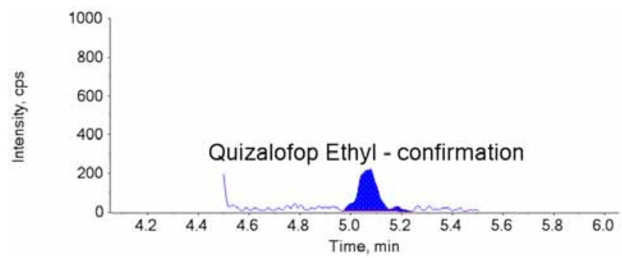
Sample Name: ARA-09-15-10-144-0001A12 + 0.01 µg/g
Data File: ARA-09-15-10 Q13.wiff
Sample Index: 22
Acquired: 6/10/2010 8:18:55 AM
Analyte Peak Area: 451
Analyte Ion Transition: 343.1/108.1 amu



Sample Name: ARA-09-15-10-144-0001A12 + 0.01 µg/g
Data File: ARA-09-15-10 Q13.wiff
Sample Index: 22
Acquired: 6/10/2010 8:18:55 AM
Analyte Peak Area: 5615
Analyte Ion Transition: 373.2/299.0 amu



Sample Name: ARA-09-15-10-144-0001A12 + 0.01 µg/g
Data File: ARA-09-15-10 Q13.wiff
Sample Index: 22
Acquired: 6/10/2010 8:18:55 AM
Analyte Peak Area: 1173
Analyte Ion Transition: 373.2/91.0 amu



Sample Name: ARA-09-15-10-144-0001A12 + 0.01 µg/g
Data File: ARA-09-15-10 Q13.wiff
Sample Index: 22
Acquired: 6/10/2010 8:18:55 AM
Analyte Peak Area: 6027
Analyte Ion Transition: 343.1/270.9 amu

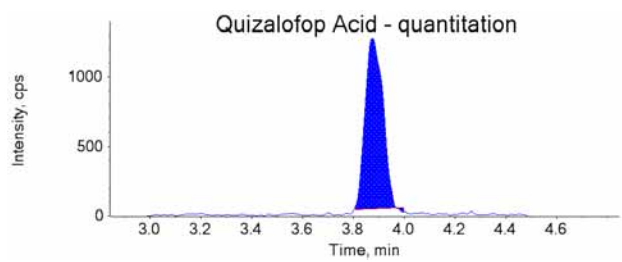
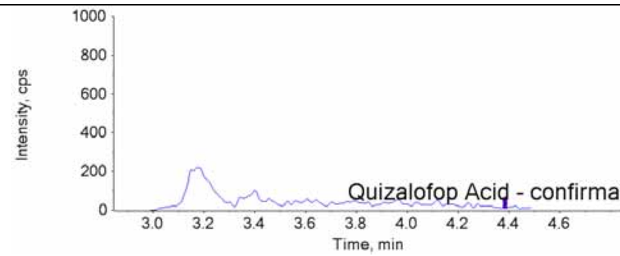
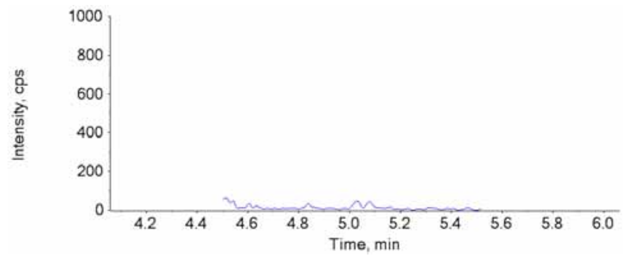


Figure 142. Typical Chromatogram of Quizalofop Analysis, Corn Meal 0.01 µg/g Recovery

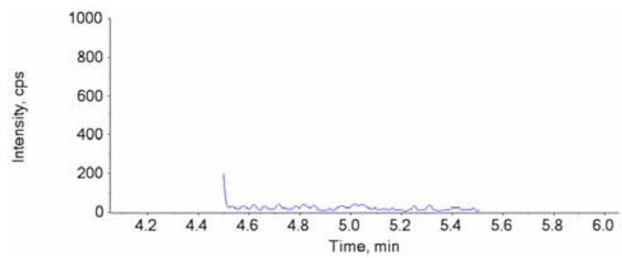
Sample Name: ARA-09-15-10-144-0003A1
Data File: ARA-09-15-10 Q13.wiff
Sample Index: 47
Acquired: 6/10/2010 1:30:56 PM
Analyte Peak Area: 0
Analyte Ion Transition: 343.1/108.1 amu



Sample Name: ARA-09-15-10-144-0003A1
Data File: ARA-09-15-10 Q13.wiff
Sample Index: 47
Acquired: 6/10/2010 1:30:56 PM
Analyte Peak Area: 0
Analyte Ion Transition: 373.2/299.0 amu



Sample Name: ARA-09-15-10-144-0003A1
Data File: ARA-09-15-10 Q13.wiff
Sample Index: 47
Acquired: 6/10/2010 1:30:56 PM
Analyte Peak Area: 0
Analyte Ion Transition: 373.2/91.0 amu



Sample Name: ARA-09-15-10-144-0003A1
Data File: ARA-09-15-10 Q13.wiff
Sample Index: 47
Acquired: 6/10/2010 1:30:56 PM
Analyte Peak Area: 0
Analyte Ion Transition: 343.1/270.9 amu

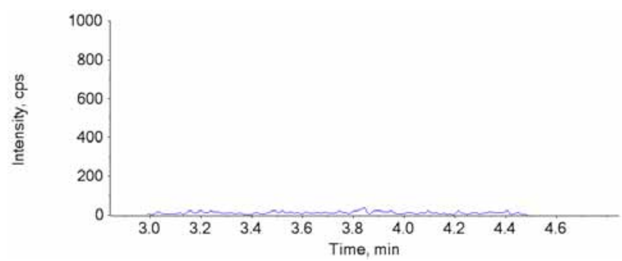
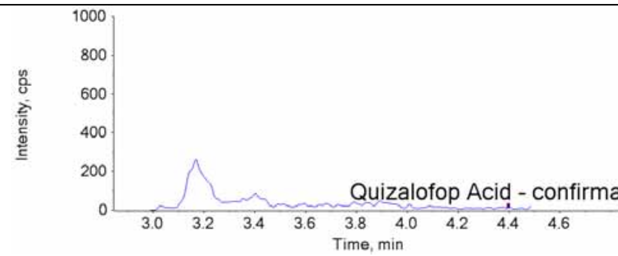
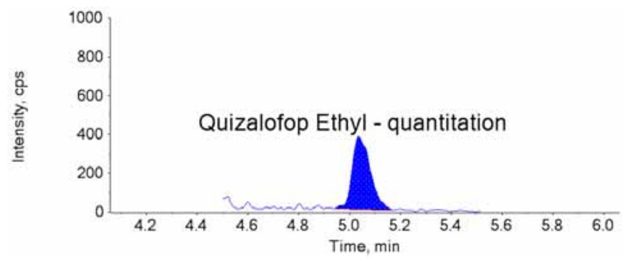


Figure 143. Typical Chromatogram of Quizalofop Analysis, Treated Corn Meal (144-0003)

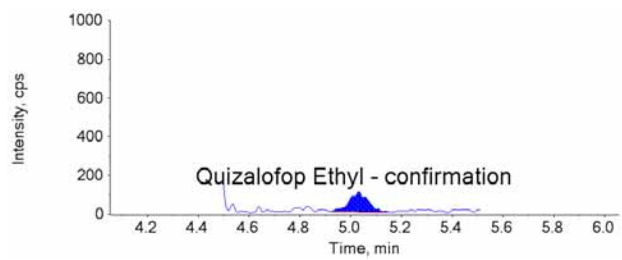
Sample Name: ARA-09-15-10-144-0004A1
Data File: ARA-09-15-10 Q13.wiff
Sample Index: 49
Acquired: 6/10/2010 1:55:52 PM
Analyte Peak Area: 0
Analyte Ion Transition: 343.1/108.1 amu



Sample Name: ARA-09-15-10-144-0004A1
Data File: ARA-09-15-10 Q13.wiff
Sample Index: 49
Acquired: 6/10/2010 1:55:52 PM
Analyte Peak Area: 1871
Analyte Ion Transition: 373.2/299.0 amu



Sample Name: ARA-09-15-10-144-0004A1
Data File: ARA-09-15-10 Q13.wiff
Sample Index: 49
Acquired: 6/10/2010 1:55:52 PM
Analyte Peak Area: 532
Analyte Ion Transition: 373.2/91.0 amu



Sample Name: ARA-09-15-10-144-0004A1
Data File: ARA-09-15-10 Q13.wiff
Sample Index: 49
Acquired: 6/10/2010 1:55:52 PM
Analyte Peak Area: 870
Analyte Ion Transition: 343.1/270.9 amu

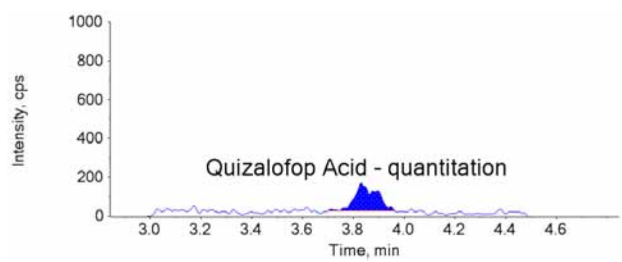
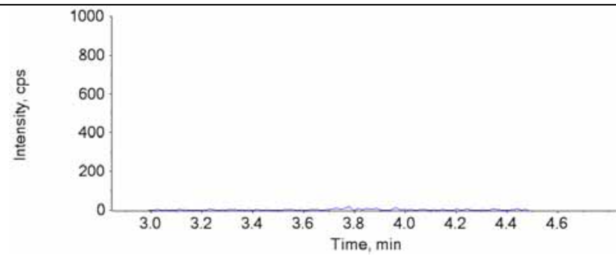
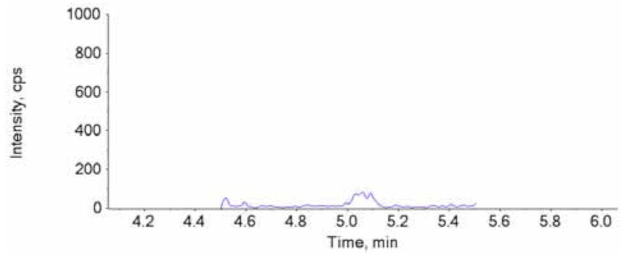


Figure 144. Typical Chromatogram of Quizalofop Analysis, Treated Corn Meal (144-0004)

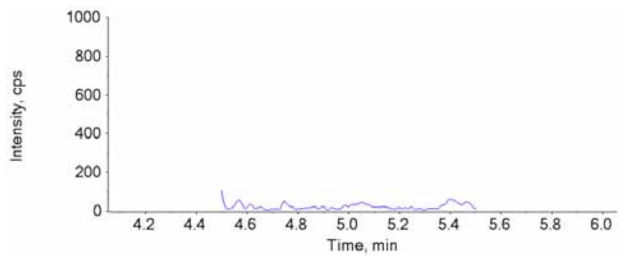
Sample Name: ARA-09-15-10-143-0001A9
Data File: ARA-09-15-10 Q15.wiff
Sample Index: 2
Acquired: 6/10/2010 7:37:34 PM
Analyte Peak Area: 0
Analyte Ion Transition: 343.1/108.1 amu



Sample Name: ARA-09-15-10-143-0001A9
Data File: ARA-09-15-10 Q15.wiff
Sample Index: 2
Acquired: 6/10/2010 7:37:34 PM
Analyte Peak Area: 0
Analyte Ion Transition: 373.2/299.0 amu



Sample Name: ARA-09-15-10-143-0001A9
Data File: ARA-09-15-10 Q15.wiff
Sample Index: 2
Acquired: 6/10/2010 7:37:34 PM
Analyte Peak Area: 0
Analyte Ion Transition: 373.2/91.0 amu



Sample Name: ARA-09-15-10-143-0001A9
Data File: ARA-09-15-10 Q15.wiff
Sample Index: 2
Acquired: 6/10/2010 7:37:34 PM
Analyte Peak Area: 0
Analyte Ion Transition: 343.1/270.9 amu

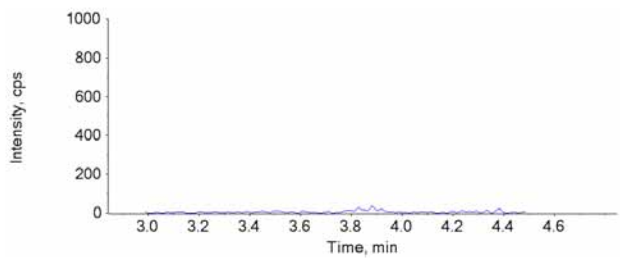
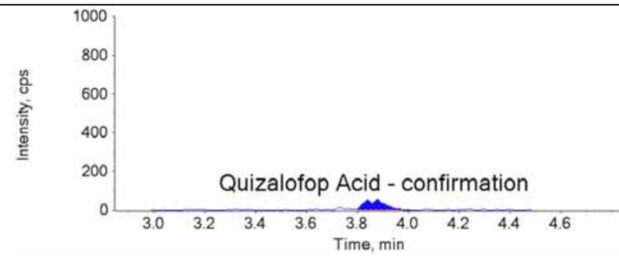
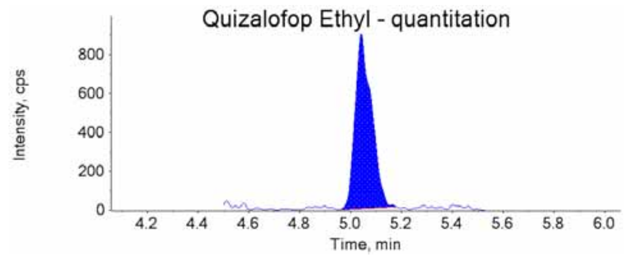


Figure 145. Typical Chromatogram of Quizalofop Analysis, Control Corn Oil, Wet Processed (143-0001)

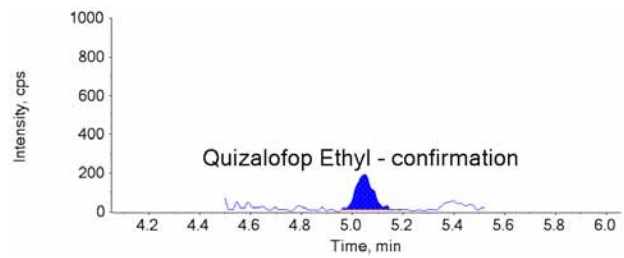
Sample Name: ARA-09-15-10-143-0001A11 + 0.01 µg/g
Data File: ARA-09-15-10 Q15.wiff
Sample Index: 9
Acquired: 6/10/2010 9:04:56 PM
Analyte Peak Area: 274
Analyte Ion Transition: 343.1/108.1 amu



Sample Name: ARA-09-15-10-143-0001A11 + 0.01 µg/g
Data File: ARA-09-15-10 Q15.wiff
Sample Index: 9
Acquired: 6/10/2010 9:04:56 PM
Analyte Peak Area: 4044
Analyte Ion Transition: 373.2/299.0 amu



Sample Name: ARA-09-15-10-143-0001A11 + 0.01 µg/g
Data File: ARA-09-15-10 Q15.wiff
Sample Index: 9
Acquired: 6/10/2010 9:04:56 PM
Analyte Peak Area: 852
Analyte Ion Transition: 373.2/91.0 amu



Sample Name: ARA-09-15-10-143-0001A11 + 0.01 µg/g
Data File: ARA-09-15-10 Q15.wiff
Sample Index: 9
Acquired: 6/10/2010 9:04:56 PM
Analyte Peak Area: 3399
Analyte Ion Transition: 343.1/270.9 amu

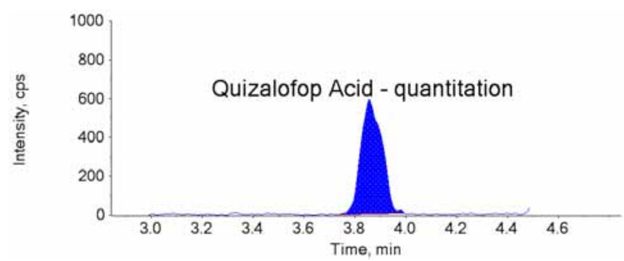
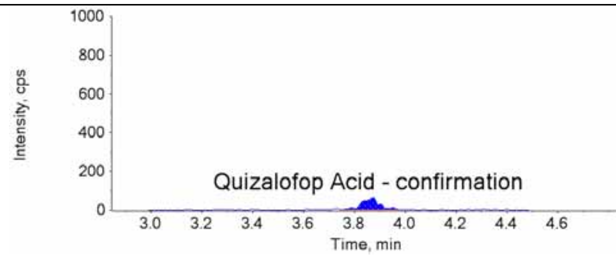
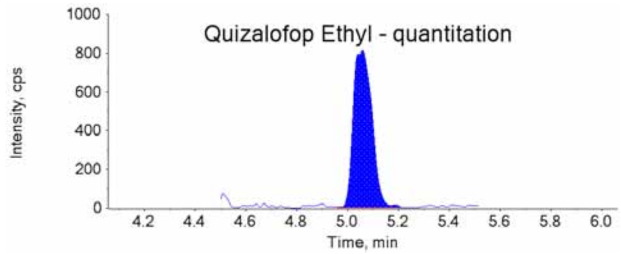


Figure 146. Typical Chromatogram of Quizalofop Analysis, Corn Oil (Wet Processed) 0.01 µg/g Recovery

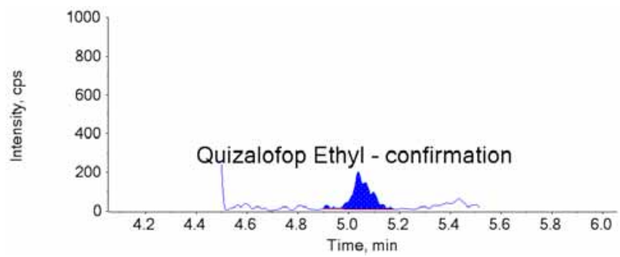
Sample Name: ARA-09-15-10-143-0001A12 + 0.01 µg/g
Data File: ARA-09-15-10 Q15.wiff
Sample Index: 10
Acquired: 6/10/2010 9:17:23 PM
Analyte Peak Area: 304
Analyte Ion Transition: 343.1/108.1 amu



Sample Name: ARA-09-15-10-143-0001A12 + 0.01 µg/g
Data File: ARA-09-15-10 Q15.wiff
Sample Index: 10
Acquired: 6/10/2010 9:17:23 PM
Analyte Peak Area: 4160
Analyte Ion Transition: 373.2/299.0 amu



Sample Name: ARA-09-15-10-143-0001A12 + 0.01 µg/g
Data File: ARA-09-15-10 Q15.wiff
Sample Index: 10
Acquired: 6/10/2010 9:17:23 PM
Analyte Peak Area: 877
Analyte Ion Transition: 373.2/91.0 amu



Sample Name: ARA-09-15-10-143-0001A12 + 0.01 µg/g
Data File: ARA-09-15-10 Q15.wiff
Sample Index: 10
Acquired: 6/10/2010 9:17:23 PM
Analyte Peak Area: 3806
Analyte Ion Transition: 343.1/270.9 amu

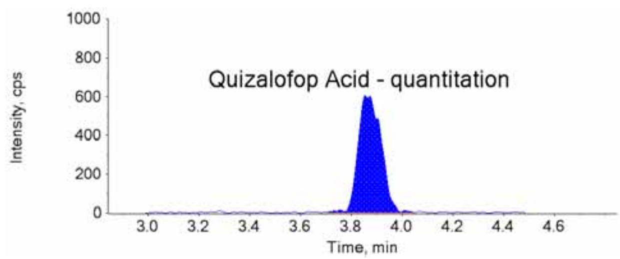
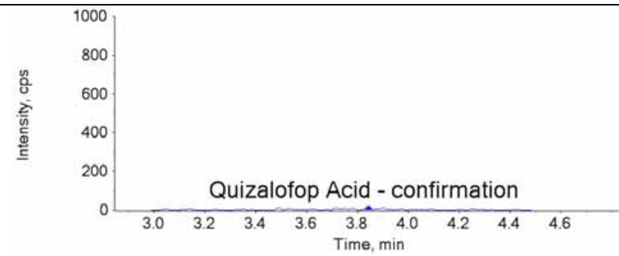
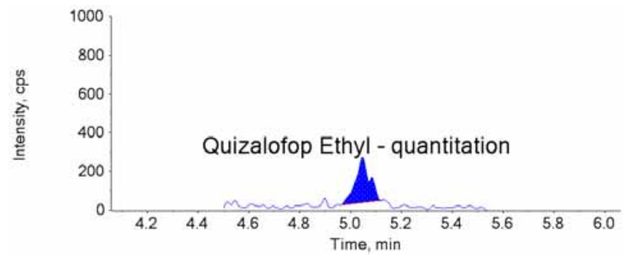


Figure 147. Typical Chromatogram of Quizalofop Analysis, Corn Oil (Wet Processed) 0.01 µg/g Recovery

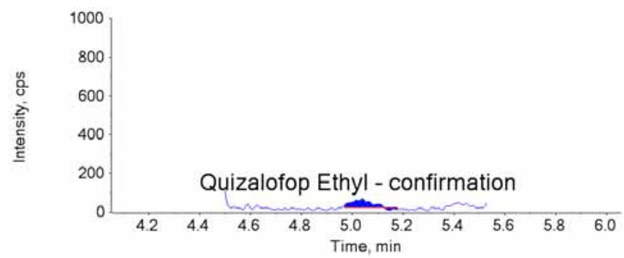
Sample Name: ARA-09-15-10-143-0003A1
Data File: ARA-09-15-10 Q15.wiff
Sample Index: 30
Acquired: 6/11/2010 1:26:53 AM
Analyte Peak Area: 25
Analyte Ion Transition: 343.1/108.1 amu



Sample Name: ARA-09-15-10-143-0003A1
Data File: ARA-09-15-10 Q15.wiff
Sample Index: 30
Acquired: 6/11/2010 1:26:53 AM
Analyte Peak Area: 902
Analyte Ion Transition: 373.2/299.0 amu



Sample Name: ARA-09-15-10-143-0003A1
Data File: ARA-09-15-10 Q15.wiff
Sample Index: 30
Acquired: 6/11/2010 1:26:53 AM
Analyte Peak Area: 254
Analyte Ion Transition: 373.2/91.0 amu



Sample Name: ARA-09-15-10-143-0003A1
Data File: ARA-09-15-10 Q15.wiff
Sample Index: 30
Acquired: 6/11/2010 1:26:53 AM
Analyte Peak Area: 391
Analyte Ion Transition: 343.1/270.9 amu

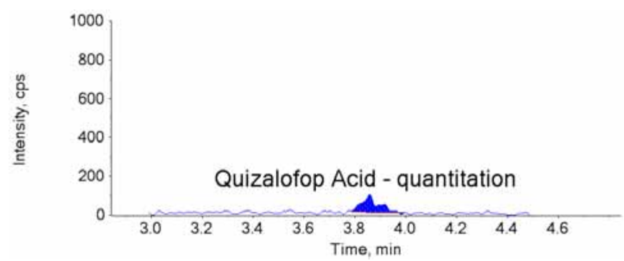
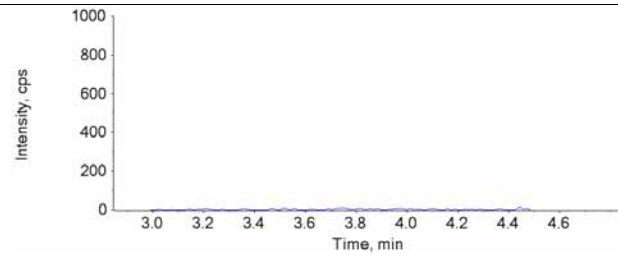
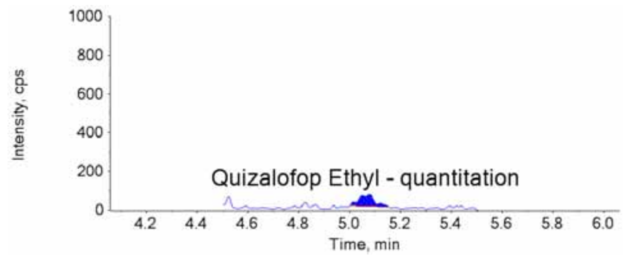


