

# **Supporting document 2**

# ASSESSMENT OF MICROBIOLOGICAL HAZARDS ASSOCIATED WITH THE FOUR MAIN MEAT SPECIES

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## **Executive Summary**

As part of Food Standards Australia New Zealand's proposal to assess whether a Primary Production and Processing Standard for Meat and Meat Products was required, FSANZ identified hazards that may be found in meat, where in the meat supply chain they may be introduced into the animal or the meat and where in the supply chain they may be controlled.

This report identifies hazards (both identified and potential) that may be associated with meat from the four main meat species (cattle, sheep, goats and pigs), and lists pathogenic microorganisms that, if unmanaged, present or may potentially present a risk to public health. The information has been derived from industry data, microbiological analyses and published scientific data. The document does not attempt to document the severity of illness presented by these hazards, nor does it determine the likelihood of their occurrence in the final meat product or characterise the risk they may present. The report does however review meat associated foodborne disease evidence in Australia.

A range of potential hazards have been identified along the production and primary processing chain. Limited, if any, prevalence and incidence data is available for these hazards in meat. Given the lack of epidemiological evidence also available, it would suggest that the likelihood of these hazards causing illness from consumption of meat is quite low.

The principal microbiological hazards associated with the four main animal species are:

Animal	Principal microbiological hazard
Cattle	Pathogenic Escherichia coli, Salmonella spp., Campylobacter jejuni and C. coli,
Sheep	Pathogenic Escherichia coli and Salmonella spp.
Goats	Pathogenic Escherichia coli and Salmonella spp.
Pigs	Salmonella spp., Yersinia enterocolitica and Y. pseudotuberculosis, Toxoplasma gondii, Campylobacter jejuni and C. coli.

During the animal production phase, there are a number of key inputs and activities which influence the manner in which hazards may be introduced or amplified. They are summarised below:

Input and/ or activity	Comment	Step in chain where control may be applied
Animal Health	Pathogens may exist in the animal with or without exhibiting clinical signs	Animals with clinical signs of disease or illness are identified and managed at:  • Dispatch from farm/saleyard  • Arrival at abattoir  • Ante-mortem inspection  Without clinical signs, potential hazards may be identified and managed at:  • Slaughter to minimise contamination from external surfaces or internal spillage  • Post-mortem inspection
Stress	Animals may be more susceptible to infection and/or have increased faecal	<ul><li>Minimise exposure of animals to stress during:</li><li>Transport</li><li>Lairage</li></ul>

Input and/ or activity	Comment	Step in chain where control may be applied
	shedding. Pathogens colonise the gut	
Feed	Feed has the potential to introduce pathogens into the gut or environment	Management of input of manure and fertiliser onto pasture Control supplements Oversight of ensilage operations
Water	Contributes to internal and external contamination	Access of animals to suitable drinking water
Environment and management of biosecurity	Pathogens may contaminate external surfaces of animal, or can lead to ingestion or infection of the animal	Pasture management Vermin and pest control Good agricultural practices Sound animal husbandry

During the primary processing stage there are two main sources of contamination to the meat carcass:

- External contamination: from the animal (hide, skin, fleece, hooves, faeces, etc) and the environment (including personnel), and
- Internal contamination: during evisceration and dressing operations and where the spillage of gastrointestinal tract contents occurs.

The burden of illness that may be attributed to meat and meat products was assessed by evaluating OzFoodNet outbreak data. Sixty-six outbreaks of foodborne illness associated with meat products in Australia were reported to OzFoodNet between January 2003 and June 2008. More recent data drawn from published OzFoodNet reports¹ indicate 42 meat-associated outbreaks were reported between June 2008 and December 2011. While the data demonstrates the occurrence of outbreaks involving meat, they are usually due to dishes containing a meat product. Attribution to a specific meat source is either limited or difficult to establish with any confidence. Where meat products have been implicated in foodborne illness, generally these were further processed products and the most common causative microorganisms were *Salmonella* serotypes, *Clostridium perfringens* and *Staphylococcus aureus*. The undercooking of meat and temperature abuse after cooking were the major causes of meat-associated outbreaks.

The findings of this assessment are consistent with the significant body of evidence that exists for the Australian domestic meat industry indicating that domestically-reared red meat (cattle, sheep, goats) and pigs, processed under existing standards, present a low risk to public health. Also evidenced is that industry personnel are mature in their knowledge and management of food safety risks.

Considerable data are available to support the safety of meat and meat products produced from beef, sheep and pork in Australia. The evidence suggests that Australian meat from these species has a low microbial load and generally low prevalence of pathogens. Many of the pathogens listed in this assessment occur infrequently or not at all on Australian meat.

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<sup>&</sup>lt;sup>1</sup> OzFoodNet Annual (2008, 2009, 2010) and Quarterly (2011) reports available at: http://www.ozfoodnet.gov.au/internet/ozfoodnet/publishing.nsf/Content/reports-1

## **Background**

Food Standards Australia New Zealand (FSANZ) has responsibility for protecting the health and safety of consumers through the development of food standards. The FSANZ Act requires FSANZ, when developing or varying standards, to have regard to "the need for standards to be based on risk analysis using the best available scientific evidence".

The development and application of a Primary Production and Processing Standard for Meat and Meat Products will be dependent on an analysis of the public health and safety risks, economic and social factors and current regulatory an industry practices. The analysis of the public health and safety risks will be based on a comprehensive scientific assessment of public health hazards associated with the consumption of meat.

FSANZ uses a number of methodologies to assess hazards, including risk profiling, quantitative and qualitative assessments and scientific evaluations. The methodology utilised depends on the purpose of the assessment and on the availability, quality and quantity of data.

The assessment will consider all stages in the meat supply chain, from the growing environment through to primary processing. In undertaking the assessment, FSANZ will utilise available information including current microbiological and chemical surveillance data, epidemiological data, consumption data and existing published and unpublished risk assessments from a variety of sources.

## Introduction

## **Purpose**

The purpose of this assessment document is to provide a review of the inputs and key stages of the meat supply chain for cattle, sheep, goats and pigs.

In the process of undertaking this work, the following questions are being addressed:

- What are the factors (including inputs, practices and activities and environmental factors, etc) which influence hazards at each step of the meat supply chain?
- What are the food safety hazards associated with each factor of the meat supply chain?

The hazards associated with each step in the supply chain are described and listed in a series of tables. The outputs of this evaluation will also facilitate the identification of any significant gaps in knowledge, and assist in identifying the requirement for any further risk assessment work.

## Scope

The assessment considers all stages of the meat supply chain, from the animal production environment up to the end of primary processing (*ie*: post-abattoir carcass or boning room) for the four main meat species; cattle, sheep, goats and pigs.

This assessment will identify both recognised and potential hazards but not food safety-related market access hazards as defined below:

- Recognised hazards are those where epidemiological data exists to support illness occurring as a result of consuming meat or meat products.
- Potential hazards are those hazards which may present a food safety risk from consumption of meat and meat products, but where no epidemiological evidence exists.
- Market access related hazards are those potential hazards related to food safety which are technical requirements to trade, *ie:* generic *E. coli* and Total Viable Counts.

# **Existing assessments**

A number of comprehensive scientific assessments have been undertaken in Australia on the microbiological hazards that may be found in the major meat species and the risk posed to consumers from consumption of meat and meat products. These include scientific assessments and risk-profiles generated by Meat and Livestock Australia and Australian Pork Limited.

In 2008, FSANZ commissioned a review of the domestic meat supply chain<sup>1</sup> which indicated that some sectors of the meat industry, such as domestically reared red meat (cattle, sheep and goats) and pigs are fairly mature in their knowledge and management of food safety risks.

<sup>&</sup>lt;sup>1</sup> Unpublished report, "Information, collation and review of risk assessments on meat and meat products", South Australian Research and Development Institute

Key findings of the report included:

- Considerable evidence exists supporting the microbiological and chemical safety of meat and meat products from commonly consumed species (beef, sheep and pork).
- In large part, meat associated outbreaks are a consequence of post cooking contamination or post cooking temperature abuse.
- The review of quantitative risk assessments indicates that control strategies employed closer to the consumer are more likely to have a direct and major effect on foodborne hazards.

The review notes that a large body of Australian, peer-reviewed work on red meat processing has been published over a number of decades, culminating in three national baseline studies on beef and sheep meat. These include analysis of indicator organisms such as Total Count, *Enterobacteriaceae*, Coliforms/*E. coli*, *Staphylococcus aureus* and the pathogens: *Campylobacter*, *Listeria*, *Salmonella* and Enterohaemorrhagic *E. coli* (EHEC). State based surveys have also been undertaken focused exclusively on domestic abattoirs and Very Small Plants.

The *E. coli* and *Salmonella* Monitoring (ESAM) program provides a database of over 300,000 test results for beef, sheep and pig carcasses processed at export establishments. ESAM data suggests that Australian meat from these species has a low microbial load and generally low prevalence of pathogens.

These Australian peer-reviewed and ESAM data indicate that standards of hygiene during slaughter and processing of beef, sheep and pigs in Australia are at least equal to those of major trading partners and competitors.

#### **Epidemiological Evidence**

The public health burden presented by meat and meat products in Australia was determined by examination of the epidemiological evidence assembled by OzFoodNet (Appendix 1).

The OzFoodNet Outbreak Register shows that between January 2003 and June 2008 there were 66 outbreaks associated with meat in Australia. More recent data drawn from published OzFoodNet reports<sup>2</sup> indicate 42 meat-associated outbreaks were reported between June 2008 and December 2011. The majority of outbreaks were due to dishes containing a meat product. Unfortunately attribution to a specific meat source is complex as outbreaks are usually reported as being a result of consuming a "mixed dish". Where meat products have been implicated in foodborne illness, these were generally further processed product with the causative microorganisms being *Salmonella* serotypes, *Clostridium perfringens* and *Staphylococcus aureus*. Undercooking of meat and temperature abuse after cooking are major factors in outbreaks.

<sup>&</sup>lt;sup>2</sup> OzFoodNet Annual (2008, 2009, 2010) and Quarterly (2011) reports available at: <a href="http://www.ozfoodnet.gov.au/internet/ozfoodnet/publishing.nsf/Content/reports-1">http://www.ozfoodnet.gov.au/internet/ozfoodnet/publishing.nsf/Content/reports-1</a>

Sources of foodborne illness are determined through epidemiological and/or microbiological analysis during outbreak investigations. Critical for the generation of good data is the ability to quickly identify an outbreak and initiate an investigation in order to attribute illness to a particular food. Difficulties exist because of:

- Time delays in recognition or notification of an outbreak;
- Food recall biases when attempting to gather food consumption histories;
- Long exposure windows for specific pathogens (e.g. Listeria monocytogenes);
- Reluctance of individuals to participate in investigations;
- Inability to trace food products to their source;
- Inability to obtain representative food samples for microbiological analysis; and
- A lack of precision in methods for sample analysis and pathogen identification.

It is important to recognise that outbreak data only represents a small proportion of actual cases of foodborne illness, as many outbreaks go unrecognised and/or unreported to health authorities. People do not always seek medical attention for mild forms of gastroenteritis, medical practitioners do not always collect specimens for analysis, and not all foodborne illnesses require notification to health authorities. Furthermore, most gastrointestinal illness occurs as sporadic cases with no obvious association with each other, and it can be very difficult to identify a source of infection from an investigation of a single case.

## 1. Cattle Production in Australia

#### Introduction

Traditionally, cattle production in Australia has been based upon extensive farming systems, which range from the harsh, dry climates of the north to the cooler, wetter, green pastures of southern Australia. Significant differences exist between climatic and geographical conditions, and on the species of animal grown and the production practices employed. Furthermore, beef production systems are evolving from extensive to semi-intensive and intensive units across the Australian landscape.

The Australian herd is over 28 million head of cattle, which produce around 3 million tonnes of beef and veal per annum (ABARE 2011 figures)<sup>3</sup>.

#### **Cattle Production**

The organization of beef cattle production in Australia continues to advance, reflecting improved knowledge and changing market demands. Producers are switching to cow-calf operations, producing young cattle for feedlots or the live export trade and reducing production of grass fed animals.

Within the milder climatic conditions of Southern Australia, breeds such as *Bos Taurus* are grown predominately on pasture in the mountains and plains. While in the north, native pastures such as tropical grasses, scrub land and legumes prevail and these are more suited to breeds such as *Bos indicus*. Under these conditions cattle graze on extensive open-range holdings. Extensively reared cattle entering the marketplace are generally between 15-24 months of age with average slaughter weight (dressed carcass) in excess of 260kg (ABARE, 2011). The major inputs during production are feed and water, with supplementary feeding at certain times of the year or during drought.

Importantly, there has been an increasing trend in recent years towards finishing cattle on feedlots. In 2001, approximately 26 percent of beef was finished in feedlots in south-east Queensland and New South Wales. Feedlots provide some advantages over traditional extensive cattle production, including enhanced control over quality and attributes of the carcass. Over 700 accredited cattle feedlots existed in 2009.

Until receipt at the feedlot yards, cattle finished on feedlots are initially subjected to the same production methods and inputs as extensively reared cattle. Once in the feedlot environment, cattle are more contained, restricted in their movements, are at higher stocking rates and exposed to greater environmental influences (*i.e.* environmental conditions including heat). This can cause the animal to experience an increased level of stress which may increase pathogen carriage and load, potentially increasing contamination on carcasses from any ingesta spilled during processing.

Lower slaughter ages are adopted for specialized beef systems. For example calves range from 'bobby' calves slaughtered within a few days of birth, to specially fed heavier veal calves. Bobby calves present special needs, as they are quickly separated from the cow and artificially fed, then transported on the fifth day to the slaughterhouse. Cull cow and live animals rejected from export disposition are other sub-sections of the beef industry in Australia.

The key steps in the production and processing of cattle are summarised in Figure 1.

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<sup>3</sup> Available from

http://www.daff.gov.au/abares/publications\_remote\_content/publication\_series/australian\_commodity\_statistic s?sq\_content\_src=%2BdXJsPWh0dHAIM0EIMkYIMkYxNDMuMTg4LjE3LjIwJTJGYW5yZGwlMkZEQUZGU2V ydmljZSUyRmRpc3BsYXkucGhwJTNGZmlkJTNEcGJfYWdjc3RkOWFiY2MwMDIyMDEyXzEyYS54bWwmYW xsPTE%3D

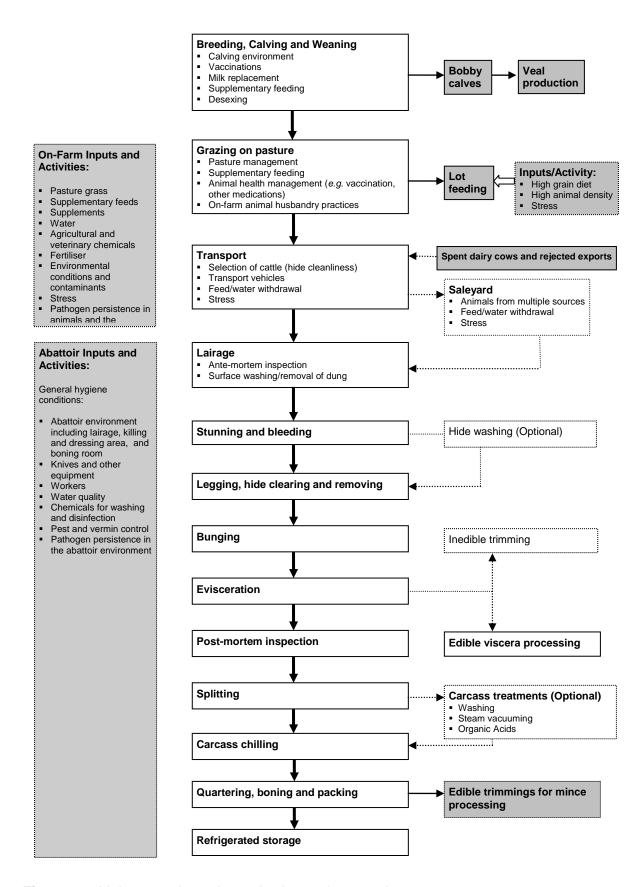


Figure 1: Major steps in cattle production and processing

## **Abattoir Operations**

Regardless of the production method utilised, once the animal is received at the abattoir gate and enters lairage, slaughtering operations are undertaken using very similar processing steps.

Minor differences may exist depending on the plant's capabilities and design but the main steps remain the same. Others factors which may influence abattoir operations include: single species or multiple species plant; age of plant; chain speed; export or domestic; and different slaughtering practices.

### **Hazard Identification**

The following tables outline the microbiological hazards that may be encountered along the cattle production and processing chain. Separate tables address the extensive and feedlot primary production methods, bobby calf production and the transport and slaughter operations.

## (a) Extensive Cattle Production

Inpu	ut/Activity	Comment	
1.	Animal Production (i	including calving, health status, zoonoses	)
1.1	Growing the cattle to market condition	Cattle may carry pathogens with or without	ut exhibiting any clinical signs.
(Ani	imal health status of	<b>Notes</b> : The following hazards may be fou of cattle:	nd in the gastrointestinal tract and exterior surfaces
the	cattle)		
		Foodborne pathogens more commonly as.	sociated with cattle include;
		Campylobacter spp.	
		Clostridium spp.	
		Pathogenic E. coli	
		Listeria monocytogenes	
		Salmonella spp.	
		Yersinia enterocolitica	
		Mycobacterium bovis	
		Brucella abortus	
		Other potential foodborne pathogens asso Yersinia pseudotuberculosis	ociated with cattle include:
		Mycobacterium avium subsp. paratuberci	doria
		Cryptosporidium parvum and C. muris	uosis Giardia lamblia
		Sarcocystis hominis	Toxoplasma gondi
		Taenia saginata	TSE agent
		Tuena sagnaia	Tob agent
		<b>Note</b> : Carrier status includes the followin	g states:
		<ul> <li>Diseased animals due to infection</li> </ul>	
		<ul> <li>Super-shedder (i.e. high levels of shed in high levels in their faece</li> </ul>	of pathogens are present in the animal's gut and are
			ent in the animal's gut contents and are therefore
		<ul> <li>Carrier (i.e. pathogens are prese shedding the bacteria into the er</li> </ul>	nt in organs but not gut contents therefore not
			ciated with handling, which could potentially be
		transmitted via meat consumption.	3,
		Notes: Examples include:	
		<ul> <li>Anthrax (Bacillus anthracis)</li> </ul>	
		<ul> <li>Melioidosis (Burkholderia pseu</li> </ul>	domallei)
		■ Q Fever (Coxiella burnetii)	
2.		es pasture, grains, concentrates and silag	
2.1	Pasture	A range of pathogens may be present in so	oil which can contaminate cattle.