Figure 148. Typical Chromatogram of Quizalofop Analysis, Treated Corn Oil, Wet Processed (143-0003)

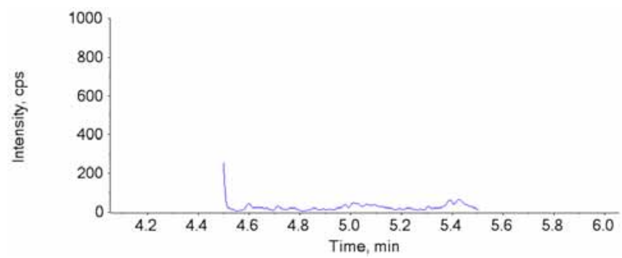
Sample Name: ARA-09-15-10-143-0004A1
Data File: ARA-09-15-10 Q15.wiff
Sample Index: 31
Acquired: 6/11/2010 1:39:20 AM
Analyte Peak Area: 0
Analyte Ion Transition: 343.1/108.1 amu



Sample Name: ARA-09-15-10-143-0004A1
Data File: ARA-09-15-10 Q15.wiff
Sample Index: 31
Acquired: 6/11/2010 1:39:20 AM
Analyte Peak Area: 272
Analyte Ion Transition: 373.2/299.0 amu



Sample Name: ARA-09-15-10-143-0004A1
Data File: ARA-09-15-10 Q15.wiff
Sample Index: 31
Acquired: 6/11/2010 1:39:20 AM
Analyte Peak Area: 0
Analyte Ion Transition: 373.2/91.0 amu



Sample Name: ARA-09-15-10-143-0004A1
Data File: ARA-09-15-10 Q15.wiff
Sample Index: 31
Acquired: 6/11/2010 1:39:20 AM
Analyte Peak Area: 81
Analyte Ion Transition: 343.1/270.9 amu

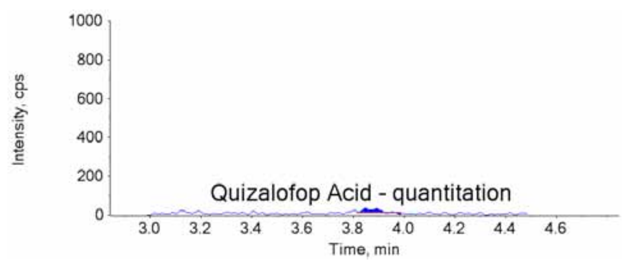
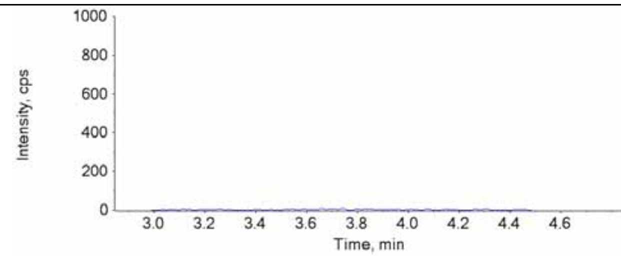
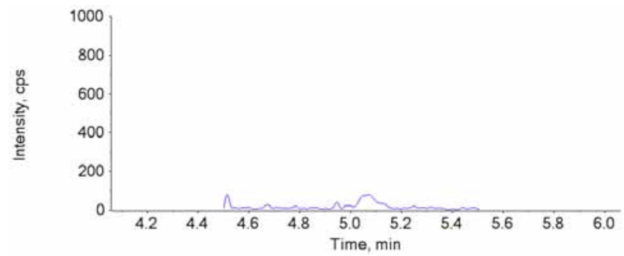


Figure 149. Typical Chromatogram of Quizalofop Analysis, Treated Corn Oil, Wet Processed (143-0004)

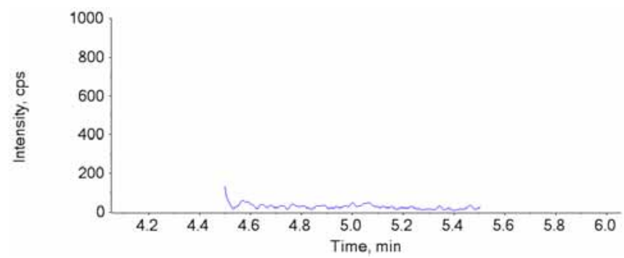
Sample Name: ARA-09-15-10-147-0001A9
Data File: ARA-09-15-10 Q15.wiff
Sample Index: 3
Acquired: 6/10/2010 7:50:03 PM
Analyte Peak Area: 0
Analyte Ion Transition: 343.1/108.1 amu



Sample Name: ARA-09-15-10-147-0001A9
Data File: ARA-09-15-10 Q15.wiff
Sample Index: 3
Acquired: 6/10/2010 7:50:03 PM
Analyte Peak Area: 0
Analyte Ion Transition: 373.2/299.0 amu



Sample Name: ARA-09-15-10-147-0001A9
Data File: ARA-09-15-10 Q15.wiff
Sample Index: 3
Acquired: 6/10/2010 7:50:03 PM
Analyte Peak Area: 0
Analyte Ion Transition: 373.2/91.0 amu



Sample Name: ARA-09-15-10-147-0001A9
Data File: ARA-09-15-10 Q15.wiff
Sample Index: 3
Acquired: 6/10/2010 7:50:03 PM
Analyte Peak Area: 0
Analyte Ion Transition: 343.1/270.9 amu

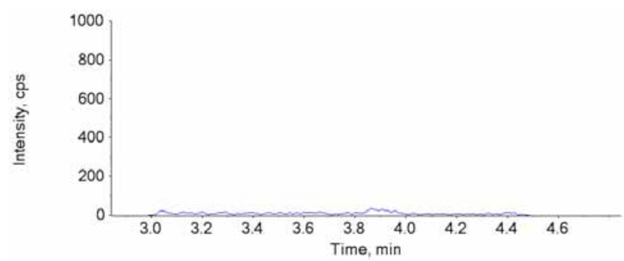
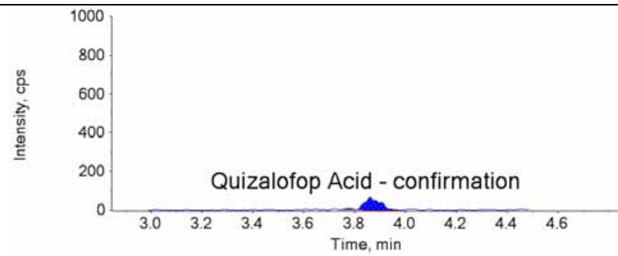
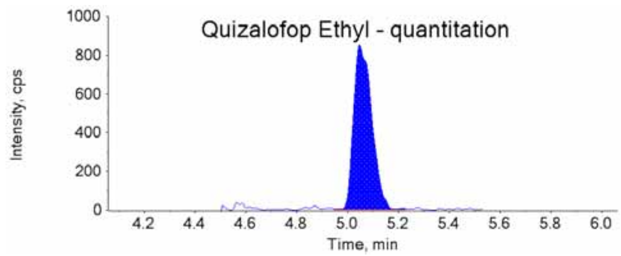


Figure 150. Typical Chromatogram of Quizalofop Analysis, Control Corn Oil, Dry Processed (147-0001)

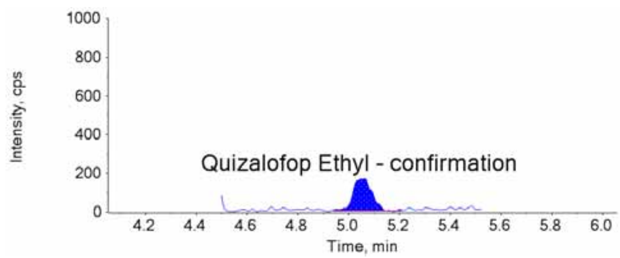
Sample Name: ARA-09-15-10-147-0001A11 + 0.01 µg/g
Data File: ARA-09-15-10 Q15.wiff
Sample Index: 12
Acquired: 6/10/2010 9:42:20 PM
Analyte Peak Area: 296
Analyte Ion Transition: 343.1/108.1 amu



Sample Name: ARA-09-15-10-147-0001A11 + 0.01 µg/g
Data File: ARA-09-15-10 Q15.wiff
Sample Index: 12
Acquired: 6/10/2010 9:42:20 PM
Analyte Peak Area: 4275
Analyte Ion Transition: 373.2/299.0 amu



Sample Name: ARA-09-15-10-147-0001A11 + 0.01 µg/g
Data File: ARA-09-15-10 Q15.wiff
Sample Index: 12
Acquired: 6/10/2010 9:42:20 PM
Analyte Peak Area: 895
Analyte Ion Transition: 373.2/91.0 amu



Sample Name: ARA-09-15-10-147-0001A11 + 0.01 µg/g
Data File: ARA-09-15-10 Q15.wiff
Sample Index: 12
Acquired: 6/10/2010 9:42:20 PM
Analyte Peak Area: 3530
Analyte Ion Transition: 343.1/270.9 amu

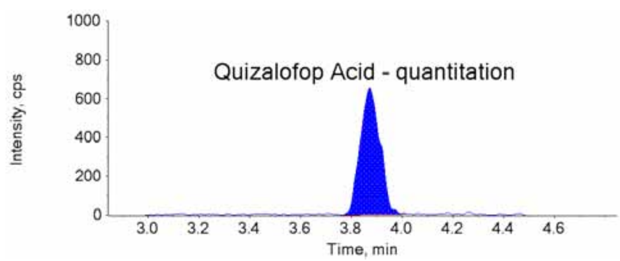
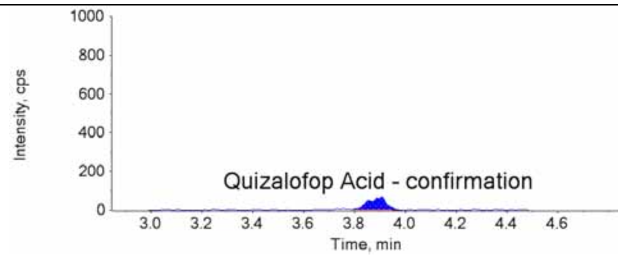
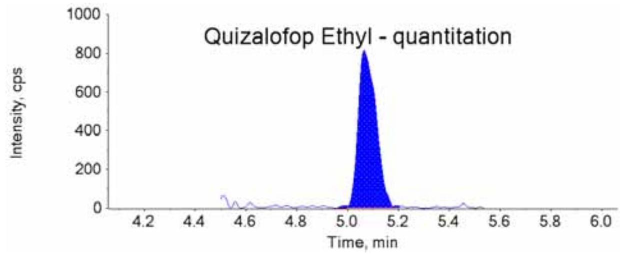


Figure 151. Typical Chromatogram of Quizalofop Analysis, Corn Oil (Dry Processed) 0.01 µg/g Recovery

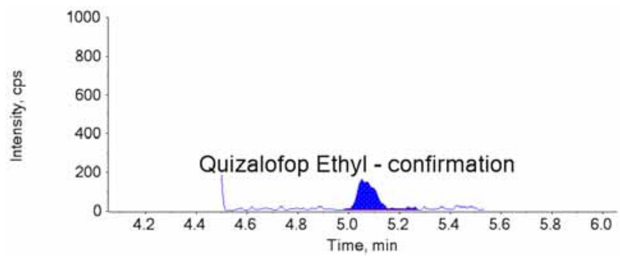
Sample Name: ARA-09-15-10-147-0001A12 + 0.01 µg/g
Data File: ARA-09-15-10 Q15.wiff
Sample Index: 13
Acquired: 6/10/2010 9:54:48 PM
Analyte Peak Area: 320
Analyte Ion Transition: 343.1/108.1 amu



Sample Name: ARA-09-15-10-147-0001A12 + 0.01 µg/g
Data File: ARA-09-15-10 Q15.wiff
Sample Index: 13
Acquired: 6/10/2010 9:54:48 PM
Analyte Peak Area: 3980
Analyte Ion Transition: 373.2/299.0 amu



Sample Name: ARA-09-15-10-147-0001A12 + 0.01 µg/g
Data File: ARA-09-15-10 Q15.wiff
Sample Index: 13
Acquired: 6/10/2010 9:54:48 PM
Analyte Peak Area: 851
Analyte Ion Transition: 373.2/91.0 amu



Sample Name: ARA-09-15-10-147-0001A12 + 0.01 µg/g
Data File: ARA-09-15-10 Q15.wiff
Sample Index: 13
Acquired: 6/10/2010 9:54:48 PM
Analyte Peak Area: 3908
Analyte Ion Transition: 343.1/270.9 amu

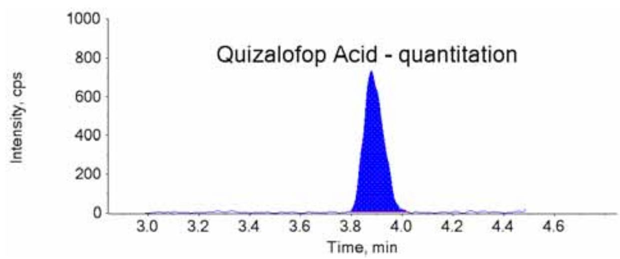
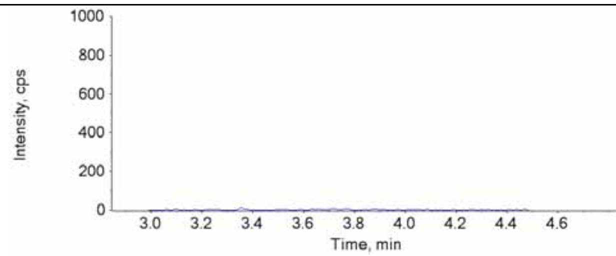
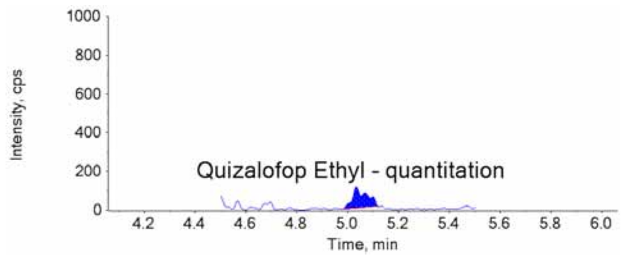


Figure 152. Typical Chromatogram of Quizalofop Analysis, Corn Oil (Dry Processed) 0.01 µg/g Recovery

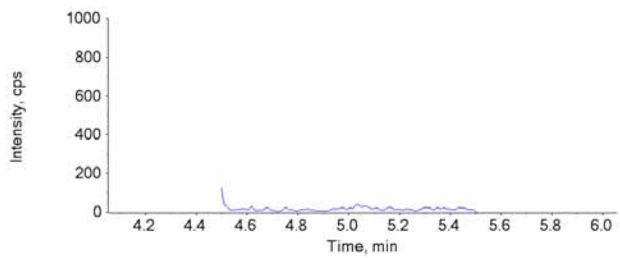
Sample Name: ARA-09-15-10-147-0003A1
Data File: ARA-09-15-10 Q15.wiff
Sample Index: 32
Acquired: 6/11/2010 1:51:49 AM
Analyte Peak Area: 0
Analyte Ion Transition: 343.1/108.1 amu



Sample Name: ARA-09-15-10-147-0003A1
Data File: ARA-09-15-10 Q15.wiff
Sample Index: 32
Acquired: 6/11/2010 1:51:49 AM
Analyte Peak Area: 421
Analyte Ion Transition: 373.2/299.0 amu



Sample Name: ARA-09-15-10-147-0003A1
Data File: ARA-09-15-10 Q15.wiff
Sample Index: 32
Acquired: 6/11/2010 1:51:49 AM
Analyte Peak Area: 0
Analyte Ion Transition: 373.2/91.0 amu



Sample Name: ARA-09-15-10-147-0003A1
Data File: ARA-09-15-10 Q15.wiff
Sample Index: 32
Acquired: 6/11/2010 1:51:49 AM
Analyte Peak Area: 93
Analyte Ion Transition: 343.1/270.9 amu

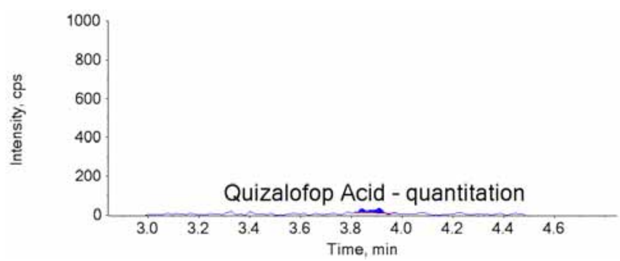
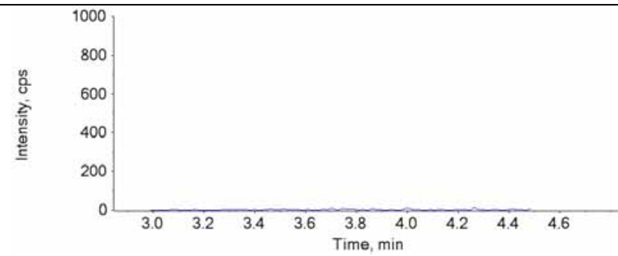
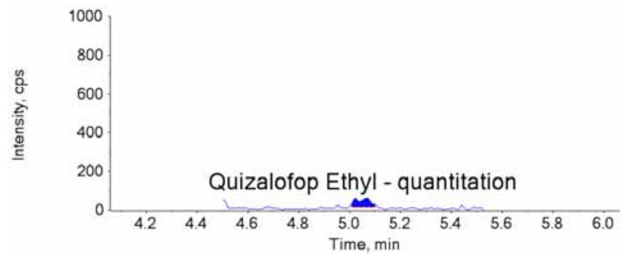


Figure 153. Typical Chromatogram of Quizalofop Analysis, Treated Corn Oil, Dry Processed (147-0003)

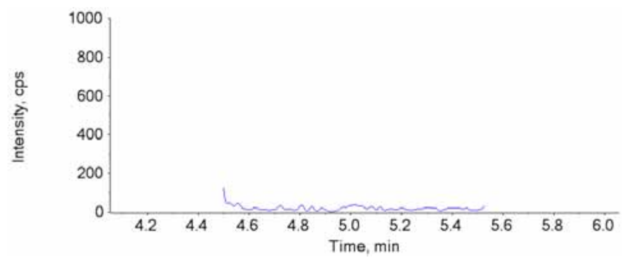
Sample Name: ARA-09-15-10-147-0004A1
Data File: ARA-09-15-10 Q15.wiff
Sample Index: 33
Acquired: 6/11/2010 2:04:17 AM
Analyte Peak Area: 0
Analyte Ion Transition: 343.1/108.1 amu



Sample Name: ARA-09-15-10-147-0004A1
Data File: ARA-09-15-10 Q15.wiff
Sample Index: 33
Acquired: 6/11/2010 2:04:17 AM
Analyte Peak Area: 179
Analyte Ion Transition: 373.2/299.0 amu



Sample Name: ARA-09-15-10-147-0004A1
Data File: ARA-09-15-10 Q15.wiff
Sample Index: 33
Acquired: 6/11/2010 2:04:17 AM
Analyte Peak Area: 0
Analyte Ion Transition: 373.2/91.0 amu



Sample Name: ARA-09-15-10-147-0004A1
Data File: ARA-09-15-10 Q15.wiff
Sample Index: 33
Acquired: 6/11/2010 2:04:17 AM
Analyte Peak Area: 0
Analyte Ion Transition: 343.1/270.9 amu

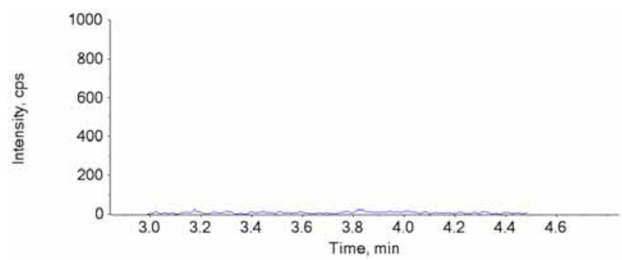
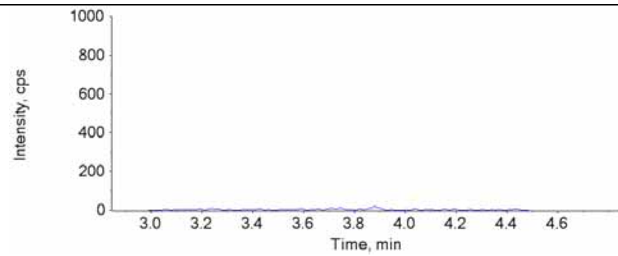
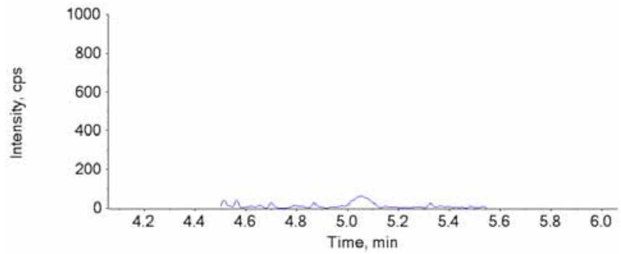


Figure 154. Typical Chromatogram of Quizalofop Analysis, Treated Corn Oil, Dry Processed (147-0004)

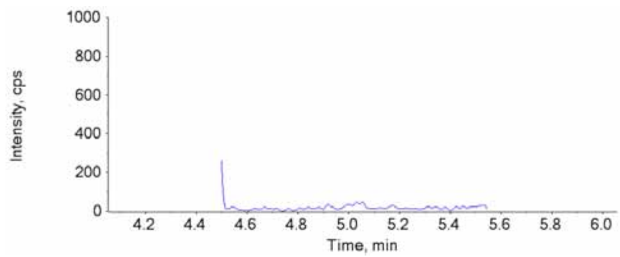
Sample Name: ARA-09-15-10-142-0001A9
Data File: ARA-09-15-10 Q15.wiff
Sample Index: 4
Acquired: 6/10/2010 8:02:33 PM
Analyte Peak Area: 0
Analyte Ion Transition: 343.1/108.1 amu



Sample Name: ARA-09-15-10-142-0001A9
Data File: ARA-09-15-10 Q15.wiff
Sample Index: 4
Acquired: 6/10/2010 8:02:33 PM
Analyte Peak Area: 0
Analyte Ion Transition: 373.2/299.0 amu



Sample Name: ARA-09-15-10-142-0001A9
Data File: ARA-09-15-10 Q15.wiff
Sample Index: 4
Acquired: 6/10/2010 8:02:33 PM
Analyte Peak Area: 0
Analyte Ion Transition: 373.2/91.0 amu



Sample Name: ARA-09-15-10-142-0001A9
Data File: ARA-09-15-10 Q15.wiff
Sample Index: 4
Acquired: 6/10/2010 8:02:33 PM
Analyte Peak Area: 0
Analyte Ion Transition: 343.1/270.9 amu

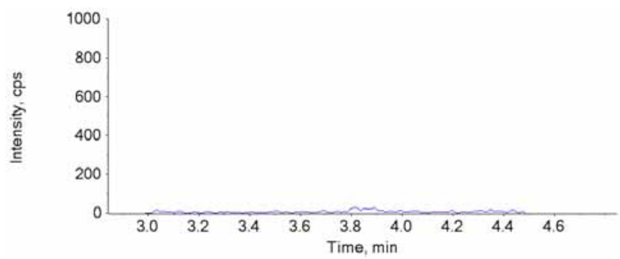
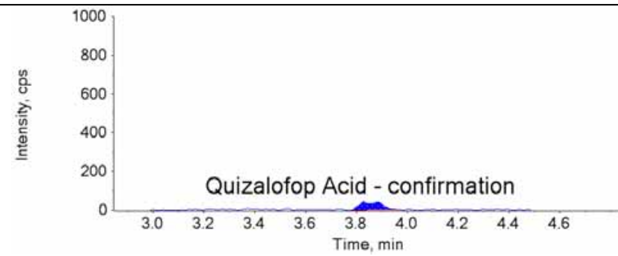
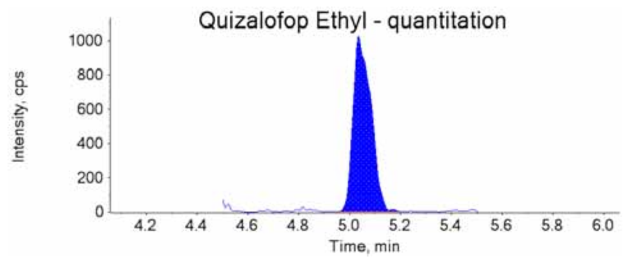