Inpu	t/Activity	Comment
(Wat	ter/Soil/Faeces)	Note: Pathogens include:
		Bacillus, Clostridium, L. monocytogenes, Salmonella and pathogenic E. coli
		A range of pathogens may be present in irrigation water which can contaminate pasture. Irrigation water includes water from natural waterways or recycled water.
		Notes: Pathogens include; Pathogenic E. coli, Campylobacter, Salmonella, Cryptosporidium, Giardia.
		Pasture may be directly contaminated with pathogens excreted in cattle faecal matter, which
		may persist.
		Pathogens from contaminated pasture may be transferred to the external surfaces of cattle (hide) or the gut through consumption of contaminated pasture.
		Notes: Routes of pasture contamination include: Directly deposited from animals or through overland water runoff.
2.2	Pasture	Pasture may be contaminated with pathogens in effluents that are applied as soil fertilisers (ie
		manure and slurry).
(Effl	uents)	
		<b>Notes:</b> Effluents may be contaminated with pathogens that originate from cattle's gastrointestinal tracts and excreted in their faeces. Some pathogens may be able to survive during manure and slurry manufacturing processes and may be persistent for extended periods in the manure and slurry.
2.3	Feeds	Animal feed including roughage (e.g. hay and silage), grain, concentrates and supplements
(Incl	uding roughages,	may be contaminated with pathogens, which may result in a route of pathogen transmission to animals.
	ns, concentrates,	No. Date of the Land
supp	lements)	Notes: Pathogens detected include:  Salmonella spp. in protein meal, haylage and vegetable based feeds  E. coli O157:H7 in forages and alfalfa
		Cl. perfringens in mixed animal feeds
		Cl. botulinum in haylage, silage, pasture, brewer's grains and mixed feed Parasites
		Pathogens may remain in silage as a result of inappropriate ensiling processes and be transmitted to cattle when silage is consumed.
		<b>Notes</b> : Under the optimal ensiling process, harvested forage is stored under moist anaerobic conditions, the lactobacilli flourish, which causes a decrease in pH, and other bacterial populations including pathogens will decrease. However, inappropriately prepared, stored or used silage will allow pathogens to survive and possibly multiply. If forage's moisture content is too high, appropriate fermentation by lactobacilli may not be occur, consequently the secondary fermentation by <i>Clostridium spp</i> . may take place.
		Pathogens such as <i>Listeria monocytogenes</i> , <i>Bacillus</i> spp., pathogenic <i>E. coli</i> and <i>Clostridia</i> spp. are reportedly detected in silage.
2.4	Meat and bone meal (MBM)	Feeding ruminant by-products or materials which may contain TSE agents may contaminate cattle.
	centrates and lements	<b>Notes</b> : A ruminant feed ban is currently in place in Australia. Australia continues to be free of the transmissible spongiform encephalopathies (TSEs).
3. Di	inking Water (includi	ng town, reticulated, ground, surface and run-off water)
3.1	Consumption of	Water may be a source of microbiological contamination for stock.
	town/reticulated	
	water	<b>Notes</b> : Low likelihood of pathogens being present, but cross-contamination may result in drinking water contaminating stock <i>e.g.</i> pathogenic <i>E. coli, Salmonella</i> spp., <i>Campylobacter</i> spp.
3.2	Consumption of	Unprotected groundwater is prone to faecal contamination from livestock, wild animals,
	groundwater	domestic pets and humans which may contain a wide range of pathogens and may contaminate cattle.
		<b>Notes</b> : Pathogens may include pathogenic <i>E. coli, Salmonella</i> spp., <i>Campylobacter</i> spp.
3.3	Consumption of surface water and run-off water	Natural waterways in pasture (e.g. creeks, rivers and dams) may be contaminated with pathogens which could then be a source of microbial contamination of cattle.
		<b>Notes</b> : Natural waterways in pasture may be contaminated with pathogens, originating from agriculture, industrial or municipal wastewater discharged to the upper course of waterways.

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ental sources.
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# (b) Intensive (Feedlot) Production

Inpu	nt/Activity	Comment
1.	1. Animal Production (including calving, health status, zoonoses)	
1.1	Receipt of cattle	Disease transmission between animals due to mixing animals of different origins or higher animal density in the feedlot pen.
		Notes: Mixing of animals from different origins and social groups at markets contributes to the risk of contaminating animals with foodborne pathogens. Due to higher animal density, the lot feeding animals are more susceptible to a range of respiratory diseases, which may not be zoonoses but may reduce animals' natural immune system. As a result, the animals may become more susceptible to other pathogens, such as food-borne pathogens.

Innu	t/Activity	Comment
1.2	Growing the cattle	Cattle may carry pathogens with or without exhibiting any clinical signs.
1.4	to market condition	cause may carry paulogens with or without exhibiting any chilled signs.
		Refer Extensive Cattle Table
(Animal health and carrier status of the cattle)		Stress may impact on the animal's natural defence mechanisms resulting in an increased susceptibility to pathogens. Stress also causes increased pathogen shedding in the faeces. Feedlot cattle may be susceptible to higher stress levels.
2.	Animal Feed (include	Notes: Stressors in feedlot cattle may include:  High animal stocking rates Grouping unfamiliar animals together Handling practices particular to the feedlot – transport from farm to feedlot, moving between pens and associated injuries Unclean environment including dirty and dusty floor, drinking water and pens Mixing sick animals with healthy ones Extreme climate conditions specific to the feedlot (eg there may be no shade available for animals) Competition of feed and water Feed and water changes when introduced to the feedlot spasture, grains, concentrates and silage)
2.1	Pasture	Not applicable once animal is in feedlot environment
(XXI a d	tor/Soil/Essess	
2.2	ter/Soil/Faeces) Pasture	Not applicable once animal is in feedlot environment
2.2	Lastare	The applicable once animal is in Jeculot cirrionness.
	uents)	
2.3	Feeds (including	Animal feed including roughage (e.g. hay and silage), grain, concentrates and supplements
	roughages, grains, concentrates,	may be contaminated with pathogens, which may result in a route of pathogen transmission to animals.
	supplements)	annuais.
		Refer Extensive Cattle Table
2.4	Silage	Pathogens may remaining in silage as a result of inappropriate ensiling processes and be transmitted to cattle when silage is consumed.
		Refer to Extensive Cattle Table
2.5	Meat and bone meal (MBM)	Ruminant by-products or materials being fed to cattle
Conc	centrates and	Refer to Extensive Cattle Table
	lements	
		uding town, reticulated, ground, surface and run-off water)
3.1	Consumption of	Water may be a source of microbiological contamination for stock.
	town/reticulated	Defen to Extensing Cattle Table
3.2	water Consumption of	Refer to Extensive Cattle Table Refer to Extensive Cattle Table
	groundwater	· · · · · · · · · · · · · · · · · · ·
3.3	Consumption of	Refer to Extensive Cattle Table
	surface water and run-off water	
3.5	Consumption of	A range of pathogens may remain in untreated or treated recycled water. The waste water
	recycled water	treatment may not be sufficient to inactivate some pathogens.
		Defen Entangina Catala Tabla
4.	Animal Husbandry P	Refer Extensive Cattle Table ractices (including veterinary chemicals, handling practices)
4.1	Animal Husbandry  Animal husbandry	Stress may impact on the animal's natural defence mechanisms resulting in an increased
	practices	susceptibility to pathogens. Stress also causes increased pathogen shedding in the faeces.
	75.74	Refer Extensive Cattle Table
4.2	Medication of cattle	Therapeutic and other use of antimicrobials on cattle may lead to the emergence of resistant microorganisms.
		microorganisms.
		Refer Extensive Cattle Table
5.		ng premises, building and equipment, personnel)
5.1	Environmental contamination of the	Stock may become directly contaminated by pathogens derived from environmental sources.
	environment	Refer Extensive Cattle Table

Input/Activity	Comment
	Microbiological contamination of exterior surfaces of cattle from the environment of the feedlot production system.
	Notes: Animal's hides, hooves and feed may be visibly and microbiologically contaminated by soil and build-up of animal faeces on the feedlot floor.  Water may be highly contaminated by the exterior surface of cattle as a large number of animals access a limited number of water troughs in a feedlot pen.

# (c) Bobby Calf Production

Inpu	ıt/Activity	Comment
1.	Animal Production (i	ncluding calving, health status, zoonoses)
1.1	Calving	Calving may result in microbial contamination of the newborn calf and the calving environment.
		There may be vertical transmission of foodborne pathogens from sick mother.
		<b>Notes</b> : The following pathogens may be transmitted vertically, found in contaminated artificial formula/milk for calf, and/or found in pregnant cow and new born calves (with or without clinical signs) with higher prevalence than in mature cattle:
		Brucella abortus EHEC (O157:H7)
		Campylobacter spp. Listeria monocytogenes Clostridium spp. Corynebacterium ulcerans Salmonella spp.
1.2	Growing the cattle	Cattle may carry pathogens with or without exhibiting any clinical signs.
1.2	to market condition	cutto may early participants with or without extinorting any entired signs.
	00 11111 1100 0011011011	Refer Extensive Cattle Table
(Ani	mal health and	Newborn animals are more susceptible to particular pathogens.
	ier status of the	
cattl	/	
2.	Animal Feed (include	es pasture, grains, concentrates and silage)
	Pasture	Not applicable to bobby calves
_	ter/Soil/Faeces)	
	Pasture	Not applicable to bobby calves
	uents)	
	Feeds	Contamination of artificial formula/milk for calf.
	uding roughages, ns, concentrates,	Notes: Pathogens may be found in contaminated artificial formula/milk for calves either from
	olements)	the formula itself or via cross contamination from preparation utensils.
2.4		Not directly applicable to bobby calves. Cross contamination from preparation utensils may
	5g.	occur
2.5	MBM	Not directly applicable to bobby calves. Cross contamination from preparation utensils may
Cone	centrates and	occur.
supp	olements	
3.		uding town, reticulated, ground, surface and run-off water)
3.1	Consumption of	Water may be a source of microbiological contamination for stock.
	town/reticulated	
	water	Refer Extensive Cattle Table
3.2	Consumption of	Unprotected groundwater is prone to faecal contamination from livestock, wild animals,
	groundwater	domestic pets and humans which may contain a wide range of pathogens and may contaminate cattle.
		Refer Extensive Cattle Table
3.3	Consumption of	Natural waterways in pasture (e.g. creeks, rivers and dams) may be contaminated with
	surface water and	pathogens which could then be a source of microbial contamination of cattle.
	run-off water	Refer Extensive Cattle Table
3.5	Consumption of	A range of pathogens may remain in untreated or treated recycled water. The waste water
3.3	recycled water	treatment may not be sufficient to inactivate some pathogens.
	100 cica muci	
		Refer Extensive Cattle Table

Inpu	ıt/Activity	Comment
4.	Animal husbandry pr	ractices (including veterinary chemicals, handling practices)
4.1	I.1 Animal husbandry Stress may impact on the animal's natural defence mechanisms resulting in an increased	
	practices	susceptibility to pathogens. Stress also causes increased pathogen shedding in the faeces.
		Refer to Extensive Cattle Table
4.2	Medication of cattle	Refer to Extensive Cattle Table.
5.	Environment (including premises, building and equipment, personnel)	
5.1	Environmental	Stock may become directly contaminated by pathogens derived from environmental sources.
	contamination of the	
	farming	Refer to Extensive Cattle Table
	environment	

# (d) Transport, Saleyards, Lairage, Slaughter and Carcass Dressing Operations

Activity	Comment
All or most activities –	
transport and saleyards	Contamination, injury or other matters that could impact on the health or suitability of cattle for meat production occur because personnel lack skills and knowledge to implement practices
transport and saleyards	that avoid injury to cattle, assess suitability for slaughter or other matters that could impact on
	the safety or suitability of cattle for meat production or the meat.
All or most activities-	Contamination, injury or other matters that could impact on the health or suitability of cattle
lairage, slaughter and	for meat processing occur because personnel lack skills and knowledge to implement practices
carcass dressing	that avoid injury to cattle, assess suitability for slaughter or other matters that could impact on
operations.	the safety or suitability of cattle for meat processing
operations.	Contamination from personnel involved in slaughter and meat processing
	Contamination from premises and equipment
	Contamination from premises and equipment and personnel
1. Preparation and Tran	sport to Market/Abattoir
	Dirty cattle may increase the likelihood of pathogen contamination onto carcass from hides
handling operations	during the slaughtering and dressing process.
	61
(according to the	<b>Notes:</b> Surface bacterial counts can rise, as the hide becomes dirtier. A range of foodborne
dirtiness)-	pathogens may exist in the animal's exterior surfaces such as the hooves, hide and skin, fair or
	fleece.
	The hide dirtiness is influenced by a number of factors, such as: extensively or intensively
	produced (including whether housed), age, coat length, clipping, journey time, feeding regime.
1.2 Transport	Pathogens may contaminate cattle via cross-contamination from the transport vehicle.
	<b>Notes:</b> Foodborne pathogens can be detected in the transport vehicle prior to loading cattle.
	Pathogen prevalence on hides may be affected by: type of vehicle (ie single or double deck),
	floor type (ie metal or wooden), bedding (non or straw bedding), cleanliness of the truck,
	cleanliness of animals and the distance travelled.  Stress in livestock occurs more frequently during the period between leaving the farm and
	slaughter (ie transportation). Such stresses may increase human pathogen shedding by
	livestock, and also increase pathogen loads within the animal or herd.
	investock, and also increase pathogen loads within the alimital of field.
	<b>Notes:</b> The prevalence of pathogens in a herd may increase due to the host's weakened
	immune system. Pathogen loads being shed by the individual animal may increase. Stress
	may be caused prior to and during transport by: feed and water deprivation, mixing with
	unfamiliar animals, confined space (ie trucks), distance travelled, climatic change, changes in
	feed.
	Persistent pathogens in animals and the transport vehicle may be transmitted to other animals
	when comingled.
	<b>Notes:</b> Some foodborne pathogens can survive lengthy periods of time in animals and the
	environment during transport.
	Pathogens include: Salmonella spp., EHEC, Listeria monocytogenes.

Activity		Comment
1.3	Feed Curfew	Pathogen loads in the animal may increase when they are deprived of feed and water prior to and during transportation. Extended time in lairage off feed may also increase pathogen load in the animal.
		<b>Notes</b> : Feed deprivation (both reduced and interrupted) may: trigger the growth of pathogens in the rumen of livestock; change microflora in the rumen and lower digestive tract (e.g. colon) due to a changed pH level; decrease the animal's ability to eliminate the pathogen from the rumen.
2.	Saleyards	
2.1	Holding and processing	Fransfer of pathogens between animals in saleyard pens due to the common livestock marketing system mixing animals from multiple sources.
		Increased chance of infection in younger animals.
		<b>Note:</b> Younger animals are more susceptible to infectious agents, may be infected with higher loads of pathogens compared to mature animals and are more likely to attend the marketing activities.
		<b>Issue:</b> Increased pathogen shedding due to stresses associated with marketing activities.
		<b>Note:</b> Stressors include: excessive transportation; deprivation of feed and water; over crowding; unfamiliar feed; mixed with unfamiliar animals.
3.	Lairage	
3.1	Lairage environment	Microbiological contamination of lairage environment by animals and subsequent transfer to other cattle in the pen.
		Notes: The following bacterial pathogens have been detected in lairage environment and include:  • E. coli O157 • Salmonella
3.2	Water	<ul> <li>Campylobacter</li> <li>Use of untreated water for cleaning of the lairage environment may introduce pathogenic</li> </ul>
3.2	water	microorganisms.
3.3	Ante-mortem	Diseased, downer and dying animals may get through to slaughter.
		<b>Notes</b> : Identification of animals that may not be displaying symptoms of disease or conditions which would make them unfit for human consumption, and/or may compromise the integrity of the slaughterhouse.
		Microbiological contamination of lairage environment by animals and subsequent transfer to other cattle in the pen.
		<b>Notes</b> : The following bacterial pathogens have been detected in lairage environment and include:
		<ul> <li>E. coli O157 detected: in all steps in lairage, pen side rails, Salmonella detected: in knocking box, on hide, in environment Campylobacter detected: on hide post-transit</li> </ul>
4.	Slaughtering Operation	
4.1	Cattle washing	Excessive levels of soil, dust and faeces on animal hide represent a source of contamination.
		<b>Notes</b> : Bacterial pathogens have been detected after pre-slaughter wash on hide sites (inside hind leg, bung, flap and brisket) and residue of faecally contaminated hide after washing prior to slaughter.
4.2	Stunning and	Contamination of the slaughtering and processing environment.
	bleeding	<b>Notes</b> : Stunning method (including immobilisation) should ensure adverse effects such as blood-splash and fractures are avoided.
		The following bacterial pathogens have been detected on cattle post-stunning & bleeding:  pathogenic <i>E. coli</i> (including O157, non-O157 and STEC)  Salmonella,  Staphylococcus (coagulase positive)
		Captive bolt may be a source of contamination either from transfer of external contaminants to internal organs, or through re-use of captive bolt between animals.

Activ	vity	Comment
4.3	Carcass hide	High microbial levels on carcasses.
٦.٥	washing	Tingii inicrobiai ieveis on carcasses.
	J	Notes: E. coli O157 detected pre & post carcass washing
(also	occurs post	Salmonella detected pre & post carcass washing
4.4	trimming) Legging, hide	Opportunity for cross contamination between hide and carcass.
	clearing and hide	officers, or construction and the construction and
	removal	<b>Notes</b> : Pathogenic bacteria detected on animals prior to hide removal. Isolates include:
		<ul> <li>Pathogenic E. coli</li> <li>Enterobacteriaceae</li> </ul>
		Salmonella
		Notes: Pathogenic bacteria detected on carcasses post hide removal. Isolates include:  • Pathogenic <i>E. coli</i>
		Salmonella
		■ L. monocytogenes
		<ul> <li>Coagulase-positive Staphylococcus</li> </ul>
		Notes: Contamination of carcass via microorganisms in air
4.5	Bunging	Opportunity for faecal leakage onto carcass and into processing environment.
		Notes. Dath again hastoric associated with hymning settle include.
		Notes: Pathogenic bacteria associated with bunging cattle include;  Pathogenic <i>E. coli O157:H7</i>
		■ Salmonella
		■ Enterobacteriaceae.
		Notes: Washing pre-evisceration carcasses pre or post bunging can affect the carcass
		contamination from the rectum. Pooling in the rectal area from wash solution can influence
		carcass contamination
4.6	Evisceration	Opportunity for faecal contamination of utensils and slaughtering environment if carried out incorrectly.
		incorrectly.
		<b>Notes</b> : Pathogenic bacteria detected on carcass pre-evisceration include:
		Pathogenic E. coli
		<ul><li>Enterobacteriaceae</li><li>Salmonella spp.</li></ul>
		Mycobacterium avium subsp. paratuberculosis
		Notes. Dethis herteric detected
		Notes: Pathogenic bacteria detected on carcass post-evisceration include:  • Campylobacter spp.
		Coagulase-positive Staphylococcus
		■ Pathogenic E. coli O157:H7
		Notes: Pathogenic bacteria detected on utensils & within the slaughtering environment
		include:
		Coagulase-positive Staphylococcus
		<ul> <li>Pathogenic E. coli</li> <li>L. monocytogenes</li> </ul>
		Potential for pathogens in faeces or gastrointestinal tract to contaminate carcass.
		Notes: Pathogenic bacteria detected in faeces of slaughtered cattle post-evisceration include:  Pathogenic E.coli O157 [H7 & H- (predominant)]
		■ Salmonella spp.
		■ Campylobacter spp.
		L. monocytogenes
		<b>Notes:</b> Pathogenic bacteria detected in faeces of slaughtered cattle post-evisceration include:
		■ Pathogenic <i>E. coli O157:H7</i>
4.7	Post mentem	<ul> <li>Salmonella spp.</li> <li>Macroscopic evidence of disease or faecal contamination of the carcass.</li> </ul>
4.7	Post mortem	iviacioscopic evidence of disease of faecai contamination of the carcass.
		Potential for growth of any contaminating pathogens.
		Pathogenic organisms may be present in offal.
		Notes: Campylobacter spp. in liver.

Activ	vity	Comment
4.8	Trimming	Carcass contamination.
		Notes: Pathogenic bacteria detected on carcass post-trimming include:  E. coli O157 Salmonella Campylobacter Listeria  Notes: Pathogenic bacteria detected on carcass post-splitting include:
		■ E. coli O157:H7
4.9	Carcass washing (optional)	Excess microbial levels on carcasses. May also provide a moist environment for pathogens to survive.
		Notes: Pathogenic bacteria reported on carcasses post-washing include:  Mycobacterium avium subsp. paratuberculosis  Coagulase-positive Staphylococcus pathogenic E. coli (including E. coli O157)  Washing may introduce contaminants that may be subsequently passed to the carcass.
4.10	Storage	Notes: Cryptosporidium parvum Opportunity for outgrowth of pathogens.
		Notes: Pathogenic bacteria detected on chilled carcasses include:  pathogenic E. coli Salmonella spp: Listeria monocytogenes  Opportunity for cross-contamination between carcasses.
4.11	Quartering, boning and packing	Opportunity for cross-contamination.  Notes: Pathogenic bacteria detected on meat in boning room include:  Staphylococcus B. cereus E. coli O157:H7 Salmonella spp. L. monocytogenes  Beef Trimmings used to make ground beef may contain pathogenic bacteria.  Notes: Isolates detected include: pathogenic E. coli Salmonella spp. S. aureus Salmonella spp Campylobacter spp. (C.jejuni; C.coli) L. monocytogenes;
4.12	Storage of packed meat	Notes: Pathogenic bacteria detected on equipment used in the boning process.  Opportunity for outgrowth of pathogens

#### References

Anderson RJ, Walker RL, Hird DW, Blanchard PC (1997) Case-control study of an outbreak of clinical disease attributable to *Salmonella menhaden* infection in eight dairy herds. Journal of the American Veterinary Medical Association 210(3):528–530

Arthur TM, Barkocy-Gallagher GA, Rivera-Betancourt M, Koohmaraie M (2002) Prevalence and characterization of non-O157 Shiga toxin-producing *Escherichia coli* on carcasses in commercial beef cattle processing plants. Appl Environ Microbiol 68(10):4847–4852

Arthur TM, Bosilevac JM, Brichta-Harhay DM, Kalchayanand N, Shackelford SD, Wheeler TL, Koohmaraie M (2007a) Effects of a minimal hide wash cabinet on the levels and prevalence of *Escherichia coli* O157:H7 and *Salmonella* on the hides of beef cattle at slaughter. Journal of Food Protection 70(5):1076–1079

Arthur TM, Bosilevac JM, Brichta-Harhay DM, Guerini MN, Kalchayanand N, Shackelford SD, Wheeler TL, Koohmaraie M (2007b) Transportation and lairage environment effects on prevalence, numbers, and diversity of *Escherichia coli* O157:H7 on hides and carcasses of beef cattle at processing. Journal of Food Protection 70(2):280–286

Arthur TM, Bosilevac JM, Brichta-Harhay DM, Kalchayanand N, King DA, Shackelford SD, Wheeler TL, Koohmaraie M (2008a) Source tracking of *Escherichia coli* O157:H7 and *Salmonella* contamination in the lairage environment at commercial US beef processing plants and identification of an effective intervention. Journal of Food Protection 71(9):1752–1760

Arthur TM, Brichta-Harhay DM, Bosilevac JM, Guerini MN, Kalchayanand N, Wells JE, Shackelford SD, Wheeler TL, Koohmaraie M (2008b) Prevalence and characterization of *Salmonella* in bovine lymph nodes potentially destined for use in ground beef. Journal of Food Protection 71(8):1685–1688

Arthur TM, Bosilevac JM, Nou XW, Shackelford SD, Wheeler TL, Kent MP, Jaroni D, Pauling B, Allen DM, Koohmaraie M (2004) *Escherichia coli* O157 prevalence and enumeration of aerobic bacteria, *Enterobacteriaceae*, and *Escherichia coli* O157 at various steps in commercial beef processing plants. Journal of Food Protection 67(4):658–665

Aslam M, Service C (2006) Antimicrobial resistance and genetic profiling of *Escherichia coli* from a commercial beef packing plant. Journal of Food Protection 69(7):1508–1513

Aslam M, Greer GG, Nattress FM, Gill CO, McMullen LM (2004a) Genotypic analysis of *Escherichia coli* recovered from product and equipment at a beef-packing plant. Journal of Applied Microbiology 97(1):78–86

Aslam M, Greer GG, Nattress FM, Gill CO, McMullen LM (2004b) Genetic diversity of *Escherichia* coli recovered from the oral cavity of beef cattle and their relatedness to faecal *E. coli*. Letters in Applied Microbiology 39(6):523–527

Aslam M, Nattress F, Greer G, Yost C, Gill C, McMullen L (2003) Origin of contamination and genetic diversity of *Escherichia* coli in beef cattle. Appl Environ Microbiol 69(5):2794–2799

Avery LM, Killham K, Jones DL (2005) Survival of *E. coli* O157:H7 in organic wastes destined for land application. Journal of Applied Microbiology 98(4):814–822

Avery SM, Small A, Reid CA, Buncic S (2002) Pulsed-field gel electrophoresis characterization of Shiga toxin-producing *Escherichia coli* O157 from hides of cattle at slaughter. Journal of Food Protection 65(7):1172–1176

Avery SM, Liebana E, Hutchison ML, Buncic S (2004) Pulsed field gel electrophoresis of related *Escherichia coli* O157 isolates associated with beef cattle and comparison with unrelated isolates from animals, meats and humans. International Journal of Food Microbiology 92(2):161–169

Bacon RT, Sofos JN, Belk KE, Hyatt DR, Smith GC (2002) Prevalence and antibiotic susceptibility of *Salmonella* isolated from beef animal hides and carcasses. Journal of Food Protection 65(2):284–290

Bacon RT, Belk KE, Sofos JN, Clayton RP, Reagan JO, Smith GC (2000) Microbial populations on animal hides and beef carcasses at different stages of slaughter in plants employing multiple-sequential interventions for decontamination. Journal of Food Protection 63(8):1080–1086

Barham AR, Barham BL, Johnson AK, Allen DM, Blanton JR, Miller MF (2002) Effects of the transportation of beef cattle from the feedyard to the packing plant on prevalence levels of *Escherichia coli* O157 and *Salmonella* spp. Journal of Food Protection 65(2):280–283

Barkocy-Gallagher GA, Arthur TM, Rivera-Betancourt M, Nou XW, Shackelford SD, Wheeler TL, Koohmaraie M (2003) Seasonal prevalence of Shiga toxin-producing *Escherichia coli*, including O157:H7 and non-O157 serotypes, and *Salmonella* in commercial beef processing plants. Journal of Food Protection 66(11):1978–1986

Barlow RS, Gobius KS, Desmarchelier PM (2006) Shiga toxin-producing *Escherichia coli* in ground beef and lamb cuts: results of a one-year study. International Journal of Food Microbiology 111(1):1–5

Beach JC, Murano EA, Acuff GR (2002a) Prevalence of *Salmonella* and *Campylobacter* in beef cattle from transport to slaughter. Journal of Food Protection 65(11):1687–1693

Beach JC, Murano EA, Acuff GR (2002b) Serotyping and antibiotic resistance profiling of *Salmonella* in feedlot and nonfeedlot beef cattle. Journal of Food Protection 65(11):1694–1699

Bell RG (1997) Distribution and sources of microbial contamination on beef carcasses. Journal of Applied Microbiology 82(3):292–300

Blanco J, Blanco M, Blanco JE, Mora A, Gonzalez EA, Bernardez MI, Alonso MP, Coira A, Rodriguez A, Rey J, Alonso JM, Usera MA (2003) Verotoxin-producing *Escherichia coli* in Spain: prevalence, serotypes, and virulence genes of O157:H7 and Non-O157 VTEC in ruminants, raw beef products, and humans. Experimental Biology and Medicine 228(4):345–351

Bosilevac JM, Guerini MN, Brichta-Harhay DM, Arthur TM, Koohmaraie M (2007) Microbiological characterization of imported and domestic boneless beef trim used for ground beef. Journal of Food Protection 70(2):440–449

Breuer T, Benkel DH, Shapiro RL, Hall WN, Winnett MM, Linn MJ, Neimann J, Barrett TJ, Dietrich S, Downes FP, Toney DM, Pearson JL, Rolka H, Slutsker L, Griffin PM, Investigation Team (2001) A multistate outbreak of *Escherichia coli* O157:H7 infections linked to alfalfa sprouts grown from contaminated seeds. Emerging Infectious Diseases 7(6):977–982

Brichta-Harhay DM, Guerini MN, Arthur TM, Bosilevac JM, Kalchayanand N, Shackelford SD, Wheeler TL, Koohmaraie M (2008) *Salmonella* and *Escherichia coli* O157:H7 contamination on hides and carcasses of cull cattle presented for slaughter in the United States: an evaluation of prevalence and bacterial loads by Immunomagnetic separation and direct plating methods. Appl Environ Microbiol 74(20):6289–6297

Brownlie LE, Grau FH (1967) Effect of food intake on the growth and survival of *Salmonellas* and *Escherichia coli* in the bovine rumen. Journal of General Microbiology 46:125–134

Buncic S, McKinstry J, Reid CA, Anil MH (2002) Spread of microbial contamination associated with penetrative captive bolt stunning of food animals. Food Control 13(6-7):425–430

Burfoot D, Whyte R, Tinker D, Howell M, Hall K, Holah J, Smith D, White R, Baker D, McIntosh J (2006) Importance of airborne contamination during dressing of beef and lamb carcasses. Journal of Food Protection 69(12):2828–2836

Carney E, O'Brien SB, Sheridan JJ, McDowell DA, Blair IS, Duffy G (2006) Prevalence and level of *Escherichia coli* O157 on beef trimmings, carcasses and boned head meat at a beef slaughter plant. Food Microbiology 23(1):52–59

Childs KD, Simpson CA, Warren-Serna W, Bellenger G, Centrella B, Bowling RA, Ruby J, Stefanek J, Vote DJ, Choat T, Scanga JA, Sofos JN, Smith GC, Belk KE (2006) Molecular characterization of *Escherichia coli* O157:H7 hide contamination routes: feedlot to harvest. Journal of Food Protection 69(6):1240–1247

Connor KM, Quirie MM, Baird G, Donachie W (2000) Characterization of United Kingdom isolates of *Corynebacterium pseudotuberculosis* using pulsed-field gel electrophoresis. J Clin Microbiol 38(7):2633–2637

Cuesta Alonso EP, Gilliland SE, Krehbiel CR (2007) Incidence and toxin production ability of *Escherichia coli* O157:H7 isolated from cattle trucks. Journal of Food Protection 70(10):2383–2385

Davies MH, Hadley PJ, Stosic PJ, Webster SD (2000) Production factors that influence the hygienic condition of finished beef cattle. Veterinary Record 146(7):179—+

Davis MA, Hancock DD, Rice DH, Call DR, DiGiacomo R, Samadpour M, Besser TE (2003) Feedstuffs as a vehicle of cattle exposure to *Escherichia coli* O157:H7 and *Salmonella enterica*. Veterinary Microbiology 95(3):199–210

Desmarchelier PM, Higgs GM, Mills L, Sullivan AM, Vanderlinde PB (1999) Incidence of coagulase positive *Staphylococcus* on beef carcasses in three Australian abattoirs. International Journal of Food Microbiology 47(3):221–229

Dewell GA, Simpson CA, Dewell RD, Hyatt DR, Belk KE, Scanga JA, Morley PS, Grandin T, Smith GC, Dargatz DA, Wagner BA, Salman MD (2008a) Impact of transportation and lairage on hide contamination with *Escherichia coli* O157 in finished beef cattle. Journal of Food Protection 71(6):1114–1118

Dewell GA, Simpson CA, Dewell RD, Hyatt DR, Belk KE, Scanga JA, Morely PS, Grandin T, Smith GC, Dargatz DA, Wagner BA, Salman MD (2008b) Risk associated with transportation and lairage on hide contamination with *Salmonella enterica* in finished beef cattle at slaughter. Journal of Food Protection 71(11):2228–2232

Diez-Gonzalez F, Callaway TR, Kizoulis MG, Russell JB (1998) Grain feeding and the dissemination of acid-resistant *Escherichia coli* from cattle. Science 281(5383):1666–1668

Elder RO, Keen JE, Siragusa GR, Barkocy-Gallagher GA, Koohmaraie M, Laegreid WW (2000) Correlation of enterohemorrhagic *Escherichia coli* O157 prevalence in feces, hides, and carcasses of beef cattle during processing. Proceedings of the National Academy of Sciences of the United States of America 97(7):2999–3003

Enokimoto M, Kubo M, Bozono Y, Mieno Y, Misawa N (2007) Enumeration and identification of *Campylobacter* species in the liver and bile of slaughtered cattle. International Journal of Food Microbiology 118:259–263

Eustace I, Midgley J, Giarrusso C, Laurent C, Jenson I, Sumner J (2007) An alternative process for cleaning knives used on meat slaughter floors. International Journal of Food Microbiology 113(1):23–27

Fegan N, Vanderlinde P, Higgs G, Desmarchelier P (2004a) Quantification and prevalence of *Salmonella* in beef cattle presenting at slaughter. Journal of Applied Microbiology 97(5):892–898

Fegan N, Vanderlinde P, Higgs G, Desmarchelier P (2004b) The prevalence and concentration of *Escherichia coli* O157 in faeces of cattle from different production systems at slaughter. Journal of Applied Microbiology 97(2):362–370

Fegan N, Higgs G, Vanderlinde P, Desmarchelier P (2005a) An investigation of *Escherichia coli* O157 contamination of cattle during slaughter at an abattoir. Journal of Food Protection 68(3):451–457

Fegan N, Vanderlinde P, Higgs G, Desmarchelier P (2005b) A study of the prevalence and enumeration of *Salmonella enterica* in cattle and on carcasses during processing. Journal of Food Protection 68(6):1147–1153

Fenlon DR (1986) Rapid quantitative assessment of the distribution of listeria in silage implicated in a suspected outbreak of listeriosis in calves. Veterinary Record 118:240–242

Fenlon DR, Wilson J, Donachie W (1996) The incidence and level of *Listeria monocytogenes* contamination of food sources at primary production and initial processing. Journal of Applied Bacteriology 81:641–650

Fox JT, Shi X, Nagaraja TG (2008a) *Escherichia coli* O157 in the rectoanal mucosal region of cattle. Foodborne Pathogens and Disease 5(1):69–77

Fox JT, Renter DG, Sanderson MW, Nutsch AL, Shi X, Nagaraja TG (2008b) Associations between the presence and magnitude of *Escherichia coli* O157 in feces at harvest and contamination of preintervention beef carcasses. Journal of Food Protection 71(9):1761–1767

Fremaux B, Prigent-Combaret C, Vernozy-Rozand C (2008) Long-term survival of Shiga toxin-producing *Escherichia coli* in cattle effluents and environment: an updated review. Veterinary Microbiology 132(1-2):1–18

Garber LP, Wells SJ, Hancock DD, Doyle MP, Tuttle J, Shere JA, Zhao T (1995) Risk factors for fecal shedding of *Escherichia coli* O157:H7 in dairy calves. Journal of the American Veterinary Medical Association 207(1):46–49

Garcia MM, Lior H, Stewart RB, Ruckerbauer GM, Trudel JR, Skljarevski A (1985) Isolation, characterization, and serotyping of *Campylobacter jejuni* and *Campylobacter coli* from slaughter cattle. Appl Environ Microbiol 49(3):667–672

Gill CO, Jones T, Bryant J, Brereton DA (2000) The microbiological conditions of the carcasses of six species after dressing at a small abattoir. Food Microbiology 17(2):233–239

Gill COA, McGinnis JC (1999) Improvement of the hygienic performance of the hindquarters skinning operations at a beef packing plant. International Journal of Food Microbiology 51(2-3):123–132

Glickman LT, McDonough PL, Shin SJ, Fairbrother JM, Ladue RL, King SE (1981) Bovine Salmonellosis attributed to *Salmonella* anatum-contaminated haylage and dietary stress. Journal of the American Veterinary Medical Association 178(12):1268–1272

Grau FH (1987) Prevention of microbial contamination in the export beef abattoir. In: Smulders FJM (ed) Elimination of pathogenic organisms from meat and poultry. Elsevier Science Publishers B.V., Amsterdam, The Netherlands, p. 221–233

Grau FH, Smith MG (1974) Salmonella contamination of sheep and mutton carcasses related to pre-slaughter holding conditions. Journal of Applied Bacteriology 37(1):111–116

Gray DF, Lewis PF (1958) Bonemeal as a source of bovine salmonellosis. Australian Veterinary Journal(November):345–351

Haines H, Bobbitt J, Simons J, Rowland D, Coates K (2000) A review of process interventions aimed at reducing contamination of cattle carcasses. Project Number FLOT.213. Meat and Livestock Australia,

Hancock DD, Besser TE, Kinsel ML, Tarr PI, Rice DH, Paros MG (1994) The prevalence of *Escherichia coli* O157:H7 in dairy and beef cattle in Washington State. Epidemiology and Infection 113(2):199–207

Hansson IB (2001) Microbiological meat quality in high- and low-capacity slaughterhouses in Sweden. Journal of Food Protection 64(6):820–825

Heir E, Lindstedt BA, Rotterud OJ, Vardund T, Kapperud G, Nesbakken T (2004) Molecular epidemiology and disinfectant susceptibility of *Listeria monocytogenes* from meat processing plants and human infections. International Journal of Food Microbiology 96(1):85–96

Jericho KWF, Ho J, Kozub GC (2000) Aerobiology of a high-line speed cattle abattoir. Journal of Food Protection 63(11):1523–1528

Keel K, Brazier JS, Post KW, Weese S, Songger JG (2007) Prevalence of PCR ribotypes among *Clostridium difficile* isolates from pigs, calves, and other species. J Clin Microbiol 45(6):1963–1964

Maciorowski KG, Herrera P, Jones FT, Pillai SD, Ricke SC (2007) Effects on poultry and livestock of feed contamination with bacteria and fungi. Animal Feed Science and Technology 133(1-2):109–136

Madden RH, Murray KA, Gilmour A (2007) Carriage of four bacterial pathogens by beef cattle in Northern Ireland at time of slaughter. Letters in Applied Microbiology 44(2):115–119

Mcevoy JM, Doherty AM, Sheridan JJ, Blair IS, McDowell DA (2003a) The prevalence of *Salmonella* spp. in bovine faecal, rumen and carcass samples at a commercial abattoir. Journal of Applied Microbiology 94(4):693–700

Mcevoy JM, Duffy G, Moriarty EM, Lowery CJ, Sheridan JJ, Blair IS, McDowell DA (2005) The prevalence and characterisation of *Cryptosporidium* spp. in beef abattoir water supplies. Water Research 39(15):3697–3703

Mcevoy JM, Doherty AM, Sheridan JJ, Thomson-Carter FM, Garvey P, McGuire L, Blair IS, McDowell DA (2003b) The prevalence and spread of *Escherichia coli* O157:H7 at a commercial beef abattoir. Journal of Applied Microbiology 95(2):256–266

McGee P, Bolton DJ, Sheridan JJ, Earley B, Leonard N (2001) The survival of *Escherichia coli* O157:H7 in slurry from cattle fed different diets. Letters in Applied Microbiology 32(3):152–155

McGrath JF, Patterson JT (1969) Meat hygiene: the pre-slaughter treatment of fatstock. Vet Rec 85:521-524

Meadus WJ, Gill CO, Duff P, Badoni M, Saucier L (2008) Prevalence on beef carcasses of *Mycobacterium avium* subsp *paratuberculosis* DNA. International Journal of Food Microbiology 124(3):291–294

Meat and Livestock Australia (2003a) *Through-chain risk profile for the Australian red meat industry. Part 1: Risk Profile.* In: Meat and Livestock Australia. eds. Report No. PRMS.038c, Sydney.