Figure 155. Typical Chromatogram of Quizalofop Analysis, Control Corn Starch (142-0001)

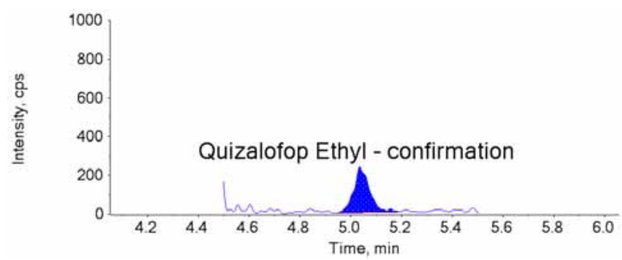
Sample Name: ARA-09-15-10-142-0001A11 + 0.01 µg/g
Data File: ARA-09-15-10 Q15.wiff
Sample Index: 16
Acquired: 6/10/2010 10:32:17 PM
Analyte Peak Area: 245
Analyte Ion Transition: 343.1/108.1 amu



Sample Name: ARA-09-15-10-142-0001A11 + 0.01 µg/g
Data File: ARA-09-15-10 Q15.wiff
Sample Index: 16
Acquired: 6/10/2010 10:32:17 PM
Analyte Peak Area: 5012
Analyte Ion Transition: 373.2/299.0 amu



Sample Name: ARA-09-15-10-142-0001A11 + 0.01 µg/g
Data File: ARA-09-15-10 Q15.wiff
Sample Index: 16
Acquired: 6/10/2010 10:32:17 PM
Analyte Peak Area: 1025
Analyte Ion Transition: 373.2/91.0 amu



Sample Name: ARA-09-15-10-142-0001A11 + 0.01 µg/g
Data File: ARA-09-15-10 Q15.wiff
Sample Index: 16
Acquired: 6/10/2010 10:32:17 PM
Analyte Peak Area: 2773
Analyte Ion Transition: 343.1/270.9 amu

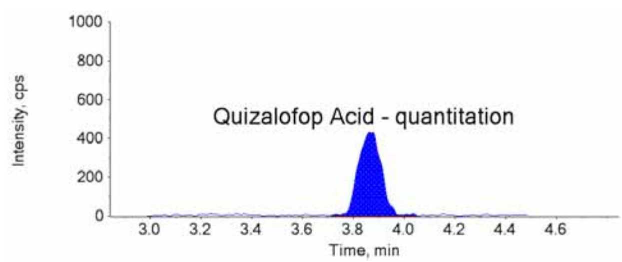
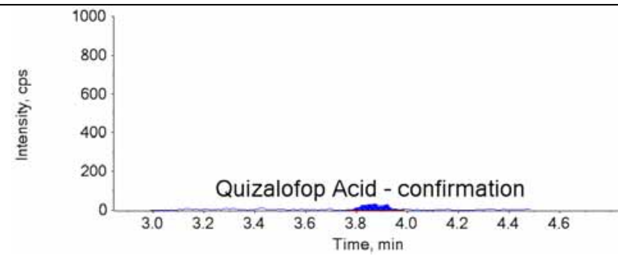
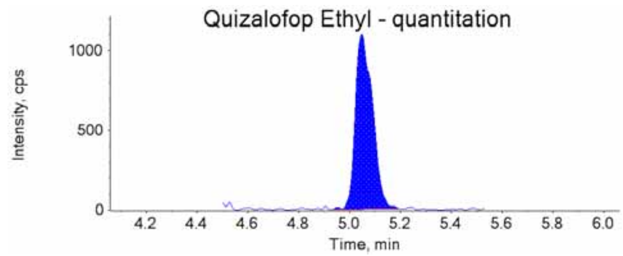


Figure 156. Typical Chromatogram of Quizalofop Analysis, Corn Starch 0.01 µg/g Recovery

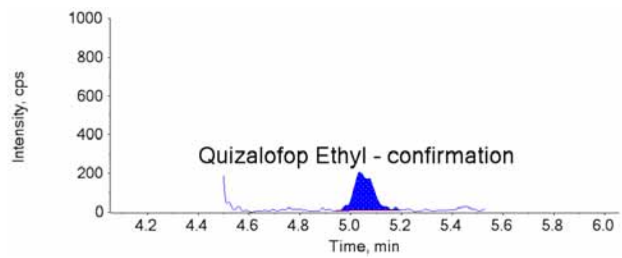
Sample Name: ARA-09-15-10-142-0001A12 + 0.01 µg/g
Data File: ARA-09-15-10 Q15.wiff
Sample Index: 17
Acquired: 6/10/2010 10:44:45 PM
Analyte Peak Area: 223
Analyte Ion Transition: 343.1/108.1 amu



Sample Name: ARA-09-15-10-142-0001A12 + 0.01 µg/g
Data File: ARA-09-15-10 Q15.wiff
Sample Index: 17
Acquired: 6/10/2010 10:44:45 PM
Analyte Peak Area: 5190
Analyte Ion Transition: 373.2/299.0 amu



Sample Name: ARA-09-15-10-142-0001A12 + 0.01 µg/g
Data File: ARA-09-15-10 Q15.wiff
Sample Index: 17
Acquired: 6/10/2010 10:44:45 PM
Analyte Peak Area: 1042
Analyte Ion Transition: 373.2/91.0 amu



Sample Name: ARA-09-15-10-142-0001A12 + 0.01 µg/g
Data File: ARA-09-15-10 Q15.wiff
Sample Index: 17
Acquired: 6/10/2010 10:44:45 PM
Analyte Peak Area: 2846
Analyte Ion Transition: 343.1/270.9 amu

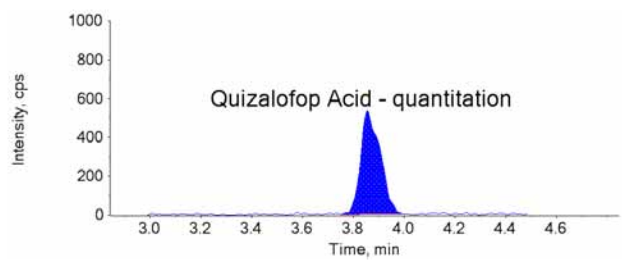
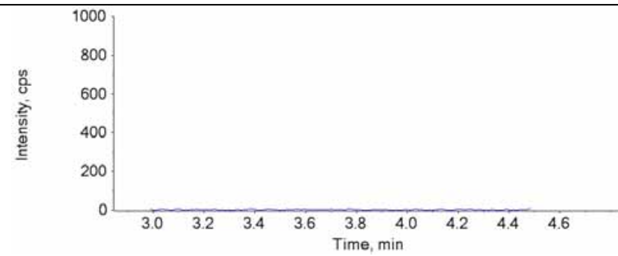
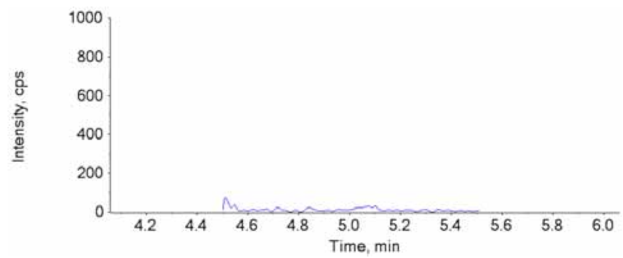


Figure 157. Typical Chromatogram of Quizalofop Analysis, Corn Starch 0.01 µg/g Recovery

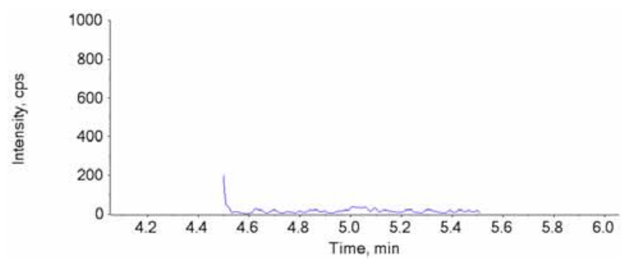
Sample Name: ARA-09-15-10-142-0003A1
Data File: ARA-09-15-10 Q15.wiff
Sample Index: 39
Acquired: 6/11/2010 3:19:05 AM
Analyte Peak Area: 0
Analyte Ion Transition: 343.1/108.1 amu



Sample Name: ARA-09-15-10-142-0003A1
Data File: ARA-09-15-10 Q15.wiff
Sample Index: 39
Acquired: 6/11/2010 3:19:05 AM
Analyte Peak Area: 0
Analyte Ion Transition: 373.2/299.0 amu



Sample Name: ARA-09-15-10-142-0003A1
Data File: ARA-09-15-10 Q15.wiff
Sample Index: 39
Acquired: 6/11/2010 3:19:05 AM
Analyte Peak Area: 0
Analyte Ion Transition: 373.2/91.0 amu



Sample Name: ARA-09-15-10-142-0003A1
Data File: ARA-09-15-10 Q15.wiff
Sample Index: 39
Acquired: 6/11/2010 3:19:05 AM
Analyte Peak Area: 0
Analyte Ion Transition: 343.1/270.9 amu

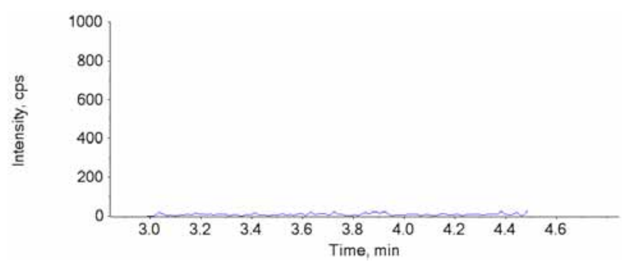
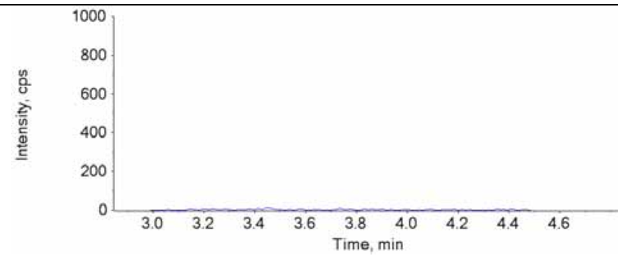
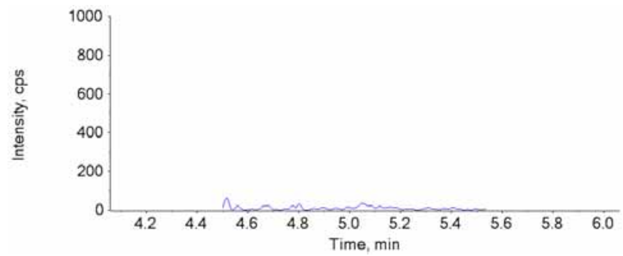


Figure 158. Typical Chromatogram of Quizalofop Analysis, Treated Corn Starch (142-0003)

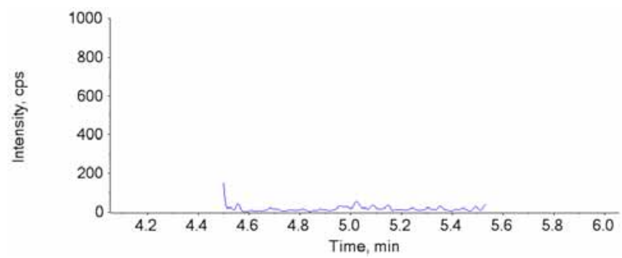
Sample Name: ARA-09-15-10-142-0004A1
Data File: ARA-09-15-10 Q15.wiff
Sample Index: 40
Acquired: 6/11/2010 3:31:34 AM
Analyte Peak Area: 0
Analyte Ion Transition: 343.1/108.1 amu



Sample Name: ARA-09-15-10-142-0004A1
Data File: ARA-09-15-10 Q15.wiff
Sample Index: 40
Acquired: 6/11/2010 3:31:34 AM
Analyte Peak Area: 0
Analyte Ion Transition: 373.2/299.0 amu



Sample Name: ARA-09-15-10-142-0004A1
Data File: ARA-09-15-10 Q15.wiff
Sample Index: 40
Acquired: 6/11/2010 3:31:34 AM
Analyte Peak Area: 0
Analyte Ion Transition: 373.2/91.0 amu



Sample Name: ARA-09-15-10-142-0004A1
Data File: ARA-09-15-10 Q15.wiff
Sample Index: 40
Acquired: 6/11/2010 3:31:34 AM
Analyte Peak Area: 0
Analyte Ion Transition: 343.1/270.9 amu

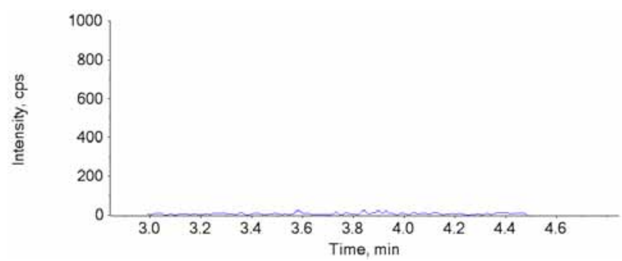


Figure 159. Typical Chromatogram of Quizalofop Analysis, Treated Corn Starch (142-0004)

Calibration Data

Linear with 1/x Weighting

Slope =	0.1079
Intercept =	0.0068
r^2 =	0.9995

Standard Concentration (ng/mL)	Injection Number	Analyte Peak Area	ISTD Peak Area	Quantitation Ratio	Response Factor	Calculated Concentration	Percent of Theoretical
0.15	1	6615	306962	0.022	0.1437	0.13657	91
0.5	10	17499	301735	0.058	0.1160	0.47444	95
1	19	37126	304093	0.122	0.1221	1.06863	107
5	28	167751	295605	0.567	0.1135	5.19777	104
10	37	348435	314033	1.110	0.1110	10.22311	102
20	46	680682	304498	2.235	0.1118	20.66078	103
35	55	1103669	292476	3.774	0.1078	34.92020	100
50	62	1544024	291938	5.289	0.1058	48.96849	98

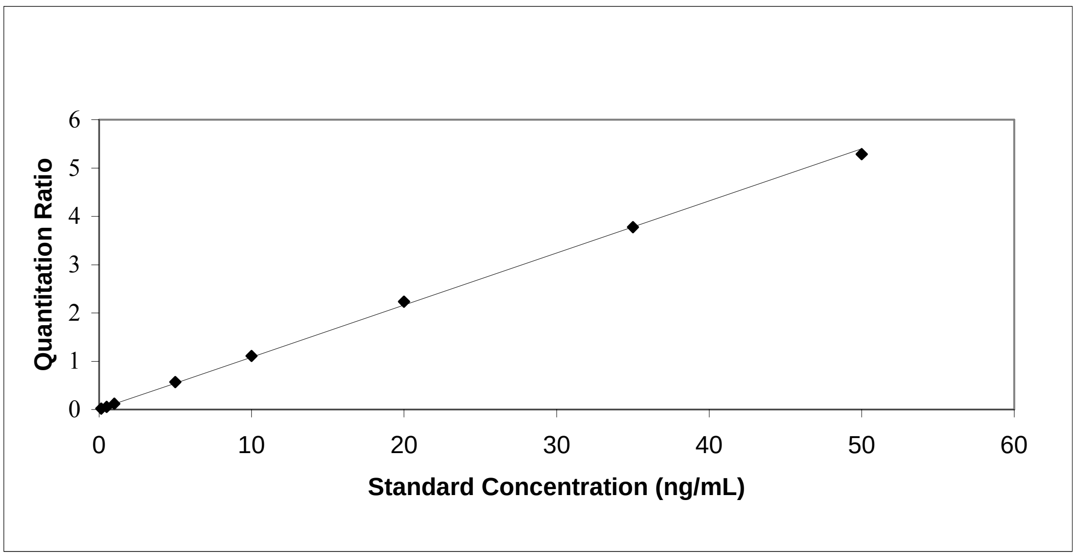


Figure 160. Typical Calibration Curve for the Quantitative Determination of 2,4-D in Corn from Set ARA-09-15-10 D16

Calibration Data

Linear with 1/x Weighting

Slope =	0.7101
Intercept =	0.0450
r^2 =	0.9994

Standard Concentration (ng/mL)	Injection Number	Analyte Peak Area	ISTD Peak Area	Quantitation Ratio	Response Factor	Calculated Concentration	Percent of Theoretical
0.15	1	5482	39182	0.140	0.9327	0.13371	89
0.5	10	15608	38207	0.409	0.8170	0.51198	102
1	19	29239	38261	0.764	0.7642	1.01290	101
5	28	137266	37557	3.655	0.7310	5.08385	102
10	37	273292	36222	7.545	0.7545	10.56223	106
20	46	533398	36792	14.498	0.7249	20.35382	102
35	55	887540	35507	24.996	0.7142	35.13897	100
50	62	1288660	37101	34.734	0.6947	48.85254	98

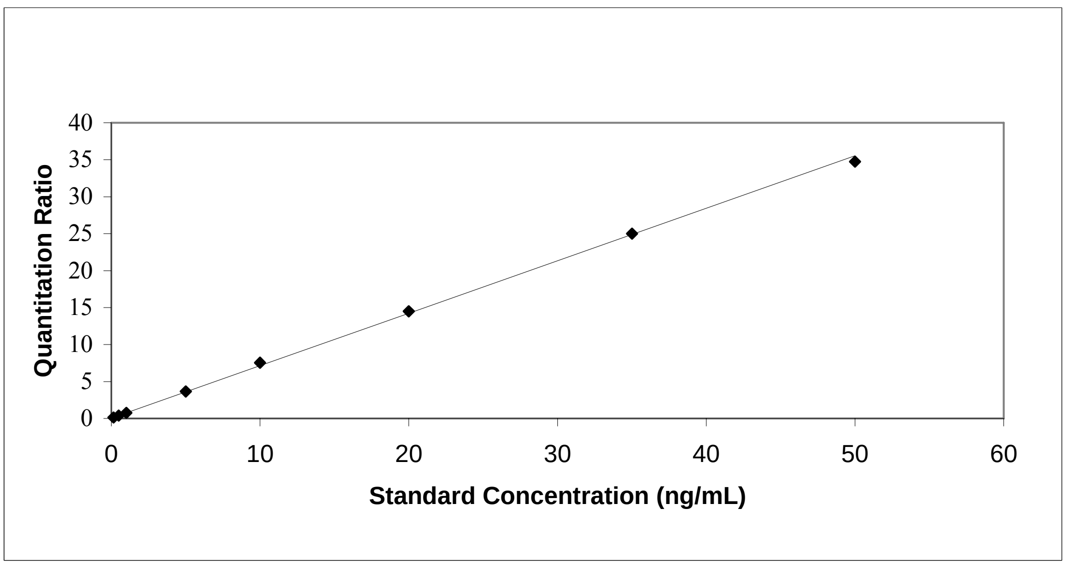


Figure 161. Typical Calibration Curve for the Quantitative Determination of 2,4-DCP in Corn from Set ARA-09-15-10 D16

Calibration Data

Linear with 1/x Weighting

Slope =	20558
Intercept =	27.6469
r^2 =	0.9990

Standard Concentration (ng/mL)	Injection Number	Analyte Peak Area	Response Factor	Calculated Concentration	Percent of Theoretical
0.1	1	1853	18530	0.08879	89
0.25	9	5520	22080	0.26716	107
5	24	103453	20691	5.03089	101
10	32	207320	20732	10.08327	101
20	40	437718	21886	21.29046	106
35	48	701880	20054	34.14002	98
50	55	1016611	20332	49.44940	99

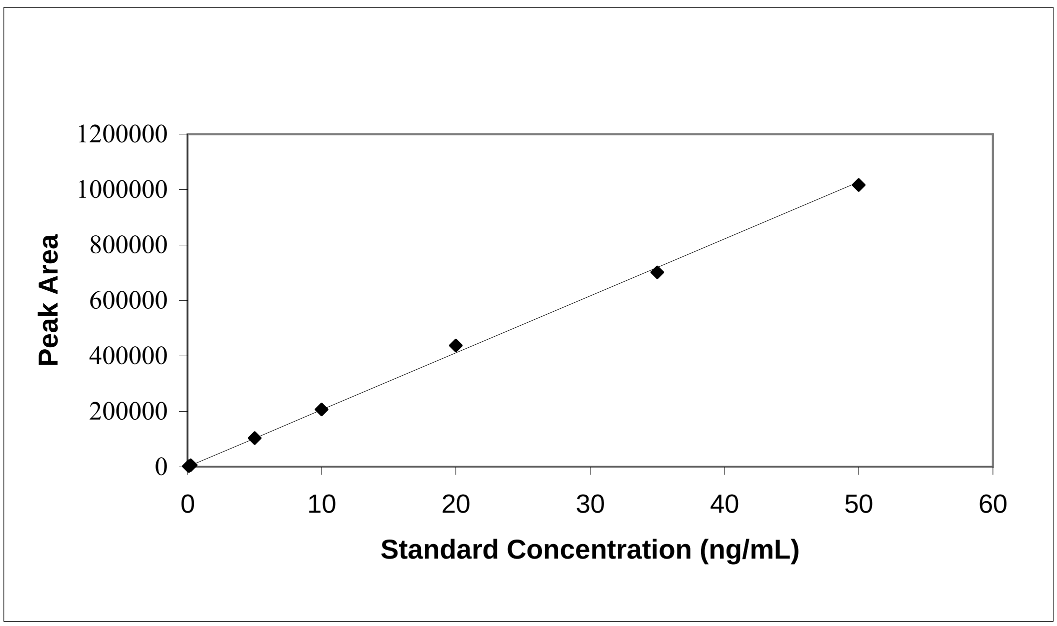


Figure 162. Typical Calibration Curve for the Quantitative Determination of Quizalofop Acid in Corn from Set ARA-09-15-10 Q02

Calibration Data

Linear with 1/x Weighting

Slope =	15158
Intercept =	264.4343
r^2 =	0.9997

Standard Concentration (ng/mL)	Injection Number	Analyte Peak Area	Response Factor	Calculated Concentration	Percent of Theoretical
0.1	1	1528	15280	0.08336	83
0.25	9	4529	18116	0.28134	113
5	24	78725	15745	5.17621	104
10	32	154598	15460	10.18171	102
20	40	303815	15191	20.02587	100
35	48	520700	14877	34.33423	98
50	55	762212	15244	50.26728	101

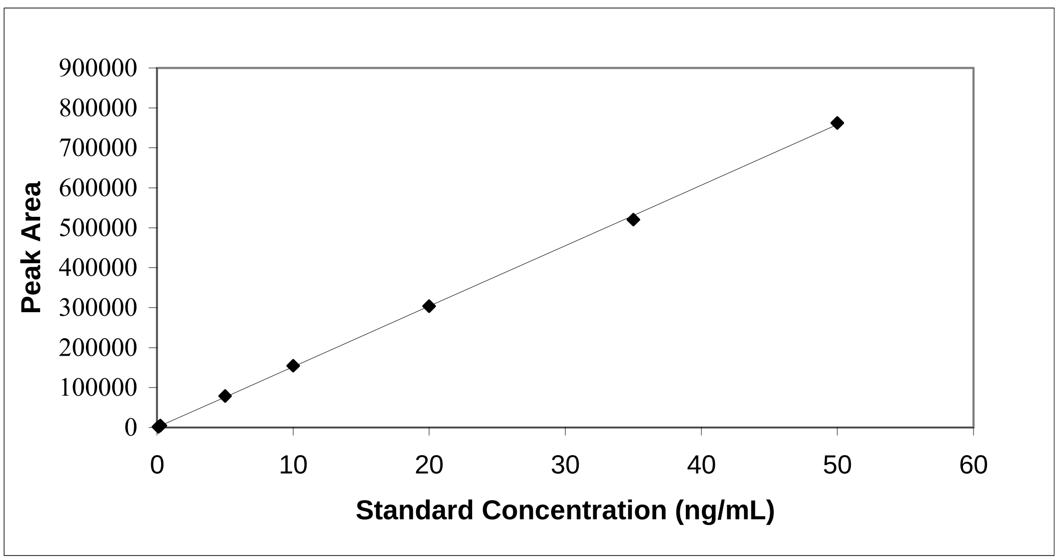


Figure 163. Typical Calibration Curve for the Quantitative Determination of Quizalofop-Ethyl in Corn from Set ARA-09-15-10 Q02