Meat and Livestock Australia (2003b) *Through-chain risk profile for the Australian red meat industry. Part 2: Technical information.* In: Meat and Livestock Australia. eds. Report No. PRMS.038c., Meat and Livestock Australia, Sydney.

Moriarty EM, Duffy G, Mcevoy JM, Caccio S, Sheridan JJ, McDowell D, Blair IS (2005a) The effect of thermal treatments on the viability and infectivity of *Cryptosporidium parvum* on beef surfaces. Journal of Applied Microbiology 98(3):618–623

Moriarty EM, Mcevoy JM, Lowery CJ, Thompson HP, Finn M, Sheridan JJ, Blair IS, McDowell DA, Duffy G (2005b) Prevalence and characterisation of *Cryptosporidium* species in cattle faeces and on beef carcases at slaughter. Veterinary Record 156(6):165–168

Murphy C, Carroll C, Jordan KN (2006) Environmental survival mechanisms of the foodborne pathogen *Campylobacter jejuni*. Journal of Applied Microbiology 100(4):623–632

National Advisory Committee on Microbiological Criteria for Foods (1993) Generic HACCP for raw beef. Food Microbiology 10(6):449–488

Nel S, Lues JFR, Buys EM, Venter P (2004) Bacterial populations associated with meat from the deboning room of a high throughput red meat abattoir. Meat Science 66(3):667–674

Nicholson FA, Groves SJ, Chambers BJ (2005) Pathogen survival during livestock manure storage and following land application. Bioresource Technology 96(2):135–143

Phillips, D., Sumner, J., Alexander, J.F. and Dutton, K.M. (2001a) Microbiological quality of Australian beef. *J Food Prot.* 64(5):692-696.

Phillips, D., Jordan, D., Morris, S., Sumner, J. and Jenson, I. (2005) *Microbological quality of Australian beef and sheepmeat - results of the industry's third national abattoir study*. In: Meat and Livestock Australia. eds. Meat and Livestock Australia.

Phillips, D., Jordan, D., Morris, S., Jenson, I. and Sumner, J. (2006a) A national survey of the microbiological quality of beef carcasses and frozen boneless beef in Australia. *Journal of Food Protection* 69(5):1113-1117.

Pirs T, Ocepek M, Rupnik M (2008) Isolation of *Clostridium difficile* from food animals in Slovenia. J Med Microbiol 57(6):790–792

Pointon, A. (2007) *Toxoplasma gondii in meat and meat products*. Report No. A.MFS.0113, Meat and Livestock Australia.

Pointon, A., Sumner, J., Delcenserie, V. and Slade, J. (2007) Information, collation and review of risk assessments on meat and meat products. (Unpublished Work).

Sumner, J., Petrenas, E., Dean, P., Dowsett, P., West, G., Wiering, R. and Raven, G. (2003) Microbial contamination on beef and sheep carcases in South Australia. *International Journal of Food Microbiology* 81(3):255-260.

Rangel JM, Sparling PH, Crowe C, Griffin PM, Swerdlow DL (2005) Epidemiology of *Escherichia coli* O157: H7 outbreaks, United States, 1982-2002. Emerging Infectious Diseases 11(4):603–609

Reicks AL, Brashears MM, Adams KD, Brooks JC, Blanton JR, Miller MF (2007) Impact of transportation of feedlot cattle to the harvest facility on the prevalence of *Escherichia coli* O157:H7, *Salmonella*, and total aerobic microorganisms on hides. Journal of Food Protection 70:17–21

Rodriguez-Palacios A, Stampfli HR, Duffield T, Peregrine AS, Trotz-Williams LA, Arroyo LG, Brazier JS, Weese JS (2006) *Clostridium difficile* PCR ribotypes in calves, Canada. Emerging Infectious Diseases 12(11):1730–1736

Shale K, Lues JFR, Venter P, Buys EM (2005) The distribution of *Staphylococcus* sp on bovine meat from abattoir deboning rooms. Food Microbiology 22(5):433–438

Stolle FA (1986) Rodding in the West-Berlin slaughterhouse - A possible method of improving hygiene in slaughterhouses or additional labor expenditure in the modern cattle slaughtering procedure. British Veterinary Journal 142(1):30–35

Stopforth JD, Lopes M, Shultz JE, Miksch RR, Samadpour M (2006a) Microbiological status of fresh beef cuts. Journal of Food Protection 69(6):1456–1459

Stopforth JD, Lopes M, Shultz JE, Miksch RR, Samadpour M (2006b) Location of bung bagging during beef slaughter influences the potential for spreading pathogen contamination on beef carcasses. Journal of Food Protection 69(6):1452–1455

Troutt HF, Osburn BI (1997) Meat from dairy cows: possible microbiological hazards and risks. Revue Scientifique et Technique de Office International des Epizooties 16(2):405–414

Uzal FA, Connole MD, O'Boyle D, Dobrenov B, Kelly WR (1999) *Mortierella wolfii* isolated from the liver of a cow in Australia. Veterinary Record 145(9):260–261

Vanderlinde PB, Shay B, Murray J (1998) Microbiological quality of Australian beef carcass meat and frozen bulk packed beef. J Food Prot 61(4):437–443

Vanderlinde PB, Fegan N, Mills L, Desmarchelier PM (1999) Use of pulse field gel electrophoresis for the epidemiological characterisation of coagulase positive *Staphylococcus* isolated from meat workers and beef carcasses. International Journal of Food Microbiology 48(2):81–85

Vanselow BA, Krause DO, McSweeney CS (2005) The Shiga toxin-producing *Escherichia coli*, their ruminant hosts, and potential on-farm interventions: a review. Australian Journal of Agricultural Research 56(3):219–244

Vanselow BA, Hornitzky MA, Walker KH, Eamens GJ, Bailey GD, Gill PA, Coates K, Corney B, Cronin JP, Renilson S (2007) *Salmonella* and on-farm risk factors in healthy slaughter-age cattle and sheep in eastern Australia. Australian Veterinary Journal 85(12):498–502

Wang G, Zhao T, Doyle MP (1996) Fate of enterohemorrhagic *Escherichia coli* O157:H7 in bovine feces. Appl Environ Microbiol 62(7):2567–2570

Zhao T, Doyle MP, Shere J, Garber L (1995) Prevalence of enterohemorrhagic *Escherichia coli* O157:H7 in a survey of dairy herds. Appl Environ Microbiol 61(4):1290–1293

# 2. Sheep Production in Australia

#### Introduction

The prime lamb industry is concentrated in New South Wales, Western Australia and Victoria with the main outputs being lamb meat and mutton. In addition, there are live sheep exports into the Middle East market. While large volumes of industry outputs are exported, Australians continue to consume large volumes of lamb meat.

#### **Lamb and Mutton Production**

Primary production of lambs and sheep are predominantly based on extensive production systems. The most efficient way to produce lambs is on quality pasture with at least 30% legume content ideal. The major inputs during primary production are feed and water, with some supplement feeding undertaken to achieve target growth rates. Cereal grains tend to be the most cost-effective form of feed supplementation.

Importantly, there is also an increasing trend towards finishing lambs in feedlot environments. Prior to receipt at the feedlot yards, lambs finished on feedlots are initially subjected to the same production methods and inputs as extensively reared animals. Once in the feedlot environment, lambs are more contained, restricted in their movements, are at higher stocking rates and exposed to greater environmental influences (*i.e.* environmental conditions including heat).

The Australian sheep industry has developed integrity systems to verify and assure the food safety status, to improve meat quality and to ensure the traceability of livestock. This is through all sectors of the sheepmeat industry, from the farm through to feedlots, transport, saleyards, and processing plants.

The key steps in the production and processing of sheep are summarised in Figure 2.

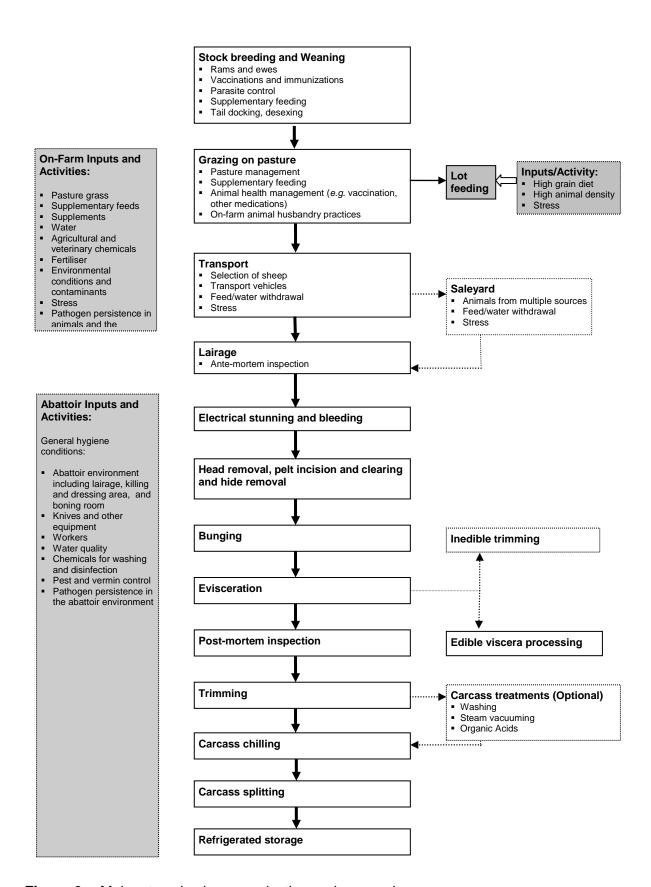


Figure 2: Major steps in sheep production and processing

## **Abattoir Operations**

Production and slaughtering operations are undertaken using very similar processing steps.

Minor differences may exist depending on the plant's capabilities and design but the main steps remain the same. Others factors which may influence abattoir operations include: single species or multiple species plant; age of plant; chain speed; export or domestic; and different slaughtering practices.

## **Hazard Identification**

The following tables outline the microbiological hazards that may be encountered along the entire sheep production and processing chain.

## (a) Extensive Sheep Production

Input/Activity	Comment	
1. Animal Production (including sourcing animals, birthing, health status, zoonoses etc)		
1.1 Growing the sheep to market condition	Increased pathogen load in lambs finished in a feedlot environment	
(Animal health and carrier	<b>Notes</b> : Feedlot lambs may be subject to increased stress and environmental conditions which may increase pathogen load in the animal.	
status of the sheep)	Sheep may carry pathogens with or without exhibiting any clinical signs.	
	<b>Notes</b> : The following hazards may be found in the gastrointestinal tract and exterior surfaces of sheep:	
	Foodborne pathogens which have been more commonly associated with sheep include; Salmonella spp. Pathogenic E. coli (EHEC)	
	Other possible foodborne pathogens associated with sheep meat include: Campylobacter jejuni	
	Yersinia enterocolitica Yersinia pseudotuberculosis	
	Cryptosporidium parvum	
	Toxoplasma gondii	
	Cryptosporidium parvum  Sheep may carry pathogens normally associated with handling, which could potentially be	
	transmitted via meat consumption.	
	Notes: Examples include:	
	Burkholderia pseudomallei(Melioidosis)	
	Coxiella burnetii (Q Fever) Bacillus anthracis (Anthrax)	
	Buctius animacis (Animax)	
2. Animal Feed (includes )	pasture, grains, concentrates and silage)	
2.1 Pasture	A range of pathogens may be present in soil which can contaminate sheep.	
(Water/Soil/Faeces)	Refer to Extensive Cattle Table	
2.2 Pasture	Pasture may be contaminated with pathogens in effluents that are applied as soil fertilisers	
	(i.e. manure and slurry).	
(Effluents)		
2.3 Feeds	Refer to Extensive Cattle Table  Animal feed including roughage (e.g. hay and silage), grain, concentrates and supplements	
(Including roughages,	may be contaminated with pathogens, which may result in a route of pathogen transmission	
grains, concentrates,	to animals.	
supplements)		
	Refer to Extensive Cattle Table	
2.4 Silage	Pathogens may remaing in silage as a result of inappropriate ensiling processes and be	
	transmitted to cattle when silage is consumed.	
	Refer to Extensive Cattle Table	

Input/Activity		Comment
2.5	Meat and bone meal (MBM)	Feeding ruminant by-products or materials which may contain TSE agents may contaminate sheep.
	centrates and elements	<b>Notes</b> : A ruminant feed ban is currently in place in Australia. Australia continues to be free of the transmissible spongiform encephalopathies (TSEs).
3.		ing town, reticulated, ground, surface and run-off water)
3.1	Consumption of town/reticulated water	Water may be a source of microbiological contamination for stock.  Refer to Extensive Cattle Table
3.2	Consumption of groundwater	Unprotected groundwater may be contaminated by faecal matter from livestock, wild animals, domestic pets and humans which may contain a wide range of pathogens and may contaminate sheep.  Refer to Extensive Cattle Table
3.3	Consumption of surface water and run-off water	Natural waterways in pasture (e.g. creeks, rivers and dams) may be contaminated with pathogens which could then be a source of microbial contamination of sheep.  Refer to Extensive Cattle Table
3.4	Consumption of recycled water	A range of pathogens may remain in untreated or treated recycled water. The waste water treatment may not be sufficient to inactivate some pathogens.  Refer to Extensive Cattle Table
4.	Animal husbandry prac	tices (including veterinary chemicals, handling practices)
4.1	Animal husbandry practices	Stress may impact on the animal's natural defence mechanisms resulting in an increased susceptibility to pathogens. Stress also causes increased pathogen shedding in the faeces.  *Refer to Extensive Cattle Table*
4.2	Medication of sheep	Therapeutic and other use of antimicrobials on sheep may lead to the emergence of resistant microorganisms.  Refer to Extensive Cattle Table
5.	Environment (including	premises, building and equipment, personnel)
5.1	Environmental contamination of the farming environment	Stock may become directly contaminated by pathogens derived from environmental sources.  *Refer to Extensive Cattle Table*

# (b) Transport, Saleyards, Lairage, Slaughter and Carcass Dressing Operations

Activ	vity	Comment
All o	or most activities –	Refer to Cattle Transport Table
transport and saleyards		
All o	or most activities-	Refer to Cattle Transport Table
laira	ge, slaughter and	
carca	ass dressing	
oper	ations.	
1.	Preparation and Trai	nsport to Market/Abattoir
1.1	Selection of sheep	Refer to Cattle Transport Table
	and handling	
	operations	
	(according to the	
	dirtiness)-	
1.2	Transport	Refer to Cattle Transport Table
1.3	Feed Curfew	Refer to Cattle Transport Table
2.	2. Saleyards	
2.1	Holding and	Refer to Cattle Transport Table
	processing	
3.	•	
3.1	Ante-mortem	Refer to Cattle Transport Table
		Microbiological contamination of lairage environment by animals and subsequent transfer to
		other sheep in the pen.

Activity		Comment
		Notes: The following pathogens have been reported to be detected in the lairage environment (international and domestic:literature)  • Yersinia pseudotuberculosis  • Yersinia enterocolitica  • Campylobacter spp.  • Pathogenic E. coli  • Cryptosporidium parvum
4.	Slaughtering Operat	
4.1	Sheep washing	Excessive levels of soil, dust and faeces on animal fleece represent a source of contamination.  Notes: Washing increased aerobic plate count levels on clean shorn, dirty shorn, clean
4.0	G 1	woolly and dirty woolly
4.2	Stunning and bleeding	Refer to Cattle Transport Table
	biceunig	<b>Notes</b> : Cutting of the oesophagus may contaminate the neck, head and blood with ruminal contents.
		<b>Notes:</b> Experimental simulation in sheep demonstrates the potential transfer of marker organisms detected in blood, liver, spleen, lung, kidney, lymph nodes, deep muscle and on carcass surface.
		Contamination to the surrounding environment.
		<b>Notes:</b> Experimental simulation in sheep demonstrates the potential transfer of marker organisms to the air, and slaughter man hands and apron after stunning
4.3	Pelt incision &	Opportunity for cross contamination between pelt and carcass.
	cleaning	<b>Notes</b> : Pelt removal by mechanical means may allow dirt, dust and hairs to contaminate the carcass
		<b>Notes:</b> Conventional dressing systems may increase carcass contamination as sheep is hung by hind legs and cuts are made on hindquarters, hence the pelt is pulled from the hind/anus region over the carcass. With inverted dressing the sheep is hung by the forelegs and pelt is puller from the forequarter down to the anus.
4.4	Bunging	Opportunity for faecal leakage onto carcass and into processing environment.
		<b>Notes</b> : Washing pre-evisceration carcasses pre or post bunging can affect the carcass contamination from the rectum. Pooling in the rectal area from wash solution can influence carcass contamination.
4.5	Evisceration	Opportunity for faecal contamination of utensils and slaughtering environment if carried out incorrectly.
		Potential for pathogens in faeces or gastrointestinal tract to contaminate carcass.
		Notes: Pathogens detected post evisceration include:
		<ul> <li>Pathogenic E. coli</li> <li>Campylobacter jejuni/coli</li> </ul>
		<ul> <li>Campylobacter jejuni/coli</li> </ul>
		• Campylobacter spp.
4.6	Post mortem	Salmonella spp.  Refer to Cattle Transport Table
7.0	1 ost mortem	Pathogenic organisms may be present in edible offal.
		<ul> <li>Notes: Potentially pathogenic bacteria has been detected on sheep offal and includes:</li> <li>Salmonella spp. in liver; diaphragmatic muscle and abdominal muscle</li> <li>Lamb livers found to contain initial surface flora which included: Bacillus, Staphylococcus.</li> </ul>
4.7	Trimming	Carcass contamination.
		Notes: Pathogenic bacteria detected on carcass post-trimming include:  Pathogenic <i>E. coli</i> Salmonella spp.  Listeria spp.
4.8	Carcass washing	Excess microbial levels on carcasses.
	onal)	

Activ	ity	Comment
		Notes: May provide a moist environment for pathogens to survive. Pathogenic bacteria detected on carcass post-washing include:  Pathogenic E. coli Y. enterocolitica Salmonella spp.
4.9	Storage	Refer to Cattle Transport Table
4.10	Quartering, boning and packing	Opportunity for cross-contamination.  Notes: Pathogenic bacteria detected on meat in boning room.
4.11	Storage of packed meat	Opportunity for outgrowth of pathogens if stored above minimum temperatures for growth

#### References

Barlow RS, Gobius KS, Desmarchelier PM (2006) Shiga toxin-producing *Escherichia coli* in ground beef and lamb cuts: results of a one-year study. International Journal of Food Microbiology 111(1):1–5

Biss ME, Hathaway SC (1995) Microbiological and Visible Contamination of Lamb Carcasses According to Preslaughter Presentation Status - Implications for HACCP. Journal of Food Protection 58(7):776–783

Biss ME, Hathaway SC (1998) A HACCP-based approach to hygienic slaughter and dressing of lamb carcasses. New Zealand Veterinary Journal 46(5):167–172

Branham LA, Carr MA, Scott CB, Callaway TR (2005) *E. coli* O157 and *Salmonella* spp. in white-tailed deer and livestock. Current Issues in Intestinal Microbiology 6(2):25–29

Buncic S, McKinstry J, Reid CA, Anil MH (2002) Spread of microbial contamination associated with penetrative captive bolt stunning of food animals. Food Control 13(6-7):425–430

Chapman PA, Siddons CA, Malo ATC, Harkin MA (1997) A 1-year study of *Escherichia coli* O157 in cattle, sheep, pigs and poultry. Epidemiology and Infection 119(2):245–250

Chapman PA, Malo ATC, Ellin M, Ashton R, Harkin MA (2001) *Escherichia coli* O157 in cattle and sheep at slaughter, on beef and lamb carcasses and in raw beef and lamb products in South Yorkshire, UK. International Journal of Food Microbiology 64(1-2):139–150

Djordjevic SP, Hornitzky MA, Bailey G, Gill P, Vanselow B, Walker K, Bettelheim KA (2001) Virulence properties and serotypes of Shiga toxin-producing *Escherichia coli* from healthy Australian slaughter-age sheep. J Clin Microbiol 39(5):2017–2021

Duffy EA, Belk DE, Sofos JN, LeValley SB, Kain ML, Tatum JD, Smith GC, Kimberling CV (2001) Microbial contamination occurring on lamb carcasses processed in the United States. Journal of Food Protection 64(4):503–508

European Food Safety Authority (2007) Monitoring and identification of human enteropathogenic *Yersinia* spp. The EFSA Journal

Fegan N, Desmarchelier P (1999) Shiga toxin-producing *Escherichia coli* in sheep and pre-slaughter lambs in eastern Australia. Letters in Applied Microbiology 28(5):335–339

Fenlon DR (1986) Rapid quantitative assessment of the distribution of listeria in silage implicated in a suspected outbreak of listeriosis in calves. Veterinary Record 118:240–242

Fenlon DR, Wilson J (1998) The quantitative assessment of *Listeria monocytogenes* growth in a laboratory ensiling system allowing limited aerobic spoilage. Grass and Forage Science 53:292–295

Fenlon DR, Wilson J, Donachie W (1996) The incidence and level of *Listeria monocytogenes* contamination of food sources at primary production and initial processing. Journal of Applied Bacteriology 81:641–650

Field R, Kalchayanand N, Rozbeh M, Andersen M (1991) Influence of a pelt puller on microbiological quality of

lamb. Sheep Research Journal 7(3):24-26

Gill CO, Delacy KM (1982) Microbial spoilage of whole sheep livers. Appl Environ Microbiol 43(6):1262-1266

Gill CO, Baker LP (1998) Assessment of the hygienic performance of a sheep carcass dressing process. Journal of Food Protection 61(3):329–333

Hadley PJ, Holder JS, Hinton MH (1997) Effects of fleece soiling and skinning method on the microbiology of sheep carcases. Veterinary Record 140(22):570–574

Jones K, Howard S, Wallace JS (1999) Intermittent shedding of thermophilic *Campylobacters* by sheep at pasture. Journal of Applied Microbiology 86(3):531–536

Kiermeier, A. and Pointon, A. (2005) *Processing variables and microbiological quality in sheep processing*. Report No. PRMS.082, Meat and Livestock Australia.

Kelly CA, Lynch B, McIoughlin AJ (1980) The microbiological quality of Irish lamb carcasses. Irish Journal of Food Science and Technology 4(2):125–132

Kudva IT, Hunt CW, Williams CJ, Nance UM, Hovde CJ (1997) Evaluation of dietary influences on *Escherichia coli* O157:H7 shedding by sheep. Appl Environ Microbiol 63(10):3878–3886

Kumar SB, Mandal LN, Sinha BK (1977) A short note on *Aeromonas* strains isolated from market goat meat and its public health importance. Science and Culture 43(4):186–187

Meat and Livestock Australia (2003a) Australian Sheepmeat: Naturally safe., Meat and Livestock Australia, Sydney, Australia.

. Accessed 1 October 2003a

Meat and Livestock Australia (2003b) Through-chain risk profile for the Australian red meat industry. Part 1: Risk Profile. PRMS.038c. Sydney

Meat and Livestock Australia (2003c) Through-chain risk profile for the Australian red meat industry. Part 2: Technical information. PRMS.038c. Meat and Livestock Australia, Sydney

Molla W, Molla B, Alemayehu D, Muckle A, Cole L, Wilkie E (2006) Occurrence and antimicrobial resistance of *Salmonella* serovars in apparently healthy slaughtered sheep and goats of central Ethiopia. Tropical Animal Health and Production 38(6):455–462

Narasimha RD, Ramesh BS (1992) The microbiology of sheep carcasses processed in a modern Indian abattoir. Meat Science 32(4):425–436

Orden JA, Ruiz-Santa-Quiteria JA, Blanco M, Blanco JE, Mora A, Cid D, Gonzalez EA, Blanco J, De la Fuente R (2003) Prevalence and characterization of Vero cytotoxin-producing *Escherichia coli* isolated from diarrhoeic and healthy sheep and goats. Epidemiology and Infection 130(2):313–321

Phillips D, Sumner J, Alexander JF, Dutton KM (2001) Microbiological quality of Australian sheep meat. Journal of Food Protection 64(5):697–700

Quilez J, Torres E, Chalmers RM, Hadfield SJ, del Cacho E, Sanchez-Acedo C (2008) *Cryptosporidium* genotypes and subtypes in lambs and goat kids in Spain. Appl Environ Microbiol 74(19):6026–6031

Rajmalliah N, Sreenivas RM, Dhananjaya RB, Ramakrishna RG (1989) Occurrence of *Salmonella* in slaughtered sheep and goats in Hyderabad. Indian Journal of Comparative Microbiology, Immunology and Infectious Diseases 10(4):193–196

Shane SM, Montrose MS (1985) The occurrence and significance of *Campylobacter jejuni* in man and animals. Veterinary Research Communications 9(1):167–198

Sierra ML, Gonzalezfandos E, Garcialopez ML, Fernandez MCG, Prieto M (1995) Prevalence of *Salmonella*, *Yersinia*, *Aeromonas*, *Campylobacter*, and cold-growing *Escherichia coli* on freshly dressed lamb carcasses. Journal of Food Protection 58(11):1183–1185

Slee KJ, Button C (1990a) Enteritis in sheep, goats and pigs due to *Yersinia pseudotuberculosis* infection. Australian Veterinary Journal 67(9):320–322

Slee KJ, Button C (1990b) Enteritis in sheep and goats due to *Yersinia enterocolitica* infection. Australian Veterinary Journal 67(11):396–398

Slee KJ, Skilbeck NW (1992) Epidemiology of *Yersinia pseudotuberculosis* and *Y. enterocolitica* infections in sheep in Australia. J Clin Microbiol 30(3):712–715

Stanley KN, Wallace JS, Currie JE, Diggle PJ, Jones K (1998) Seasonal variation of thermophilic *Campylobacters* in lambs at slaughter. Journal of Applied Microbiology 84(6):1111–1116

Stern NJ (1980) Effect of boning, electrical-stimulation and medicated diet on the microbiological quality of lamb cuts. Journal of Food Science 45(6):1749–1752

Sumner J, Raven G, Givney R (2004) Have changes to meat and poultry food safety regulation in Australia affected the prevalence of *Salmonella* or of salmonellosis? International Journal of Food Microbiology 92(2):199–205

Vanderlinde PB, Shay B, Murray J (1999) Microbiological status of Australian sheep meat. Journal of Food Protection 62(4):380–385

Woldemariam E, Molla B, Alemayehu D, Muckle A (2005) Prevalence and distribution of *Salmonella* in apparently healthy slaughtered sheep and goats in Debre Zeit, Ethiopia. Small Ruminant Research 58(1):19–24

## 3. Goat Production in Australia

#### Introduction

Goat meat production in Australia involves a combination of strategies: the harvesting of rangeland goats; the breeding and production from rangeland goats; and the processing of farmed goats. The majority of goat meat is derived from rangeland goat populations, and these animals provide landholders with a source of goats suitable for cross-breeding with the main meat species such as Boer goats.

The term 'rangeland' describes goats that roam and are raised on natural grasslands, shrub lands, deserts and alpine areas. Supply chain development over recent years has helped improve the quality and consistently of rangeland goats, with animals drafted according to market specifications before being consigned for slaughter. Saleyards are rarely used and this ensures that goats are consigned direct from property of origin to slaughter, thus minimising transport and stress.

This utilisation of rangeland populations has allowed expansion of the domestic goat herd and supported demand for a more consistent supply of goat meat.

There are an estimated 2.6 million rangeland goats, distributed across all Australian states and territories. Rangeland goats are a complex management problem, because they are both a major environmental pest and a commercial resource, providing a source of income to farmers who muster them for sale.

#### **Goat Production**

The majority of goats slaughtered in Australia are derived from harvesting operations. Feral goats are present over much of Australia, with the largest numbers found in the semi-arid pastoral areas of Western Australia, western New South Wales, southern South Australia, and central and south-western Queensland.

Rangeland goats are harvested by mustering by motorcycle or horse with the aid of dogs or with light aircraft, taking advantage of the tendency for these goats to aggregate into larger herds. Goats may also be trapped at water, with traps consisting of a goat-proof fence surrounding a water point that is entered through one-way gates or ramps.

Pre-slaughter management can have a significant impact on the marketability of goat meat. It involves management practices at the point of capture or on-farm, through to slaughter. Mustering, drafting, loading, trucking, handling, noise, strange surroundings and mixing with other stock are all associated with the marketing process, and poor management of these pre-slaughter operations can reduce liveweights and carcass weights; impact on meat yields, meat quality and safety; and increase mortalities, injuries and condemnations.

Australia commenced exporting goat meat in 1952 and is the world's largest supplier of chilled and frozen goat meat. The principal export markets are the United States, Taiwan, Malaysia, Korea, Singapore, and Canada.

The key steps in the production and processing of goats are summarised in Figure 3.

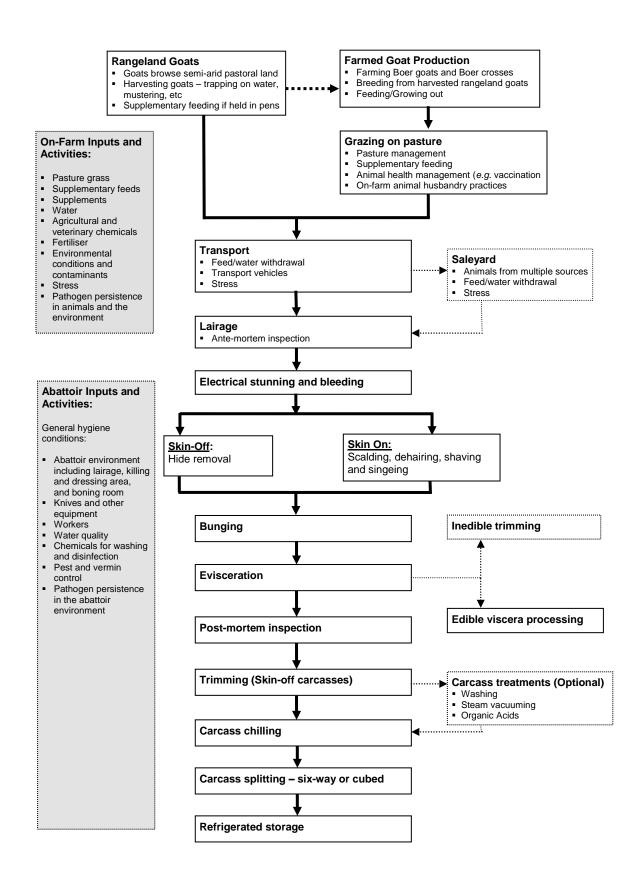


Figure 3: Major steps in goat harvesting, production and processing

## **Abattoir Operations**

Production and slaughtering operations are undertaken using very similar processing steps.

Minor differences may exist depending on the plant's capabilities and design but the main steps remain the same. Others factors which may influence abattoir operations include: single species or multiple species plant; age of plant; chain speed; export or domestic; and different slaughtering practices.

#### **Hazard Identification**

The following tables outline the microbiological hazards that may be encountered along the entire goat production and processing chain.

## (a) Goat Production (Rangeland and farmed production)

Inpu	t/Activity	Comment
1.	<b>Animal Production (</b>	including sourcing animals, birthing, health status, zoonoses etc)
1.1	Trapping	Increased pathogen load in the animal
	Rangeland Goats	
	o .	<b>Notes:</b> Goats are trapped on water and held for up to 3 days. Fed hay. Once sufficient numbers
		are obtained, and then they're transported to slaughter. Feed curfew applies prior to loading.
		Exempt NLIS tagging requirement.
1.2	Growing the goat to	Higher pathogen load (Salmonella spp.) reported in rangeland goats
	market condition	
(Aniı	mal health and	
carri	er status of the goat)	
		Goats may carry pathogens with or without exhibiting any clinical signs.
		<b>Notes</b> : The following hazards may be found in the gastrointestinal tract and exterior surfaces of
		goats:
		Foodborne pathogens more commonly associated with goat meat include;
		Salmonella spp.
		Pathogenic E. coli (including O157)
		Other possible foodborne pathogens associated with goat meat include:
		Campylobacter jejuni
		Yersinia enterocolitica
		Yersinia pseudotuberculosis
		Cryptosporidium parvum
		Toxoplasma gondii
		Goat may carry pathogens normally associated with handling, which could potentially be transmitted via meat consumption.
		Notes: Examples include:
		Burkholderia pseudomallei (Melioidosis)
		Leptospira spp. (Leptospirosis)
		Coxiella burnetii (Q Fever)
		Age of the animal influences susceptibility of the animal to pathogens.
		<b>Notes:</b> Young kids (Capretto) have a carcase weight between 6 -12 kg (Hot Standard Carcass
		Weight) and may be more susceptible to pathogens, as may Chevon (no more than two-tooth
		and with no male secondary sexual characteristics)
2.	Animal Feed (include	es pasture, grains, concentrates and silage)
2.1	Pasture	A range of pathogens may be present in soil which can contaminate goats.
(Wat	er/Soil/Faeces)	
		Refer to Extensive Cattle Table
2.2	Pasture	Pasture may be contaminated with pathogens in effluents that are applied as soil fertilisers (i.e.
		manure and slurry).
(Efflu	uents)	
		Refer to Extensive Cattle Table

Inpu	t/Activity	Comment	
2.3 Feeds		Animal feed including roughage (e.g. hay and silage), grain, concentrates and supplements may	
	1000	be contaminated with pathogens, which may result in a route of pathogen transmission to	
(Including roughages,		animals.	
grains, concentrates,			
supplements)		Refer to Extensive Cattle Table	
2.4	Silage	Pathogens may remaining in silage as a result of inappropriate ensiling processes and be	
	8	transmitted to cattle when silage is consumed.	
		, and the second	
		Refer to Extensive Cattle Table	
2.5	Meat and bone	Refer to Extensive Cattle Table	
	meal (MBM)		
		<b>Notes</b> : A ruminant feed ban is currently in place in Australia. Australia continues to be free of	
Conc	centrates and	the transmissible spongiform encephalopathies (TSEs).	
supp	lements		
3.	<b>Drinking Water (inc</b>	luding town, reticulated, ground, surface and run-off water)	
3.1	Consumption of	Water may be a source of microbiological contamination for stock.	
	town/ reticulated		
	water	Refer to Extensive Cattle Table	
		Unprotected groundwater is prone to faecal contamination from livestock, wild animals,	
	groundwater	domestic pets and humans which may contain a wide range of pathogens and may contaminate	
		goats.	
Refer to Extensive Cattle Table			
3.3	Consumption of	Natural waterways in pasture (e.g. creeks, rivers and dams) may be contaminated with	
surface water and pathogens which could then be a source of microbial contamination of goats		pathogens which could then be a source of microbial contamination of goats.	
run-off water			
		Refer to Extensive Cattle Table	
3.4			
recycled water treatment may not be sufficient to inactivate some pathogens.		treatment may not be sufficient to inactivate some pathogens.	
_		Refer to Extensive Cattle Table	
4.		ractices (including veterinary chemicals, handling practices)	
4.1	Animal husbandry	Stress may impact on the animal's natural defence mechanisms resulting in an increased	
	practices	susceptibility to pathogens. Stress also causes increased pathogen shedding in the faeces.	
		Notes: Costs and in montaylon named and goets, amount to be montaylonly assessable to atmoss	
		<b>Notes</b> : Goats and in particular rangeland goats, appear to be particularly susceptible to stress conditions.	
		Collutions.	
		Pathogen growth and shedding by animals may be encouraged by a range of on-farm husbandry	
		practices stressors. These include: mustering, drenching, restraining for veterinary check-ups	
		including vaccination, restraining for transport preparation, desexing, dehorning, ear-marking,	
		housing, competition for feed and water, extreme climate changes.	
4.2	Medication of goats	Therapeutic and other use of antimicrobials on goats may lead to the emergence of resistant	
7.2		microorganisms.	
		Refer to Extensive Cattle Table	
5.	Environment (includ	ling premises, building and equipment, personnel)	
5.1	Environmental Environmental	Stock may become directly contaminated by pathogens derived from environmental sources.	
	contamination of	Ty Financial and the second and the	
	the farming	Refer to Extensive Cattle Table	
	environment		
		l	

# (b) Transport, Saleyards, Lairage, Slaughter and Carcass Dressing Operations

Activity		Comment	
All or most activities –		Refer to Cattle Transport Table	
transport and saleyards		Rejer to Came Transport Labe	
All or most activities-		Refer to Cattle Transport Table	
lairage, slaughter and		and the same and t	
carcass dressing			
operations.			
1.		sport to Market/Abattoir	
1.1	Selection of goat and	Dirty goats may increase the likelihood of pathogen contamination onto carcass from hides	
	handling operations	during the slaughtering and dressing process.	
	(according to the		
	dirtiness)-	<b>Notes</b> : Rangeland goats sent directly to slaughter after being collected may have increased hide	
		dirtiness.	
		Surface bacterial counts can rise, as the hide becomes dirtier. A range of foodborne pathogens	
		may exist in the animal's exterior surfaces such as the hooves, hide and skin, hair or fleece.	
		The hide dirtiness is influenced by a number of factors, such as: extensively or intensively	
		produced (including whether housed), age, coat length, clipping, journey time, feeding regime.	
1.2	Transport	Refer to Cattle Transport Table	
		Stress in livestock occurs more frequently during the period between leaving the farm and	
		slaughter ( <i>i.e.</i> transportation). Such stresses may increase human pathogen shedding by	
		livestock, and also increase pathogen loads within the animal or herd.	
		Nickey Co. d. 1.1. dill.d.d. Till. 1. C. d. d. d. 1.1.	
		<b>Notes:</b> Goats are particularly susceptible to stress. The prevalence of pathogens in a herd may	
		increase due to the host's weakened immune system.	
		Pathogen loads being shed by the individual animal may increase. Stress may be caused prior to	
		and during transport by: feed and water deprivation, mixing with unfamiliar animals, confined space (i.e. trucks), distance travelled, climatic change, changes in feed.	
13	Food Curfow	Refer to Cattle Transport Table	
2.			
	Holding and	Refer to Cattle Transport Table	
2.1	processing	Refer to Cuite Trunsport Tubic	
3.	Lairage		
3.1	Ante-mortem	Refer to Cattle Transport Table	
		Microbiological contamination of lairage environment by animals and subsequent transfer to	
		other goats in the pen.	
		<b>Notes</b> : The following bacterial pathogens have been detected in the lairage environment:	
		■ Pathogenic E. coli	
		■ Salmonella spp.	
		<ul> <li>Campylobacter jejuni</li> </ul>	
_	a	Cryptosporidium parvum	
4.	Slaughtering Operation		
4.1	Goat washing	Refer to Cattle Transport Table	
4.2	Stunning and	Refer to Cattle Transport Table	
	bleeding	Opportunity for gross contamination from in casts willed during 11-1-1-1	
		Opportunity for cross contamination from ingesta spilled during bleedout.	
12	Carcass hide	Refer to Cattle Transport Table	
4.3	washing (also occurs	Rejet to Cuttle Trunsport Tuote	
	post trimming)		
Skir	n-On	Contamination of the carcass from scald tank.	
	a Scalding, dehairing,	Containment of the cureups from sound think.	
	shaving and singeing	<b>Notes:</b> Scald tank water may redistribute pathogen contamination from hair and blood (if head	
		has been removed) onto external surfaces of the goat or into neck wound.	
		Contamination of carcass from residual hair.	
		Notes: Salmonella is ubiquitous on goat hair.	
		Notes: Salmonella is ubiquitous on goat hair.  Temperature of scald tank water and/or transition time in tank may be insufficient to	