Linear with 1/x Weighting
Equations

```
              µg/g Found = ((((Area/IS Area)-Intercept)/Slope) x UC x DF x MF) - Control µg/g Found
        Uncorrected µg/g = ((((Area/IS Area)-Intercept)/Slope) x UC x DF x MF)
                    where:
                     Area = Analyte Peak Area
                  IS Area = Internal Standard Peak Area
                       UC = Unit Conversion = 0.001 (ng to µg)
                       DF = Dilution Factor
                       MF = Method Factor = (FV x EV)/(AF x NSW)
                       MF = (0.5 x 100)/(1 x 5) = 20 mL/g
                       FV = Final Volume = 0.5 mL
                       EV = Extraction Volume = 100 mL
                       AF = Aliquot Factor = 1 mL
                      NSW = Nominal Sample Weight = 5 g

        Percent Recovery = (µg/g Found/µg/g Added) x 100
```

Example Calculations

	Fortified Control Data	Field Sample Data
Set ID =	ARA-09-15-10 D16	ARA-09-15-10 D16
Analyte =	2,4-D - quantitation	2,4-D - quantitation
Sample ID =	ARA-09-15-10-011-0001A4 + 0.01 µg/g	ARA-09-15-10-139-0002A2
Injection Number =	58	45
Area =	19594	0
IS Area =	344637	380088
Intercept	0.0068	0.0068
Slope	0.1079	0.1079
DF =	1	1
µg/g Added =	0.01	Not Applicable
Control µg/g Found =	0	Not Applicable
Set Average % Recovery =	91	91

Fortified Control Example

```
              µg/g Found = ((((19594/344637) - 0.0068)/0.1079) x 0.001 x 1 x 20) - 0
                         = 0.0093

        Percent Recovery = (0.0093/0.01) x 100
                         = 93
```

Field Sample Example

```
        Uncorrected µg/g = ((((0/380088) - 0.0068)/0.1079) x 0.001 x 1 x 20)
                         = ND

           Reported µg/g = ND
```

Figure 164. Example Calculations for the Quantitative Determination of 2,4-D in Corn from Set ARA-09-15-10 D16

Linear with 1/x Weighting

Equations

```
          µg/g Found = (((Area-Intercept)/Slope) x UC x DF x MF) - Control µg/g Found
   Uncorrected µg/g = (((Area-Intercept)/Slope) x UC x DF x MF)
              where:
               Area = Analyte Peak Area
                 UC = Unit Conversion = 0.001 (ng to µg)
                 DF = Dilution Factor
                 MF = Method Factor = (FV x EV)/(AF x NSW)
                 MF = (1 x 100)/(0.5 x 5) = 40 mL/g
                 FV = Final Volume = 1 mL
                 EV = Extraction Volume = 100 mL
                 AF = Aliquot Factor = 0.5 mL
                NSW = Nominal Sample Weight = 5 g

   Percent Recovery = (µg/g Found/µg/g Added) x 100
```

Example Calculations

	Fortified Control Data	Field Sample Data
Set ID =	ARA-09-15-10 Q02	ARA-09-15-10 Q02
Analyte =	Quizalofop Acid - quantitation	Quizalofop Acid - quantitation
Sample ID =	ARA-09-15-10-020-0001A10 + 0.01 µg/g	ARA-09-15-10-020-0002A1
Injection Number =	5	26
Area =	5172	0
Intercept	27.6469	27.6469
Slope	20558	20558
DF =	1	1
µg/g Added =	0.01	Not Applicable
Control µg/g Found =	0	Not Applicable
Set Average % Recovery =	100	100

Fortified Control Example

```
         µg/g Found = (((5172 - 27.6469)/20558) x 0.001 x 1 x 40) - 0
                    = 0.01

   Percent Recovery = (0.0100/0.01) x 100
                    = 100
```

Field Sample Example

```
   Uncorrected µg/g = (((0 - 27.6469)/20558) x 0.001 x 1 x 40)
                    = ND

      Reported µg/g = ND
```

Figure 165. Example Calculations for the Quantitative Determination of Quizalofop Acid in Corn from Set ARA-09-15-10 Q02

Appendix D – Protocol and Revisions

Ag Research Associates

1730 Denham Road
Sycamore, GA 31790

Study Title: **Magnitude of the Residue of 2,4-D and Quizalofop-P-ethyl in/on Herbicide Tolerant Field Corn Containing the Aryloxyalkanoate Dioxygenase-1 (AAD-1) Gene**

Study Number: **ARA-09-15-10**

Test Guidelines: OPPTS 860.1500 Crop Field Trials with Decline
OPPTS 860.1520 Processed Feed/Food
PMRA Directive 98-02

Sponsor Company: Dow AgroSciences, LLC
9330 Zionsville Road
Indianapolis, Indiana 46268

DAS Study ID: 090052

Proposed Study Dates:
Experimental Start: May, 2009
Experimental Termination: January, 2010
Study Completion: May, 2010

Quality Assurance Review: Gene Burnett Date: 17 May 2009
Gene Burnett
Ag Research Associates

Approval Signatures:

Sponsor: D. Fonseca Date: May 15, 2009
Diego Fonseca
Dow AgroSciences

Sponsor Study Monitor: Steve W. Rosser Date: 15-May-2009
Steve W. Rosser
Dow AgroSciences

Study Director: John M. Becker Date: 19 May 09
John M. Becker
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Analysis Facility: Dow AgroSciences, LLC
9330 Zionsville Road
Indianapolis, IN 46268

1. INTRODUCTION

The purpose of this study is to determine the magnitude of residues of 2,4-D and quizalofop-P-ethyl herbicides in/on field corn raw agricultural commodities (forage, grain, and stover) and processed fractions (aspirated grain fraction, starch, meal, grits, flour, refined oil from wet and dry process) when a transgenic hybrid corn line containing the inserted *AAD-1* gene, a genetic modification added to increase tolerance to certain herbicides, is treated with commercially available 2,4-D and quizalofop-P-ethyl herbicides. The samples of raw agricultural commodities and processed fractions produced in this study may also be analyzed for selected metabolites of 2,4-D or quizalofop-P-ethyl which will be specified in an amendment to the protocol that will provide details on the analytical phase of the study.

The data to be generated are based upon the requirements of the USEPA Pesticide Assessment Guideline OPPTS 860.1500 (**Reference 1**), OPPTS 860.1520 Processed Feed/Food (**Reference 2**), and Canadian Pesticide Management Regulatory Agency Directive 98-02, Section 9 (**Reference 3**). This study will be conducted according to USEPA FIFRA Good Laboratory Practice Standards, Title 40 Code of Federal Regulations Part 160 (**Reference 4**). Exceptions to GLP compliance will be documented in the raw data and listed in the GLP compliance statement of the final study report.

The active ingredients in the test substances are 2,4-D and quizalofop-P-ethyl. 2,4-D [Weedar 64, a soluble liquid concentrate end-use product containing nominally 38.9% (w/w) acid equivalent (AE) of 2,4-dichlorophenoxyacetic acid, dimethylamine salt] is a broad-spectrum herbicide for the control of broadleaf weed in a variety of crops. Quizalofop-P-ethyl [Assure II, an emulsifiable concentrate end-use product containing nominally 10.3% (w/w) of ethyl(R)-2-[4-(6-chloroquinoxalin-2-yl oxy)- phenoxy] propionate] is a broad-spectrum herbicide for the control of grasses in a variety of crops.

The test system will be a transgenic hybrid corn line containing the inserted *AAD-1* gene planted in plots that will be treated with 2,4-D and quizalofop-P-ethyl. The *AAD-1* gene expresses the aryloxyalkanoate dioxygenase-1 (AAD-1) protein which provides increased tolerance of corn to 2,4-D and quizalofop-P-ethyl herbicides. There are two Events of AAD-1 that are to be evaluated in separate treated plots in this study. One treated plot is to be planted with a hybrid corn line containing AAD-1 Event DAS-40278-9 and the other treated plot is to be planted with a hybrid corn line containing AAD-1 Event DAS-40474-7. Due to a limited supply of transgenic seed, the untreated control plots are to be planted with a hybrid that has a genetic background representative of the transgenic lines used in the treated plots, but does not have gene to express the AAD-1 protein.

Field trials will be managed by Ag Research Associates (ARA) at twenty five (25) locations: two in Region 1 (New Jersey and Pennsylvania), two in Region 2 (Georgia), seventeen in Region 5 (Missouri, Nebraska, Kansas, Iowa, Illinois, Indiana, Ohio, Michigan, Minnesota), two in Region 6 (Oklahoma and Texas), and two in Canada Region 5 (Ontario). There will be 20 trials to collect the RAC samples, 3 trials to collect RAC samples at five sampling intervals after the last application (RAC + Decline trials), and two trials to collect RAC samples and bulk grain for food/feed processing (RAC + Processing trials). Each RAC and RAC + Decline trial location will consist of one untreated plot and two treated plots, one treated plot for each AAD-1 Event. The Processing trials will have 5 plots; one untreated plot to collect RAC and bulk samples, two 1X-rate treated plots to collect RAC samples, and two 2X-rate treated plots to collect the bulk grain sample for processing. Each treated plot will receive three (3) applications of the 2,4-D test substance and one (1) application of the quizalofop-P-ethyl test substance with each application occurring separately at a specified growth stage.

The analytical phase will be conducted in the Sponsor's analytical facility. Further details on the analytical phase of the study, including analytical method and analytes, will be added in an amendment to the protocol.

2. OBJECTIVES

- The purpose of this study is to determine the magnitude of residues of 2,4-D and quizalofop-P-ethyl, and any specified metabolites in/on field corn forage, grain, and stover, and processed fractions when herbicide tolerant transgenic hybrid lines of field corn containing either AAD-1 Event DAS-40278-9 or AAD-1 Event DAS-40474-7 are treated with the test substances.
- This study will determine residues in the RACs, residue decline at intervals after the last treatment, and residues in processed feed/food commodities.
- The study will be conducted to meet data requirements for crop residue studies outlined in USEPA OPPTS Guidelines 860.1500, Crop Field Trials **(Reference 1)** and OPPTS 860.1520 Processed Feed/Food (**Reference 2**), and Pesticide Management Regulatory Agency Directive 98-02 (**Reference 3**), according to USEPA FIFRA GLPS (**Reference 4**).

3. TEST SYSTEM

Crop: Field corn

Variety: Two transgenic hybrid lines of field corn each containing a different Event of the inserted *AAD-1* gene will be used as the test system with each hybrid line planted in separate treated plots. The *AAD-1* gene expresses the aryloxyalkanoate dioxygenase-1 (AAD-1) protein which provides increased tolerance of corn to 2,4-D and quizalofop-P-ethyl herbicides. One treated plot is to be planted with a hybrid corn line containing AAD-1 Event DAS-40278-9 and the other treated plot is to be planted with a hybrid corn line containing AAD-1 Event DAS-40474-7. Due to a limited supply of transgenic seed, the untreated control (unsprayed) plots are to be planted with a hybrid that has a genetic background representative of the transgenic lines used in the treated plots, but does not have genes to express the AAD-1 protein.

Documentation of the seed identification and characterization for both the seed used in untreated control and treated plots will be included in the study file.

RAC Crop commodities: Field corn forage, grain, and stover

Processed feed/food commodities: Field corn aspirated grain fraction, starch, refined oil (wet milling), meal, grits, flour, and refined oil (dry milling)

Justification: This test system was selected to establish the level of residues in a commercially viable transgenic herbicide tolerant line of field corn in order to meet requirements in OPPTS 860.1500.

4. TEST SUBSTANCES

One test substance will be Weedar 64®, a commercially available product containing 2,4-D as the active ingredient formulated as a soluble concentrate (SL) of 2,4-dichlorophenoxyacetate, dimethylamine salt.

The second test substance will be Assure II®, a commercially available product containing quizalofop-P-ethyl [ethyl(R)-2-[4-(6-chloroquinoxalin-2-yl oxy)- phenoxy] propionate] as the active ingredient.

An archived sample(s) of each test substance shall be retained by the Sponsor.

Active Ingredient Common Name:	2,4-D
Active Ingredient Chemical Name:	2,4-Dichlorophenoxyacetic acid, dimethylamine salt
CAS Number:	94-75-7
Product Name:	Weedar 64
Concentration of AE [a, b]	454 g AE/L (acid equivalent)
Percent of AE [a, b]:	39.1% (w/w) acid equivalent (AE)
Formulation density:	1.1601 g/mL at 20°C
Formulation Type:	SL (Soluble Liquid Concentrate)
Sponsor TSN Number:	TSN026491-0010
Lot Number:	ILG-C-004
Re-certification Date:	12-Feb-2011

Active Ingredient Common Name:	Quizalofop-P-ethyl
Active Ingredient Chemical Name:	Ethyl(R)-2-[4-(6-chloroquinoxalin-2-yl oxy)-phenoxy] propionate
CAS Number:	76578-14-8
Product Name:	Assure II Herbicide
Concentration of AI [b, c]:	105 g/L (a.i., active ingredient)
Percent of AI [b, c]:	10.3% (w/w), a.i.
Formulation density:	1.0223 g/mL at 20°C
Formulation Type:	EC (Emulsifiable Concentrate)
Sponsor TSN Number:	TSN020252-0004
Lot Number:	APR06YZ059
Re-certification Date:	05-Mar-2011

(a) **Application rate for 2,4-D is based on the acid equivalent (AE) concentration of 2,4-D** rather than on the concentration of the a.i., 2,4-D, dimethylamine salt

(b) Details of the test material purity assay will be included in the study file.

(c) **Application rate for quizalofop-P-ethyl is based on the active ingredient (AI) concentration**, not on the acid equivalent (AE)

4.1. **Characterization of Test Substances and Analytical Standard(s):**

Characterization of the test substances, as required by USEPA FIFRA GLPS, 40 CFR 160.105(a), was conducted by the Sponsor and these records as well as samples of the test substances and analytical standards are being retained by the Sponsor.

4.2. **Retention of Test Substance Container(s):**

The test substance container(s) must be retained for the duration of the study [40 CFR Part 160.105 (c)]. After all applications are made, the test substance container(s) will be returned to the Sponsor (refer to Section 7.5). Any remaining test substance should be discarded in a responsible manner according to local regulations, and the container triple rinsed before returning to the Sponsor. Records will be maintained in the field trial notebook documenting the use, transfer, if any, disposition of test substance, and return of the container. Documentation will include the date and weight or volume of test substance received and withdrawn at each distribution or disposal.

4.3. **Storage of Test Substances:**

The test substances will be stored in accordance with the Certificate of Analysis or the instructions on the substance container label. During the treatment period, the test substances will be stored in a location where temperatures do not exceed 50°C (~120° F) and are maintained above 0°C. A record of test substance storage conditions will be maintained at least from the period of receipt until final application.

4.4 **Stability of Test Substances:**

Test substance application solutions will not be retained for more than 8 hours to minimize concerns about stability in the spray tank. Stability of the test substances shall be determined by the Sponsor as required by the USEPA FIFRA GLPs, 40

CFR 160.105(e). The Sponsor will also conduct any re-certification needed before the completion of the study.

4.5. **Safe Handling:**

A Material Safety Data Sheet (MSDS) and/or other available information necessary for proper and safe handling, shipping, and storage of the test substances will be made available to study personnel. Facility SOPs regarding personal protective equipment (PPE) and handling research test materials will be followed.

5. REGULATORY COMPLIANCE

The transgenic field hybrid corn lines to be used in the treated plots are considered regulated articles by the USDA and CFIA. The trials and plant materials generated from them are to be managed in a manner that complies with USDA or CFIA requirements.

There is a minimum of 660 feet physical isolation distance (200 meters for Canada sites) required from the regulated trial to other corn, except that regulated Dow AgroSciences (DAS) corn may be planted in a separate trial within 660 feet (200 meters in Canada) at the same site. In the U.S., there is also a requirement of a 10 ft. bare soil buffer around the perimeter of the trial. In addition, the trial will be surrounded by a minimum of 6 rows (or 15 ft.) of a commercially available corn hybrid of similar relative maturity. The border will be continuous around the outside perimeter of the trial. The planted buffer provides some animal control, but an electrified fence is suggested in areas likely to have animals such as deer and raccoons.

6. FIELD PHASE

6.1. **Field Trial Notebook**:

A field trial notebook containing guidance and forms for recording raw data and documenting pertinent events during the conduct of the field phase will be provided by ARA. Please read the protocol and become familiar with the forms in the ARA field trial notebook.

The field trial notebook will be maintained as the one thorough, original record-keeping document for each field trial. Relevant sections and data for the field trial notebook will be completed, or notation made as to why data are not available or relevant. If other forms are needed and used to supplement the forms supplied within the ARA field trial notebook, the forms on which the original observations were recorded must be added into the field trial notebook as the original raw data. The field notebook and all original data will be documented and maintained according to GLPs and field facility SOPs. At the conclusion of the field phase of the study, the Principal Field Investigators (PFIs) will complete the field trial notebook and submit the notebook to the Study Director's Testing Facility (ARA).

If you have any questions regarding the protocol or any of the forms in the field trial notebook, please do not hesitate to contact the Study Director. If the Study Director is unavailable, call the Study Director Management, David Stephens on (229) 821-0689 (mobile).

6.2. Test Site Locations:

The Study Director and Sponsor identified appropriate test sites and located Principal Field Investigators (PFIs) in the States and Regions given in the table below in which field corn is commonly grown. The test site location will be documented in the field trial notebook.

Field Trial Number	Field Investigator	Research Company	NAFTA Region	State / Prov.	County	Trial Type
NJ-01	Dan Ramsdell	Crop Management Strategies	1	NJ	Hunterdon	RAC
PA-02	Dan Ramsdell	Crop Management Strategies	1	PA	Lehigh	RAC
GA-03	Chris Cromer	Ag Research Associates	2	GA	Tift	RAC + Processing[a]
GA-04	Chris Cromer	Ag Research Associates	2	GA	Turner	RAC + Decline[b]
IL-05	Sue Dorsey	SGS Alvey Ag Research	5	IL	Bureau	RAC
IL-06	Tim Boeker	SGS Alvey Ag Research	5	IL	Clinton	RAC
IN-07	John Bailey	SGS Alvey Ag Research	5	IN	Parke	RAC
MO-08	Matt Cordell	Ag Research Associates	5	MO	Jasper	RAC
MO-09	Matt Cordell	Ag Research Associates	5	MO	Henry	RAC
IL-10	Matt Cordell	Ag Research Associates	5	IL	Schuyler	RAC
MI-11	Chad Harris	Ag Research Associates	5	MI	Lenawee	RAC + Decline[b]
OH-12	Chad Harris	Ag Research Associates	5	OH	Fulton	RAC
IN-13	Chad Harris	Ag Research Associates	5	IN	Lagrange	RAC
NE-14	Matt Krause	Ag Research Associates	5	NE	York	RAC + Processing[a]
NE-15	Denny Stamm	Ag Research Associates	5	NE	Polk	RAC
KS-16	Denny Stamm	Ag Research Associates	5	KS	Republic	RAC
IA-17	David Bennett	Bennett Agricultural Research	5	IA	Jefferson	RAC
MO-18	David Bennett	Bennett Agricultural Research	5	MO	Adair	RAC
IA-19	Kyle Johnson	Eurofins Agroscience Services	5	IA	Winneshiek	RAC
MN-20	Kyle Johnson	Eurofins Agroscience Services	5	MN	Fillmore	RAC
MO-21	Nathan Goldschmidt	Shoffner Farm Research	5	MO	Stoddard	RAC
OK-22	Tim Case	Ag Research Associates	6	OK	Marshall	RAC + Decline[b]
TX-23	Tim Case	Ag Research Associates	6	TX	Denton	RAC
ON-24	Jamie Parnell	Vaughn Ag Research Services	5	Ontario	Branchton[c]	RAC
ON-25	Jamie Parnell	Vaughn Ag Research Services	5	Ontario	Dutton[c]	RAC

(a) The Processing trial will require sufficient space to allow large untreated and 2X-rate treated plots to collect the bulk grain samples.
(b) The Decline trial will require sufficient space to allow at least 5 sampling intervals for all three commodities plus an additional interval for corn forage.
(c) Town names given for Canada sites rather than County name.

6.3. Test Site Layout, Isolation Buffers, and Preparation:

The PFI should select a test site that is relatively uniform in soil characteristics, well-drained, have proper levels of nutrients, and receive agronomic management, such as fertilizer, pesticides, and irrigation (as needed) to the entire test site, typical for the location to produce a commercial crop. The test site must be large enough to accommodate the test plots described in Section 6.5. The plot area will be large enough to allow the test substance to be applied accurately and in a manner that represents a commercial application technique (broadcast spray with a boom sprayer).

Each RAC and RAC + Decline test site will have one untreated plot and two treated plots (one for a transgenic corn line containing AAD-1 Event DAS-40278-9 and the other for a corn line containing AAD-1 Event DAS-40474-7). The RAC + Processing trials will require 5 plots, an untreated plot (large enough to collect the RAC samples and a bulk grain sample), two 1X-rate treated plots (for RAC samples with one plot for each of the two Events, as described above), and two 2X-rate treated plots, again with one plot for each of the two Events, as described above. The untreated plot and the two 2X-treated plots need to be large enough to yield the bulk treated grain sample for processing. The treated and untreated plots should be clearly identified by labeled stakes or other means which contain a unique identifier (such as trial number, plot designation, and seed identification).

All **treated plot(s)** in RAC, RAC + Decline and RAC + Processing sites will be planted with transgenic lines of corn containing the *AAD-1* gene, with one plot being planted with a line containing AAD-1 Event DAS-40278-9 and the other plot being planted to a line containing AAD-1 Event DAS-40474-7. **The untreated control (unsprayed) plot at all sites (RAC, RAC + Decline and RAC + Processing) are to be planted with a non-transgenic hybrid** that has a genetic background representative of the transgenic lines used in the treated plots, but does not have genes to express the AAD-1 protein.

As indicated in Section 6.4, buffer zones between plots are required to avoid cross-contamination due to spray drift or run-off. There will be an untreated **buffer zone** of at least 50 feet between the untreated and treated plots (including plots to which 2,4-D or quizalofop-P-ethyl is applied in an adjacent trial). Also, there is to be a minimum buffer zone of at least 25 ft. between treated plots. Additionally, the untreated plot should be located upslope and/or upwind (for the prevailing wind) from the treated plot and have little slope to minimize the potential for contamination.

As described in Section 5, to meet USDA or CRIA requirements, there is a minimum of 660 feet physical isolation distance (200 meters for Canada sites) required from the regulated trial to other corn, except other regulated DAS corn that may be used in a separate trial at the same site. In the U.S., there is also a requirement of a 10 ft. bare soil buffer around the perimeter of the trial. In addition, the trial will be surrounded by a minimum of 6 rows (or 15 ft.) of a commercially available corn hybrid of similar relative maturity. The border will be continuous around the outside perimeter of the trial.

Details of the test site, including accurate test site and plot diagrams, must be documented in the field trial notebook. The location of the trial site with distances from permanent markers, such as major roads, intersections, towns, and/or GPS coordinates will be documented. The location of the plots, with distances between plots, and separation buffers must be documented.

Records and identification of planted field corn seed will be documented in the field trial notebook. The method of soil preparation and equipment used will be recorded in the field trial notebook.

6.4. **Test Site History and Prevention of Contamination**:

The 3-year pesticide use history of the test area containing the research plots should be assured prior to planting of the transgenic corn seed and will be recorded in the field trial notebook. Do not locate plots where products containing 2,4-D or quizalofop-P-ethyl have been applied within the previous 18 months of the planting date.

Test sites will be located to avoid the possibility of inadvertent contamination of the test substances from an adjacent research trial, agricultural field, residential, or commercial area. As indicated in Section 6.3, buffer zones between plots are required to avoid cross-contamination due to spray drift or run-off. There will be an untreated **buffer zone** of at least 50 feet between the untreated and treated plots (including plots to which 2,4-D or quizalofop-P-ethyl is applied in an adjacent trial). Also, there is to be a minimum buffer zone of at least 25 ft. between treated plots. Additionally, the untreated plot should be located to minimize the potential for contamination from the treated plots (upslope and/or upwind depending on trial site).