Activity Comment	
Skin-off	Refer to Cattle Transport Table
4.4b Legging, hide clearing and hide removal	Contamination of the carcass.
removar	<b>Notes:</b> Contamination of the carcass can occur via cross-contamination from hide and/or equipment
4.5 Bunging	Opportunity for faecal leakage onto carcass and into processing environment
	<b>Notes</b> : Washing pre-evisceration carcasses pre or post bunging can affect the carcass contamination from the rectum. Pooling in the rectal area from wash solution can influence carcass contamination
4.6 Evisceration	Refer to Cattle Transport Table
Potential for pathogens in faeces or gastrointestinal tract to contaminate carcass	
4.7 Post mortem	Refer to Cattle Transport Table
	Pathogenic organisms may be present in edible offal.
4.8 Trimming	Refer to Cattle Transport Table
4.9 Carcass washing (Optional)	Refer to Cattle Transport Table
4.10 Storage	Refer to Cattle Transport Table
4.11 Quartering, boning and packing	Opportunity for cross-contamination
and packing	Notes: Cross-contamination can occur from food handlers and/or equipment
4.12 Storage of packed meat	Refer to Cattle Transport Table

#### References

Branham LA, Carr MA, Scott CB, Callaway TR (2005) *E. coli* O157 and *Salmonella* spp. in white-tailed deer and livestock. Current Issues in Intestinal Microbiology 6(2):25–29

Das MS, Roy DK, Das S (1990) Occurence of salmonellae in slaughtered pigs, goat meat, meat handlers and slaughtered-house workers. Journal of Communicable Diseases 22:39–42

Dontorou A, Papadopolou C, Filioussis G, Apostolou I, Economou V, Kansouzidou A, Levidiotou S (2004) Isolation of a rare *Escherichia coli* O157:H7 strain from farm animals in Greece. Comparative Immunology Microbiology and Infectious Diseases 27(3):201–207

Duffy L, Barlow R, Fegan N, Vanderlinde P (2008) Prevalence and serotypes of *Salmonella* associated with goats at two Australian abattoirs. Letters in Applied Microbiology Ahead of print:1–5

European Food Safety Authority (2007) Monitoring and identification of human enteropathogenic *Yersinia* spp. The EFSA Journal

Molla W, Molla B, Alemayehu D, Muckle A, Cole L, Wilkie E (2006) Occurrence and antimicrobial resistance of *Salmonella* serovars in apparently healthy slaughtered sheep and goats of central Ethiopia. Tropical Animal Health and Production 38(6):455–462

Pritchard GC, Willshaw GA, Bailey JR, Carson T, Cheasty T (2000) Verocytotoxin-producing *Escherichia coli* O157 on a farm open to the public: outbreak investigation and longitudinal bacteriological study. Veterinary Record 147(10):259–264

Quilez J, Torres E, Chalmers RM, Hadfield SJ, del Cacho E, Sanchez-Acedo C (2008) *Cryptosporidium* genotypes and subtypes in lambs and goat kids in Spain. Appl Environ Microbiol 74(19):6026–6031

Rajmalliah N, Sreenivas RM, Dhananjaya RB, Ramakrishna RG (1989) Occurrence of *Salmonella* in slaughtered sheep and goats in Hyderabad. Indian Journal of Comparative Microbiology, Immunology and Infectious Diseases 10(4):193–196

SEIMIYA YM, SASAKI K, SATOH C, Takahashi M, YAEGASHI G, IWANE H (2005) Caprine enteritis associated with *Yersinia pseudotuberculosis* infection. The Journal of Veterinary Medical Science 67(9):887–890

Shane SM, Montrose MS (1985) The occurrence and significance of *Campylobacter jejuni* in man and animals. Veterinary Research Communications 9(1):167–198

Slee KJ, Button C (1990) Enteritis in sheep, goats and pigs due to *Yersinia pseudotuberculosis* infection. Australian Veterinary Journal 67(9):320–322

Vanderlinde, P., Duffy, P. and Barlow, S. (2003) Salmonella ecology in goat and goat meat. Report No. PRMS.027, Food Science Australia report for Meat and Livestock Australia.

Woldemariam E, Molla B, Alemayehu D, Muckle A (2005) Prevalence and distribution of *Salmonella* in apparently healthy slaughtered sheep and goats in Debre Zeit, Ethiopia. Small Ruminant Research 58(1):19–24

# 4. Pig Production in Australia

#### Introduction

Pork production occurs predominantly in the grain belts of Australia reflecting the reliance on grain as the major source of pig feed. Hence the quantity of pork produced in each state is linked to the size of the major grain growing regions, but is also influenced by proximity to major population centres.

In contrast to most other meat products, a significant proportion of pig meat consumed in Australia is imported. In 2012-13, imports accounted for around 47 percent of total pig meat consumption, and at least 70 percent of the bacon, ham and smallgoods consumed in Australia.

Australian pork is also exported to markets in Singapore, Hong Kong, Japan and New Zealand.

## **Pig Production**

The Australian pig industry comprises approximately 1500 pig producers producing around 4.7 million pigs annually (*personal communication, APL*). Pig production systems range from extensive outdoor farms to intensive operations.

The vast majority of pigs are intensively reared, using all-in all-out production strategies. This enhances disease management and enables producers to better meet market specifications. These all-in all-out systems generally use weekly batch farrowing methods, where sows are placed into groups to allow matings and farrowings to occur at distinct weekly intervals, making grouped movement and marketing of pigs more easily managed. Such systems make extensive use of artificial insemination.

In recent times there has been increasing use of off-site grow-out facilities, rather than single site farrow-to-finish operations. This minimises the transfer of infectious diseases from breeders to market pigs and also reduces stress. Under these production arrangements, there has been greater use of lower-cost 'shelter' facilities that group-house pigs on bedding (straw or rice hulls) rather than traditional sheds.

The use of outdoor production is increasing, practiced with sows and litters in southern Australia, with grower pigs usually brought into sheds or shelters after weaning.

Once grown to market size, pigs are taken to abattoirs for processing.

Average slaughter weights for Australian pigs are increasing as a result of genetic improvement, changing processor requirements, and industry efforts to achieve greater production efficiencies at farm and processing levels.

The key steps in the production and processing of pigs are summarised in Figure 4.

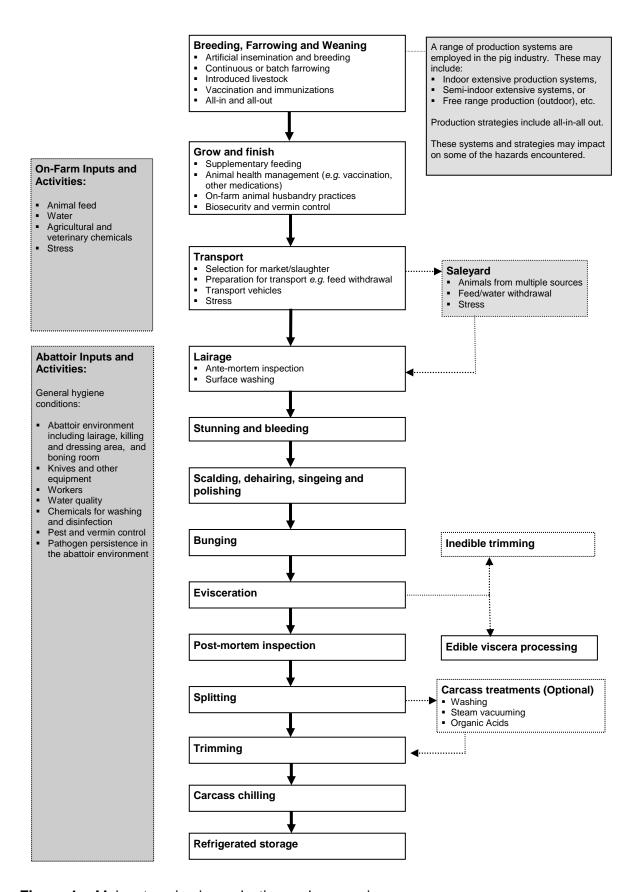


Figure 4: Major steps in pig production and processing

# **Abattoir Operations**

Most pigs in Australia are slaughtered in dedicated pig processing facilities.

Minor differences may exist depending on the plant's capabilities and design but the principal processing steps remain the same. Factors which may influence abattoir operations include: age of plant; chain speed; and whether the plant is an export registered facility.

## **Hazard Identification**

The following tables outline the microbiological hazards that may be encountered along the entire pig production and processing chain.

# (a) Pig Production

Input/ Activity	Comment	
1. Animal Production (in	ncluding birthing, health status, zoonoses)	
1.1 Growing the pigs to market condition	Pigs may carry pathogens with or without exhibiting any clinical signs.	
(Animal health status of	<b>Notes</b> : The following hazards may be found in the gastrointestinal tract and exterior surfaces of pigs:	
the pig)	1 6	
	Foodborne pathogens which have been more commonly associated with pigs include:  Salmonella spp.  Yersinia enterocolitica  Toxoplasma gondii	
	Campylobacter spp. (C. jejuni, C. coli) Listeria monocytogenes	
	Other possible foodborne pathogens associated with pigs include:  Y. pseudotuberculosis  Clostridium perfringens	
	Clostridium botulinum and Cl. difficile	
	Cryptosporidium parvum and C. suis	
	Pathogenic E. coli	
	Giardia lamblia	
	Sarcocystis suihominis	
	Staphylococcus aureus	
	Streptococcus suis Taenia solium and T. asiatica	
	Tuenta sonum ana 1. astanea	
	Notes: Carrier status includes the following states:  Animals showing clinical signs of disease due to infection with a pathogen  Super-shedder (i.e. high levels of pathogens are present in the animal's gut and are shed in high levels in their faeces)  Shedder (i.e. pathogens are present in the animal's gut contents and are therefore	
	shed in faeces)  Carrier (i.e. pathogens are present in organs but not gut content, therefore are not shed in faeces)	
	<b>Notes:</b> Different herd types and different production systems may have an impact on the microbiological status of the animals.	
	<b>Notes:</b> The prevalence of pathogens in the existing herd may increase when new stock is introduced.	
2. Animal Feed (includes	s pasture, grains, concentrates, meal etc)	
2.1 Pasture	A range of pathogens may be present in soil which can contaminate pigs.	
(Water/Soil/Faeces)	Refer Extensive Cattle Table	
	(outdoor production only)	
	<b>Notes:</b> For outdoor production systems, contamination may arise as a result of access to wild animals, birds and carrion. Pigs are known to readily eat both dead and living rodents and other wildlife including insects. Rodents, wildlife, flies and cockroaches can act as both	

Input/ Activity		Comment
T and January		vectors and reservoirs for pathogens in the farming environment. Carrion can be a reservoir
		of anaerobic bacterial pathogens.
		Important to note that pigs will have supplements beyond just pasture
2.2 Pasture		Refer Extensive Cattle Table
	I ustui v	
(Effl	uents)	(outdoor production only)
		Pasture may be contaminated with pathogens in effluents that are applied as soil fertilisers (ie manure and slurry.
2.3 (Incl	Feeds uding grains, meal,	Feeds including grain, meal, pellets and supplements may be contaminated with pathogens, which may result in a pathogen transmission to animals.
	ts, supplements)	<b>Notes</b> : Pigs are omnivores and therefore consume a wide range of feeds. Some studies indicate an association between pathogen infection and the feeding of particular ingredients, such as animal origin ingredients and by-product meal.
		Notes: The form in which the feed is presented may play a significant role in the pathogen prevalence in pigs.  Salmonella has been reported in stockfeed. Serovars and prevalence reported differ
		depending on type of feed.  A higher Salmonella sero-prevalence has been associated with feeding pelleted rations
		to finishers and feeding whey.
		<b>Notes:</b> Feed may become contaminated with pathogens during transport, storage or within the farm feeding system.
2.4	Silage	Not applicable to pigs.
2.5	Meat and bone meal	Feeding of meat and bone meal may be a source of TSE agents which may contaminate pigs.
	(MBM)	Notes: Meat and bone meal is permitted in pig rations.
Conc	centrates and	rotes. Weat and boile mear is permitted in pig rations.
	lements	
3.1	Drinking Water (inclu Consumption of	ding town, reticulated, ground, surface and run-off water)  Water may be a source of microbiological contamination for stock
3.1	town/reticulated	water may be a source of inicrobiological containmation for stock
	water	Refer to Extensive Cattle Table
3.2	Consumption of groundwater	Unprotected groundwater is prone to faecal contamination from livestock, wild animals, domestic pets and humans which may contain a wide range of pathogens and may contaminate pigs
		Refer to Extensive Cattle Table
3.3	Consumption of	Refer to Extensive Cattle Table  Natural waterways (e.g. creeks, rivers and dams) may be contaminated with pathogens
	surface water and run-off water	which could be a source of microbial contamination of pigs.
		Refer to Extensive Cattle Table
		(outdoor production only)
3.4	Consumption of	Refer to Extensive Cattle Table
4.	recycled water Animal Husbandry Pr	cactices (including veterinary chemicals, handling practices)
4.1	Stress caused by	Stress may impact on the animal's natural defence mechanisms resulting in an increased
	animal husbandry	susceptibility to pathogens. Stress also causes increased pathogen shedding in the faeces.
conditions, changes in feed types and watering, handling and transport of new animals into existing herds, weaning, unfamiliar noise and smel		Refer Extensive Cattle Table
		Notes: Stressors include grouping unfamiliar animals together, changes in climate conditions, changes in feed types and watering, handling and transport of pigs, introduction
		of new animals into existing herds, weaning, unfamiliar noise and smells, high stocking
4.2	Medication of pigs	densities, restraining, husbandry practices.  Incorrect use of therapeutics and other antimicrobials may lead to the emergence of resistant microorganisms.
		microorganisms.
		Refer Extensive Cattle Table

Input/ Activity		Comment	
**		<b>Notes</b> : <i>Salmonella Typhimurium</i> DT 104 with multi-resistance to ampicillin, streptomycin, tetracyclines, chloramphenicol and spectinomycin is endemic in overseas pork industry. No reports of DT 104 within the Australian domestic pork industry.	
5. Environment (including housing systems, premises, buildings and equipment, personnel)			
5.1	Housing types	Types of housing may influence the types of pathogens that pigs may carry or be contaminated with.	
		<b>Notes</b> : Factors influencing pathogen status include type of separation between units, type of pens, possibility of snout contact between pens, type of floor including whether dry or	
		straw-bedded floor, partitions close-fitted to floor, quarantine facility, hygienic-lock	
		facilities.	
5.2	Environmental	Pigs may become directly contaminated by pathogens derived from environmental sources.	
	contamination of the		
	farming	<b>Note</b> : Some foodborne pathogens are ubiquitous in the farming environment, while others	
	environment	may be introduced into the farming environment by poor biosecurity practices via visitors,	
		vehicles, rodents, wild animals, pet animals, carrions, houseflies and other insects such as	
		cockroaches.	

# (b) Transport, Saleyards, Lairage, Slaughter and Carcass Dressing Operations

Input/Activity	Comment	
All or most activities - transport and saleyards	Contamination, injury or other matters that could impact on the health or suitability of pigs for meat production occur because personnel lack skills and knowledge to implement	
	practices that avoid injury to pigs, assess suitability for slaughter or other matters that could impact on the safety or suitability of pigs for meat production or the meat.	
All or most activities- lairage, slaughter and carcass dressing operations.	Contamination, injury or other matters that could impact on the health or suitability of pigs for meat production occur because personnel lack skills and knowledge to implement practices that avoid injury to pigs, assess suitability for slaughter or other matters that could impact on the safety or suitability of pigs for meat production or the meat.	
	Contamination from personnel involved in slaughter and meat production	
	Contamination from premises and equipment	
	Contamination from premises and equipment and personnel	
1. Preparation and Transport to Market/Abattoir		
1.1 Selection of pigs and	Dirty pigs may increase the likelihood of pathogen contamination onto carcass from external	
handling operations	surfaces during the slaughtering and dressing process.	
(According to dirtiness)	<b>Notes:</b> Skin dirtiness is influenced by a number of factors, such as; production system	
(recording to direction)	(intensive, extensive, sheds with bedding systems), age, journey time, feeding regime.	
1.2 Transport vehicles	Pathogens may contaminate pigs via cross-contamination from the transport vehicle.	
	<b>Notes</b> : Transport vehicle may be contaminated with pathogens from previous loads. The washing procedures used for the vehicle may be insufficient for effective pathogen elimination.	
	Stress during transportation and associated handling may result in increase shedding of pathogens in faeces. Stress may also induce non-shedding carrier animals to start shedding.  Notes: Stress factors include noise, smells, mixing with unfamiliar pigs from other rearing pens or farms, high stocking densities, feed and water deprivation, transportation time, change in environment including temperature.	
1.3 Feed Curfew	Pathogen load in the animal may increase when they are deprived of feed and water prior to and during transportation. Extended time in lairage off feed may also increase pathogen load in the animal.	
	<b>Notes</b> : There was reported correlation with feed withdrawal times with the number of pathogens in the caecal content. APIQ requires pigs to be slaughtered between 6 – 24 hours after they have been removed from feed to minimise possible <i>Salmonella</i> contamination of the carcass. May also reduce vomiting during transport.	
2. Saleyards		
2.1 Holding and processing	Pathogen transfer between animals in saleyard pens due to mixing animals from multiple sources.	