6.5. **Test Plots**:

At each RAC-only and RAC + Decline test site there will be three plots; one untreated plot planted with non-transgenic seed (with genetic background similar to the transgenic corn lines, but lacking the inserted *AAD-1* gene), and two treated plots, each planted with a transgenic line of field corn seed (one plot planted with a line containing AAD-1 Event DAS-40278-9 and the other plot being planted with a line containing AAD-1 Event DAS-40474-7). At the Processing test site there will be five plots; one untreated plot planted with non-transgenic seed (sized to yield RAC and bulk samples), two 1X-rate treated plots for RAC samples only (one plot planted with a line containing AAD-1 Event DAS-40278-9 and the other plot being planted with a line containing AAD-1 Event DAS-40474-7), and two 2X-rate treated plots for use in producing a bulk seed sample for processing (one plot planted with a line containing AAD-1 Event DAS-40278-9 and the other plot being planted with a line containing AAD-1 Event DAS-40474-7). At the Processing sites the untreated control plot and each of the 2X rate treated plots need to be large enough to produce ~1000 lb bulk grain sample for use in processing.

The treated RAC plots should be of sufficient size to produce duplicate samples of the respective sample type. **Additionally, there is to be a forage sample collected at each of two timings in RAC plots and at an early timing and the 5 specified decline sample intervals in the RAC + Decline plots.** The untreated plot should be of sufficient size to produce a single sample at each sampling interval of all three commodities. The designated bulk seed plots will require only a single ~1000 lb sample of seed.

Minimum RAC individual sample requirements from the plots are as follows:

- Forage: minimum 1 kg from at least 12 plants from separate areas of the plot. Divide each stem with leaves attached into 3 approximately equal lengths.

Take top, middle and bottom portions, respectively, from each of three groups of four stems ensuring that parts of all 12 stems are included in the sample

- Grain: minimum 1 kg from at least 12 separate areas of the plot
- Stover: minimum 1 kg from at least 12 plants from separate areas of the plot. Stems divided and composited into a sample as described for forage.

The Decline trial will require additional plot area to allow five (5) collection intervals for each crop commodity from each plot. Corn forage is also to be sampled at an additional interval in addition to the five decline intervals.

The Processing trial will require sufficient plot area to yield single bulk grain samples (about 1000 pounds each) from both the untreated (T1 plot) and each 2X-rate treated plots (T4 and T5 plots).

The treated and untreated plots should be clearly identified by labeled stakes or other means which contain a unique identifier (such as trial number and plot designation).

6.6. **Receipt of Regulated Seed**:

The transgenic field corn seed lines for this study are considered regulated articles by the USDA and CFIA. Therefore, proper receipt, handling, working, and storage areas must be available in accordance to USDA or CFIA requirements. The work area should be easily isolated from non-regulated areas and afford easy clean-up of regulated material. **Confirm that documentation and identification** was received with the seed packets/containers and they match what is expected for this study (AAD-1 Event DAS-40278-9 and AAD-1 Event DAS-40474-7). Properly log the sample into the field trial notebook. When the plots are planted, **record the supplied seed identification information in the field trial notebook** to confirm the identity of the seed planted in each plot.

6.7. **Test Site and Crop Maintenance and Irrigation**:

Plots should be maintained in accordance with prevailing commercial agronomic practices for field corn, including tillage, maintenance pesticides, fertilizer, and irrigation as needed to produce a viable crop. As needed, the plots will be maintained relatively free of weeds using a registered herbicide (not containing 2,4-D or other phenoxy-type herbicides, or quizalofop-P-ethyl) at labeled rates. If needed to control insects or disease, the plots can be treated with a registered insecticide or fungicide at labeled rates. Any applications of maintenance chemicals will be made to all plots (untreated and treated). Tilling of rows, work alleys, buffers, and edges is acceptable. Perform all operations to the untreated plot first to avoid the potential of contamination. Document all procedures and equipment used to maintain the plots.

The need for irrigation and fertilizer will be evaluated on an on-going basis and applied as needed during the growing season to maintain a viable crop. When needed, irrigation should be applied at a rate to prevent flow of water from one plot to another in order to avoid the potential for cross-contamination. The date and amount of irrigation, if any, will be documented in the field trial notebook.

6.8. **Meteorological Monitoring**:

Weather data will be collected and documented in the field trial notebook. These data may be from an on-site weather station, or obtained from nearby government, academic, or commercial weather data sources. Weather data should be supplied as a **summary spreadsheet** including the parameters noted below. The meteorological raw data from an on-site station should be available as part of the facility records. Weather data from an off-site station need not be collected in accordance with GLPs.

- Daily air temperature (minimum, maximum)
- Daily rainfall and supplemental irrigation, if any
- Monthly average maximum and minimum temperature and monthly total rainfall compared to historical averages

The following data will be recorded in the field trial notebook **for each application**:

- Wind speed, at the time of application
- Wind direction, at the time of application
- Presence of soil surface moisture
- Presence of plant surface moisture (dew)

6.9. **Test Site Soil Information**:

Soil characterization (soil class or type name/designation), organic matter (%OM), and cation exchange capacity (CEC) for the test site area will be recorded in the field trial notebook. Characterization information can be obtained from a previously available USGS soil survey or characterization analysis, but if a characterization analysis is used, it should be from within the last 5 years. This information need not be generated in this study and does not have to be GLP-compliant.

7. TEST SUBSTANCE APPLICATION

There will be four (4) broadcast herbicide applications with a boom sprayer to each treated plot, two applications of 2,4-D, followed by one treatment of quizalofop-P-ethyl, followed by a final application of 2,4-D to simulate a worst-case commercial herbicide treatment. The Processing trials will have additional exaggerated rate (2X) treatments for each transgenic line (5 total plots).

Application No. 1 is to be an application of 2,4-D applied at pre-emergence. Application No. 4 (the last of the 4 applications) is to be an application of 2,4-D applied over-the-top to corn in approximately the V8 growth stage (or corn that is approximately 48 inches tall, whichever comes first). The timings for Application No. 2 (the second application of 2,4-D) and Application No. 3 (the application of quizalofop-P-ethyl) are to be scheduled relative to the date on which it is expected that the corn will be in approximately the V8 stage (or approximately 48 inches tall) and Application No. 4 is to be applied. **Application No. 2 (the second over-the-top application of 2,4-D) is to be**

applied at 12 days (± 1 day) before Application No. 4. Application No. 3 (the over-the-top application of quizalofop-P-ethyl) is to be applied at 5 days (±1 day) before Application No. 4. Monitor the growth and development of the crop closely in order to be able to predict at least 12 days in advance of the crop reaching the V8 stage or 48 inch height (whichever comes first) so that the proper timings for Application No. 2 and Application No. 3 can be carried out. Once the target date for Application No. 4 is determined, the schedule for Application Nos. 2 and 3 can be set relative to that date. The timings for Application No. 2 and Application No. 3 are being based on the expected timing for Application No. 4 rather than targeting specific growth stages at Application Nos. 2 and 3 in order to attain the desired intervals between applications. It is expected that the growth stage of the corn at the timing for Application No. 2 and Application No. 3 will vary somewhat depending on how quickly the corn is expected to grow over the approximate 12 day interval between Application No. 2 and Application No. 4. Under average to good growing conditions, it is thought that corn will likely be at approximately the V4 growth stage at Application No. 2 and at approximately the V6 growth stage at Application No. 3. However, the timing for Application No. 2 and Application No. 3 should be scheduled relative to the expected date for Application No. 4 rather than based on target growth stages. Record both the date and growth stage of the crop at each of the four applications in the Field Trial Notebook. All applications that take place following crop emergence are **to be foliar over-the-top applications. A non-ionic adjuvant, such as X-77, at approximately 0.25% (v/v) will be added in the spray mixture with each application of 2,4-D. A crop oil concentrate, such as Agri-Dex, at approximately 1.25% (v/v) will be added in the spray mixture with quizalofop-P-ethyl. Unexpected results or phytotoxic effects, along with application information, must be transmitted (e.g., phone call, e-mail) to the Study Director immediately after observation of such a field event.**

7.1. **Application Equipment and Calibration**:

The route of administration is a backpack, tractor-, or ATV-mounted boom sprayer with adequate agitation and output in order to simulate a typical commercial application. Spray pressure and nozzles should be selected to provide a uniform spray avoiding drift and misting. Spray nozzle type and pressure established at the beginning of each application will be recorded in the field notebook.

Application equipment will be calibrated no more than 2 days prior to the application event, preferably on the day of application. Calibration will be performed according to the Standard Operating Procedure (SOP) of the Field Investigator. Calibration activities, calculation of application rate, and spray delivery results will be recorded in the field trial notebook.

7.2 **Spray Volume, Adjuvant, and Mixing Instructions**:

Spray equipment should be thoroughly clean before mixing the test substance.

Use a spray volume equivalent to approximately 200 - 400 L/ha (~20 - 40 gallons/acre) for thorough coverage of the treated plot area. **Use a non-ionic surfactant (NIS), such as X-77 or equivalent, at approximately 0.25% (v/v) in the spray mixture with 2,4-D. For application of quizalofop-P-ethyl, use a crop oil concentrate (COC), such as Agri-Dex or equivalent, at approximately 1.25% (v/v) in the spray mixture.** Add the required amount of test substance, water, and non-ionic surfactant or crop oil concentrate for application of 2,4-D or quizalofop-P-ethyl, respectively, following Field Facility SOPs to prepare the tank mix solution. Use a sprayer with a mix tank that has continuous agitation or can be mixed manually by shaking to ensure thorough mixing. Use mechanical agitation for a minimum of 1 minute, or manually invert mix-tank at least 20 times to ensure full dispersal of the test substance. If manual mixing is necessary, repeat the mixing operation if more than 15 minutes elapses before application starts. Details of agitation type and the mixing time will be documented in the field trial notebook. Discard the application solution if it is not applied within 8 hours of mixing.

7.3 **Application Rate and Timing:**

For each treated plot, there will be a total of 4 (four) applications, as follows.

In this study the application rate for 2,4-D is expressed on an acid equivalent (a.e.) basis, so the target application rate is in units of lb a.e./A or g a.e./ha. The application rate for quizalofop-P-ethyl is expressed on an active ingredient (a.i.) basis, so the target application rate is in units of lb a.i./A or g a.i./ha. Note that in Section 4 the test substance information for 2,4-D presents test substance concentration on an a.e. basis and the test substance information for quizalofop-P-ethyl presents test substance concentration on an a.i. basis.

In the RAC and RAC + Decline treated plots there will be 2 broadcast spray applications of 2,4-D, each at approximately 1.0 pound acid equivalent/acre (~1120 g acid equivalent/ha), one application of quizalofop-P-ethyl at approximately 0.082 pound AI/acre (~92 g/ha), and one application of 2,4-D at approximately 1.0 pound acid equivalent/acre (~1120 g acid equivalent/ha). Each application should fall within the range of 95 -110% of the target rate.

For the 2X-rate Processing treated plots, there will be 2 broadcast spray applications of 2,4-D each at approximately 2.0 pound acid equivalent/acre (~2240 g acid equivalent/ha), one application of quizalofop-P-ethyl at approximately 0.164 pound AI/acre (~184 g/ha), and a final application of 2,4-D at approximately 2.0 pound acid equivalent/acre (~2240 g acid equivalent/ha). Again, each application should fall within the range of 95 -110% of the target rate.

The 4 applications will occur as follows:

- Application No. 1 will be 2,4-D with NIS applied at preemergence.
- Application No. 2 will be 2,4-D with NIS applied 12 days (± 1 day) **before Application No. 4.**
- Application No. 3 will be quizalofop-P-ethyl with COC applied 5 days (± 1 day) **before Application No. 4**.
- Application No. 4 will be 2,4-D with NIS applied to corn at approximately the V8 growth stage or when the corn is approximately 48 inches tall, whichever comes first.

A summary of Treatments and Applications is presented in the table in Section 7.4. Additional description of the four applications is also given near the beginning of Section 7. As indicated near the beginning of Section 7, **the growth and development of the crop will need to be monitored closely in order to be able to estimate at least 12 days in advance when the crop will reach the V8 stage or 48 inch height (whichever comes first) so that the proper timings for Application No. 2 and Application No. 3 can be carried out.** Once the target date for Application no. 4 is selected, the schedule for Application Nos. 2 and 3 can be set relative to that date (i.e. Application No. 2 applied 12 days (± 1 day) before Application No. 4 and Application No. 3 applied 5 days (± 1 day) before Application No. 4).

All applications are to be made broadcast with a boom sprayer and those that take place following crop emergence are to be foliar over-the-top applications. Adjust the sprayer nozzles to give a uniform application. Application of the test substance will be to the entire treated plot area and spray passes must not overlap. Do not use a single nozzle hand wand applicator. The growth stage of the crop is to be determined based on approximately 50 to 100% of the crop being at the specified stage. The Study Director must be notified if the application timing varies more than one growth stage (for 95% of crop) from the target stage. Detailed records of the spray mixture, application and environmental data (at application) will be documented in the field trial notebook. Tank mix samples are not required.

7.4 Treatment Schedule:

Plot Designation	Application Number	Test Substance	Test Substance (lb /A) a, b, c	Test Substance (g /ha) a, b, c	Target Application Timing d, e
Untreated Control – Non-transgenic corn Plot (T1)					
Treatment 1 Untreated	0	None	None	None	None
Herbicide Treated Corn Line AAD-1 Event DAS-40278-9 Plot (T2)					
Treatment 2	1	2,4-D	1.0	1120	Pre-emergence
Treatment 2	2	2,4-D	1.0	1120	12 days (± 1 day) before Application No. 4
Treatment 2	3	Quizalofop-P-ethyl	0.082	92	5 days (± 1 day) before Application No. 4
Treatment 2	4	2,4-D	1.0	1120	V8 or 48 in., whichever comes first
Herbicide Treated Corn Line AAD-1 Event DAS-40474-7 Plot (T3)					
Treatment 3	1	2,4-D	1.0	1120	Pre-emergence
Treatment 3	2	2,4-D	1.0	1120	12 days (± 1 day) before Application No. 4
Treatment 3	3	Quizalofop-P-ethyl	0.082	92	5 days (± 1 day) before Application No. 4
Treatment 3	4	2,4-D	1.0	1120	V8 or 48 in., whichever comes first
2X Herbicide Treated Corn Line AAD-1 Event DAS-40278-9 Plot (T4) –for **Processing Trials ONLY**					
Treatment 4	1	2,4-D	2.0	2240	Pre-emergence
Treatment 4	2	2,4-D	2.0	2240	12 days (± 1 day) before Application No. 4
Treatment 4	3	Quizalofop-P-ethyl	0.164	184	5 days (± 1 day) before Application No. 4
Treatment 4	4	2,4-D	2.0	2240	V8 or 48 in., whichever comes first
2X Herbicide Treated Corn Line AAD-1 Event DAS-40474-7 Plot (T5) –for **Processing Trials ONLY**					
Treatment 5	1	2,4-D	2.0	2240	Pre-emergence
Treatment 5	2	2,4-D	2.0	2240	12 days (± 1 day) before Application No. 4
Treatment 5	3	Quizalofop-P-ethyl	0.164	184	5 days (± 1 day) before Application No. 4
Treatment 5	4	2,4-D	2.0	2240	V8 or 48 in., whichever comes first

(a) 2,4-D application rates are based on acid equivalent (a.e.), so rate is in units of lb a.e./A or g a.e./ha. Quizalofop-P-ethyl application rates are based on active ingredient (a.i.), so rate is in units of lb a.i./A or g a.i./ha.

(b) 2,4-D is to be applied with a non-ionic surfactant (NIS) at approximately 0.25% (v/v)in the spray mixture. Quizalofop-P-ethyl is to be applied with a crop oil concentrate (COC) at approximately 1.25% (v/v) in the spray mixture.

(c) The rates indicated are target rates; the application rate calculated from sprayer output, test substance used, carrier, nozzle spacing, and plot pass times should be within 95 to 110% of the target rate.

(d) All applications are to be applied broadcast with a boom sprayer and after the crop has emerged are to be applied as foliar over-the-top applications.

(e) Application No. 1 is applied preemergence. Application No. 4 is to be targeted at approximately the V8 growth stage or when the corn is approximately 48 in. tall, whichever comes first. Monitor the growth and development of the crop in order to estimate at least 12 days in advance the date at which the corn is expected to reach the V8 stage or 48 in. tall (whichever comes first) and set this as the date targeted for Application No. 4. Once the target date for Application No. 4 is selected, the schedule for Application Nos. 2 and 3 can be set. Application No. 2 is to be applied 12 days (± 1 day) before Application No. 4 and Application No. 3 is to be applied 5 days (± 1 day) before Application No. 4.

7.5. Storage and Return of the Test Material Container(s):

The test substance **container(s)** must be retained for at least the duration of the study and will be **returned to the Sponsor**. After the completion of the final application event, dispose of any remaining test substance (properly rinse container), and return the **test substance container(s) to the Sponsor**. The shipping address to return the containers to the Sponsor is:

Dow AgroSciences, LLC
Attn: Test Substance Coordinator
9330 Zionsville Road
Building 304
Indianapolis, IN 46268-1054
Phone Number: (317) 337-4955

8. RESIDUE SAMPLE COLLECTION

8.1 Sample Identification:

Each sample will be assigned a unique number that will be used throughout the study for identification and tracking purposes. The Sample ID numbers will be generated by the Sponsor and printed on labels. Use the pre-printed labels provided by DAS to identify each sample. If pre-printed labels are not available, label the sample bag or container using a permanent marker with Study Number, Trial Number, Treatment Number, sample type (commodity), Sample ID Number, sampler's signature or initials, and date collected.

8.2 Sample Description:

Field corn Forage – Field corn forage is the whole aerial portion of the field corn plant typically collected at the late dough to early dent stage. However in this study, corn forage RAC samples are to be collected at two timings.

The first timing is to be targeted at 40 days (± 2 days) after the final application of 2,4-D at the V8 stage or 48 in. corn, whichever comes first (application no. 4 in Section 7.4). This sample timing is intended to be earlier than "typical" and the forage samples collected at this timing are expected to not yet have reached the late dough growth stage.

Corn forage RAC samples are also to be collected at a second timing targeted at the "typical" late-dough growth to early-dent stage. For this sample timing, the actual number of days after the final application of 2,4-D will be dependent on the rate of corn maturation to reach the typical growth stage rather than a pre-assigned number of days.

For each corn forage sample, 12 plants located randomly across the plot should be cut approximately 1 to 2 inches above the ground. Any excess soil may be brushed from the stalk and/or leaves, <u>but do not wash</u>. The 12 plants collected for each sample should then be divided into 3 groups of 4 plants each. Each corn plant should be cut into 3 approximately equal length sections **with the leaves and ears attached** (top, middle and bottom section). Collect the top, middle, and bottom sections of the plants, respectively, from each of the three groups of four plants and composite these sections into a single corn forage sample. The composite sample should contain a part of each of the 12 collected plants and include 4 bottom sections, 4 middle sections and 4 top sections with each section coming from a separate corn plant.

Field corn Stover – Field corn stover RAC is the dried, mature plant stalk remaining after collection and removal of the grain or whole ear (cob + grain). Stover samples should be collected on the same day as the grain. The stover and grain samples should be collected when the plants reach maturity suitable for commercial harvest, which typically will be when the plants have dried to approximately 80 to 85% dry matter. For the Decline trials, stover should be collected before, at and after typical maturity, as specified in Section 8.4.

For each corn stover sample, 12 plants located randomly across the plot should be cut approximately 1 to 2 inches above the ground **and the ears removed**. The 12 plants collected for each sample should then be divided into 3 groups of 4 plants each. Each corn plant should be cut into 3 approximately equal lengths sections with the leaves attached (top, middle and bottom section). Collect the top, middle and bottom sections of the plants, respectively, from each of the three groups of four plants and composite these sections into a single corn forage sample. The composite sample should contain a part of each of the 12 collected plants and include 4 bottom sections, 4 middle sections and 4 top sections with each section coming from a separate corn plant.

Field corn Grain – Field corn grain RAC is the seed removed from the cob and husk. Grain samples should be collected once the plant and ears have senesced and dried and the corn first reaches maturity suitable for commercial harvest. The grain samples should be collected on the same day that the stover samples are collected. For the Decline trials, grain should be collected before, at and after typical maturity, as specified in Section 8.4.

Each RAC grain sample should be at least 1 kg from at least 12 plants from different areas across the plot. Each bulk grain sample should be about 1000 pounds (~450 kg).

8.3 **Sample Replicates**:

At each sampling event, collect one (1) composite sample from the untreated plot (untreated T1) and **two (2) independently collected composite RAC samples or Decline samples from the treated plots (T2 and T3). Collect a single bulk grain sample for processing from each of the untreated (T1) and 2X-rate treated plots (T4 and T5). In addition to the bulk seed samples collected from the treated plots, collect duplicate "From-the-Field" RAC samples of seed from the bulk seed treated plots (T4 and T5).** Ensure that each sample is representative of the entire plot by collecting samples randomly throughout the plot and avoid collecting samples near plot edges. Do not collect one sample and split it to provide the requested replicate or sample amount.

8.4 **Sample Collection and Timing**:

At each sampling event, collect one composite sample from the untreated plot (control) and **two independently collected composite samples from the treated plots** (except a single treated bulk grain sample will be collected for processing).

Care must be taken to avoid the possibility of any contamination of the samples. Gloves should be worn during sampling and should be changed between collection of the untreated and treated samples. Samples from the untreated plot will be collected prior to collecting samples from the treated plot. Ensure that each sample is representative of the entire plot. **Do not collect one sample and split it to provide the requested replicate or sample amount.** Do not sample plants at the periphery of the plot (row ends or perimeter rows), where treatment may not have been uniform.

At the time of collection, a sufficient amount of field corn forage, grain, or stover will be collected from the untreated and treated plots to generate the necessary samples. At a minimum, subsamples should be collected from 12 plants from different areas within each plot and composited to comprise a sample representative of the entire plot to meet the weight requirements below. Duplicate, but independently collected, treated samples of field corn forage, grain, and stover RACs should be collected. Single untreated and treated bulk samples of field corn grain should be collected for processing.

Samples should be collected as follows:

For the RAC-only Trials:

- Each treated sample of **forage** should consist of a composite sample of a minimum 1 kg from at least 12 plants located randomly across the plot which should be cut approximately 1 to 2 inches above the ground. The 12 plants collected for each sample should be divided into 3 groups of 4 plants each. Each corn plant should be cut into 3 approximately equal lengths sections with the leaves attached (top, middle and bottom section). Collect the top, middle and bottom sections of the plants, respectively, from each of the three groups of four plants and composite these sections into a single corn forage sample. The composite sample should contain a part of each of the 12 collected plants and include 4 bottom sections, 4 middle sections and 4 top sections with each section coming from a separate corn plant.

  In this study, corn forage RAC samples are to be collected at two timings. The first timing is to be targeted at 40 days (± 2 days) after the final application of 2,4-D at the V8 stage or 48 in. corn, whichever comes first (Application No. 4 in Section 7.4). This sample timing is intended to be early and the forage samples collected at this timing are expected to not yet have reached the late dough growth stage. Corn forage RAC samples are also to be collected at a second timing targeted at the late dough to early dent stage of grain development. For this sample timing, the actual number of days after the final application of 2,4-D will be dependent on the time required for the corn to reach the late dough / early dent growth stage rather than a pre-assigned number of days. For purposes of sample label documentation, samples collected at this timing have been arbitrarily designated as being collected at 60 days after the final application of 2,4-D (application no. 4), but the samples should be collected at the time when the corn reaches the late dough / early dent stage and the actual harvest date recorded.

- Each treated field corn **stover** sample should consist of a composite sample af a minimum 1 kg collected from at least 12 plants located randomly across the plot. These plants should be cut approximately 1 to 2 inches above the ground **and the ears removed**. The 12 plants collected for each sample should then be divided into 3 groups of 4 plants each. Each corn plant should be cut into 3 approximately equal lengths sections with the leaves attached (top, middle and bottom section). Collect the top, middle and bottom sections of the plants, respectively, from each of the three groups of four plants and composite these sections into a single corn forage sample. The composite sample should contain a part of each of the 12 collected plants and include 4 bottom sections, 4 middle sections and 4 top sections with each section coming from a separate corn plant.

  Stover samples should be collected on the same day as the grain. The stover samples should be collected when the plants first reach maturity suitable for commercial harvest, which typically will be when the plants have dried to approximately 80 to 85% dry matter. The actual number of days after the final application of 2,4-D until the corn is sufficiently mature for harvest of stover and grain will be dependent on the time required for the corn to reach maturity rather than a pre-assigned number of days. For purposes of sample label documentation, samples collected at this timing have been arbitrarily designated as being collected at 90 days after the final application of 2,4-D (Application No. 4), but the samples should be collected at the time when the corn first reaches maturity suitable for commercial harvest and the actual date of sample collection recorded.

- Each treated sample of **grain** should consist of minimum 1 kg (removed from the cob and husk) collected from at least 12 plants from separate areas within the plot. Grain samples should be collected once the plant and ears have senesced and dried and the corn first reaches maturity suitable for commercial harvest.

  The grain samples should be collected on the same day that the stover samples are collected. As with the stover samples, the actual number of days after the final application of 2,4-D until the corn is sufficiently mature for harvest of grain will be dependent on the time required for the corn to reach maturity rather than a pre-assigned number of days. For purposes of sample label documentation, samples collected at this timing have been arbitrarily designated as being collected at 90 days after the final application of 2,4-D (Application No. 4), but the samples should be collected at the time when the corn first reaches maturity suitable for commercial harvest and the actual harvest date recorded.

- The untreated plot (T1) should be harvested on the same days as the treated plots to yield a sample of forage, stover, and grain. The untreated control samples should be collected in the same manner and be a similar size as described above for treated samples. Although two independently collected samples of each commodity are required from each treated RAC plot, only a single sample is required from the untreated plot.

For the RAC + Decline Trials

- Field corn forage samples will be collected in the same manner and will be a similar size as described above for RAC-only Trials (i.e. see above Section for RAC-only Trials; each sample is to consist of a composite sample of a minimum 1 kg from at least 12 plants). In this study, corn forage RAC samples are to be collected at two timings. The first timing is to be targeted at 40 days (± 2 days) after the final application of 2,4-D at the V8 stage or 48 in. corn, whichever comes first (Application No. 4 in Section 7.4). Corn forage RAC samples are also to be collected at a second timing targeted at the late dough to early dent growth stage. For this sample timing, the actual number of days after the final application of 2,4-D will be dependent on the time required for the corn to reach the late dough to early dent growth stage rather than a pre-assigned number of days. For purposes of sample label documentation, RAC samples collected at this timing have been arbitrarily designated as being collected at 60 days after the final application of 2,4-D (Application No. 4), but the samples should be collected at the time when the corn reaches the late dough / early dent stage and the actual sample date recorded. For Decline samples, corn forage should also be collected 14 and 7 days (± 2 days) before the corn is expected to be at the late dough to early dent stage [i.e. 14 and 7 days (± 2 days) before the RAC sample is collected] and then at 7 and 14 days (± 2 days) after the RAC sample collection. Therefore, corn forage will be collected at a total of 6 intervals at the RAC + Decline sites -- 40 DAT after Application No. 4, 14 days before RAC sample, 7 days before RAC sample, RAC sample collected once corn reaches the late dough to early dent stage, 7 days after the RAC sample collection, and 14 days after the RAC sample collection. For purposes of sample labeling, as mentioned above, the RAC sample is arbitrarily being assigned a harvest timing of 60 DAT after Application No. 4. Therefore, the four additional forage decline samples that are to be collected will be indicated arbitrarily on sample labels as 46, 53, 67 and 74 DAT. However, the actual collection dates for these four samples relative to Application No. 4 are to be based on the timing when the forage is expected to be at the correct maturity for a forage RAC sample and then the decline samples collected at 14 and 7 days (± 2 days) before the RAC sample is collected and then at 7 and 14 days (± 2 days) after the RAC sample collection. Record the actual date of sample collection.
- Field corn stover samples will be collected in the same manner and will be a similar size as described above for RAC-only Trials (i.e. see above Section for RAC-only Trials; each sample is to consist of a composite sample of a minimum 1 kg from at least 12 plants with ears removed). The actual number of days after the final application of 2,4-D until the corn is sufficiently mature for harvest of RAC stover and grain will be dependent on the time required for the corn to reach maturity rather than a pre-assigned number of days. For purposes of sample label documentation, RAC samples collected at this timing have been arbitrarily designated as being collected at 90 days after the final

application of 2,4-D (Application No. 4), but the RAC samples should be collected at the time when the corn first reaches maturity suitable for commercial harvest and the actual date of sample collection recorded. Corn grain samples should be collected on the same day that the stover samples are collected. At sites where Decline + RAC samples are collected, samples of corn stover should be collected at a total of five timings: 14 and 7 days (± 2 days) before the corn is expected to be mature for normal harvest, at a timing when the corn first reaches maturity suitable for commercial harvest (i.e. the RAC sample), and then at 7 and 14 days (± 2 days) after the RAC sample collection. For purposes of sample labeling, as mentioned above, the RAC sample is arbitrarily being assigned a harvest timing of 90 DAT after Application No. 4. Therefore, the four additional stover decline samples that are to be collected will be indicated arbitrarily on sample labels as 76, 83, 97 and 104 DAT. However, the actual collection dates for these four samples relative to Application No. 4 are to be based on the timing when the stover is expected to be at the correct maturity for a stover RAC sample and then the decline samples collected at 14 and 7 days (± 2 days) before the RAC sample is collected and then at 7 and 14 days (± 2 days) after the RAC sample collection. Record the actual date of sample collection.

- Field corn grain samples will be collected in the same manner and will be a similar size as described above for RAC-only Trials (i.e. see above Section for RAC-only Trials; minimum 1 kg of grain removed from the cob and husk and collected from at least 12 plants from separate areas within the plot). The grain samples should be collected on the same day that the stover samples are collected. As with the stover samples, the actual number of days after the final application of 2,4-D until the corn is sufficiently mature for harvest of grain will be dependent on the time required for the corn to reach maturity rather than a pre-assigned number of days. For purposes of sample label documentation, samples collected at this timing have been arbitrarily designated as being collected at 90 days after the final application of 2,4-D (Application No. 4), but the samples should be collected at the time when the corn first reaches maturity suitable for commercial harvest and the actual harvest date recorded. At sites where Decline + RAC samples are collected, samples of corn grain should be collected at a total of five timings: 14 and 7 days (± 2 days) before the corn is expected to be mature for normal harvest, at a timing when the corn first reaches maturity suitable for commercial harvest (i.e. the RAC sample), and then at 7 and 14 days (± 2 days) after the RAC sample collection. For purposes of sample labeling, as mentioned above, the RAC sample is arbitrarily being assigned a harvest timing of 90 DAT after Application No. 4. Therefore, the four additional stover decline samples that are to be collected will be indicated arbitrarily on sample labels as 76, 83, 97 and 104 DAT. However, the actual collection dates for these four samples relative to Application No. 4 are to be based on the timing when the stover is expected to be at the correct maturity for a stover RAC sample and then the decline samples collected at 14 and 7 days (± 2 days) before the RAC sample is collected and then at 7 and 14 days (± 2 days) after the RAC sample collection. Record the actual date of sample collection.

- Forage, stover, and grain should be harvested from the untreated plot on the same days that commodity is harvested from the treated plot. The untreated control samples should be collected in the same manner and be a similar size as described previously for the RAC samples. Although two independently collected samples of each commodity are required from each treated plot at each sample timing, only a single sample is required from the untreated plot at each sample timing.

For the RAC + Processing Trials:

- Field corn forage samples will be collected from the untreated plot T1 and the 1X-treated plots, T2 and T3. The corn forage samples are to be collected in the same manner and will be a similar size as previously for RAC-only Trials (i.e. see above Section for RAC-only Trials; each sample is to consist of a composite sample of a minimum 1 kg from at least 12 plants). In this study, corn forage RAC samples are to be collected at two timings. The first timing is to be targeted at 40 days (± 2 days) after the final application of 2,4-D at the V8 stage or 48 in. corn, whichever comes first (Application No. 4 in Section 7.4). Corn forage RAC samples are also to be collected at a second timing targeted at the late dough to early dent growth stage. For this sample timing, the actual number of days after the final application of 2,4-D will be dependent on the time required for the corn to reach the late dough to early dent growth stage rather than a pre-assigned number of days. For purposes of sample label documentation, samples collected at this timing have been arbitrarily designated as being collected at 60 days after the final application of 2,4-D (Application No. 4), but the samples should be collected at the time when the corn reaches the late dough to early dent stage and the actual sample date recorded.
- Field corn RAC stover samples will be collected on the same day as RAC grain samples and will be collected from the untreated plot T1 and the 1X-treated plots, T2 and T3. Field corn stover samples will be collected in the same manner and will be a similar size as described above for RAC-only Trials (i.e. see above Section for RAC-only Trials; each sample is to consist of a composite sample of a minimum 1 kg from at least 12 plants with ears removed). The actual number of days after the final application of 2,4-D until the corn is sufficiently mature for harvest of stover and grain will be dependent on the time required for the corn to reach maturity rather than a pre-assigned number of days. For purposes of sample label documentation, samples collected at this timing have been arbitrarily designated as being collected at 90 days after the final application of 2,4-D (Application No. 4), but the samples should be collected at the time when the corn first reaches maturity suitable for commercial harvest and the actual date of sample collection recorded.
- Field corn grain RAC samples will be collected on the same day as the RAC stover samples and will be collected from the untreated plot T1 and the 1X-treated plots, T2 and T3. Field corn grain samples will be collected in the same manner and will be a similar size as described above for RAC-only Trials (i.e. see above Section for RAC-only Trials; minimum 1 kg of grain removed from the cob and husk and collected from at least 12 plants from separate areas

within the plot). The actual number of days after the final application of 2,4-D until the corn is sufficiently mature for harvest of stover and grain will be dependent on the time required for the corn to reach maturity rather than a pre-assigned number of days. For purposes of sample label documentation, samples collected at this timing have been arbitrarily designated as being collected at 90 days after the final application of 2,4-D (Application No. 4), but the samples should be collected at the time when the corn first reaches maturity suitable for commercial harvest and the actual date of sample collection recorded.

- Bulk field corn grain samples (about 1000 pounds each) will be collected at "typical" commercial maturity from both the untreated plot (T1) and each of the 2X-treated plots (T4 and T5). A single bulk grain sample from each plot is required. As with the RAC grain samples described above, for purposes of sample label documentation, samples collected at this timing have been arbitrarily designated as being collected at 90 days after the final application of 2,4-D (Application No. 4), but the samples should be collected at the time when the corn first reaches maturity suitable for commercial harvest and the actual date of sample collection recorded.
- Corn "From-the-Field" RAC grain samples will also be collected from the **bulk** 2X-treated plots (T4 and T5) at "typical" commercial harvest and on the same date that the bulk grain samples are collected. Duplicate "From-the-Field" grain samples are required from each of the 2X-treated plots (T4 and T5). These samples are to be collected in the same manner and have the same size requirement as described above for RAC-only corn grain samples (i.e. see above Section for RAC-only Trials; minimum 1 kg of grain removed from the cob and husk and collected from at least 12 plants from separate areas within the plot). As with the RAC and bulk samples of grain for processing, for purposes of sample label documentation, samples collected at this timing have been arbitrarily designated as being collected at 90 days after the final application of 2,4-D (Application No. 4), but the samples **should be collected on the same date that the bulk seed samples are collected** and the actual date of sample collection recorded.

8.5 **Sample Packaging and Signage**:

The transgenic field corn samples from this study are considered regulated articles by the USDA and CFIA. Therefore, packaging and signage must be used in order to store and ship the samples. Three layers of containment are required, which typically includes a heavy, 5-mil plastic bag containing the sample, over-bagged with a labeled cloth residue bag, within a shipping box or plastic cooler. Samples must be clearly marked as regulated material, so it is advisable to place sample bags within boxes immediately upon storage with signs indicating the contents are "Regulated Plant Materials".

For each sample, weigh the amount of commodity placed in the bag/container and record the weight in the field trial notebook.

8.6 Estimated Number of Field Samples:

A summary of expected samples for the RAC, RAC + Decline, and RAC + Processing trials is presented in the table below:

Trial No.	Region	Researcher	Field corn Forage [a]		Field corn Stover		Field corn Grain		Total
			Control	Treated	Control	Treated	Control	Treated	
NJ-01	1	Dan Ramsdell	2	8	1	4	1	4	20
PA-02	1	Dan Ramsdell	2	8	1	4	1	4	20
GA-03	2	Chris Cromer	2	8	1	4	2 [b]	10 [b]	27 [b]
GA-04	2	Chris Cromer	6	24	5	20	5	20	80 [c]
IL-05	5	Sue Dorsey	2	8	1	4	1	4	20
IL-06	5	Tim Boeker	2	8	1	4	1	4	20
IN-07	5	John Bailey	2	8	1	4	1	4	20
MO-08	5	Matt Cordell	2	8	1	4	1	4	20
MO-09	5	Matt Cordell	2	8	1	4	1	4	20
IL-10	5	Matt Cordell	2	8	1	4	1	4	20
MI-11	5	Chad Harris	6	24	5	20	5	20	80 [c]
OH-12	5	Chad Harris	2	8	1	4	1	4	20
IN-13	5	Chad Harris	2	8	1	4	1	4	20
NE-14	5	Matt Krause	2	8	1	4	2 [b]	10 [b]	27 [b]
NE-15	5	Denny Stamm	2	8	1	4	1	4	20
KS-16	5	Denny Stamm	2	8	1	4	1	4	20
IA-17	5	David Bennett	2	8	1	4	1	4	20
MO-18	5	David Bennett	2	8	1	4	1	4	20
IA-19	5	Kyle Johnson	2	8	1	4	1	4	20
MN-20	5	Kyle Johnson	2	8	1	4	1	4	20
MO-21	5	Nathan Goldschmidt	2	8	1	4	1	4	20
OK-22	6	Tim Case	6	24	5	20	5	20	80 [c]
TX-23	6	Tim Case	2	8	1	4	1	4	20
ON-24	5A	Jamie Parnell	2	8	1	4	1	4	20
ON-25	5A	Jamie Parnell	2	8	1	4	1	4	20
		Total	**62**	**248**	**37**	**148**	**39**	**160**	**694**

(a) Corn forage samples are to be collected at two timings - 40 days (± 3 days) after application number 4, and then also when corn reaches the late-dough growth stage. For the decline trials, forage RAC samples are to be collected at 5 intervals - two additional timings before and two timings after the "typical" sample collection interval at the late dough stage.

(b) Includes RAC samples plus one bulk control grain (T1) and two bulk 2X-treated grain samples (one bulk grain sample each from T4 and T5) for processing, plus four "From-the-Field" grain RAC samples collected from T4 and T5 (duplicate 1 kg samples of grain from each of T4 and T5).

(c) Includes decline samples of field corn forage at 6 total timings: at approximately 40 days (± 3 days) after application number 4, at 14 and 7 days (± 2 days) before the corn is expected to be at the late-dough stage, at a timing when the corn reaches the late dough stage (i.e., the RAC sample), and then at 7 and 14 days (± 2 days) after the RAC sample collection. Corn stover and grain are collected at 5 timings: 14 and 7 days (± 2 days) before the corn is expected to reach commercial mature harvest, at a timing when the corn reaches mature commercial harvest stage (i.e., RAC sample timing), and then at 7 and 14 days (± 2 days) after the RAC sample collection.

8.7. **Sample Integrity and Handling**:

Field corn forage, grain, and stover RAC and decline samples must be chilled (cooler with blue, dry or wet ice, or freezer) within 4 hours of collection and placed in a freezer (with temperature monitoring, maintained on average below 0°C, typically near -20 °C) within 8 hours of collection in order to maintain their integrity.

See Section 8.9 for instructions on Sample Handling, Storage and Shipping of the bulk seed samples to be used in processing.

Document the collection, handling, and time for samples to be taken from the field to storage in the field trial notebook. The temperature of the storage unit must be monitored and documented.

8.8 **Sample Storage:**

The transgenic field corn samples from this study are considered regulated articles by the USDA and CFIA. Therefore, samples must be properly stored in a manner to prevent mixing with non-regulated material. All regulated seed and plant parts must be clearly marked and stored in a restricted-access area segregated from non-regulated material. Typically, this requires a separate, locked freezer unit, or isolated area within a large freezer room. The storage unit or area must be clearly marked as a regulated materials storage or work area.

8.9 **Field Sample Shipping**:

The transgenic field corn commodities from this study are considered regulated articles by the USDA and CFIA. Special Labeling is required to ship regulated materials. Therefore, proper documentation and notifications must be made in order to ship them. The Study Director and/or Sponsor Monitor will provide the necessary documentation to ship samples. As indicated in Section 8.5, three levels of packaging containment are required for shipping all samples.

RAC and Decline samples should be shipped frozen to the Sponsor's sample preparation facility, as soon after collection as possible, by ARA freezer truck with temperature monitoring capability. If another carrier, or overnight air shipment with dry ice is required, contact the Study Director for authorization, and arrange shipment to avoid delivery on a weekend or holiday. Do not ship samples from different studies in the same box unless approved by the Study Director. Treated and control samples (packaged in individual residue bags) may be shipped in the same container provided they are frozen and each sample bag has been over-bagged so there is no danger of contamination and seepage if thawing occurs.

The bulk samples should be shipped (ambient or refrigerated) to the Processing Facility (see Section 9) ideally within 72 hours of collection and delivered within approximately 7 days of harvest; avoid delivery on a weekend or holiday. If more than approximately 72 hours are required before the samples can be shipped to the Processing Facility, place the samples in a refrigerated unit (not a freezer) for storage until they can be picked up for shipment and notify the Study Director.

To coordinate shipping, contact ARA prior to shipping samples;

Mr. Terry Blackburn
Ag Research Associates, LLC
Sample Shipping and Storage
Phone number; (252) 437-3114
Mobile number; (252) 458-4571
FAX number; (252) 437-3114
e-mail; terry.blackburn@agresearchassociates.com

Ship RAC and Decline samples to the following address, unless the Study Director indicates an alternate preparation facility.

Dow AgroSciences LLC
Sample Management
9330 Zionsville Road, 306, Cb/020
Indianapolis, Indiana 46268
Phone; (317) 337-3622
FAX; (317) 337-3888

Note: Prior to delivering samples to Dow AgroSciences, send notification of sample shipment (specify protocol number, study director, carrier, expected arrival date, shipment identification number, and USDA notification number) **by e-mail to eclsampman@dow.com**. If e-mailing is not possible, call (317) 337-3622 or fax (317) 337-3888 with the information.

8.10 **Crop Destruction, Volunteer Monitoring, and Test Site Decommissioning**:

After the last sampling has been completed, destroy any/all remaining transgenic field corn and any buffer zone non-transgenic field corn so that it will not be used for human or animal feed or allowed to persist or propagate in the environment. Remaining vegetative material should be incorporated into the soil and left to decompose by the elements. For Canadian sites, follow CFIA guidelines regarding crop destruct / incorporation. All machinery that may retain reproductive parts of the transgenic crop should be cleaned after use. Document the date and method of crop destruction and equipment cleanout on an appropriate form in the field trial notebook.

Monitoring for volunteer plants in the transgenic plots will be required in 2010 as part of the requirements of the USDA and CFIA. However, this requirement for decommissioning is not part of the requirement for this Magnitude of Residue study and will not be part of this study protocol. Refer to the USDA release permit or CFIA documentation for full information on requirements for volunteer monitoring and decommissioning. The Sponsor will manage this process independent of this study.

8.11 **Field Trial Summary**:

At the conclusion of the field phase of the study, a Field Trial Summary must be prepared for each field trial by the respective PFI, or delegate, and submitted to the

Study Director or Testing Facility. The PFI should review the field trial notebook and any other raw data or facility records to summarize the trial and ensure completeness of the data package. If the field trial is being conducted by a non-ARA subcontractor, it is the responsibility of the subcontractor to have the Field Trial Notebook and Field Trial Summary audited by the subcontractor's QAU. A template of the DAS EUREKA summary table format or the ARA Field Trial Summary will be made available to the PFI by the Study Director or Sponsor Monitor.

Within **approximately one month after the last sample shipment**, send by secure service the Field Trial Summary and supporting Field Trial Notebook and data package (complete GLP field trial notebook paper forms containing original raw field data and documentation) to the Study Director or Testing Facility for review. Keep a legible, verified copy of the field trial notebook and any other raw data as a backup in case of loss or damage during transit.

9. FOOD/FEED COMMODITY PROCESSING

The preparation of food/feed commodities from the field corn grain bulk samples will be conducted by GLP Technologies, Navasota, TX. The Sample Processing Principal Investigator, Dick Dusek, is responsible for ensuring that the processing is conducted in accordance with the protocol, GLPs, and GLP Technologies' SOPs.

The control bulk field corn grain sample and the two treated bulk grain samples (one from each of two treated transgenic corn lines) from each trial (2 trials) will each be processed simulating commercial practices for field corn processed fractions. Three "from the processor" whole field corn grain sample from each bulk sample (the RAC) will be collected just prior to processing. The required field corn commodities from each bulk sample are: "from the processor" whole field corn grain, aspirated grain fraction, starch, refined oil (wet milling), meal, grits, flour, and refined oil (dry milling). In addition to the 3 samples of "from the processor" whole grain sample collected prior to cleaning or processing, two samples of each processed fraction from each bulk grain sample are to be collected (one sample of aspirated grain fraction may be collected if there is insufficient material for two samples).

9.1 Sample Processing Facility:

GLP Technologies
Attn: Dick Dusek
22723 State Hwy 6 South
Navasota, Texas 77868
Telephone: (936) 825-2184
FAX: (936) 825-7929
E-Mail: ddusek@glptech.net

9.2 Sample Processing Procedures:

The sample processing will be conducted according to GLP Technology SOPs for preparation of field corn processed commodities. The field corn processing procedures includes cleaning of the bulk field corn grain (screening and aspiration), a wet-milling process for starch and oil generation, and a dry-milling process for generation of grits, meal, flour, and oil. The processing procedures include grain prep/cleaning, grain conditioning, extrusion/expansion, oil extraction with hexane, and refining of crude oil. The detailed procedures for commodity generation will be included in the Processing study file and summarized in the Procession Fractions report.

9.3 **Expected Processed Food/Feed Samples:**

There will be 3 bulk samples of grain per trial with one bulk grain sample collected from the untreated control plot (T1) and then a bulk grain sample collected from each of two treated plots (T4 and T5). As mentioned previously, three "from the processor" corn grain samples from each bulk sample (the RAC) will be collected just prior to cleaning and processing. Except for the aspirated grain fraction, two samples of each processed fraction are to be collected from each bulk grain sample.

The expected whole grain samples prior to cleaning and processing and the processed commodities samples from each trial and approximate sample size for each sample are:

- Whole field corn grain RAC prior to processing – minimum 1 kg (~2 lbs). [3 samples each from T1, T4 and T5 – total of 9 grain samples]
- Aspirated grain fractions – all available [1 sample each from T1, T4 and T5]
- Starch – minimum 1 liter or 1kg (~2 lbs) [2 samples each from T1, T4 and T5]
- Refined oil from wet milling – ~1 liter [2 samples each from T1, T4 and T5]
- Meal – minimum 1 kg (~2 lbs.) [2 samples each from T1, T4 and T5]
- Grits – minimum 1 kg (~2 lbs.) [2 samples each from T1, T4 and T5]
- Flour – minimum 1 kg (~2 lbs.) [2 samples each from T1, T4 and T5]
- Refined oil from dry milling – ~1 liter [2 samples each from T1, T4 and T5]

Treated field corn processed commodities remaining after the required samples are collected must be disposed of following requirements for regulated material.

9.4 **Sample Identification**:

Each sample will be assigned a unique number that will be used throughout the study for identification and tracking purposes. The Sample ID numbers will be generated by the Sponsor and printed on labels. Use the pre-printed labels provided to properly identify each sample. If pre-printed labels are not available, label the sample bag or container using a permanent marker with Study Number, Trial Number, Treatment Number, sample type (commodity), Sample ID Number, sampler's signature or initials, and date collected.

9.5 **Processed Sample Handling, Storage, and Shipping**:

Handle samples in a way to avoid the possibility of contamination between the untreated and treated samples. Samples should be placed into properly labeled containers and stored in a freezer as soon after preparation / collection as possible. The temperature of the freezer in which samples are stored must be monitored and documented and must be maintained below 0°C (typically, approximately minus-20°C).