Input/Activity		Comment		
		Notes: Saleyards constitute a very small percentage of the domestic farmed pig industry.		
3.	Lairage	Discould demonstrate the second define and second s		
3.1	Ante- mortem	Diseased, downer and dying animals may get through to slaughter.		
		<b>Notes</b> : Identification of animals that may be displaying symptoms of disease or conditions that would make them unfit for human consumption, and/or may compromise the integrity of the slaughterhouse		
		Time held in lairage may increase in pathogen load within the animal.		
		<b>Notes</b> : Time pigs are held in lairage prior to slaughter can affect the pathogen load in the gastrointestinal tract. There was a reported correlation with feed and water withdrawal times with the number of pathogens in the caecal content in pigs. 'Carrier pigs' ( <i>i.e.</i> pigs which are infected but not shedding) may start shedding during lairage.		
		The lairage environment can become contaminated which may be transferred to pigs.		
		<b>Notes</b> : Transfer of potential pathogens can occur between animals via physical contact <i>eg</i> . skin soiled with faeces and dust or through oral & nasal contact. The following pathogens have been identified in faeces or rectal samples of animals in lairage:		
		Cleaning and disinfection of the lairage pen may not effectively reduce pathogen load.		
		<ul> <li>Notes: The following pathogens have been identified in the lairage environment:</li> <li>Salmonella spp.</li> <li>Salmonella spp.</li> </ul>		
		Yersinia enterocolitica		
4. Slaughtering Operations				
4.1	Pig washing	Excessive levels of soil, dust and faeces on animals represent a source of contamination.  Washing may not remove all microorganisms from the skin or may spread localised contamination.		
		Notes: Microorganisms detected on pigs post-washing include: <ul><li>Salmonella spp.</li></ul>		
4.2	Stunning & bleeding	Contamination of the slaughtering and processing environment		
		<b>Notes</b> : Stunning method should ensure adverse effects such as blood-splash and fractures are avoided.		
		The following pathogens have been detected on pigs post-bleeding: <ul><li>Salmonella spp.</li></ul>		
		<ul><li>Listeria spp. (L. monocytogenes)</li><li>Coagulase-positive Staphylococcus aureus</li></ul>		
		Contamination of animals from abattoir environment		
		Notes Misses and described in the shortest stronging to the discount of the di		
		Notes: Microorganisms detected in the abattoir stunning & bleeding area include:  • Yersinia enterocolitica		
		<ul> <li>Listeria monocytogenes</li> </ul>		
		Salmonella spp.  Mathicillin resistant Stanbylococcus auraus		
		Methicillin resistant Staphylococcus aureus     Sticking may internalise surface bacterial pathogens		
4.3	Scalding	Scald tank may not sufficiently reduce pathogen load on carcass.		
	Ü	Notes: Microorganisms detected on pigs post-scalding include:		
		Salmonella spp.  Granden idia Stanlar and a second a second and a second a second and a second		
Coagulase positive Staphylococcus aureus  Contamination of carcase from scald tank environment		Coagulase positive Staphylococcus aureus  Contamination of carcase from scald tank environment.		
		<b>Notes</b> : Scald tank is a potential source of bacterial contamination if temperature drops or the level of organic matter is high.		
l				

Input/Activity		Comment			
4.4 Dehairing		Dehairing process may redistribute existing bacterial contamination more evenly over the			
		carcass.			
		Notes: Microorganisms detected on pigs post-dehairing include:			
		■ Salmonella spp.			
		■ Coagulase positive Staphylococcus aureus			
		Contamination of the carcass from the dehairing equipment.			
		<b>Notes:</b> Dehairing equipment may force faeces out of the anus, contaminating the equipment			
		and carcass			
4.5	Cincoina				
4.5	Singeing	Pathogen contamination may remain on carcass post singeing especially in skin folds, ears or hair follicles.			
4.6	D !! ! !				
4.6	Polishing	The polishing process may redistribute existing bacterial contamination on the skin more evenly over the carcass.			
		evenly over the calcuss.			
		Notes: Microorganisms detected on pigs post-polishing include:			
		<ul><li>Staphylococcus aureus</li><li>Salmonella spp.</li></ul>			
		Listeria monocytogenes			
		Contamination of animals from abattoir polishing environment			
4.7	Pre-evisceration	Washing may spread localised microorganisms on the skin to other areas of the carcass			
washing may spread localised inicroorganishs on the skill to other are					
		<b>Notes:</b> Microorganisms detected on pigs post-evisceration washing include:			
4.8	Bunging	<ul> <li>Salmonella spp.</li> <li>Opportunity for faecal leakage onto carcass and into processing environment.</li> </ul>			
	Dunging .	opportunity for faceur readings onto careago and into processing environment.			
		<b>Notes</b> : Faeces contains potentially hazardous bacteria which include:			
		<ul><li>Listeria spp.</li><li>Salmonella spp.</li></ul>			
		Toxoplasma gondii			
		<ul> <li>Campylobacter jejuni/coli</li> </ul>			
		Yersinia enterocolitica  Cross contamination between carcasses and bunging equipment and environment.			
		Cross containmation between careasses and bunging equipment and environment.			
		Notes: Microorganisms detected on bunging equipment include:			
4.9	Caraga anarina	<ul> <li>Salmonella spp. detected on the rectal pistol (used prior to evisceration)</li> <li>Cross contamination from equipment to carcasses</li> </ul>			
4.9	Carcase opening	Cross contamination from equipment to carcasses			
		Notes: Microorganisms detected in carcase-opening environment include:			
4 10	Evisceration	<ul> <li>Salmonella spp. detected on knife blades</li> <li>Opportunity for faecal contamination of carcasses, utensils and slaughtering environment if</li> </ul>			
4.10	Evisceration	carried out incorrectly.			
		<b>Notes:</b> Potential pathogens identified in pigs which may cause carcass contamination if			
		evisceration is carried out incorrectly include:  Salmonella spp.			
		■ Toxoplasma gondii			
		Campylobacter jejuni/coli			
		<ul><li>Listeria spp.</li><li>Yersinia enterocolitica</li></ul>			
		Torsula emerocolinea			
4.11	Post-mortem	Macroscopic evidence of disease or faecal contamination of the carcass.			
		Incision of tissues during post-mortem inspection may be a source of contamination for the			
		slaughter house environment and the carcasses			
		<b>Notes:</b> Microorganisms detected in tissues which may be inspected during post-mortem include:			
		include:  Salmonella spp.			
		■ Campylobacter spp.			
		Yersinia enterocolitica			

Input/Activity	Comment		
	<b>Notes:</b> A study in Australia demonstrated similar level of contamination occurred when using either traditional (incision) and risk-based (visual) post-mortem inspection.		
	Pathogenic organisms may be present in edible offal.		
	Notes: Pathogens detected in pig offal include:  - Yersinia enterocolitica - Listeria spp Salmonella spp - Campylobacter spp.		
	Notes: Contaminated equipment/environment may transfer microorganisms to edible offal		
4.12 Trimming	Carcass contamination.		
	Notes: An opportunity to remove tissue and any other contamination, however some contamination may be missed and remain on carcass  Coagulase positive <i>S. aureus</i> was detected on neck, belly, back and ham of carcasses		
4.13 Washing	Washing may introduce or spread existing contamination over the carcass. It may also provide a moist environment for pathogens to survive.		
	Notes: Microorganisms detected post-washing include:  Coagulase positive S. aureus  Yersinia enterocolitica  S. aureus  Salmonella spp Listeria monocytogenes		
4.15 Storage	Opportunity for outgrowth of pathogens		
	Refer to Cattle Transport Table  Notes: Carcass cooling rate depend on size, air temperature and flow rate and position of the carcase in the cooling chamber.		
4.16 Splitting, Boning, packing	Contamination of carcass during the splitting, boning and packaging process		
	<b>Notes</b> : Opportunity for cross-contamination between carcasses/portions and the processing environment		
	Possible microbiological contaminants include:  Listeria monocytogenes S. aureus Salmonella spp Clostridium perfringens Yersinia enterocolitica Campylobacter spp.		
4.18 Storage of packed meat	Potential for outgrowth of pathogens.  Refer to Cattle Transport Table		

#### References

Alban L, Stärk KDC (2005) Where should the effort be put to reduce the *Salmonella* prevalence in the slaughtered swine carcass effectively? Preventive Veterinary Medicine 68(1):63–79

Autio T, Sateri T, Fredriksson-Ahomaa M, Rahkio M, Lunden J, Korkeala H (2000) *Listeria monocytogenes* contamination pattern in pig slaughterhouses. Journal of Food Protection 63(10):1438–1442

Berends BR, Snijders JMA, van Logtestijn JG (1993) Efficacy of current EC meat inspection procedures and some proposed revisions with respect to microbiological safety: a critical review. Vet Rec 133(17):411–415

Bolton DJ, Pearce RA, Sheridan JJ, Blair IS, McDowell DA, Harrington D (2002) Washing and chilling as critical control points in pork slaughter hazard analysis and critical control point (HACCP) systems. Journal of Applied

Microbiology 92(5):893-902

Botteldoorn N, Heyndrickx M, Rijpens N, Grijspeerdt K, Herman L (2003) *Salmonella* on pig carcasses: positive pigs and cross contamination in the slaughterhouse. Journal of Applied Microbiology 95(5):891–903

Craven JA, Hurst DB (1982) The effect of time in lairage on the frequency of *Salmonella* infection in slaughtered pigs. Journal of Hygiene Cambridge 88:107–111

Das MS, Roy DK, Das S (1990) Occurrence of salmonellae in slaughtered pigs, goat meat, meat handlers and slaughtered-house workers. Journal of Communicable Diseases 22:39–42

de Neeling AJ, van den Broek MJM, Spalburg EC, van Santen-Verheuvel MG, Dam-Deisz WDC, Boshuizen HC, van de Giessen AW, van Duijkeren E, Huijsdens XW (2007) High prevalence of methicillin resistant *Staphylococcus aureus* in pigs. Veterinary Microbiology 122(3-4):366–372

European Food Safety Authority (2007) Monitoring and identification of human enteropathogenic *Yersinia* spp. The EFSA Journal

Fedorka-Cray PJ, Collins Kelley L, Stabel TJ, Gray JT, Laufer JA (1995) Alternative routes of invasion may affect pathogenesis of *Salmonella typhimurium* in swine. Infection and Immunity 63(7):2658–2664

Fedorka-Cray PJ, Hogg A, Gray JT, Lorenzen K, Valasquez J, Von Behren P (1997) Feed and feed trucks as sources of *Salmonella* contamination in swine. Swine Health and Production 5(5):189–193

Fredriksson-Ahomaa M, Björkroth.J., Hielm S, Korkeala H (2000) Prevalence and characterization of pathogenic *Yersinia enterocolitica* in pig tonsils from different slaughterhouses. Food Microbiology 17(1):93–101

Gill CO, Dussault F, Holley RA, Houde A, Jones T, Rheault N, Rosales A, Quessy S (2000) Evaluation of the hygienic performances of the processes for cleaning, dressing and cooling pig carcasses at eight packing plants. International Journal of Food Microbiology 58(1-2):65–72

Gobat PF, Jemmi T (1990) Epidemiological studies on *Listeria spp* in slaughterhouses. Fleischwirtschaft 70(12):1448–1450

Grosspietsch R, Einschutz K, Jaeger D, Fries R (2006) Survey on the hygienic status of plastic doors of a pig abattoir. Journal of Food Protection 69(11):2738–2741

Gurtler M, Alter T, Kasimir S, Linnebur M, Fehlhaber K (2005) Prevalence of *Yersinia enterocolitica* in fattening pigs. Journal of Food Protection 68(4):850–854

Hamilton, D., Bobbitt, J., Pointon, A., Lester, S., Coates, K. and Dahl, J. (1999) *Benchmarking the Salmonella Status of Australian Pig Herds*. Report No. Final report DAS 34/1014, PRDC, Appendix 8.

Hamilton, D., Bobbitt, J.L., Dahl, J., Coates, K., Lester, S. & Pointon, A.M. (2000a): Risk Factors for within herd *Salmonella* infection of pigs in Australia. In *Proceedings of the 16th Congress of the International Pig Veterinary Society*. Cargill, C., and McOrist, S. (eds). Melbourne, Australia: IPVS, p. 204.

Hamilton, D., Smith, P. and Pointon, A. (2007) National Salmonella and E. coli Monitoring (ESAM) data from Australian pig carcases from 2000 to 2006. *In: Proceedings of 7th International Symposium on the Epidemiology and Control of Foodborne Pathogens in Pork, 7th International Symposium on the Epidemiology and Control of Foodborne Pathogens in Pork, Verona, Italy.* pp129-132.

Hamilton.D.R., Holds, G., Bobbitt, J., Kiermeier, A., Holyoake, P., Fahy, T., Davos, D., Heuzentoeder, M., Lester, S. and Pointon, A. (2005) Case studies of the ecology of Salmonella infection across major Australian pig production systems, including bedding-rearing systems. 19 February 9 A.D.

Kijlstra A, Jongert E (2008) Control of the risk of human toxoplasmosis transmitted by meat. International Journal for Parasitology 38(12):1359–1370

Knudtson LM, Hartman PA (1993) Enterococci in pork processing. Journal of Food Protection 56(1):6-9

Kotula AW (1987) Control of extrinsic and intrinsic contamination of pork. In: Smulders FJM (ed) Elimination of pathogenic organisms from meat and poultry. Elsevier Science Publishers B.V., Amsterdam, The Netherlands, p. 181–201

Kotula AW, Emswilerrose BS (1988) Airborne microorganisms in a pork processing establishment. Journal of Food Protection 51(12):935–937

Lake R, Hudson A, Cressey P (2002) Risk profile: *Toxoplasma gondii* in red meat and meat products. FW0138. Institute of Environmental Science & Research Limited, Christchurch, New Zealand. <a href="http://www.nzfsa.govt.nz/science/risk-profiles/toxoplasma-gondii-in-red-meat.pdf">http://www.nzfsa.govt.nz/science/risk-profiles/toxoplasma-gondii-in-red-meat.pdf</a>

Laukkanen R, Martinez PO, Siekkinen KM, Ranta J, Maijala R, Korkeala H (2008) Transmission of *Yersinia pseudotuberculosis* in the pork production chain from farm to slaughterhouse. Appl Environ Microbiol 74(17):5444–5450

Leblanc D, Ward P, Gagné MJ, Poitras E, Müller P, Trottier YL, Simard C, Houde A (2007) Presence of hepatitis E virus in a naturally infected swine herd from nursery to slaughter. International Journal of Food Microbiology 117(2):160–166

Li W, She R, Wei H, Zhao J, Wang Y, Sun Q, Zhang Y, Wang D, Li R (2009) Prevalence of hepatitis E virus in swine under different breeding environment and abattoir in Beijing, China. Veterinary Microbiology 133(1-2):75–83

Linton AH, Hinton MH (1987) Prevention of microbial contamination of redmeat in the antemortem phase: epidemiological aspects. In: Smulders FJM (ed) Elimination of pathogenic organisms from meat and poultry. Elsevier Science Publishers B.V., Amsterdam, The Netherlands, p. 9–25

Lo Fo Wong D, Hald T, Wolf P, Swanenburg M (2002) Epidemiology and control measures for *Salmonella* in pigs and pork. Livestock Production Science 76(3):215–222

Lo Fo Wong DMA, Dahl J, Stege H, van der Wolf PJ, Leontides L, von Altrock A, Thorberg BM (2004) Herd-level risk factors for subclinical *Salmonella* infection in European finishing-pig herds. Preventive Veterinary Medicine 62(4):253–266

Mannion C, Egan J, Lynch BP, Fanning S, Leonard N (2008) An investigation into the efficacy of washing trucks following the transportation of pigs - a *Salmonella* perspective. Foodborne Pathogens and Disease 5(3):261–271

Martín-Peláez S, Martín-Orúe SM, Pérez JF, Fàbrega E, Tibau J, Gasa J (2008) Increasing feed withdrawal and lairage times prior to slaughter decreases the gastrointestinal tract weight but favours the growth of cecal *Enterobacteriaceae* in pigs. Livestock Science 119(1-3):70–76

Martín-Peláez S, Peralta B, Creus E, Dalmau A, Velarde A, Pérez JF, Mateu E, Martín-Orúe SM (2009) Different feed withdrawal times before slaughter influence caecal fermentation and faecal *Salmonella* shedding in pigs. The Veterinary Journal In Press, Corrected Proof

Miller MF, Carr MA, Bawcom DB, Ramsey CB, Thompson LD (1997) Microbiology of pork carcasses from pigs with differing origins and feed withdrawal times. Journal of Food Protection 60(3):242–245

Nesbakken T (1988) Enumeration of *Yersinia enterocolitica* O:3 from the porcine oral cavity, and its occurrence on cut surfaces of pig carcasses and the environment in a slaughterhouse. International Journal of Food Microbiology 6(4):287–293

Nesbakken T, Eckner K, Hoidal HK, Rotterud OJ (2003) Occurrence of *Yersinia enterocolitica* and *Campylobacter* spp. in slaughter pigs and consequences for meat inspection, slaughtering, and dressing procedures. International Journal of Food Microbiology 80(3):231–240

Nollet N, Maes D, De Zutter L, Duchateau L, Houf K, Huysmans K, Imberechts H, Geers R, de Kruif A, Van Hoof J (2004) Risk factors for the herd-level bacteriologic prevalence of *Salmonella* in Belgian slaughter pigs. Preventive Veterinary Medicine 65(1-2):63–75

Pearce RA, Sheridan JJ, Bolton DJ (2006) Distribution of airborne microorganisms in commercial pork slaughter processes. International Journal of Food Microbiology 107(2):186–191

Pearce RA, Bolton DJ, Sheridan JJ, McDowell DA, Blair IS, Harrington D (2004) Studies to determine the critical control points in pork slaughter hazard analysis and critical control point systems. International Journal of Food Microbiology

Pedersen KB (1979) Occurence of *Yersinia enterocolitica* in the throat of swine. Contributions to Microbiology and Immunology 5:253–256

Roberts TA, Hudson WR (1987) Contamination prevention in the meat plant: the standpoint of an importing country. In: Smulders FJM (ed) Elimination of pathogenic organisms from meat and poultry. Elsevier Science Publishers B.V., Amsterdam, The Netherlands, p. 235–250

Saidealbornoz JJ, Knipe CL, Murano EA, Beran GW (1995) Contamination of pork carcasses during slaughter, fabrication, and chilled storage. Journal of Food Protection 58(9):993–997

Shane SM, Montrose MS (1985) The occurrence and significance of *Campylobacter jejuni* in man and animals. Veterinary Research Communications 9(1):167–198

Skovgaard N (1987) Prevention of microbial contamination in the ante mortem phase: the SPF (specific pathogen-free) concept. In: Smulders FJM (ed) Elimination of pathogenic organisms from meat and poultry. Elsevier Science Publishers B.V., Amsterdam, The Netherlands, p. 39–56

Spescha C, Stephan R, Zweifel C (2006) Microbiological contamination of pig carcasses at different stages of slaughter in two European Union-approved abattoirs. Journal of Food Protection 69(11):2568–2575

Stärk K, Wingstrand A, Dahl J, Møgelmose V, Lo Fo Wong DMA (2002) Differences and similarities among experts' opinions on *Salmonella enterica* dynamics in swine pre-harvest. Preventive Veterinary Medicine 53(1-2):7–20

Warriner K, Aldsworth TG, Kaur S, Dodd CER (2002) Cross-contamination of carcasses and equipment during pork processing. Journal of Applied Microbiology 93(1):169–177

Warriss PD (2003) Optimal lairage times and conditions for slaughter pigs: a review. Veterinary Record 153(6):170–176

Wehebrink T, Kemper N, Beilage EG, Krieter J (2008) Prevalence of *Campylobacter spp.* and *Yersinia spp.* in the pig production. Berliner und Munchener Tierarztliche Wochenschrift 121(1-2):27–32

Williams LP, Newell KW (1970) Salmonella excretion in joy-riding pigs. American Journal of Public Health and the Nation's Health 60(5):926–929

Zewde BM, Robbins R, Abley MJ, House B, Morrow WEM, Gebreyes WA (2009) Comparison of swiffer wipes and conventional drag swab methods for the recovery of *Salmonella* in swine production systems. Journal of Food Protection 72(1):142–146

Zheng DM, Bonde M, Sørensen JT (2007) Associations between the proportion of *Salmonella* seropositive slaughter pigs and the presence of herd level risk factors for introduction and transmission of *Salmonella* in 34 Danish organic, outdoor (non-organic) and indoor finishing-pig farms. Livestock Science 106(2-3):189–199

# Summary

The microbiological status of meat is influenced by factors along the entire meat supply chain. While a vast array of microbiological hazards could potentially contaminate the carcass, only a small number of these pathogens may present a risk to consumers if unmanaged. The hazard tables list a wide range of microbiological hazards that may be found on the carcasses originating from cattle, sheep, goats and pigs.