Processed feed/food commodity samples should be shipped to the DAS Sample Management facility. Ship the frozen samples by freezer truck with temperature monitoring capability or, if requested by Study Director, by overnight air shipment with dry ice. Treated and control samples (packaged in individual containers) may be shipped together provided they are frozen and each sample container has been over-bagged in plastic bags so there is no danger of contamination and seepage if thawing occurs. Three levels of packaging containment are required for shipping of all samples. **Ship samples to the following address, unless the Study Director indicates an alternate preparation facility.**

Dow AgroSciences LLC
Sample Management
9330 Zionsville Road, 306, Cb/020
Indianapolis, Indiana 46268
Phone; (317) 337-3622
FAX; (317) 337-3888

Note: Prior to delivering samples to Dow AgroSciences, send notification of sample shipment (specify protocol number, study director, carrier, expected arrival date, shipment identification number, and USDA notification number) **by e-mail to eclsampman@dow.com**. If e-mailing is not possible, call (317) 337-3622 of fax (317) 337-3888 with the information.

9.6 **Excess Sample Destruction and Cleanup**:

Unless otherwise specified by the Study Director, all remaining transgenic bulk grain and/or processed commodities from the transgenic materials will be disposed of in a manner so that transgenic grain or the processed products produced from it will not be used for human or animal feed and will not be allowed to persist or propagate in the environment. All machinery that may retain any seed or crop commodity should be cleaned after use. Document the date(s) and method(s) of grain/seed and excess commodity destruction and equipment cleanup on an appropriate form in the processing phase data file.

9.7 **Sample Processing Summary**:

At the conclusion of the Sample Processing phase, a Sample Processing Summary must be prepared by the Sample Processing Principal Investigator or delegate, and sent to the Study Director. The Sample Processing Summary report will include, at a minimum, handling and preparation procedures and fractionation and/or material balance information. The Sample Processing Summary and raw data will be reviewed by the Sample Processing Facility QAU and verified to be compliant with GLPs, or exceptions noted, including when audits/inspections were performed. The final Sample Processing Summary with raw data will be completed and sent to the Study Director within approximately six (6) weeks after all processed fractions are shipped to Dow AgroSciences.

10. SAMPLE HOMOGENIZATION FOR ANALYSIS

Upon receipt at the Sponsor's Sample Preparation laboratory, the samples will be inspected for damage, identified, and checked against the shipping documentation according to DAS SOP(s). Samples will be stored frozen at approximately -20°C. Samples will be prepared by DAS Sample Management in accordance with applicable DAS SOPs for the crop and commodity to provide a homogeneous sample for analysis. After chopping/grinding and homogenization, subsamples will be prepared for the analytical laboratory.

The remaining "retain for storage" sample will be stored frozen at the DAS Sample Management facility. If needed, additional subsample can be provided to the analytical laboratory upon request. Samples will be retained in frozen storage at least until the study is completed, discontinued, or terminated. After that time, sample disposal is at the discretion of the Sponsor and will be under the authorization of the Sponsor's QAU in accordance with USEPA FIFRA GLPs, 40 CFR Part 160.195(c).

11. ANALYTICAL PHASE

Details of the Analytical Phase will be added by Protocol Amendment. The overall conduct of the Analytical Phase is the responsibility of the Principal Analyst at the Sponsor's analytical facility. Unless specified otherwise in the study protocol, the procedures to be followed for the analysis of samples generated from this study will be those outlined in DAS SOP ECL-37. The analytical method to be used to analyze the samples and other details concerning the analytical phase of the study, such as the analytical standards, will be added by amendment to this protocol. If needed, the selected metabolites of 2,4-D, which may also be analyzed for in samples of raw agricultural commodities and processed fractions, will be specified in the protocol amendment.

11.1 **Analytical Summary**:

The analytical laboratory will prepare an Analytical Summary including the analysis methodology, equipment performance records, analytical results for samples, including fortified sample recoveries, and untreated and treated sample residue results. The DAS EUREKA analytical summary table format will be used to tabulate the final crop commodity residue results and other key analytical information, but is not intended to be part of reporting of study results under GLP and as such does not need to be QA audited. The analytical laboratory's on-site Quality Assurance Unit (QAU) will audit the analyses and the Summary. The Analytical Summary will include a signed statement from the Principal Analyst attesting to the GLP compliance status of the analytical phase of the study. The Analytical Summary will be appended to the final study report.

12. BIAS AND CONTAMINANTS

No experimental bias is expected during the study. Measures to control bias include composite samplings, replicated treated samples, and separate analysis of the replicates. No contaminants are expected during this study and measures will be taken to avoid contamination, such as cleaning equipment, careful collection of crop samples, and using known maintenance products.

If crop injury or another unexpected plant response occurs, document the type and duration in the field trial notebook and notify the Study Director.

13. DATA EVALUATION AND STATISTICAL ANALYSIS

Data analysis, interpretation, and reporting will be conducted according to pertinent regulatory agency guidelines. Results of sample analyses will be entered into a spreadsheet format and common statistics (e.g., mean, median, standard deviation, relative standard deviations, and/or regression analysis) will be performed with spreadsheet functions. Potential outliers, if any, will be determined using a recognized statistical method. Examples or written descriptions of all non-standard calculations (not within spreadsheet software) will be included in the study file, and in the final study report if these data are reported.

14. RECORDS TO BE MAINTAINED

All raw data must be retained in the study records. Original "raw data" (or exact, verified copies) include completed field trial notebook forms, original observations, analytical data, spreadsheets, paper work sheets, pertinent emails, memoranda, and notes, phone logs, and activities of the study that are necessary for the reconstruction and evaluation of the study. Any pertinent data generated in addition to that requested will be inserted into the study file. "Raw Data" may include photographs, slides, videotapes, computer files (magnetic media), and recorded data from automated instruments.

Indelible ink must be used for handwritten study information, signatures, and dates.

15. QUALITY ASSURANCE

ARA's QAU will provide overall assurance of compliance to the Good Laboratory Practice Standards (GLPs) for this study. Any field facility and the analytical laboratory may undergo a quality assurance facility inspection by the ARA QAU if it is considered necessary to assure that facilities, equipment, personnel, procedures, and records conform to GLPs.

GLP compliance inspections of the field phase will be performed by the local field trial QAU at a frequency to ensure the integrity of the trial. In-progress inspections will typically include a critical event, such as calibration of application devices, application of the test substance, or representative sample collection. However, when a PFI has multiple trials, a critical phase audit should be done on at least one trial but it is not necessary to perform a critical phase audit of every trial. An audit of the final field trial notebook and field phase summary is required for each trial. QAU inspection reports will be forwarded to the Study Director promptly for review and signature.

The Processing Facility QAU will conduct GLP compliance inspection(s) and audit(s) of the feed/food sample processing phase at a frequency to ensure the integrity of the phase. The Processing Facility QAU will audit the processing raw data file and the final Processed Fractions summary report.

The DAS QAU will conduct GLP compliance inspections and audits of sample preparation and the analytical phase at a frequency to ensure the integrity of the phase. The DAS QAU will audit the analytical raw data file and the final Analytical Summary.

Any field, processing, and/or analytical phase Summary(s) may be reviewed by the ARA QAU to ensure that the required methods and standard operating procedures were followed, and for overall protocol and GLP compliance. Results from these QAU inspections will be reported to the Field, Processor, or Principal Analyst, as appropriate, the Study Director, and the Study Director's management.

The Final Study Report will be reviewed by the ARA QAU to ensure that overall the protocol was followed and the study was conducted in compliance with the GLPs, or exceptions will be noted. A signed statement will be included in each of the phase summaries and the Final Study Report specifying when audits/inspections were performed and when they were reported to the Study Director and the Study Director's management.

16. GOOD LABORATORY PRACTICE

The study will be conducted in accordance with the EPA FIFRA Good Laboratory Practice Standards, Title 40 Code of Federal Regulations Part 160 (**Reference 4**).

When modifications to this protocol are deemed necessary, an amendment (notification of change before the event occurs) to the protocol will be written. The amendment will include a description of the change, the reason for the change, the effect of the change on the study, and the effective date of the change. Depending on the phase of the study, the Principal Field Investigator or Principal Analyst should contact the Study Director notifying the necessary modifications and discussing the amendment. The Protocol Amendment will be signed and dated by the Study Director and the Sponsor Monitor, and if appropriate the Principal Field Investigator or Principal Analyst, and appended to the protocol. Any amendments to the protocol will be promptly documented by the Study Director and sent to study participants.

Deviations (notification of change after the event occurs) from this protocol or SOPs will be recorded in the field trial notebook by the Principal Field Investigator, or analytical laboratory notebook by the Principal Analyst, and will be submitted for approval by the Study Director. Any deviations from the protocol or SOPs must be reported to the Study Director as soon as possible after the deviation occurs. The deviation notification form will include a description of the change and the reason for the deviation. The Study Director will assess the effect and impact of the deviation on the study. The completed deviation will include a description of the change, the reason for the change, the effect of the change on the study, and will be approved by the Study Director.

17. FINAL STUDY REPORT

A final study report presenting the field, processing, and analytical data will be generated in accordance with the USEPA PR 86-5 **(Reference 5)** and FIFRA GLPs requirements, 40 CFR Part 160.185. The Study Director will consolidate field, processing, and analytical activities and key data collected under this protocol within one final report. The field and analytical summaries will be appended to the final study report. The final report of the study will be reviewed by ARA QAU, approved by the Sponsor, and issued by the Study Director.

18. RECORDS RETENTION

All original raw data generated except for facility raw data, such as calibration logs, chemical storage, and freezer temperature logs, will be organized and transferred to the Sponsor's archives (Dow AgroSciences, Indianapolis, IN), as a separate raw data package for archival after study completion. Facility records will be maintained by the corresponding facility. Verified copies of facility records may be requested.

After completion of the study, the final report, protocol, and all raw data and records will be shipped to the Sponsor Study Monitor, or his/her designee, and retained in the Sponsor's archives in Indianapolis, Indiana.

19. REFERENCES

1. Residue Chemistry Test Guidelines, OPPTS 860.1500, Crop Field Trials, EPA 712-C-95-183, August 1995.
2. Residue Chemistry Test Guidelines, OPPTS 860.1520, Processed Food/Feed, EPA 712-C-96-184, August 1996.
3. Pest Management Regulatory Agency, Health Canada, Residue Chemistry Guidelines, Section 9: Crop Field Trials, Regulatory Directive Dir98-02, June 1998.
4. U.S. Environmental Protection Agency. 1989. Pesticide Programs; Good Laboratory Practice Standards; Final Rule (40 CFR, Part 160). *Federal Register, Vol.* 54, No. 158: 34052-34074.
5. U.S. Environmental Protection Agency. 1986. PR Notice 86-5. Standard format for data submitted under the Federal Herbicide, Fungicide and Rodenticide Act (FIFRA) and certain provisions of the Federal Food, Drug and Cosmetic Act (FFDCA). 17 pages.

Ag Research Associates

1730 Denham Road
Sycamore, GA 31790

PROTOCOL AMENDMENT

Study Number: ARA-09-15-10 **Amendment Number:** 01

Study Title: Magnitude of the Residue of 2,4-D and Quizalofop-P-ethyl in/on Herbicide Tolerant Field Corn Containing the Aryloxyalkanoate Dioxygenase-1 (AAD-1) Gene

Description of Change:

1) The County location of Trials GA-03 and GA-04 will be switched (see below). Trial GA-03, the RAC + Processing trial, will be located in Turner County, Georgia. Trial GA-04, the RAC + Decline trial, will be located in Tift County, Georgia. Chris Cromer will remain as the PFI for both trials.

Field Trial Number	Field Investigator	Research Company	NAFTA Region	State / Prov.	County	Trial Type
GA-03	Chris Cromer	Ag Research Associates	2	GA	Turner	RAC + Processing[a]
GA-04	Chris Cromer	Ag Research Associates	2	GA	Tift	RAC + Decline[b]

Reason for Changes:

1) The Turner County Georgia farm location is better capable of accommodating the large size of the plots required for the Processing sample, and the required physical separation from other corn.

Effect of Changes:

1) No negative impact is expected. Both Counties are listed on the APHIS Permit for "release" of the transgenic seed.

Effective Date: 08Jun09

Approved By:

Study Director: John M. Becker
John M. Becker, PhD
Ag Research Associates
Date: 08 Jun 09

Sponsor Study Monitor: Steve W. Rosser
Steve W. Rosser, PhD
Dow AgroSciences
Date: 10 - Jun - 2009

Ag Research Associates
1730 Denham Road
Sycamore, GA 31790

PROTOCOL AMENDMENT

Study Number: ARA-09-15-10 **Amendment Number:** 02

Study Title: Magnitude of the Residue of 2,4-D and Quizalofop-P-ethyl in/on Herbicide Tolerant Field Corn Containing the Aryloxyalkanoate Dioxygenase-1 (AAD-1) Gene

Description of Change:

1) The Principal Field Investigator for Trial IA-17 is changed from David Bennett to Jason Niekamp (protocol Sections 6.2 and 8.6). Both PFIs are employed by Bennett Agricultural Research Corporation and the location of the trial is not affected.

2) The Principal Analyst for the study will be Gary Schelle of Dow AgroSciences (9330 Zionsville Road, 306 / 2A, Indianapolis, IN; Phone # 317 337-3636).

Reason for Changes :

1) The responsibility for this field trial in Iowa was transferred to Jason Niekamp.

2) The responsibility for the analytical phase was not previously determined, but has been assigned to Gary Schelle.

Effect of Changes:

1) No negative impact is expected. Jason Niekamp is a qualified PFI for this field trial.

2) No negative impact. The PA is named to oversee the analytical phase.

Effective Date: 15Jul09

JC 21-Sep-2010

Approved By:

Study Director: John M. Becker Date: 15 Jul 09
John M. Becker, PhD
Ag Research Associates

Sponsor Study Monitor: Steve W. Rosser Date: 16-Jul-2009
Steve W. Rosser, PhD
Dow AgroSciences

Ag Research Associates

1730 Denham Road
Sycamore, GA 31790

PROTOCOL AMENDMENT

Study Number: ARA-09-15-10 **Amendment Number:** 03

Study Title: **Magnitude of the Residue of 2,4-D and Quizalofop-P-ethyl in/on Herbicide Tolerant Field Corn Containing the Aryloxyalkanoate Dioxygenase-1 (AAD-1) Gene**

Description of Changes:

1) The sample shipping coordinator is changed from Mr. Terry Blackburn to Mr. Tim Case of ARA (Protocol Section 8.9). Contact and scheduling for ARA freezer truck shipping should be directed to Mr. Case at 806 248-7975 (office phone), 806-277-0164 (mobile phone) or tim.case@agresearchassociates.com (email).

Reason for Changes:

1) Mr. Tim Case has assumed responsibility for sample shipping activities for ARA.

Effect of Changes:

1) There is no negative impact. Tim Case will be the point-of-contact for sample shipment to the DAS Sample Management lab.

Effective Date: 06Aug09

Approved By:

Study Director: John M. Becker — Date: 07Aug09
John M. Becker, PhD
Ag Research Associates

Sponsor Study Monitor: Steve W. Rosser — Date: 12-Aug-2009
Steve W. Rosser, PhD
Dow AgroSciences

Ag Research Associates

1730 Denham Road
Sycamore, GA 31790

PROTOCOL AMENDMENT

Study Number: ARA-09-15-10 **Amendment Number:** 04

Study Title: Magnitude of the Residue of 2,4-D and Quizalofop-P-ethyl in/on Herbicide Tolerant Field Corn Containing the Aryloxyalkanoate Dioxygenase-1 (AAD-1) Gene

Description of Change:

1) The bulk grain samples for processing from Trials GA-03 and NE-14 can also be shipped frozen [in addition to ambient or refrigerated according to the protocol (within 72 hours for receipt within 7 days)] to the Sample Processing Facility (Protocol Section 8.9). If the bulk samples will be shipped frozen, they should be frozen within 72 hours of collection. Ambient or refrigerated samples should still be delivered within 7 days of collection; however frozen samples should be delivered within 30 days. The requirement to notify the Study Director if refrigerated samples cannot be shipped within 72 hours is removed.

Reason for Change:

1) The Sample Processing Facility will store the bulk samples frozen, so freezing samples prior to shipping is acceptable. Ambient or refrigerated shipping according to the protocol with expedited delivery is acceptable, without SD notification.

Effect of Change:

1) There is no negative impact. Shipment of frozen bulk samples is acceptable for subsequent Processing and is likely to preserve any possible residues.

Effective Date: 29Sep09

Approved By:

Study Director: John M. Becker Date: 29Sep09
John M. Becker, PhD
Ag Research Associates

Sponsor Study Monitor: Steve W. Rosser Date: 05-Oct-2009
Steve W. Rosser, PhD
Dow AgroSciences

Ag Research Associates

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PROTOCOL AMENDMENT

Study Number: ARA-09-15-10 **Amendment Number:** 05

Study Title: **Magnitude of the Residue of 2,4-D and Quizalofop-P-ethyl in/on Herbicide Tolerant Field Corn Containing the Aryloxyalkanoate Dioxygenase-1 (AAD-1) Gene**

Description of Change:

1) The Lead Quality Assurance for the Study is changed from Gene Burnett to Jane W. Brown from Ag Research Associates. Jane's contact information is;

   Jane W. Brown
   Ag Research Associates, LLC
   7 Grant Court
   Hamilton, NJ 08619
   Phone; (609) 587-4625

Reason for Change:

1) The Lead QA Auditor/Manager responsibilities were reassigned to Jane Brown of ARA.

Effect of Change:

1) There is no negative impact on the study. Jane Brown is qualified and competent to serve as Lead QA for this study.

Effective Date: 20Nov09

Approved By:

Study Director: John M. Becker Date: 21Nov09
John M. Becker, PhD
Ag Research Associates

Sponsor Study Monitor: Steve W. Rosser Date: 24-Nov-2009
Steve W. Rosser, PhD
Dow AgroSciences

Ag Research Associates
1730 Denham Road
Sycamore, GA 31790

PROTOCOL AMENDMENT

Study Number: ARA-09-15-10 **Amendment Number:** 06

Study Title: Magnitude of the Residue of 2,4-D and Quizalofop-P-ethyl in/on Herbicide Tolerant Field Corn Containing the Aryloxyalkanoate Dioxygenase-1 (AAD-1) Gene

Description of Change:

1) Clarification is provided for the Food/Feed Commodity Processing phase (Section 9).

   a. For aspirated grain fraction (AGF) generation, an approximate 650-pound sample of RAC grain from each treatment (T1, T4, T5) will be prepared for aspiration, dust will be generated and aspirated from the bulk grain, dust will be classified (by size), ash content determined, and then dust recombined (after discussion with Study director) into the AGF sample. A single AGF sample will be about 1 kg, or all available mass, from each treatment.

   b. After generation of the AGF (after aspiration and screening) and prior to separation for milling, cleaned whole grain will be sampled in triplicate from each treatment. This is an added commodity and is in addition to the pre-AGF, "from the processor" whole grain RAC sample already specified in the protocol. Each cleaned grain sample will weigh approximately 1 kg.

   c. "Refined oil" from each wet- and dry-milling process will be alkali-refined. Each alkali-refined oil sample will be approximately 400g (versus about 1 liter currently noted in protocol) and duplicate samples will be collected from each treatment.

   d. For Sample Destruction and Cleanup (Section 9.6), any excess vegetable oil will be discarded according to Texas State regulations and not used as food or feed. Any remaining regulated whole seed (to include floor waste) will be contained and ground to devitalize the seed so that it cannot persist or propagate. Any excess commodity byproducts, including the ground, devitalized seed, will be composted at a local public landfill and not used as food or feed. Appropriate signage with USDA permit number will be displayed at the Processing facility.

Reason for Change:

1) Details about the procedures, samples, and sample size for the food/feed commodity processing phase are clarified.

Effect of Change:

1) There is no negative impact on the study. Details on the procedures followed by GLP Technologies and samples expected from the Food/Feed Commodity Processing phase are clarified.

Effective Date: 30Nov09

Approved By:

Study Director: John M. Becker Date: 30Nov09
John M. Becker, PhD
Ag Research Associates

Sponsor Study Monitor: for S. Rosser Date: 03 Dec. 2009
Steve W. Rosser, PhD
Dow AgroSciences

Ag Research Associates

1730 Denham Road
Sycamore, GA 31790

PROTOCOL AMENDMENT

Study Number: ARA-09-15-10 **Amendment Number:** 07

Study Title: **Magnitude of the Residue of 2,4-D and Quizalofop-P-ethyl in/on Herbicide Tolerant Field Corn Containing the Aryloxyalkanoate Dioxygenase-1 (AAD-1) Gene**

Description of Change:

1) The Study Director Management for the Study is changed from David Stephens to Tim Case from Ag Research Associates (ARA). Tim's contact information is;

   Tim Case
   Ag Research Associates, LLC
   P.O. Box 455
   Groom, TX 79039
   Telephone; (806) 277-0164
   E-mail; tim.case@agresearchassociates.com

Reason for Change:

1) Study Director Management responsibilities within ARA were reassigned to Tim Case.

Effect of Change:

1) There is no negative impact on the study. Tim Case is responsible for oversight of the ARA Study Director.

Effective Date: 01Apr10

Approved By:

Study Director: John M. Becker Date: 01 Apr 10
John M. Becker, PhD
Ag Research Associates

Sponsor Study Monitor: Steve W. Rosser Date: 05-Apr-2010
Steve W. Rosser, PhD
Dow AgroSciences

Ag Research Associates

1730 Denham Road
Sycamore, GA 31790

PROTOCOL AMENDMENT

Study Number: ARA-09-15-10 **Amendment Number:** 08

Study Title: Magnitude of the Residue of 2,4-D and Quizalofop-P-ethyl in/on Herbicide Tolerant Field Corn Containing the Aryloxyalkanoate Dioxygenase-1 (AAD-1) Gene

Description of Change:

1) The transgenic seed line DAS-40474-7 (AAD-1 Event DAS-40474-7) will not be developed for commercialization by the Sponsor. This seed line was being tested in Treatment Plots T3 (RAC trials) and T5 (processing trials) of this study. Therefore, further effort on this seed line will be discontinued. The field and processing phase data for these treatments collected to-date will be summarized as needed, but not used and may not be fully developed or reported in comparison to the remaining seed line (Event AAD-1 DAS-40278-9 used in Treatment Plots T2 and T4). Analytical work on samples generated from plots T3 and T5 will not be done.

Reason for Change:

1) The Sponsor discontinued development of the DAS-40474-7 seed line for commercial and business reasons.

Effect of Change:

1) The final study report will include a summary of field data and processed commodities for Treatment Plots T3 and T5, but may not be fully reported, Quality Assurance audited, or GLP-compliant (noted on GLP Compliance Page as appropriate). The final study report will not include any analytical data for samples from Treatment Plots T3 and T5 since none will be generated. The final study report will include field, processing, and analytical information on the remaining seed line (Event AAD-1 DAS-40278-9) used in Treatment Plots T2 and T4.

Effective Date: 04May10

Approved By:

Study Director: John M. Becker Date: 04 May 10
John M. Becker, PhD
Ag Research Associates

Sponsor Study
Monitor: Steve W. Rosser Date: 10-May-2010
Steve W. Rosser, PhD
Dow AgroSciences

Ag Research Associates

1730 Denham Road
Sycamore, GA 31790

PROTOCOL AMENDMENT

Study Number: ARA-09-15-10 **Amendment Number:** 9

Study Title: **Magnitude of the Residue of 2,4-D and Quizalofop-P-ethyl in/on Herbicide Tolerant Field Corn Containing the Aryloxyalkanoate Dioxygenase-1 (AAD-1) Gene**

Description of Change:

The Study Director is changed from John Becker to John F. Culligan of ARA. Personnel and Contact Information on page 2 of the protocol is being modified to:

Study Director and Testing Facility:	
John "Jack" F. Culligan Ag Research Associates, LLC 1730 Denham Road Sycamore, GA 31790	Office Address and Contacts; 38 Holt Avenue Hamilton, NJ 08619-1602 Telephone: (609) 890-1136 Mobile: (609) 647-5723 FAX: (609) 890-6552 E-Mail: jack.culligan@agresearchassociates.com

Reason for Change:

Study Director responsibility within ARA was reassigned to John F. Culligan. John Becker is leaving the employ of ARA.

Effect of Change:

There is no negative impact on the study. John F. Culligan is responsible for overall conduct of the study as Study Director.

Effective Date: 24-May-2010

Approved By:

Study Director: [signature] Date: 26-May-2010
John F. Culligan
Ag Research Associates, LLC

Sponsor
Study Monitor: [signature] Date: 25-May-2010
Steve W. Rosser, PhD
Dow AgroSciences LLC

Protocol Amendment #10

Analysis Facility: Dow AgroSciences LLC
Regulatory Sciences and Government Affairs
Indianapolis Lab, USA

Test Facility: Ag Research Associates, LLC
Sycamore, GA 31790

Protocol Number: ARA-09-15-10
Amendment Number: 10 Amendment Date: 08-JUN-2010

Study Title: Magnitude of the Residue of 2,4-D and Quizalofop-P-ethyl in/on Herbicide Tolerant Field Corn Containing the Aryloxyalkanoate Dioxygenase-1 (AAD-1) Gene J.C. 08-JUN-2010

Title of Protocol Section to be Changed:

11. ANALYTICAL PHASE

Reason and Rationale:

The protocol states that the Analytical Phase will be added by Protocol Amendment in order to carry out the analytical phase of the study. This amendment supplies supplemental information as required by the protocol. Appropriate methodology and procedures used to generate the analytical data for this study are listed.

Amended Protocol Section:

Analysis of the concentrations of 2,4-D, 2,4-DCP, quizalofop ethyl ester and quizalofop acid found in/on corn raw agricultural commodities provided by the Principal Field Investigators and in/on grain and the associated processed fractions provided by the Sample Processing Principal Investigator during this study will be carried out by the Analysis Facility under local control of the Principal Analyst.

The Principal Analyst is being changed from Gary Schelle (Protocol Amendment #2) to Brian Wendelburg. The contact information for Brian Wendelburg is listed below:

Brian M. Wendelburg
Dow AgroSciences LLC
9330 Zionsville Road, Bldg. 306 / A2
Indianapolis, IN 46268
Phone: (317) 337-3649
e-mail: bwendelburg@dow.com

1. Analytical Methods

Residues of 2,4-D and the 2,4-DCP metabolite and quizalofop ethyl ester and the quizalofop acid found in/on samples of corn raw agricultural commodities and samples of the processed fractions produced in the study will be analyzed using procedures developed at the Analysis Facility. For the determination of 2,4-D and the 2,4-DCP, "Proposed Procedure for the Analysis of 2,4-D and 2,4-DCP in Corn and Soybean Matrices by LC/MS/MS" will be used. For the determination of quizalofop ethyl ester and quizalofop acid, "Proposed Procedure for the Analysis of Quizalofop Ethyl Ester and Quizalofop Acid in Corn and Soybean Matrices by LC/MS/MS" will be used. Sufficient numbers of concurrent recovery samples will be analyzed over the course of the study to validate both of the analytical methods. Instrumental settings and procedural details for these methods will be documented in the study file.

2. Analytical Standards/Internal Standards

Analyte Name	AGR/TSN Number	Percent Purity	Certification Date	Reference
2,4-D[a]	AGR275828	99.5	21-Jan-2008	FAPC07-152704
2,4-DCP[a]	AGR182992	99	02-Oct-2007	FAPC073503
2,4-D M+6 IS[b]	SCHF-006	≥98	14-Sep-2007	--[c]
2,4-DCP M+6 IS[b]	SCCJ-003	≥98	29-Oct-2002	--[c]
Quizalofop ethyl ester[a]	TSN106317	99	28-Aug-2009	FAPC09-225778
Quizalofop acid[a]	TSN106172	96	17-Nov-2009	FAPC09-227492

[a] Test Substance/ Reference Standard
[b] Stable Isotope Internal Standard
[c] Commercial source - Cambridge Isotope Laboratories, Inc.

No correction for purity will be made if the purity is greater than or equal to 96%. If the analytical standards are recertified during the course of the study and the assay is lower than 96%, the standard solutions prepared after the new assay date will be corrected for the lower purity value.

3. Procedures

a. Sample Identification

Each sample will be identified with a unique sample identifier at the Analysis Facility. This number will be used to track the sample during analysis and reporting.

b. Analytical Set

An analytical set will consist of calibration standards, a reagent blank, fortified controls, and study samples. With few exceptions, each analytical set should contain, at a minimum:

1 control sample fortified at the study limit of detection (LOD) and
1 control sample fortified at the study limit of quantitation (LOQ).

The control sample fortified at the study LOD is intended to demonstrate detection (generally, at least three times baseline) at that level and is not to be used to calculate a percent recovery of analyte from that sample. If detection is not demonstrated at that level, all study samples in the set must be reanalyzed with the following exception: if there are study samples with residue concentrations at or higher than a level at which a recovery sample was successfully analyzed, the concentrations for those study samples may be reported and do not require reanalysis, as determined by the Principal Analyst.

If a recovery sample is "lost" (due to spilling, glassware breakage, or other unavoidable event) during analysis, it is still considered to have been part of the analytical set. Acceptance of the results from that set is at the discretion of the Principal Investigator and will be approved by the Study Director.

Analytical set average recovery values should fall within 70-120% of the expected value based upon the fortification level. If the average recovery values are outside these limits, then the samples that were analyzed together with the recovery set may be reanalyzed at the discretion of the Principal Investigator with the approval of the Study Director.

Residue values obtained for field samples will be reported without correction for recovery.

c. Limit of Quantitation, Limit of Detection, and Fortification Concentrations

Limit of	Concentration
Detection	0.003 µg/g
Quantitation	0.010 µg/g

The fortification levels and minimum number of fortified control samples over the course of the study will be as follows:

1 at the LOD
6 at the LOQ
5 at a concentration to cover the high end of residues or at 10X the LOQ if no residues are detected.

d. Preparation and Maintenance of Field Samples

After receipt of the field samples from the Test Facility or processed fractions from the processing facility, the samples will be stored frozen and prepared (if required) according to Dow AgroSciences LLC SOP ECL-21. The samples may be temporarily stored refrigerated at times during preparation or analysis.

e. Statistical Methods

Means, standard deviations, and regression analysis will be used to evaluate data from these analyses. Outlier data may be statistically evaluated using appropriate outlier tests such as the student-T or Q-tests. If outlier statistics are used, it will be documented and placed in the study file and reported in the final report.

4. Amendments and Deviations

Any anticipated amendments of the protocol at the analytical facility will be communicated to the Study Director by the Principal Analyst. The Study Director will determine if an amendment is needed. Any deviations arising at the analytical facility must be communicated to the Study Director by the Principal Analyst as soon as possible after the deviation occurs.

5. Analytical Phase Summary

An analytical summary will be written by the Principal Analyst and reviewed by the Study Director. The analytical summary will include a signed statement from the Principal Analyst attesting to the GLP compliance status of the analytical phase of the study.
The DAS QAU will audit all analytical raw data and the analytical summary.

The analytical phase summary will be appended to the final study report.

Residue results will be expressed as uncorrected values (i.e., not corrected for concurrent set recoveries) or reported sample concentrations. The reported concentration is the uncorrected value with the LOD and LOQ applied. The residue concentration will be reported as ND (Not Detected) if the uncorrected concentration is below the LOD. The residue concentration will be reported as the uncorrected concentration in parentheses ( ) if the concentration is less than the LOQ but greater than or equal to the LOD. The residue concentration will be reported as the uncorrected concentration if the uncorrected concentration is equal to or greater than the LOQ. Residue concentrations reported between the LOD and LOQ have a higher degree of uncertainty than values above the LOQ.

6. Good Laboratory Practice (GLP)

The analytical facility will carry out any testing according to GLPS, as given in Section 15. QUALITY ASSURANCE, of the study protocol.

7. Quality Assurance

   Quality Assurance monitoring and reporting for the analytical phase will be the responsibility of the QAU of Dow AgroSciences LLC.

   The analytical facility QAU will also provide a statement in the analytical summary listing when inspections and audits were performed and when the findings were reported to the Study Director and Study Director Management.

8. Study File-Analytical Data

   The analytical study file will contain, at a minimum, sample preparation codes, sample chain of custody, a copy of the residue methods, analytical data sets, reference standard information (which includes TSN database printouts, verified copies of dispensation records, and preparation of standard solutions), notebooks (or verified and signed copies), final results spreadsheets, and the original analytical summary report. At the conclusion of the analytical phase of the study, the analytical summary and all of the associated raw data will then be transferred to the Sponsor's Archivist (currently June Whitney) and will be archived in the Dow AgroSciences Archives in Indianapolis, Indiana.

Impact on the Study:

It is considered that this amendment will have no adverse effect on the integrity of the study.

Approval:

Brian M. Wendelburg	04 June 2010
Principal Analyst	Date
Dow AgroSciences LLC	
Tel: (317) 337-3649	
Email: bwendelburg@dow.com	
John F. Culligan	08-Jun-2010
Study Director	Date
Ag Research Associates, LLC	
Tel: (908) 968-0736	
Email: jack.culligan@agresearchassociates.com	
Steve W. Rosser	03-Jun-2010
Study Monitor	Date
Dow AgroSciences LLC	

Ag Research Associates

1730 Denham Road
Sycamore, GA 31790

PROTOCOL DEVIATION

Study Number: ARA-09-15-10 **Deviation Number:** 01

Study Title: Magnitude of the Residue of 2,4-D and Quizalofop-P-ethyl in/on Herbicide Tolerant Field Corn Containing the Aryloxyalkanoate Dioxygenase-1 (AAD-1) Gene

Description of Changes:

1) In Trial 05, a spray solution mixing error (1000ml of the required water was omitted) occurred on the first pass of Application #2 to Treatment Plot #2, resulting in about 167% of the target rate. The error was realized at the end of the first pass and the tank-mix was re-prepared for the second pass to Treatment Plot #2. Therefore, half of the plot received the over-rate application, but half of the plot received the correct rate. The plot was of sufficient size to bisect it and collect samples from only the second half that received the correct application rate.

2) For Trial 01, the test site was established with a 10-foot bare/fallow perimeter buffer and electric fence, versus the 6 rows of corn requested in the protocol.

3) For Trial 01, Treatment Plot #2 and Treatment Plot #3 were 20 feet apart instead of a minimum of 25 feet separation as indicated in the protocol.

4) A notation about the presence of soil and plant surface moisture (dew) at the times of application was not made for Trial 11.

5) The required ~40-day forage samples for Trial 14 were not collected.

6) Stover samples for Trials 11, 21, and 25 were not 1 kg as requested in Section 8.4.

7) The Field Trial Notebook and Field Summary were not completed within about one month after the last sample shipment for several trials, including Trials 07, 11, and 19.

8) An in-progress, critical phase QA audit was not completed for Trials 04, 11 and 13 according to Section 15.

Reason for Changes:

1) The PFI added only part of the required tank-mix water, but realized the error after the first pass (due to low volume remaining in the spray tank).
2) The plot location was limited by space in order to fit the necessary treatment plots so the perimeter buffer was reduced, and electric fence was added as an animal deterrent.
3) The plot location was limited by space in order to accommodate the necessary perimeter buffer.
4) The PFI failed to record this information at application events.
5) The PFI erred and did not understand the protocol requirement for forage collection.
6) The PFI collected at least 12 plants but due to the mass of the dried stover, they were not a minimum of 1 kg.
7) The PFI submitted the Field Trial Notebook and Field Summary about 2-3 months after the last sample shipment. More time was required to complete the notebook with facility and historical information and have it audited by QAU.
8) An in-progress audit for these trials could not be scheduled and/or was not performed.

Effect of Changes:

1) There is no impact to the study. The PFI collected samples from the half of the plot that received the correct application rate.
2) There is no impact on the study. The fallow buffer and electric fence was deemed acceptable by the Sponsor as equivalent to the 6 rows of perimeter corn.
3) There is no impact on the study. Both Treatment Plots #2 and #3 received the test substance and differed only in the seed line (Event) so there is not potential for contamination.
4) Minimal impact to the study is expected. The forage samples from Trial 14 are not available for analysis. Forage samples from other trials in the study will be adequate to assess the residues in/on forage.
5) Minimal impact to the study is expected. Information about the satisfactory conditions at each application can be inferred by the existing weather data and time-of-day data that were recorded.
6) No impact to the study is expected. The sample size was representative of the entire treatment plot and of sufficient size for analysis.
7) There is no impact on the study. The delayed submission of the Field Trial Notebook and Field Summary did not affect completion of the study.
8) Minimal impact to the study is expected. The trial was completed satisfactorily by a qualified PFI and other Trials conducted by this PFI were audited during the field phase. The GLPs require a QA inspection at the study level, not for each trial.

Effective Date: 04May10

Approved By:

Study Director: John M. Becker Date: 05May10
John M. Becker, PhD
Ag Research Associates

Sponsor Study Monitor: Steve W. Rosser Date: 10-May-2010
Steve W. Rosser, PhD
Dow AgroSciences

DEVIATION FORM

GLP Technologies, 22723 State Highway 6 South, Navasota, Texas 77868

Protocol Deviation Number 02

Sponsor: Dow AgroSciences, LLC
Testing Facility: Ag Research Associates LLC
Ag Research Associates Protocol No. ARA-09-15-10
Commodity: Corn

17 May 10 JMB

Deviation from: **Protocol Section 9.3 "Expected Processed Food/Feed Samples"**

DETAILS OF DEVIATION: Protocol requested approximately 1 liter of refined oil be collected. A liter of oil weighs approximately 800 grams. Refined oil samples from the dry milled portion for both trials failed to meet this requirement. Samples from NE-14 ranged from 370-409 grams and samples from GA-03 ranged from 303-361 grams.

REASON FOR DEVIATION: Typically, softer varieties of corn are wet milled. Varieties with medium to hard endosperm are dry milled. In GLP Technologies' particular situation, we have one variety of corn to wet and dry mill. Consequently, either the wet or dry mill procedure will produce more oil.

CORRECTIVE ACTION TAKEN: Reported deviation with this form.

AFFECT ON PROCESSING: None. Wet milled samples met the requirements.

Process Principal Investigator's Signature — May 12, 2010 Date

STUDY DIRECTOR NOTIFICATION

Notified: John M. Becker (john.becker@agresearchassociates.com) on May 12, 2010

IMPACT ON STUDY: No impact on study is expected. The oil samples were adequate for analysis and were representative of the trial.

John M. Becker — 17 May 10
Study Director's Signature — Date

Original sent to Study Director for completion of "Impact on Study" and signing.

DEVIATION FORM

GLP Technologies, 22723 State Highway 6 South, Navasota, Texas 77868

Sponsor: Dow AgroSciences, LLC
Testing Facility: Ag Research Associates LLC
Ag Research Associates Protocol No. ARA-09-15-10
Commodity: Corn

Protocol Deviation Number 03
17May10 JMB

Deviation from: Protocol Section 9.7 "Sample Processing Summary"

DETAILS OF DEVIATION: Protocol requested report and raw data be completed and sent to the Study Director within approximately 6 weeks after all processed fractions are shipped to Dow. Last fraction shipment was March 2, 2010. Report and data were not completed by April 13, 2010.

REASON FOR DEVIATION: Principal Investigator error.

CORRECTIVE ACTION TAKEN: Reported deviation with this form.

AFFECT ON PROCESSING: None.

Dick Dused
Process Principal Investigator's Signature

May 12, 2010
Date

STUDY DIRECTOR NOTIFICATION

Notified: John M. Becker (john.becker@agresearchassociates.com) on May 12, 2010

IMPACT ON STUDY: No impact on study. The delayed processing phase summary did not impact completion of the study.

John M. Becker
Study Director's Signature

17 May 10
Date

Original sent to Study Director for completion of "Impact on Study" and signing.

Protocol Deviation #~~5~~ 4
① BMW 04Oct2010

Analysis Facility: Dow AgroSciences LLC
Regulatory Sciences and Government Affairs
Indianapolis Lab, USA

Test Facility: Ag Research Associates, LLC
Sycamore, GA 31790

Protocol Number: ARA-09-15-10
Deviation Number: ~~5~~ 4 Deviation Date: 01-Oct-2010
①BMW 04Oct2010

Study Title: Magnitude of the Residue of 2,4-D and Quizalofop-P-ethyl in/on Herbicide Tolerant Field Corn Containing the Aryloxyalkanoate Dioxygenase-1 (AAD-1) Gene

Title of Protocol Section Associated with Deviation:

11. ANALYTICAL PHASE
    1. Analytical Methods

Reason and Rationale:

The method for 2,4-D and 2,4-DCP outline the use of a 0.45 µm PTFE filter in Step 11. Several analytical sets used 0.2 µm PTFE filters due to availability issues.

Impact on the Study:

It is considered that this deviation will have no adverse effect on the integrity of the study. It is common to substitute equipment and supplies as needed as long as appropriate scientific evaluation is taken to ensure suitability. Recoveries for the samples which used different filters were not significantly different from those which used the filter noted in the method.

Approval:

Brian M. Wendelburg 04Oct2010

Brian M. Wendelburg
Principal Analyst
Dow AgroSciences LLC
Tel: (317) 337-3649
Email: bwendelburg@dow.com

Date

Protocol Number ARA-09-15-10
Sponsor's PTR/Ref. No. 10001686-000-90411-0001
Deviation Number ~~5~~ 4 BMW 04Oct2010

John F. Culligan
Study Director
Ag Research Associates, LLC
Tel: (908) 968-0736
Email: jack.culligan@agresearchassociates.com

Date

Steve W. Rosser

Steve W. Rosser
Study Monitor
Dow AgroSciences LLC

04-Oct-2010
Date

Protocol Number ARA-09-15-10 Page 2 of 2
Sponsor's PTR/Ref. No. 10001686-000-90411-0001
Protocol Deviation No. ~~8~~ 4
⓪ Fixed after exact copy BMW 04Oct10

01 - Oct - 2010

John F. Culligan
Study Director
Ag Research Associates, LLC
Tel: (908) 968-0736
Email: jack.culligan@agresearchassociates.com

Date

Steve W. Rosser
Study Monitor
Dow AgroSciences LLC

Date

THIS IS AN EXACT COPY OF
THE ORIGINAL DOCUMENT
BY BMW
DATE 04Oct2010

Protocol Number ARA-09-15-10
Sponsor's PTR/Ref. No. 10001686-000-90411-0001
Protocol Deviation No. ~~3~~ 4 JC. (EE) 04-OCT-2010

John F. Culligan
Study Director
Ag Research Associates, LLC
Tel: ~~(908) 968-0736~~ JC. (EE) 04-OCT-2010 (609)-890-1136
Email: jack.culligan@agresearchassociates.com

01-OCT-2010
Date

Steve W. Rosser
Study Monitor
Dow AgroSciences LLC

Date

Ag Research Associates

1730 Denham Road
Sycamore, GA 31790

PROTOCOL DEVIATION

Study Number: ARA-09-15-10 **Deviation Number:** 05

Study Title: ***Magnitude of the Residues of 2,4-D and Quizalofop-P-ethyl in/on Herbicide Tolerant Field Corn Containing the Aryloxyalkanoate Dioxygenase-1 (AAD-1) Gene***

Description of Deviations:

1. Section 6.8 of the protocol states that monthly average maximum and minimum temperatures will be collected and recorded in the field trial notebook (FTNB). For all trials, the monthly average maximum and minimum temperatures were not consistently recorded and reported during the field phase of the study (at times, only the maximum and minimum were recorded, not the average of the maximum and minimum temperatures).

2. Section 6.8 of the protocol states that the presence of soil surface moisture and plant surface moisture (dew) will be recorded at each application. For all trials, the presence of soil moisture (dew) was not consistently recorded in the FTNB.

3. Section 7.0 of the protocol states that a non-ionic adjuvant, such as X-77, at approximately 0.25% (v/v) will be added in the spray mixture at each application of 2,4-D. For Trial 06, Treatments 2 and 3 for the application performed on 26 Jun 2009, X-77 adjuvant was used at 0.18% rather than at the protocol specified 0.25%.

4. Section 7.0 of the protocol states that crop oil concentrate (COC), such as Agridex, at approximately 1.25% (v/v), will be added in the spray mixture with quizalofop-P-ethyl. For Trial 08, Treatments 2 and 3 for the application performed on 04 Aug 2009, the adjuvant should have been a crop oil concentrate rather than X-77 per protocol.

5. Section 7.3 of the protocol states that in the RAC and RAC + Decline treated plots there will be 2 broadcast spray applications of 2,4-D, each at approximately 1.0 pound acid equivalent/acre (~1120 g acid equivalent/ha), one application of quizalofop-P-ethyl at approximately 0.082 pound AI/acre (~92 g/ha), and one application of 2,4-D at approximately 1.0 pound acid equivalent/acre (~1120 g acid equivalent/ha) and each application should fall within the range of 95 -110% of the target rate. For Trial 10, Treatment 3 for the application of quizalofop-P-ethyl

performed on 08 Aug 2009 the rate is 0.077 lb ai/A, which is outside of the acceptable 95 – 110% range required by the protocol.

6. Section 7.4 of the protocol states that application 2 should be made 12 days (± 1 day) before application 4. For Trials 22 and 23, Treatment 2 the applications performed on 13 and 14 Jul 2009, respectively, were 10 days rather than the acceptable protocol range of 11 – 13 days before application 4.

7. Section 8.4 of the protocol states that the first timing of samples of forage is to be targeted at 40 days (± 2 days) after the final application of 2,4-D. At Trial 04, Trial 22, and Trial 23, the actual DAT for the 40 DAT forage samples was 30 DAT which is out of the 38 – 42 DAT range listed in the protocol.

8. Section 8.7 of the protocol states that field corn forage, grain, and stover RAC and decline samples must be chilled (cooler with blue, dry or wet ice, or freezer) within 4 hours of collection and placed in a freezer (with temperature monitoring, maintained on average below 0°C, typically near -20 °C) within 8 hours of collection in order to maintain their integrity. For Trial 18, samples 096-0001 to 098-0005 samples were not frozen within 4 hours of collection as required by the protocol but frozen in ~ 6 – 7 hours of collection.

9. Section 8.11 of the protocol states that the PFI will within approximately one month after the last sample shipment, send by secure service the field trial summary and supporting field trial notebook and data package (complete GLP field trial notebook paper forms containing original raw field data and documentation) to the Study Director or Testing Facility for review. For Trials 06 and 20, the FTNB and summary were not returned to the Study Director within one month of sample shipment.

10. In Section 8.4 of the protocol, it states that field corn stover samples will be collected on the same day as RAC grain samples and consist of a composite sample of a minimum 1 kg. In Trial 25, sample weights for the untreated and Treatment 2 stover samples were less than the required 1 kg.

11. In Section 15.0 of the protocol, it states that critical phase inspections will be conducted and reports will be forwarded to the Study Director promptly for review. Critical phase inspections for Trials 14, 15, 16, 19 and 20 were not reported to the Study Director and Study Director Management promptly as required by the protocol. There were no significant findings from these inspections, however.

12. In Section 15.0 of the protocol, it states that GLP compliance inspections will be performed by the local trial QAU at a frequency to ensure the integrity of the trial. No critical phase field inspection was performed for Trials 08, 09 and 10. The protocol requires that when a PFI (as in this case) has multiple trials, a critical phase

inspection should be done on at least one trial but it is not necessary to perform a critical phase inspection of every trial.

Reason for Deviations:

All deviations listed were a result of PFI oversight.

Effect of Deviation:

There is minimal impact to the overall study as a result of these protocol deviations.

Approved By:

Study Director: ______________________ Date: 12-Oct-2010
John F. Culligan
Ag Research Associates, LLC

Sponsor
Study Monitor: ______________________ Date: 08-Oct-2010
Steve W. Rosser, PhD
Dow AgroSciences LLC

Ag Research Associates

1730 Denham Road
Sycamore, GA 31790

PROTOCOL DEVIATION

Study Number: ARA-09-15-10 **Deviation Number:** 06

Study Title: ***Magnitude of the Residues of 2,4-D and Quizalofop-P-ethyl in/on Herbicide Tolerant Field Corn Containing the Aryloxyalkanoate Dioxygenase-1 (AAD-1) Gene***

Description of Deviation:

Several individual method recovery values for 2,4-DCP and quizalofop acid were outside the acceptable range of 70-120% listed in protocol amendment 10 (page 3 of the amendment).

Reason for Deviation:

The out of range values are being accepted as reliable method recoveries since the overall method average recovery is within the acceptable range.

Effect of Deviation:

There is minimal impact to the overall study as a result of this protocol deviation.

Approved By:

Study Director: ____________________ Date: 05-Nov-2010
John F. Culligan
Ag Research Associates, LLC

Sponsor
Study Monitor: ____________________ Date: 02-Nov-2010
Steve W. Rosser, PhD
Dow AgroSciences LLC