The principal microbiological hazards identified in the on-farm phase of meat production and after slaughtering operations include pathogenic *E. coli* and *Salmonella* spp., although there is some variation between meat species. Pathogens which have more commonly been associated with the main species are listed below:

Animal	Principal microbiological hazards		
Cattle	Pathogenic Escherichia coli, Salmonella spp., Campylobacter jejuni and C. coli,		
Sheep	Pathogenic Escherichia coli and Salmonella spp.		
Goats	Pathogenic Escherichia coli and Salmonella spp.		
Pigs	Salmonella spp., Yersinia enterocolitica, Toxoplasma gondii, Campylobacter jejuni and C. coli.		

During the animal production phase, there are a number of key inputs and activities which influence the manner in which hazards may be introduced or amplified. They are summarised below:

Input and/ or activity	Comment	Step in chain where control may be applied
Animal health	Pathogens may exist in the animal with or without exhibiting clinical signs	Animals with clinical signs of disease or illness are identified and managed at:  • Dispatch from farm/saleyard • Arrival at abattoir • Ante-mortem inspection  Without clinical signs, potential hazards may be identified and managed at: • Slaughter to minimise contamination from external surfaces or internal spillage • Post-mortem inspection
Stress	Animals may be more susceptible to infection and/or have increased faecal shedding. Pathogens colonise the gut	Minimise exposure of animals to stress during:
Feed	Feed has the potential to introduce pathogens into the gut or environment	Management of input of manure and fertiliser onto pasture Control supplements Oversight of ensilage operations
Water	Contributes to internal and external contamination	Access of animals to suitable drinking water
Environment and management of biosecurity	Pathogens may contaminate external surfaces of animal, or can lead to ingestion or infection of the animal	Pasture management Vermin and pest control Good agricultural practices Sound animal husbandry

In summary, there are two main sources of contamination to the meat carcass:

- External contamination From the animal (hide, skin, fleece, hooves, faeces, etc) and the environment, and;
- Internal contamination During evisceration and dressing operations and following spillage of gastro-intestinal tract contents.

Abattoir and slaughtering operations are currently mandated under the Australian Standard *AS4696* to ensure that meat produced for human consumption is wholesome and safe. A large number of producers in Australia adhere to a voluntary on-farm quality assurance program (Livestock Production Assurance; LPA). The accreditation system is underpinned by an on-farm property risk assessment component and utilises a voluntary National Vendor Declaration (NVD) and mandated National Livestock Identification System (NLIS) for quality assurance livestock traceability.

One additional concern has been the potential transmission of antimicrobial resistant (AMR) microorganisms to humans via food. Control measures on farm and during processing are designed to reduce the likelihood of microbial contamination of meat, irrespective of the microbial pathogens' resistance profile. The Department of Health is working with the Department of Agriculture to develop a National Antimicrobial Resistance Prevention and Containment Strategy (the National AMR Strategy) under the direction of the Antimicrobial Resistance Prevention and Containment Steering Group. That National AMR Strategy will take a One Health approach, ensuring consistent responses to AMR across sectors. The Steering Group has endorsed seven key elements of the National AMR Strategy: surveillance; international engagement; regulation; governance; communication; infection prevention and control; and research.

During the hazard assessment, a number of pathogenic (zoonotic) microorganisms were identified, and while the oral route may not be the normal route of human infection, it is plausible or potentially possible that consumers may become infected by handling raw meat, through cross-contamination, or by the ingestion of meat which has not been thoroughly cooked. In summary, leptospirosis may be controlled by vaccination of cattle and therefore presents little risk to consumers. There is limited scientific evidence attributing transmission of anthrax, melioidosis and Q fever to humans through ingestion. Available data indicates the primary mode of transmission is via inhalation or cutaneous exposure rather than through ingestion. Although ingestion is plausible as a transmission route for human infection, it is likely to be of minimal risk in Australia.

The findings of this assessment are consistent with the significant body of evidence that exists for the Australian domestic meat industry indicating that domestically-reared red meat (cattle, sheep, goats) and pigs, processed under existing standards, present a low risk to public health. Also evidenced is that industry personnel are fairly mature in their knowledge and management of food safety risks.

Further, considerable data is available to support the safety of meat and meat products produced from beef, sheep and pork in Australia. The evidence suggests that Australian meat from these species has a low microbial load and generally low prevalence of pathogens. Many of the pathogens listed in this assessment occur infrequently or not at all on Australian meat.





#### Appendix 1: Foodborne Disease Outbreaks Associated with Meat

These data are provisional and subject to change. Please quote as "OzFoodNet Unpublished Data, 2009"Please clear ALL citations of this internal brief in reports for public release.

# Prepared by: Katrina Knope, Polly Wallace, and Katie Fullerton April 2009

#### Introduction

Meat products are a common cause of foodborne outbreaks in Australia. An analysis of the OzFoodNet Outbreak Register was conducted in order to study the burden, causes and settings of these outbreaks. The OzFoodNet Outbreak Register contains data on outbreaks across Australia from January 2003 to June 2008.

## Nature of report

This report summarises outbreaks of human illness associated with meat, not including poultry, which occurred between January 2003 and June 2008.

#### **Data analysis**

This analysis was carried out in the following manner:

- Reports of outbreaks were extracted from the database using the following search terms:
- [Field: Year]: >=1 January 2003 And <= 30 June 2008</li>
- [Field: Transmission]: Foodborne Or Suspected Foodborne
- [Field: Food vehicle]: Like \*meat\* Or Like \*lamb\* Or Like \*pork\* Or Like \*bacon\* Or Like \*ham\* Or Like \*sausage\* Or Like \*steak\* Or Like \*frank\* Or Like \*beef\* Or Like \*kebab\* Or Like \*fillet\* Or Like \*roast\* Or Like \*carne\*
- [Field: Remarks]: Like \*meat\* Or Like \*lamb\* Or Like \*pork\* Or Like \*bacon\*
   Or Like \*ham\* Or Like \*sausage\* Or Like \*steak\* Or Like \*frank\* Or Like
   \*beef\* Or Like \*kebab\* Or Like \*fillet\* Or Like \*roast\* Or Like \*carne\*
- The 'Remarks' field was reviewed and where appropriate data on 'Food vehicle' were recoded to ensure consistency during analysis. Where the food vehicle field was unknown and information was found in the remarks field the food vehicle field was filled in
- Data were cleaned and recoded to provide consistent categories for data fields, including aetiological agents and food vehicles.
- Outbreaks were categorized as Meat, Dish containing meat, Suspected meat, or Suspected dish containing meat
  - Meat: outbreaks with sufficient descriptive or epidemiologic information to implicate a meat product
  - Dish containing meat: outbreaks with sufficient descriptive or epidemiologic information to implicate a dish containing meat
  - Suspected meat: outbreaks with insufficient descriptive or epidemiologic information to implicate a meat product, but high degree of investigator suspicion
  - Suspected dish containing meat: outbreaks with insufficient descriptive or epidemiologic information to implicate a dish containing meat, but high degree of investigator suspicion
- Outbreaks with only chicken as the identified food vehicle were excluded, however, outbreaks where chicken and another meat product, such as lamb or beef, were implicated were included in the analysis.
- Fish as a food vehicle was excluded from analysis.

 Data were analysed in Excel 2000 to summarise the number of people ill and hospitalised for different settings for outbreaks, mode of transmission, pathogen and implicated food vehicle.

# Outbreaks associated with meat, January 2003 to June 2008

OzFoodNet epidemiologists reported a total of 653 outbreaks of foodborne or suspected foodborne disease from January 2003 to June 2008, which represented 28% (653/2304) of all outbreaks reported. Ten percent (66/653) of these outbreaks were related to the consumption of meat or dishes containing meat, not including poultry.

In total, there were 66 meat-associated outbreaks affecting at least 1005 people, with 52 people hospitalised and no deaths. The mean number of people affected in these outbreaks was 15 people, with a range of 2 to 100 people. The largest number of meat-associated outbreaks in one year was 19 outbreaks in 2005.

Forty eight percent (32/66) of meat-associated outbreaks occurred in New South Wales, 21% (14/66) in Queensland, 14% (9/66) in Victoria, 6% (4/66) in Western Australia, 5% (3/66) in each of Northern Territory and South Australia, and 2% (1/66) in the Australian Capital Territory.

Forty three percent (29/66) of the outbreaks occurred in restaurants and 14% (9/66) were associated with takeaway food (Figure 1). Eleven percent (7/66) of the outbreaks were associated with a commercial caterer, 8% (5/66) at private residences. In 8% (5/66) of outbreaks investigators listed the setting where the food was prepared as "other unspecified settings".

An aetiological agent was identified in 55% (36/66) of the meat-associated outbreaks (Table 1). A variety of *Salmonella* serotypes were responsible for 27% (18/66) of the outbreaks, of these 12 (67%) were *Salmonella* Typhimurium. The other *Salmonella* serotypes were Anatum, Bovismorbificans, Johannesburg, Oslo, Zanzibar, and 4,12:d:-. Twelve percent of outbreaks (8/66) were due to *Clostridium perfringens*, 6% (4/66) were due to norovirus, and 5% (3/66) were due to staphylococcal toxin. There were individual outbreaks due to *Campylobacter* (not speciated), *Listeria monocytogenes*, and *Bacillus cereus*.

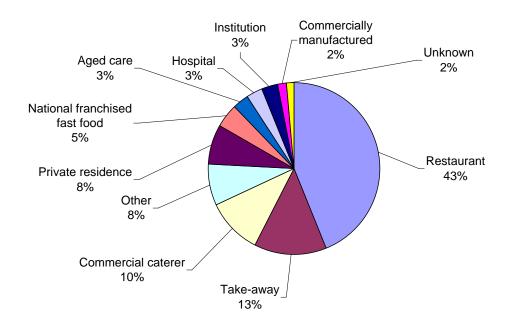
Of the 66 meat-associated outbreaks, 20% (13/66) had the food vehicle categorised as meat, 35% (23/66) had the food vehicle categorised as a dish containing meat, 17% (11/66) had the food vehicle categorised as suspected meat, and 29% (19/66) had the food vehicle categorised as suspected dish containing meat.

#### **Conclusions**

From January 2003 to June 2008 there were 66 outbreaks associated with meat in Australia. The majority of these outbreaks were due to a dish containing a meat product. Meat products cause a considerable amount of foodborne disease in Australia, particularly due to various *Salmonella* serotypes and toxin based poisonings due to *Clostridium perfringens* and *Staphylococcus aureus*. The under cooking of meat and temperature abuse after cooking are major causes of meat-associated outbreaks.

This summary is subject to at least two limitations. First, it is likely that other outbreaks thought to be caused by cross-contamination with meat or meat juices during preparation have not been captured in this summary. Cross-contamination as the cause of an outbreak is very difficult to assess and are not captured in these data. Second, it can be very difficult to categorise and summarise aggregated outbreak data by commodity. In this instance, the commodity 'meat' covers a large variety of different meat products, and, the identification of outbreaks that are due to a meat product or a dish containing a meat product is limited by the quality of the data collected. These data are often free-text, subjective summaries that do not uniformly report food vehicles by commodity type.

**Figure 1:** Settings where food was prepared in outbreaks of foodborne illness associated with meat, OzFoodNet, January 2003 to June 2008 (*n*=66).



**Table 1.** Aetiologic agent in outbreaks of foodborne illness associated with meat, OzFoodNet, January 2003 to June 2008 (n=66).

Aetiology	Outbreaks
Salmonella Typhimurium	12
Clostridium perfringens	8
Salmonella 'Other'	6
Norovirus	4
Staphylococcus aureus	2
Suspected Staphylococcal	
toxin	1
Listeria monocytogenes	1
Campylobacter	1
Bacillus cereus	1
Unknown	30
Total	66

**Table 2:** Outbreaks of foodborne illness associated with meat, excluding poultry, in OzFoodNet Sites January 2003 to June 2008 (*n*=66).

State	Year	Setting	III	Hospitalised	Category	Food Vehicle	Aetiology
ACT	2005					Roast pork on	
						bruschetta, duck and	
		Commercial Caterer	27	0	Dish containing meat	quince tartlets	Norovirus
NSW	2003	Restaurant	4	1	Meat	Pork	Salmonella 4,12:d:-
		Private Residence	6	0	Meat	Sliced soccerball ham	Unknown
						Suspected pies, beef,	
					Suspected dish	chicken, tomato &	
		Commercial Caterer	3	0	containing meat	onion	Unknown
	2004				Suspected dish		
		Hospital	5	5	containing meat	Suspected beef curry	Unknown
					Suspected dish	Suspected bacon and	
		Restaurant	20		containing meat	mushroom dish	Unknown
						Suspected bacon and	
		Restaurant	12	0	Suspected meat	ham	Unknown
		National Franchised Fast			Suspected dish	Suspected BBQ Meat	
		Food	5	1	containing meat	Lovers pizza	Unknown
							Salmonella Typhimurium
		Other	27	1	Meat	Roast pork	RDNC, 170
	2005				Suspected dish	Suspected chicken	
		Restaurant	2	0	containing meat	and bacon burgers	Unknown
		Take-Away	4	0	Dish containing meat	Roast beef and gravy	Unknown
		Restaurant	2	0	Suspected meat	Suspected beef steak	Unknown
					Suspected dish	Suspected beef	
		Restaurant	2	0	containing meat	burger	Unknown
							Suspected staphylococcal
		Restaurant	9	0	Dish containing meat	Ham pizza	toxin
		Private Residence	43	13	Meat	Lamb's liver	Salmonella Typhimurium
		Restaurant	5	0	Suspected meat	Lamb, beef	Unknown
						Suspected roasted	
		Restaurant	5	0	Suspected meat	meats	Unknown

State	Year	Setting	III	Hospitalised	Category	Food Vehicle	Aetiology
						Chicken, bacon and	
		Aged Care	10	0	Dish containing meat	mushroom sauce, rice	Clostridium perfringens
		Commercial Caterer	13	0	Dish containing meat	Beef casserole	Unknown
	2006				Suspected dish	Suspect pork in plum	Salmonella Typhimurium 170
		Restaurant	2	2	containing meat	sauce, fried ice cream	var
		Take-Away	80	0	Meat	Roast pork	Clostridium perfringens
					Suspected dish	Suspect oysters, lobsters, prawns, rainbow trout, icecream, sashimi, crab, mussels, beef	
		Postourant	13	0	•	1	Unknown
		Restaurant	13	U	containing meat	curry Suspect beef or	Unknown
					Suspected dish	chicken hamburger with salad, cheese,	
		Take-Away	4	1	containing meat	bacon	Salmonella Typhimurium
		Restaurant	24	0	Dish containing meat	Various Indian dishes - rice, beef madras, butter chicken, lamb roagn josh, vege curry	Unknown
	2007	Private Residence	8	2	Meat	Beef patties	Salmonella Typhimurium
	2007	Restaurant	14	0	Suspected dish containing meat	Raw capsicum, onions, fresh herbs, chicken and/or beef	Unknown
					Suspected dish	Suspected beef or	l
		Take-Away	4	0	containing meat	lamb kebab	Unknown
		Restaurant	9	0	Dish containing meat	Chicken stirfry or beef massaman	Unknown
		Take-Away	2	1	Dish containing meat	Meat kebab	Campylobacter
	2008	Commercial Caterer	75	0	Suspected dish	Suspected curry pumpkin, curry chicken, rice with lamb	Bacillus cereus
			75 7	0	containing meat		
		Restaurant	/	0	Dish containing meat	Suspected chilli beef	Salmonella Typhimurium U290

State	Year	Setting	III	Hospitalised	Category	Food Vehicle	Aetiology
				-		Stir fry beef with dried	
		Restaurant	4	0	Dish containing meat	hot chilli and peanut	Unknown
		Restaurant	2	0	Suspected meat	Suspected ham	Unknown
NT	2003				Suspected dish	Rice, beef and black-	
		Take-Away	5	4	containing meat	bean sauce.	Staphylococcus aureus
		Commercial Caterer	7	1	Meat	Roast meat	Salmonella Typhimurium 135
	2007	Commercial Caterer	3	0	Suspected meat	Suspect roast pork	Salmonella Oslo
QLD	2003	Restaurant	7	0	Dish containing meat	Beef burgundy	Unknown
		Other	16	0	Dish containing meat	Pasta salad with ham	Staphylococcus aureus
		Restaurant	21	2	Suspected meat	Suspected roast pork	Salmonella Typhimurium U307
	2004	National Franchised Fast					
		Food	6	0	Dish containing meat	Pizza	Clostridium perfringens
	2005					Chicken and / or lamb	
		Restaurant	14	0	Dish containing meat	guvec	Clostridium perfringens
		Restaurant	3	0	Dish containing meat	Beef rendang	Clostridium perfringens
		Aged Care	36	0	Meat	Braised steak & gravy	Clostridium perfringens
	2006				Suspected dish	Suspected lamb	
		Restaurant	6	0	containing meat	korma	Unknown
					Suspected dish	Suspected doner	
		Take-Away	4	0	containing meat	kebab	Unknown
		Restaurant	13		Dish containing meat	Chicken & lamb guvec	Clostridium perfringens
						Suspected hommus,	
						hot & spicy dip, baba	
						ghanoush dip,	
					Suspected dish	mussakka, lamb	
		Restaurant	3	1	containing meat	hotpot, lamb cutlets	Salmonella Zanzibar
						Sweet and sour pork,	
		Restaurant	8		Dish containing meat	chow mein beef	Unknown
	2007	Institution	45	0	Suspected meat	Ham; salad; bread	Norovirus
	2008	Institution	56	0	Dish containing meat		Norovirus
SA	2005	Hospital	5	5	Meat	Silverside-corned beef	Listeria monocytogenes
		National Franchised Fast			Suspected dish	Suspected chicken	l
		Food	4		containing meat	and bacon burgers	Unknown

State	Year	Setting	III	Hospitalised	Category	Food Vehicle	Aetiology
	2006					Sandwich containing	
		Restaurant	7	0	Dish containing meat	egg and ham	Salmonella Anatum
VIC	2003	Other	12	0	Meat	Spit-roasted pork	Salmonella Typhimurium 170
		Other	20	4	Meat	Spit-roasted pork	Salmonella Typhimurium 170
	2005	Restaurant	20	1	Suspected meat	Suspected roast pork	Salmonella Typhimurium 170
		Private Residence	13	0	Suspected meat	Suspected undercooked bbq meat	Salmonella Typhimurium 12
		Private Residence	10	0	Suspected dish containing meat	Suspected rice, peppers stuffed with a minced lamb filling, pieces of lamb	Unknown
	2006	Commercially			<u> </u>	Capocollo (cured	Salmonella Bovismorbificans
		Manufactured	13	4	Meat	pork)	11
		Restaurant	10	0	Suspected meat	Suspected roast meats	Unknown
	2007				Suspected dish		
		Take-Away	17	0	containing meat	Suspected meat curry	Unknown
	2008	Take-Away	14	1	Meat	Roast pork	Salmonella Johannesburg
WA	2003	Commercial Caterer	10	0	Dish containing meat	Sandwich meat	Unknown
	2004	Other	100	0	Dish containing meat	Pasta meat sauce	Clostridium perfringens
	2006	Unknown	19		Dish containing meat	Beef/salad roll	Unknown
	2007					Café meal (including	
						bolognaise sauce,	
						sliced ham, diced	
		Restaurant	26	2	Dish containing meat	chicken)	Norovirus